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132. Yukio Kameda and Tomoo Omori: Studies on Acylase Activity and Microörganisms. XVIII.*¹ A Strain of Soil Bacteria capable of resolving Lysine by its Metabolism on N²-Benzoyl or N²-Phenacyl Derivative of N⁶-Benzoyl-DL-lysine.

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In the earlier papers^{1~4}) it was reported that five strains (KT-230, KT-231, KT-232, KT-233, KT-301) of soil bacteria metabolized di-N², N⁶-benzoyl-pL-lysine (I) to produce N⁶-benzoyl-L-lysine (II) and di-N², N⁶-benzoyl-p-lysine (IV), but not metabolized N²-phenacyl-N⁶-benzoyl-pL-lysine (II), and on the other hand, that one strain (KT-311) of soil bacteria metabolized (II) to produce (III) and N²-phenacyl-N⁶-benzoyl-p-lysine (V), but not metabolized (I), but not metabolized (I), and N²-phenacyl-N⁶-benzoyl-p-lysine (V), but not metabolized (I), and KT-311 are shown in Table I.

TABLE I.	Metabolic	Activity of	Soil Bacteria	, KT-230,	KT-231,
К	Т-232, КТ	-233, KT-30)1, KT-311, 2	and KT-31	3

	KT-230, KT-231 KT-232, KT-233	KT-301	KT-311	KT-313
Benzoic acid (VI)	+	+	+	+
Di-N ² , N ⁶ -benzoyl-pl-lysine (I)	+	+		+
$Di-N^2$, N ⁶ -benzoyl-dl-lysine (I) (without NH ₄ Cl)				
$Di-N^2$, N ⁶ -benzoyl-p-lysine (IV)		—		
N ⁶ -Benzoyl-dl-lysine				
Phenylacetic acid (VII)		+	+	+
N^2 -Phenacyl-N ⁶ -benzoyl-pL-lysine (\square)			+	+
N ² -Phenacyl-N ⁶ -benzoyl- DL -lysine (II) (without NH_4C	- (1)			
N^2 -Phenacyl-N ⁶ -benzoyl-p-lysine (V)			—	

+: Within 4 days at 25° a luxuriant growth of bacteria was obtained on a culture medium with the particular organic compound as the source of carbon. This cultivation experiment was repeated 3 times in succession.

-: Almost no visible growth of bacteria was observed at 25° in 4 days. The constituents of the culture medium used in the experiments are as follows: organic substance to be tested, 0.2 g.; NH₄Cl, 0.1 g.; K₂HPO₄, 0.1 g.; MgSO₄·7H₂O, 0.05 g.; 1% CaCl₂·6H₂O, 2 drops; 1% FeCl₃·6H₂O, 1 drop; distilled water, 100 cc.; pH 7.4~7.6 (adjusted with 10% NaOH).

In the present work a new strain of soil bacteria designated KT-313 was isolated, which metabolized di-N²,N⁶-benzoyl-DL-lysine (I) and N²-phenacyl-N⁶-benzoyl-DL-lysine (II) to produce N⁶-benzoyl-L-lysine (III), di-N²,N⁶-benzoyl-D-lysine (IV) and N²-phenacyl-N⁶-benzoyl-D-lysine (V), respectively. The metabolic activity of KT-313 is shown in Table I.

From Table I, it is to be understood that KT-313 can produce N⁶-benzoyl-L-lysine (III) and di-N²,N⁶-benzoyl-D-lysine (IV) or N²-phenacyl-N⁶-benzoyl-D-lysine (V) from di-N²,N⁶-benzoyl-DL-lysine (I) or N²-phenacyl-N⁶-benzoyl-DL-lysine (II) by utilizing benzoic acid or phenylacetic acid, which was formed from di-N²,N⁶-benzoyl-L-lysine (IIIB) or N²-phenacyl-N⁶-benzoyl-L-lysine (IIIB) by the action of their acylase, as the sole

- 2) Y. Kameda, E. Toyoura, K. Matsui: This Bulletin, 7, 702 (1959).
- 3) E. Toyoura : Yakugaku Zasshi, 80, 362 (1960).

^{*1} This work was presented at the Kantō Local Meeting of the Pharmaceutical Society of Japan, October 3, 1959. Part XVII: This Bulletin, 10, 827 (1962).

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¹⁾ Y. Kameda, E. Toyoura, K. Matsui, Y. Kimura, S. Kitagawa : Nature, 182, 453 (1958).

