

in this case; the relative proportions of products of type VII and X, and of other by-products, depends very markedly upon the substitution pattern of the octalone studied: between 80% of indane in cases a and c, to 80% of tetralin in cases b and d.

The most remarkable features of this aromatization are its ease and cleanliness in the case where no angular methyl group hinders it, with HBr/AcOH, *p*-TsOH/AcOH, CF₃CO₂H or polyphosphoric acid (PPA). For example the trimethyloctalone (VIc) with PPA at 150° gives quantitatively the trimethyltetralin (VIIc) in two minutes.

*Institut de Chimie,
Université de Strasbourg,
Faculté des Sciences,
Esplanade,
Strasbourg,
France*

Philippe Bey
Florence Lederer
Guy Ourisson

Received June 16, 1965

[Chem. Pharm. Bull.]
13(9)1140~1142(1965)

UDC 612.398.145

Synthesis of 8-Hydroxypurine Nucleosides

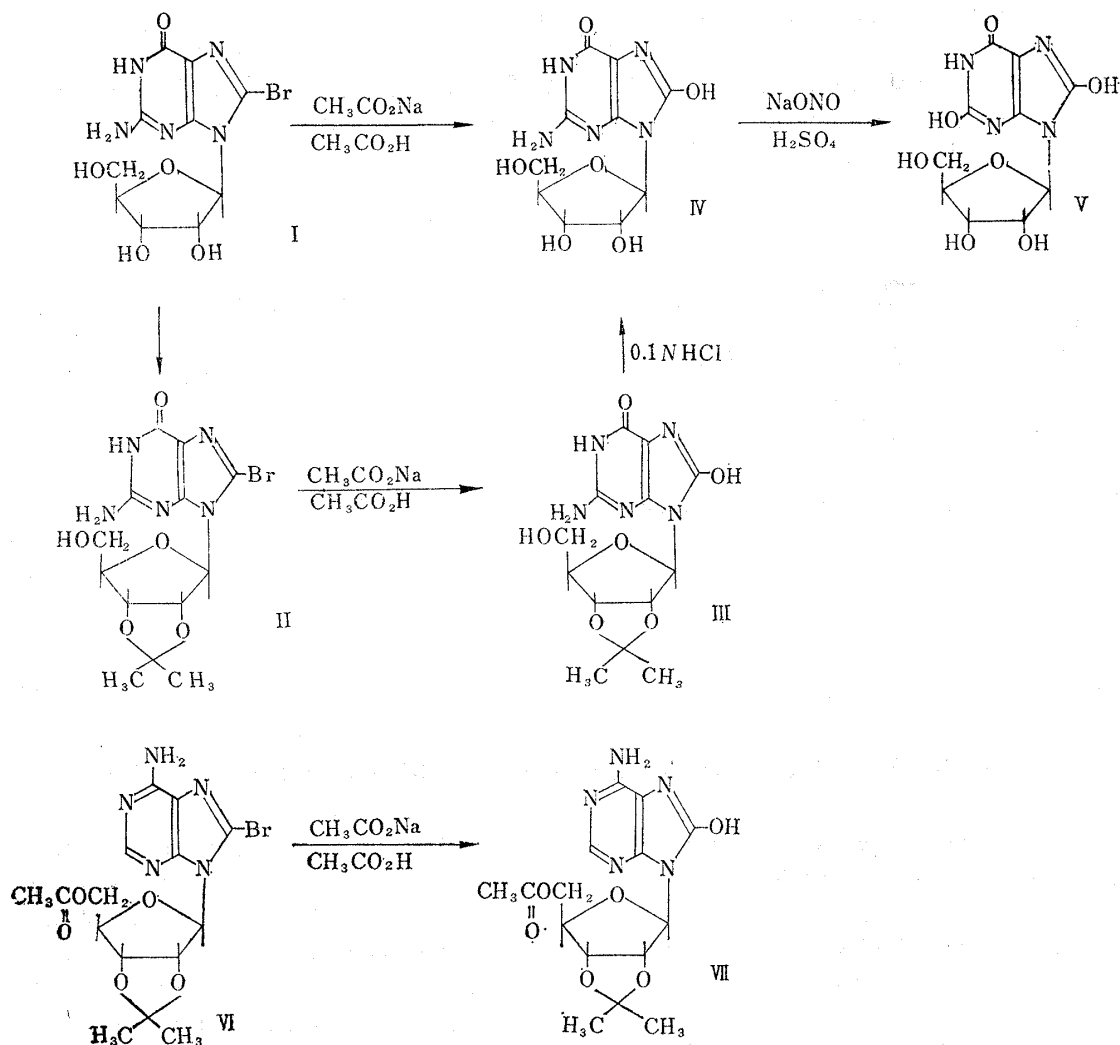
Recently, 8-hydroxynucleosides draw attention of many investigators, because of their biological interest.¹⁻³⁾ From the chemical point of view, they also are interest as the key intermediate of the synthesis of purine cyclonucleosides^{4,5)} having O-anhydro linkage.

