Chem. Pharm. Bull. 22(4) 805—808 (1974)

UDC 547.9.08:615.35.076.9

## Screening of Formosan Ferns for Phytoecdysones. I1)

Kun-Ying Yen, Ling-Ling Yang, 20) Toru Okuyama, Hiroshi Hikino, and Tsunematsu Takemoto<sup>2b)</sup>

Taipei Medical College<sup>2a)</sup> and Pharmaceutical Institute, Tohoku University<sup>2b)</sup>

(Received July 26, 1973)

Formosan ferns from 17 families, representing 50 genera, 115 species, 1 subspecies, and 3 varieties, have been screened by the bioassay for phytoecdysones. A total of 64 species and 2 varieties have been found to show distinct insect moulting hormone activity. Correlation between this hormone activity and taxonomy of ferns is discussed.

According to our earlier experience in screening plant materials of Japanese origin for the insect-moulting hormone activity, the probability of finding the activity appears to be high in ferns among the plant groups.<sup>3)</sup> In fact, a systematic survey of Japanese ferns for phytoecdysones by means of bioassay has resulted in the discovery of 170 species, 22 varieties, and 1 form showing this activity towards insects out of 283 species, 39 varieties, and 1 form belonging to 76 genera (20 families) screened. Among the active Japanese ferns, 16 species have been subjected to analysis for active principles and were found to contain 7 kinds of phytoecdysones such as ecdysone, ponasterone A, pterosterone, ecdysterone, shidasterone, lemmasterone, and ponasteroside A.<sup>1)</sup>

Taiwan is an island lying off the east coast of Asia, whose situation in the subtropical zone gives a warm temperate climate and heavy rainfall. Under these physical features, therefore, the flora of Taiwan is very rich, containing approximately 600 species of ferns.<sup>4)</sup> Our next endeavor was then directed towards the screening of ferns of Formosan origin for the insect-moulting hormone activity. Since there has been some difficulty in collecting the plant material, a limited number of ferns, *i.e.*, 115 species, 1 subspecies, and 3 varieties, have so far been made available. Some species were collected at various seasons and/or locations so that a toal of 164 specimens were screened. The screening method adopted for the present bioassay was the *Sarcophaga* test.<sup>5)</sup> The present paper deals with the result of screening ferns indigenous to Taiwan for phytoecdysones.

As shown in Table I, the present work on Formosan ferns gave a result similar to that on Japanese ferns. Thus, the screening has led to the discovery of 64 species and 2 varieties having this activity towards the insect, 6) while 51 species, 1 subspecies, and 1 variety exhibited no activity, corroborating the previous conclusion that ferns are sources of this hormonal activity with a higher probability. Among the ferns having this activity, 18 species and 2 varieties exhibited remarkable responses. The previous observation that samples of the same species but of different collecting dates and/or locations sometimes exhibited different responses, 3) was also noted in the present investigation. In particular, remarkable variation in the activity shown by Diplazium donianum was again observed. As in the previous work, 3)

<sup>1)</sup> This paper is Part XIX in the Tohoku University series on Steroids. Part XVIII: T. Takemoto, T. Okuyama, H. Jin, T. Arai, M. Kawahara, C. Konno, S. Nabetani, S. Arihara, Y. Hikino, and H. Hikino, *Chem. Pharm. Bull.* (Tokyo), 21, 2336 (1973).

<sup>2)</sup> Location: a) Taipei, Taiwan; b) Aoba-yama, Sendai.

<sup>3)</sup> H. Hikino, T. Okuyama, H. Jin, and T. Takemoto, Chem. Pharm. Bull. (Tokyo), 21, 2292 (1973).

W.-C. Shieh, "Jye-lei-chih-wu (Ferns and Fern Allies)," Provincial Taiwan Chung-Hsing University, Tai-chung, 1969.

<sup>5)</sup> T. Ohtaki, R.D. Milkman, and C.M. Williams, Biol. Bull., 135, 332 (1968).