⁴⁾ Idem: This Bulletin, 7, 789 (1959).

source of carbon and ammonia as the sole source of nitrogen. KT-313 metabolized di-N², N⁶-benzoyl-DL-lysine (I) and N²-phenacyl-N⁶-benzoyl-DL-lysine (II) to yield the following compounds respectively: N⁶-benzoyl-L-lysine (III), $[\alpha]_D^{20} + 19^\circ$, di-N², N⁶-benzoyl-D-lysine (IV), m.p. 146~147°, $[\alpha]_D^{20} + 7.5^\circ$, and N²-phenacyl-N⁶-benzoyl-D-lysine (V), m.p. 134~135°, $[\alpha]_D^{20} - 3.3^\circ$.

$$\begin{array}{cccc} C_{6}H_{5}-CO-NH-(CH_{2})_{4}-CH-COOH & \longrightarrow & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ &$$

KT-313 seemingly belongs to the *Pseudomonas* group and has the following characters: aerobic; rod-shaped; gram-negative; yield water-soluble yellowish green pigment that diffuses through the medium.

It is of interest that KT-313 hydrolyzed benzoyl ($\mathbb{II}B$) and phenacyl ($\mathbb{II}P$) derivatives of N⁶-benzoyl-L-lysine (\mathbb{II}), while KT-301 hydrolyzed ($\mathbb{II}B$), but not ($\mathbb{II}P$), and KT-311 hydrolyzed ($\mathbb{II}P$), but not ($\mathbb{II}B$).

Experimental

Preparation of Substrates—Di-N²,N⁶-benzoyl-pL-lysine (I) and N²-phenacyl-N⁶-benzoyl-pL-lysine (I) were prepared according to the procedure described previously.⁵)

Isolation and Characterization of Soil Bacteria KT-313— The constituents of the culture medium for isolation of KT-313 were as follows: N²-phenacyl-N⁶-benzoyl-pL-lysine, 0.2 g.; NH₄Cl, 0.1 g.; K₂HPO₄, 0.1 g.; MgSO₄·7H₂O, 0.05 g.; 1% CaCl₂·6H₂O, 2 drops; 1% FeCl₃·6H₂O, 1 drop; dist. H₂O, 100 cc.; pH 7.4~7.6 (adjusted with 10% NaOH). A soil sample (0.2 g.) was taken from a coppice at Yudani village in Toyama Prefecture, inoculated into 10 cc. of the above culture medium, and incubated at 25° for 6 days. If luxuriant growth of bacteria occurred, a loop of the culture fluid was transfered to a new culture medium of the same composition. Such transplantation was repeated at least 3 times. The bacterial suspension of the last generation was then planted in bouillon agar. Culture experiments were carried out in order to determine whether the microbe isolated from the agar plate could grow in the above medium.

The newly isolated soil bacteria was designated KT-313 and its metabolic activity is shown in Table I. KT-313 seemingly belongs to the *Pseudomonas* group and has the following characters; aerobic, rod-shaped, gram-negative and yields water-soluble yellowish green pigment that diffuses through the medium.

Resolution of Di-N², N⁶-benzoyl-DL-lysine (I) by the Metabolism of KT-313—100 cc. of the sterilized medium mentioned above containing 1.77 g. of di-N², N⁶-benzoyl-_{DL}-lysine in 200 cc. Erlenmeyer flask was seeded with a loop of KT-313 and incubated at 25° for 17 days. The culture thus obtained was heated at 80° for several min. and centrifuged for 10 min. at 8,000 g. to remove the insoluble mass. The supernatant was adjusted to pH 4.5 with AcOH, evaporated *in vacuo* to a small volume, acidified with HCl to pH 1.0 and extracted with AcOEt. The aqueous layer was evaporated *in vacuo* to dryness. The residue was collected by suction. Recrystallization from H₂O gave 0.5 g. (80%) of N⁶-benzoyl-L-lysine (III) as colorless leaves, m.p. 270°(decomp.); $[\alpha]_D^{20}$: +19°(c=2, 5N HCl). Anal. Calcd. for C₁₃H₁₈O₃N₂: C, 62.38; H, 7.25. Found: C, 62.51; H, 7.08.