In order to meet this requirement, many attempts were made in this laboratory to hydrolyze halogen atom substituted at 8-position. However, the acid lability of purine nucleoside and alkaline instability of 8-hydroxypurine nucleoside gave only unsatisfactory results.

We found that, when 8-bromo derivatives of adenosine and guanosine were heated in acetic acid with excess of sodium acetate, desired 8-hydroxy compounds were obtained in a good yield. 8-Bromoguanosine (I) was heated with ten fold excess of anhydrous sodium acetate (fused freshly) in glacial acetic acid at 118° for 3 hours. Resulting white solid has $\lambda_{\text{max}}^{\text{pH } 2}$ m μ : 242, 292; $\lambda_{\text{max}}^{\text{pH } 7}$ m μ : 242, 292; $\lambda_{\text{max}}^{\text{pH } 10}$ m μ : 251, 279. The similar solubility of this compound with that of sodium acetate in many solvents made it difficult to purify this specimen. In order to circumvent this difficulty 2',3'-O-isopropylidene-8-bromoguanosine (II) was reacted in acetic acid with sodium acetate and 2',3'-O-isopropylidene-8-hydroxyguanosine*¹ (III) was obtained as a hard glass. Acidic removal of isopropylidene group gave 8-hydroxyguanosine (IV), m.p. >180°(decomp.), in 38% yield. *Anal.* Calcd. for C₁₀H₁₃O₆N₅·½H₂O: C, 38.96; H, 4.54; N, 22.72. Found: C, 38.80; H, 4.72; N, 22.21. Ultraviolet absorption properties: $\lambda_{\text{max}}^{\text{pH } 2}$ m μ : 245.5, 292.5; $\lambda_{\text{max}}^{\text{pH } 7}$ m μ : 245.5, 292.5; $\lambda_{\text{max}}^{\text{pH } 10}$ m μ : 258, 280. Infrared absorption $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1720 (8-CO). Paper chromatography: Rf 0.10 (1-butanol-water, 86:14), Rf 0.69 (water, pH 10). This sample was identical in many respects with that obtained above. 8-Hydroxyguanosine

*¹ Although 8-hydroxypurine may be obtained *via* an 8-acetoxy compound, the latter compound could not be isolated.

- 1) H. S. Forrest, D. Hatfield and J. M. Lagowski: J. Chem. Soc., 1961, 963.
- 2) R. Lohrmann, J. M. Lagowski, H. S. Forrest: *Ibid.*, 1964, 452.
- 3) D. Hatfield, R. A. Greenland, H. L. Stewart, J. B. Wyngaarden: Biochim. Biophys. Acta, 91, 163 (1964).
- 4) M. Ikehara, H. Tada: J. Am. Chem. Soc., 87, 606 (1965).
- 5) M. Ikehara, H. Tada, K. Muneyama: This Bulletin, 13, 639 (1965).



was fairly stable to the acidic treatment (0.1N hydrochloric acid at 100° for 1 hour) and labile to the alkaline hydrolysis (0.1N sodium hydroxide even at room temperature) as reported in the case of uric acid riboside.¹⁾ When 8-hydroxyguanosine (IV) was reacted with nitrous acid in 20% sulfuric acid,*² a compound having ultraviolet absorption properties ($\lambda_{\max}^{\text{pH}1} \text{ m}\mu$: 232, 289; $\lambda_{\max}^{\text{pH}10} \text{ m}\mu$: 248, 303) was obtained. Paper chromatography (Rf 0.21 in 1-butanol-water, 86:14; Rf 0.64 in water, pH 10) also indicated the identical properties reported for uric acid 9-riboside.*^{3,3)}

This reaction was then applied to 8-bromo-5'-O-acetyl-2',3'-O-isopropylideneadenosine (VI) using 5 equivalents of sodium acetate. After the reflux of 2.5 hours 8-hydroxy derivative (VII) was obtained as a hard glass in the quantitative yield. Ultraviolet absorption properties : $\lambda_{\max}^{0.1N \text{ HCl}} \text{ m}\mu$: 262, 284; $\lambda_{\max}^{0.1N \text{ NaOH}} \text{ m}\mu$: 280. Infrared absorption : $\nu_{\max}^{\text{NaJol}} \text{ cm}^{-1}$: 1740 (ester CO), 1715 (8-CO). Thin-layer chromatography : Rf 0.28 (silica gel, chloroform-ethanol, 35:5). Paper chromatography : Rf 0.70 (1-butanol-water-conc. ammonia, 7:1:2). Acidic hydrolysis of this compound gave 8-hydroxyadenine, $\lambda_{\max}^{\text{pH}9} \text{ m}\mu$:

*² When the deamination reaction was carried out in dilute acetic acid, ultraviolet absorption was diminished after 2 hr. at room temperature (unpublished experiments by K. Muneyama).