<sup>6)</sup> Taxa each of which contained at least one active sample were regarded as positive.

TABLE I. The Insect Moulting Hormone Activity of Formosan Ferns

Family <sup>a</sup> )	Species <sup>b)</sup>	Chinese name	Date	Location <sup>c)</sup>	Activity
Equisetaceae	Equisetum ramosissimum Desfontaines subsp. debile Hauke	Tai-wan-mu-cheih	Aug	NT, Mt. Ho-huan	_
Lycopodiaceae	Lycopodium cernuum Linné	Quoh-shan-long	Feb	TP, An-keng	
7 - 1			Sep	TP, Yang-ming-shan	·,
	L. clavatum Linné	Shih-song	Feb	NT, Chi-tou	
	(L. clavatum Linné var. nipponicum NAKAI)				
	L. complanatum Linné	Tieh-shua-tsu	Aug	NT, Mt. Ho-huan	
	L. fargesii HERTER	Zuey-yeh-shih-song	Sep Sep	TP, Yang-ming-shan TP, Yang-ming-shan	
	L. fordii Baker	Fu's-shih-song Chwei-chi-shih-song	Sep	TP, Yang-ming-shan	
Calcainallaceae	L. phlegmaria Linné Selaginella delicatula Alston	Chuan-yuan-chian-bor	Jan	TC, Ku-kuan	
Selaginellaceae	Setaginena activatina Alston	Character and Ch	Aug	TP, Mt. Muh-tsu	_
			Aug	TP, Wu-lai	
			Sep	TP, Yang-ming-shan	
	S. doederleinii Hieronymus	Sheng-qen-chian-bor	Feb	TP, An-keng	
,	S. mollendorffii Hieronymus	Yih-yeh-chian-bor	Feb	TP, Ken-tin	_
V ( 1	• • • •		Aug	TP, Mt. Muh-tsu	-
**	S. pseudo-involvens HAYATA	Nii-mih-yeh-chian-bor	Aug	TP, Mt. Muh-tsu	-
	S. tamariscina Spring	Wann-nen-song	Feb	TP, Wu-lai	
Marattiaceae	Angiopteris lygodiifolia Rosenstock	Quan-in-tzoh-lien	Feb	NT, Chi-tou	
			Aug	TP, Mt. Muh-tsu	_
3.			Aug	TP, Wu-lai	, <del></del>
	mi i i i i i i i i i i i i i i i i i i	The sheet was such too ohim	Sep	TP, Yang-ming-shan	  ##
Osmundaceae	Plenasium banksiaefolium Prest	Tsu-chy-guo-yeh-tsu-chin	Feb Aug	TP, Wu-lai TP, Mt. Muh-tsu	<del>    </del>
Schizagages	(Osmunda banksiaefolia Kuhn)	Hai-chin-sah	Aug	TP, Mt. Mun-tsu TP, Wu-lai	11111
Schizaeaceae	Lygodium japonicum SWARTZ	Ta. Omn-san	Dec	TP, Chih-nan-kung	
	L. microstachyum Desvaux	Shia-yeh-hai-chin-sah	Aug	TP, Mt. Muh-tsu	_
	L. microstacnyum Desvaux L. scandens Swartz	Shiau-yeh-hai-chin-sah	Feb	TP, An-keng	-