The AcOEt layer was evaporated *in vacuo* to dryness. The residue was washed several times with petr. ether to remove BzOH, and recrystallized from Me₂CO and benzene to 0.7 g. (79%) of di-N²,N⁶-benzoyl-p-lysine (IV), as colorless leaves, m.p. $146 \sim 147^{\circ}$; $[\alpha]_D^{16}$: $+7.5^{\circ}(c=2, EtOH)$. Anal. Calcd. for C₂₀H₂₂O₄N₂: C, 67.78; H, 6.26. Found : C, 67.90; H, 6.11. From the petr. ether, BzOH was scarcely obtained.

⁵⁾ Y. Kameda, E. Toyoura, Y. Kimura, K. Matsui, H. Saito: Yakugaku Zasshi, 78, 759 (1958).

Resolution of N²-Phenacyl-N⁶-benzoyl-DL-lysine (II) by the Metabolism of KT-313—KT-313 was inoculated into 100 cc. of the culture medium mentioned above containing 1.84 g. of N²-phenacyl-N⁶benzoyl-DL-lysine and incubated at 25° for 5 days. The culture medium was treated according to the resolution procedure of di-N², N⁶-benzoyl-DL-lysine and afforded 0.33 g. (52.8%) of N⁶-benzoyl-Llysine (III) as colorless leaves, m.p. 270° (decomp.); $[\alpha]_D^{\circ}$: +19° (c=2, 5N HCl) (Anal. Calcd. for C₁₃H₁₈-O₃N₂: C, 62.38; H, 7.25. Found: C, 62.53; H, 6.44.), 0.85 g. (93%) of N²-phenacyl-N⁶-benzoyl-D lysine (V) as colorless leaves, m.p. 134~135°; $[\alpha]_D^{\circ}$: -3.3° (c=3, EtOH) (Anal. Calcd. for C₂₁H₂₄O₄N₂: C, 63.73; H, 7.55. Found: C, 63.93; H, 7.78), and 0.04 g. (10%) of phenylacetic acid, m.p. 74~76°. (III) was characterized as its phenylacetate (III P), m.p. 134~135°; $[\alpha]_D^{\circ}$: +3.3 (c=3, EtOH), obtained in 67.9% yield after recrystallization from Me₂CO and benzene. The mixture of (V) and (III P) melted at ca. 149°. Incidentally, N²-Phenacyl-N⁶-benzoyl-DL-lysine melts at 150~151°.

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Summary

A certain strain (KT-313) isolated from soil had the metabolic activity shown in Table I and could metabolize di-N², N⁶-benzoyl-DL-lysine (I) and N²-phenacyl-N⁶-benzoyl-DL-lysine (II) to produce N⁶-benzoyl-L-lysine (II), di-N², N⁶-benzoyl-D-lysine (IV) and N²-phenacyl-N⁶-benzoyl-D-lysine (V), respectively. KT-313 seemingly belongs to the *Pseudomonas* group. It is of interest that KT-313 hydrolyzed benzoyl (IIIB) and phenacyl (IIP) derivatives of N⁶-benzoyl-L-lysine (II), while KT-301 hydrolyzed (IIB), but not (IIP), and KT-311 hydrolyzed (IIIP), but not (IIIB).

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133. Tchan Gi Bak^{*1} and Shigeaki Kuwano: Reversible and Irreversible Inhibitions of Glutamic and Arginine Decarboxylase Activities of *Escherichia coli* by Gallic Acid and *d*-Catechin.

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Kimura, *et al.* have found that tannins and related compounds inhibit the decomposition of amino acids to form diamines ¹), hydrogen sulfide²) and indole³) by pyridoxal phosphate-dependent enzymes of *Escherichia coli*. It has also been shown that gallic acid and *d*-catechin, characteristic components of pyrogallol- and catechol-tannins respectively, inhibit the enzyme reactions through somewhat different mechanisms.

The present paper deals with the kinetic studies on the inhibition of two pyridoxal phosphate-requiring enzymes of $E.\ coli$, i.e. glutamic and arginine decarboxylases, by gallic acid and *d*-catechin. It was especially aimed to elucidate in details the differences in inhibition mechanisms due to gallic acid and *d*-catechin. Efforts were also made to show that these inhibitions are caused by both reversible and irreversible

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¹⁾ K. Kimura, S. Kuwano, H. Hikino: Yakugaku Zasshi, 78, 236 (1958).

²⁾ K. Kimura, K. Yamauchi, S. Kuwano: This Bulletin, 7, 426 (1959).

³⁾ Idem: Ibid., 7, 531 (1959).