

Nous avons inoculé intrapéritonéalement 10^6 cellules de leucémie, souche L 1210, à 6 souris pour chaque groupe—souche BDF₁ (C₅₇BL/6♀ × DBA/2♂), femelles qui sont âgées de 6—7 semaines.

Le traitement a été commencé 24 hr après l'inoculation de cellules de leucémie, et continué pendant 5 jours, une fois par jour.

Les substances ont été dissoutes dans une solution saline physiologique.

Pour le témoin, nous avons injecté la même volume de solution saline à la souris.

Les résultats sont présentés dans le tableau II.

En résumant nos résultats, le dihydronivalenol est presque aussi puissant que le nivalenol quant à l'inhibition de la synthèse des protéines des réticulocytes, et plus effectif que le nivalenol sur l'activité antileucémique, mais la toxicité aiguë du dihydronivalenol est moins grande que celle du nivalenol.

Nous voulons maintenant continuer nos recherches dans le but de mettre au point le dihydronivalenol comme médicament nouveau contre la leucémie.

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A Total Synthesis of a *dl*-Prostaglandin B₁

The prostaglandins, a family of C₂₀ prostanoid acids and biologically highly active substances having diverse pharmacological properties, have received considerable attention.¹⁾ Recently, total syntheses of prostaglandin B₁,^{2,3)} F_{1α},³⁾ and E₁⁴⁾ have been reported. Now we wish to report the total synthesis of the racemic prostaglandin B₁ in fairly short steps.

The keto acid (1) was prepared according to a slightly modified Bowman's method;⁵⁾ triethyl ethane-1,1,2-tricarboxylate⁶⁾ was transesterified to the tribenzyl ester with sodium ethoxide as catalyst, and the sodium salt of tribenzyl ester was condensed with the half acid chloride of monoethyl azelaate in refluxing benzene to give the tribenzyl oxoester. Catalytic hydrogenolysis of this ester over 10% Pd-SrCO₃ in ethyl acetate afforded the corresponding oxo tricarboxylic acid and finally decarboxylation of the acid in refluxing ethyl acetate resulted

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- 3) K.G. Holden, B. Hwang, K.R. Williams, J. Weinstock, M. Harman, and J.A. Weisbach, *Tetrahedron Letters*, **1968**, 1569; G. Just and Ch. Simonovitch, *ibid.*, **1967**, 2093.
- 4) E.J. Corey, N.H. Andersen, R.M. Carlson, J. Paust, E. Vedejs, I. Vlattas, and R.E.K. Winter, *J. Am. Chem. Soc.*, **90**, 3245 (1968); E.J. Corey, I. Vlattas, N.H. Andersen, and K. Haeding, *ibid.*, **90**, 3247 (1968).
- 5) R.E. Bowman, *J. Chem. Soc.*, **1950**, 325.
- 6) G.S. Fonken and W.S. Johnson, *J. Am. Chem. Soc.*, **74**, 832 (1952).

in the β -oxo acid (**1**) (over all yield 27%), mp 76—78°, ⁷⁾ [IR $\nu_{\max}^{\text{Nujol}}$ cm⁻¹: 3750, 3300, 1724, 1698].