*	var, microphyllum Bongard	Silve Joh Mar-Ollin-Salt	, OD	,	
Gleicheniaceae	Dicranopteris dichotoma Bernhardi	Mang-chih	Aug	TP, Mt. Muh-tsu	·
, carrier in a carrier in a	(D. linearis Underwood)		Aug	TP, Hsin-pei-tou	
Hymenophyllaceae	Vandenboschia auriculata COPELAND	Ping-jye	Feb	TP, Wu-lai	· –
Pteridaceae	Adiantum capillus-veneris Linné	Tie-shian-jye	Aug	TP, Mt. Muh-tsu	
2 1011,040040	A. caudatum Linné	Pian-yeh-tie-shian-jye	Aug	NT, Mt. Ho-huan	-
	A. diaphanum Blume	Chang-wei-tie-shian-iye	Aug	NT, Mt. Ho-huan	_
	A. flabellulatum Linné	San-yeh-tie-shian-jye	Feb	PT, Ken-tin	<del>Ⅲ</del> <del>Ⅲ</del>
	A. hispidulum SWARTZ	Mau-yeh-tie-shien-jye	Aug	NT, Lu-shan	##
	A. philippense Linné	Pan-ye-sing-tie-shien-jye	Aug	NT, Lu-shan	#
	Cheilanthes argentea Kunze	Chang-ping-fun-pei-jye	Aug	NT, Mt. Ho-huan	· · · —
	(Aleuritopteris argentea FÉE)				
	C. hirsuta Mettenius	Mau-suei-mii-jye	Aug	NT, Lu-shan	. +
	Cibolium barometz J. SMITH	Chin-gou-mau-jye	Jan	TC, Ku-kuan	_
			Aug	TP, Mt. Muh-tsu	
	Conjogramme fraxinea Diels	Chuan-yuan-fong-liau-jye	Feb	NT, Chi-tou	<del>-</del>
	C. intermedia HIERONYMUS	Hwa-fong-liau-jye	Feb	TP, An-keng	+ ~
	(C. fraxinea Diels var. intermedia C. Christ	ENSEN)	Feb	NT, Chi-tou	
	C. japonica Diels	Zu-pen-fong-liau-jye		TP, Yang-ming-shan	
	Dennstaedtia scabra Moore	Wan-jye Tsyh-ping-wan-jye	Aug Feb	TP, Wu-lai	
	D. scandens Moore	Lih-jye	Aug	TP, Mt. Muh-tsu	_
	Histiopteris incisa J. Smith	Dirijye	Aug	TP, Yang-ming-shan	_
	The bolobic alta anneilling Warrant	Tai-wan-ji-jye	Sep	TP, Yang-ming-shan	
	Hypolepis alte-gracillima HAYATA  H. punctata Mettenius	Ji-jye	Sep	TP, Yang-ming-shan	
	Microlepia hookeriana Prest.	Huk's-lin-gai-jye	Aug	TP, Yang-ming-shan	+
	M. marginata C. Christensen	Pian-yuan-lin-gai-jye	Feb	TP, Ken-tin	÷
	and growing of Carriothiopin		Aug	TP, Yang-ming-shan	_
	M. pilosula Prest.	Hor-mau-lin-gai-jye	Aug	TP, Hsin-pei-tou	
		S. 22	Aug	TP, Wu-lai	· –
	M. speluncae Moore	Zeh-dai-lin-gai-jye	Aug	TP, Hsin-pei-tou	+