*³ After completion of this experiments, we were aware of the work of Holmes, *et al.*⁶⁾ of the synthesis of uric acid 9-riboside by another route.

6) R. E. Holmes, R. K. Robins : J. Am. Chem. Soc. 87, 1772 (1962).

277; $\lambda_{\text{max}}^{\text{PH}4}$ m μ : 268; $\lambda_{\text{max}}^{\text{PH}1}$ m μ : 263, 278 (identical with those reported by Cavalieri and Bendich⁷⁾). Paper chromatography: Rf 0.26 (1-butanol-water, 86:14).

Thus a versatile method of introduction of hydroxyl group to the 8-position of purine nucleosides was established. The application of this method to the various nucleosides is in progress in this laboratory.

Authors are indebted to Mrs. Toyoko Tohma and Miss Akiko Maeda for elemental analyses.

Faculty of Pharmaceutical Sciences,
Hokkaido University,
Kita-12-jo, Nishi-5-chome, Sapporo

Morio Ikehara (池原森男)
Hiroshi Tada (多田 宏)
Kei Muneyama (宗山 敬)

Received May 8, 1965

7) L. C. Cavalieri, A. Bendich: J. Am. Chem. Soc., 72, 2587 (1950).

[Chem. Pharm. Bull.]
13(9)1142~1144(1965)

UDC 577.15.074

A Total Synthesis of Coenzyme A via Thiazoline Intermediate^{*1}

In the previous paper,¹⁾ the authors described a novel synthesis of D-pantetheine (IIIa), one of the biosynthetic intermediates of coenzyme A from D-pantothenic acid,²⁾ which comprises the interim formation of thiazoline derivative from D-pantothenonitrile (Ia) followed by acid hydrolysis thereof (hereafter referred to as the thiazoline method). This method seems to be applicable to the syntheses of other metabolites from D-pantothenic acid including coenzyme A itself. The present communication deals with the syntheses of coenzyme A and dephospho coenzyme A by the thiazoline method.

Three independent papers on the total synthesis of coenzyme A³⁻⁵⁾ have so far been published, each of which employed in principle the condensation of D-pantetheine derivatives with adenine nucleotides, differing in the procedure for pyrophosphate bond formation. The present method is characterized in building up aletheine moiety in coenzyme A structure in the last step of the synthetic route.

The key intermediates used in the present experiment are P¹-adenosine-3'-phosphate-5'P²-D-pantothenonitrile 4'-pyrophosphate (Ib) for coenzyme A, and P¹-adenosine-5'P²-D-pantothenonitrile 4'-pyrophosphate (Ic) for dephospho coenzyme A. For the preparation of Ib and Ic, it is essential at the outset to establish the synthesis of D-pantothenonitrile 4'-phosphate (V). D-Pantothenonitrile (Ia) was treated with dibenzyl phosphorochloridate in anhydrous pyridine to furnish a syrupy substance (IV), $[\alpha]_D^{25} +19.1^\circ$ (EtOH), in 73.2% yield. Catalytic hydrogenation of IV over palladium-charcoal afforded V, $[\alpha]_D^{25} +11.4^\circ$ (H₂O), IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2250 (C≡N), in 69% yield as its barium salt (C₉H₁₅O₆N₂PBa·2H₂O).

*1 A part of this communication was read at the Kanto Branch Meeting of the Pharmaceutical Society of Japan, May, 1965, Tokyo.

- 1) M. Shimizu, G. Ohta, O. Nagase, S. Okada, Y. Hosokawa: This Bulletin, 13, 180 (1965).
- 2) L. Levintow, G. D. Novelli: J. Biol. Chem., 207, 761 (1954); M. B. Hoagland, G. D. Novelli: *Ibid.*, 207, 767 (1954); G. M. Brown: J. Biol. Chem., 234, 370 (1959).
- 3) J. G. Moffatt, H. G. Khorana: J. Am. Chem. Soc., 81, 1265 (1959); 83, 663 (1961).
- 4) A. M. Michelson: Biochim. Biophys. Acta, 50, 605 (1961); 93, 71 (1964).
- 5) W. Gruber, F. Lynen: Ann., 659, 139 (1962).