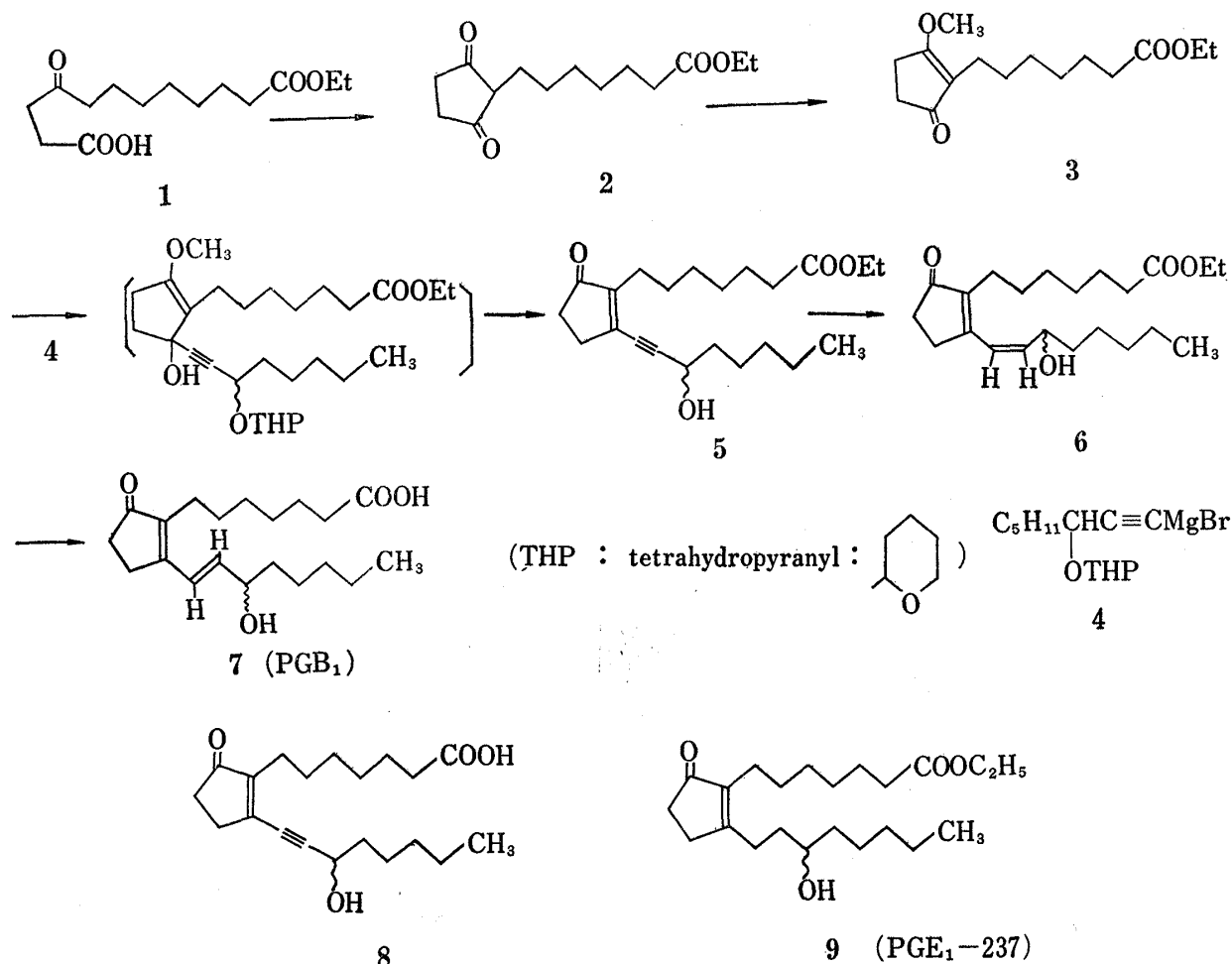


Chart 1

Cyclization ⁸⁾ of the β -oxo acid (**1**) to a cyclopentane-1,3-dione derivative (**2**) was carried out in the presence of AlCl_3 and propionyl chloride in nitrobenzene at 80° for 3.5 hr (in 63.3% yield) accompanied by a small amount of free acid, mp 150—153°. The dione (**2**), mp 94—95°, was characterized by the following physical properties; [UV $\lambda_{\max}^{\text{EtOH}}$ m μ (ϵ): 250 (16500), 270 (5700; shoulder); IR $\nu_{\max}^{\text{Nujol}}$ cm⁻¹: 2500 (enol OH), 1887, 1739; and NMR:⁹⁾ 1.18 (3H, t), 4.04 (2H, q), 2.33 (4H, s), 10.12 (H, s; enol OH)]. Treatment of **2** with diazomethane in ether gave the methyl enol ether (**3**), in 85.4% yield, [bp 163—165° (0.04 mmHg)⁷⁾ (bath temp.), UV $\lambda_{\max}^{\text{EtOH}}$ m μ (ϵ): 252 (18100); IR $\nu_{\max}^{\text{liq}}$ cm⁻¹: 1749, 1695, 1637; NMR: 1.25 (3H, t), 4.14 (2H, q), 3.96 (3H, s)].

Grignard reaction ¹⁰⁾ of **3** with an acetylenic magnesium bromide (**4**) in ether-tetrahydrofuran mixture (1:1) under reflux in nitrogen atmosphere followed by treatment with a dilute mineral acid at room temperature for 20 hr produced, after chromatography on silica gel using benzene-chloroform for elution, pure acetylenic ester (**5**), in 14% yield, as a oily substance [UV $\lambda_{\max}^{\text{EtOH}}$ m μ (ϵ): 272 (18700); IR $\nu_{\max}^{\text{liq}}$ cm⁻¹: 3344, 2232, 1733, 1695, 1613; NMR: 0.39 (3H, t;

7) All melting and boiling points are uncorrected. Satisfactory elemental analyses were obtained for all compounds unless otherwise noted.