	•		Sep	TP, Yang-ming-shan	. +
	M. strigosa Prest.	Tsu-mau-lin-gai-jye	Sep	TP, Yang-ming-shan	-
	Onychium japonicum Kunze	Zu-pen-chin-fun-jye	Jun	TY, Mt. Chiao-pan	-
	Pityrogramma calomelanos Link	Fun-yeh-jye	Apr	TC, Ku-kuan	-
	Pteridium aquilinum Kuhn	Jye	Jan	TC, Ku-kuan	1111
	var. latiusculum Underwood	mt water to a second	ъ.	and an incident	
	Pteris dispar Kunze	Tien-tsau-fong-wei-jye	Feb	TP, Yang-ming-shan	#
	P. ensiformis Burme	Jian-yeh-fong-wei-jye	Jan	TC, Ku-kuan	-
			Aug Dec	TP, Hsin-pei-tou TP, Chih-nan-kung	+ - + + + + +
the North Control of the Control	P. fauriei Hieronymus	Fuh's-fong-wei-jye	Jan	TC, Ku-kuan	71
	P. jauriei Fileronymus (P. quadriaurita Retzius)	Lan s-long-wer-lye	Feb	TP, Wu-lai	#
	(1 . Yannanana XXIII)		Aug	TC, Ku-kuan	##
	P. longipes Don	Ferng-lai-fong-wei-jye	Jan	TC, Ku-kuan	
	P. semipinnata Linné	Pan-pian-yii-lieh-fong-wei-jye	Aug	TP, Mt. Muh-tsu	_
		2y	Aug	TP, Hsin-pei-tou	+
			Dec	TP, Chih-nan-kung	+
	P. tripartita SWARTZ	San-jau-fong-wei-jye	Aug	NT, Tien-chi	+
	P. vittata Linné	Lin-gai-fong-wei-jye	Jan	TC, Ku-kuan	+
	P. wallichiana AGARDH	Wa's-fong-wei-jye	Feb	TP, Wu-lai	+ + #
	Sphenomeris chusana Copeland	Wu-jye	Feb	TP, An-keng	
		***	Aug	TP, Mt. Muh-tsu	<del>, -</del>
	(S. chinensis Maxon)				111
Davalliaceae		Tai-wan-shiau-mo-gat-jye	Jul	CY, Mt. A-li	1117
Davalliaceae	(S. chinensis Maxon)	Tai-wan-shiau-mo-gat-jye Shiau-mo-gai-jye	Jul	IL, Mt. Tai-ping	<del>111</del>
Davalliaceae	(S. chinensis Maxon) Araiostegia parvipinnula Copeland	Shiau-mo-gai-jye Dah-yeh-gu-suei-pu	Jul Jan	IL, Mt. Tai-ping TC, Ku-kuan	##
Davalliaceae	(S. chinensis MAXON) Araiostegia parvipinnula COPELAND A. perdurans COPELAND	Shiau-mo-gai-jye	Jul Jan Aug	IL, Mt. Tai-ping TC, Ku-kuan TY, Mt. Chiao-pan	##
Davalliaceae	(S. chinensis Maxon) Araiostegia parvipinnula COPELAND A. perdurans COPELAND Davallia formosana HAYATA	Shiau-mo-gai-jye Dah-yeh-gu-suei-pu	Jul Jan	IL, Mt. Tai-ping TC, Ku-kuan	## ## ## ##

	Family <sup>a</sup> )	Species <sup>b)</sup>	Chinese name	Date	Location <sup>c)</sup>	Activity <sup>d</sup> )
		N. biserrata Schott N. hirsutula Presl	Chang-yeh-shen-jye Mau-yeh-shen-jye	Feb Aug	TP, An-keng TP, Yang-ming-shan	+
				Aug	TP, Wu-lai	
	Plagiogyriaceae	Plagiogyria formosana NAKAI	Tai-wan-liu-tzu-jye	Jul	IL, Mt. Tai-ping	#
	Cyatheaceae	Cyathea hancockii Copeland	Han's-sah-lo	Feb	TP, Yang-ming-shan	#
		(Gymnosphaera denticulata COPELAND)		Sep	TP, Yang-ming-shan	# # #
		C. lepifera COPELAND	Pi-tong-shuh	Jan	TC, Ku-kuan	+
		C. podophylla COPELAND	Quei-sah-lo	Aug	TP, Yang-ming-shan	
		C. mentteniana C. Christensen et Terdieu-Blot	Tai-wan-shu-jye	Sep	TP, Yang-ming-shan	+
		C. spinulosa Wallich	Tai-wan-sah-lo	Feb	TP, Wu-lai	mu
	Aspidiaceae	Abacopteris triphylla CHING	Sin-ye-jye	Sep	TP, Yuang-tung-szu	#
	rispidiaceae	(Cyclosorus triphyllus TARDIEU -BLOT)		оср	11, 1 dang-tung-szu	++
		Acrophorus stipellatus MOORE	Nii-lin-mau-jye	Feb	NT, Chi-tou	1111
		Athyrium anisopterum CHRIST	Suh-tii-gai-jye	Aug	NT, Tien-chih	₩ ₩ ₩ ₩
		A. arisanense TAGAWA	A-li-shan-tii-gai-jye	Jul	IL, Mt. Tai-ping	##-
		A. atkinsonii BEDDOME	Yade's-tii-gai-jye	Sep	TP, Yang-ming-phan	<del>iii</del>
	. 4	Ctenitis subglandulosa CHING	Leh-mau-jye	Feb	TP, Wu-lai	
		Cyclosorus acuminatus NARAI	Mau-jye	Aug	TP, Mt. Muh-tsu	+
				Aug	TP, Wu-lai	一 + + + + + +
		C. Lange William Remarks	Wik man man ing	Sep	TP, Yang-ming-shan	+
		C. parasiticus FARWELL	Mih-mau-mau-jye	Feb Aug	TP, An-keng TP, Mt. Muh-tsu	#
	1000			Aug	TP, Wu-lai	+
		C. taiwanensis H. Ito	Tai-wan-mau-jye	Feb	TP, Wu-lai	ш
		Dietyoline griffithii Moore	We's-sen-jye	Feb	TP, Wu-lai	<del>   </del>
		var. wilfordii Moore			-,	un, ;
		(D. griffithri Moore var. pinnatifida Bedd				
		Diplazium dilatatum Blume	Koang-yeh-jih-chy-shuang-gai-jye	Aug	TP, Yang-ming-shan	+
		(Athyrium maximum COPELAND)		Sep	TP, Yang-ming-shan	+
		D. donianum TARDIEU-BLOT	Shi-ping-shuang-gai-jye	Jan	TP, Yang-ming-shan	<del>    </del>
		(Athyrium aphanoneuron OHW1)		Jul	TP, Yang-ming-shan	+
		D. esculentum SWARTZ	Quoh-gou-tsai-jye	Aug	TP, Yang-ming-shan	***
		(Athyrium esculentum Copeland)	Quon-gou-tsat-jye	Feb	TP, An-keng	₩
		D. kawakamii Hayata	Chuan-shang's-shuang-gai-jye	Sep	TP, Yang-ming-shan	
		(Athyrium procerum MILDE)	and a straint gar, 110	COP	11, 1ang-ming-anan	-
		D. mettenianum C. Christensen	Sheng-shan-shuang-gai-jye	Aug	TP, Yang-ming-shan	+
		(Athyrium mettenianum OHWI)	Q		, -(4116 111118 121111	•
		Dryopteris erythrosora O. Kuntze	Hong-bau-lin-mau-jye	Sep	TP, Yang-ming-shan	:##
		D. lepidopoda HAYATA	Hou-yeh-lin-mau-jye	Aug	NT, Mt. Ho-huan	<del>   </del>
		D. scottii Ching	Shu's-lin-mau-jye	Feb	NC, Chi-tou	
		D. sieboldii O. Kuntze	Ding-yeh-lin-mau-jye	Jan	TC, Ku-kuan	
		D. varia O. Kuntze	Nan-hai-lin-mau-jye	Feb	TP, Wu-lai	11
		Tanton haddanii Dananan	Chinah ahin ahin a	Feb	TP, Wu-lai	 ++ ++
		Lastrea beddomei Beddome L. subochthodes Tagawa	Shi-yeh-chin-shing-jye	Aug	NT, Tien-chih	111