8) For this procedure, see H. Schick, G. Lehmann, and G. Hilgetag, *Chem. Ber.*, **100**, 2973 (1967).

9) The NMR spectra were measured using Varian A-60 spectrometer and the chemical shifts are expressed in ppm units from the internal standard of tetramethylsilane in CDCl_3 solution.

10) This reaction was carried out by means of the very slow inverse addition of the Grignard solution in order to avoid attack of the ester group of **3** by the Grignard reagent.

CH₃), 1.26 (3H, t), 4.16 (2H, q), 4.68 (1H, t, CH-OH)]. Alkaline hydrolysis of **5** to the acetylenic acid (**8**) was unsuccessful but instead decomposition occurred.

Selective hydrogenation of **5** in the presence of Lindlar's catalyst¹¹⁾ with a small amount of quinoline in benzene gave after chromatography (silica gel, benzene-chloroform) a *cis*-olefinic ester (**6**), in 96% yield [UV $\lambda_{\text{max}}^{\text{EtOH}}$ m μ (ϵ): 278 (20800), IR $\nu_{\text{max}}^{\text{liq}}$ cm⁻¹: 3350, 1739, 1695, 1634]. The NMR spectrum of **6** exhibited signals at 1.26 (3H, t) and 4.15 (2H, q) due to the ethyl ester, and 6.50 (1H, d, $J=12.0$ cps) and 5.87 (1H, dd, $J=12.0$ and $J=9.5$ cps) assigned to the *cis*-vinyl proton. The *cis*-olefinic ester (**6**) was subjected to prolonged catalytic hydrogenation in the presence of a large amount of Lindlar's catalyst to produce the ethyl ester of PGE₁-237 (**9**), in quant. yield, UV $\lambda_{\text{max}}^{\text{EtOH}}$ m μ (ϵ): 237 (13700).

Treatment of **6** with 10% KOH in 50% methanol at 50° for 10 hr followed by acidification afforded a oily acidic substance, which after carefully repeated silica gel column chromatography using chloroform-ethyl acetate for elution furnished pure *dl*-prostaglandin B₁ (**7**) in waxy state in 96% yield; [UV $\lambda_{\text{max}}^{\text{EtOH}}$ m μ (ϵ): 278 (26400); IR $\nu_{\text{max}}^{\text{liq}}$ cm⁻¹: 970 1595, 1640, 1670, 1725, 3300; NMR: 6.85 (1H, d, $J=16.0$ cps), 6.25 (1H, dd, $J=16.0$ cps and $J=5.5$ cps), 6.15 (OH, COOH broadening singlet), 4.36 (1H, m, -CH-OH)]. The mass spectral analysis of **7** revealed the typical fragmentation pattern observed in the spectra of natural prostaglandins.¹²⁾

TABLE I. The Mass Spectrum of Prostaglandin B₁

<i>m/e</i>	% of base peak	<i>Isotope Abundances</i>	
189	7.5 M ⁺ -(C ₆ H ₁₂ COOH+H ₂ O)	<i>m/e</i>	% of P
191	9.5	336(P)	100
217	8.2	337(P+1)	24.6
219	100.0 M ⁺ -(C ₅ H ₁₁ +CO+H ₂ O)	338(P+2)	8.7
220	21.8 M ⁺ -(C ₅ H ₁₁ +COOH)	Calcd. % of (P+1) : 22.3	
235	22.4		
237	29.2 M ⁺ -(C ₅ H ₁₁ +CO)		
238	6.1		
247	20.4 M ⁺ -(C ₅ H ₁₁ +H ₂ O)		
265	2.7 M ⁺ -(C ₅ H ₁₁)		
289	10.2		
290	3.4 M ⁺ -(H ₂ O+CO)		
318	7.5 M ⁺ -(H ₂ O)		
336	38.8 (Parent)		
337	9.5 (P+1)		
338	4.8 (P+2)		

In Table 1 are listed some of the typical fragments in the mass spectrum of **7**. The spectrum showed a parent ion at *m/e* 336. The base peak (M⁺-117) is formed by splitting off C₆H₁₂O₂ (C₅H₁₁+CO+H₂O). Moreover, the spectrum exhibited peaks resulting from the loss of the fragment 71 (C₅H₁₁) and due to loss of the fragments 89 (C₅H₁₁+H₂O), 99 (C₅H₁₁+CO), and 116 (C₅H₁₁+COOH). The peak at 189 is formed by splitting off C₇H₁₅O₃(C₆H₁₂COOH+H₂O).

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