		Phegopteris decursive-pinnata Fée	Sie-yeh-chin-shing-jye Chyh-jour-jah-chin-shing-jye	Aug Aug	NT, Mt. Ho-huan TP, Mt. Muh-tsu	+
		(Lastrea decursive-pinnata J. Smrth)	enyn-jour-jan-enm-sning-jye	Sep	TP, Yang-ming-shan	-
		P. subaurita TAGAWA	Al-yeh-jah-chin-shing-jye	Feb	TP, Wu-lai	:#
				Aug	NT, Tien-chih	4
		Polystichopsis amabilis TAGAWA	Dah-fuh-yeh-al-jye	Aug	TP, Yang-ming-shan	
		(Rumohra amabilis CHING)		Ŭ		
		P. quadripinnata Shieh	Mau-bau-fuh-yeh-al-jye	Aug	NT, Tien-chih	·
		Polystichum nepalense C. Christensen	Zuan-gu-al-jye	Aug	NT, Tien-chih	+
	and the second	P. parvipinnulum TAGAYA	Jien-yeh-al-jye	Jul	IL, Mt. Tai-ping	++
		Tectaria kwarenkoensis C. Christensen (T. kwarenkoensis Tagawa)	Hua-lian-san-tsa-jye	Feb	TP, Wu-lai	+ +
		T. subtriphylla Copeland	San tea iva	1	Togo Town Atm	
	Blechnaceae	Blechnum niponicum Makino	San-tsa-jye Zu-pen-wu-mau-jye	Aug Jul	PT, Ken-tin IL, Mt. Tai-ping	<del>  -</del>
		B, orientale Linné	Wu-mau-jye	Aug	TP, Mt. Muh-tsu	<del>    </del>
			· • • • ·	Aug	TP, Yang-ming-shan	+
				Sep	TP, Yang-ming-shan	-
				Dec	TP, Chih-nan-kung	-
		Woodwardia japonica Smith	Zu-pen-gou-jii-jye	Aug	NT, Tien-chih	- <del>     </del> 
		W. orientalis SWARTZ	Dong-fang-gou-jii-jye	Feb	PT, Ken-tin	##
			Aug	TP, Mt. Muh-tsu	<del>    </del>	
	Aspleniaceae	Asplenium laserpitiifolium LAMARCK	Dah-yah-bai nina tia iian i	Aug	TP, Hsin-pei-tou	∰ ++
	Aspiemaceae	A. loriceum Christ	Dah-yeh-hei-ping-tie-jiau-jye Nan-hai-tie-jiau-jye	Feb Feb	TP, Wu-lai	++
		A. nidus Linné	Tai-wan-shan-su-hua	Jan	TP, Wu-lai TC, Ku-kuan	#
		A. normale Don	Sheng-ya-tie-jiau-jye	Mar	NT, Lu-shan	117
		A. trichomanes Linné	Tie-jiau-jye	Aug	NT, Tien-chih	<del>-</del>
	Polypodiaceae	Colysis elliptica Ching	Tuo-yuan-shien-jye	Jan	TC, Ku-kuan	+
		C. wrightii Ching	Lai's-shien-jye	Feb	TP, An-keng	<del>     </del>
				Feb	TP, Wu-lai	##
	Crypsinus hastatus Copeland	San-yeh-fwu-jye	Jan	TC, Ku-kuan	₩	
		Constitution Cons	Window for the	Aug	TP, Wu-lai	+ <del>-</del>
		C. quasidivaricatus Copeland	Yi-shan-fwu-jye	Aug	NT, Tien-chih	##
		Lemmaphyllum microphyllum Prest. Microsorium buergerianum Chino.	Paw-shuh-jye Po's-shing-jye	Feb	TP, Wu-lai	##
		M. fortunei Ching	Fwu's-shing-jye	Aug Jan	TP, Yang-ming-shan TC, Ku-kuan	-
		M. membranifolium CHING	Mo-yeh-shing-jye	Feb	NT, Chi-tou	
			Shwei-long-gu	Mar	NT, Lu-shan	·
		Polypodium niponicum Mettenius				
		P. taiwanianum HAYATA	Jian-yeh-shwei-long-gu	Jul	IL, Mt. Tai-ping	+
		P. taiwanianum HAYATA Pseudodrynaria coronans Ching	Yai-jiang	Jul Jan	NT, Chi-tou	+ ₩
		P. taiwanianum HAYATA		Jan Feb	NT, Chi-tou NT, Chi-tou	+
		P. taiwanianum HAYATA Pseudodrynaria coronans Ching Pyrrosia lingua FARWELL	Yai-jiang Shih-woei	Jan Feb Sep	NT, Chi-tou NT, Chi-tou TP, Yang-ming-shan	11 + # 1 + 1
	Vittariaceae	P. taiwanianum HAYATA Pseudodrynaria coronans Ching	Yai-jiang	Jan Feb	NT, Chi-tou NT, Chi-tou	+ # - + *

a) The families are arranged in the sequence presented by J. Ohwi ("Flora of Japan" Shibundo, Tokyo, 1972), and genera and species are given in alphabetical order within the families.

b) The main source of the nomenclature is the manual of Shieh, and certain different taxa have been supplemented. No attempt was made to apply to Formosan ferns about the same standards of taxonomic recognition accorded comparable taxa in the previous treatment of Japanese ferns, though many inconsistencies are doubtless detectable. Since, in comparison of the present nomenclatures of the Formosan ferns with the previous ones of the Japanese ferns, there are consequently differences in several species, synonyms recorded in the previous work are also included in parentheses in order to avoid confusion.

c) Abbreviations: CY=Chia-yi, IL=I-lan, NT=Nan-tou, PT=Ping-tung, TC=Tai-chung, TP=Tai-pai, TY=Tao-yuan
 d) Assays were carried out on Sarcophaga peregrina.<sup>5)</sup> A test solution containing a methanol extract from a dried plant meterial (10 mg) was injected into each isolated larval abd omen. The results are expressed in terms of the activity indexes which are corresponding to the following average per cent puparium formation: -=0, 0<+≤20, 20<+≤40, 40<+=60, 60<+=50, and 80<+=5100%.</li>

it was again found that families such as Equisetaceae, Lycopodiaceae, Selaginellaceae, Marattiaceae, and Schizaeaceae, whose degree of differentiation is considered to be low from the point of taxonomy, do not contain active species, and yet, among these poorly differentiated families, Osmundaceae contains a strongly active species. Although the number of plant materials presently screened is rather limited for drawing any definite conclusion, it may be said that, as has been noted before, phytoecdysones are distributed in Cyatheaceae, Aspidiaceae, and Polypodiaceae with a high probability, though these still include some species showing no activity.

For more complete understanding of the distribution of phytoecdysones in Formosan ferns, further investigation is now under way.

## Experimental

Materials—A total of 164 specimens, all growing wild, were selected. Samples from whole plants were submitted to the following extraction for bioassay. Sampling from a large fern was so made that the sample was representative of the whole plant, as completely as possible.

Preparation of Test Solutions—A dried plant material (1 g) in MeOH (20 ml) was heated under reflux for 2 hr. After filtration, the extract was concentrated under a reduced pressure to give a dried residue which was dissolved in 10% aqueous EtOH (1 ml), filtered, and the filtrate was used as the test solution.

Biological Assay—Assays were made with larvae of Sarcophaga peregrina as described by Ohtaki, et al.<sup>5)</sup> The final instar larvae, which at 24 hr after the ligation formed puparia at their anterior ends but did not pupate behind ligature, were employed as the test insects.

An aliquot of the test solution (10  $\mu$ l) was injected into each isolated larval abdomen which was kept at 26°. Ten individuals were used for each sample. The assays were evaluated 24 hr after injection when each abdomen was scored as having undergone complete, marked, slight, or no puparium formation; these responses being equated to 100, 75, 50, and 0%, respectively, for the purpose of calculating the weighed average percent puparium formation. The results in Table I are expressed in terms of the activity indices as -, +, +, +, +, and +, these indices corresponding to the following average percent puparium formation: -=0,  $0<+\leq 20$ ,  $20<+\leq 40$ ,  $40<+\leq 60$ ,  $60<+\leq 80$ , and  $80<+\leq 100\%$ .

**Acknowledgement** We are grateful to Mr. M.-T. Kao, National Taiwn University, for identification of the ferns screened.