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A New Depsidone Oxyphysodic Acid isolated from a Lichen Parmelia enteromorpha Ach.

In the thin-layer chromatographic (TLC) study of 16 species of lichens of subgenus *Hypogymnia* belonging to a genus *Parmelia* (Parmeliaceae), Nuno pointed out in 1964¹¹ that physodic acid (I) was always accompanied by an unidentified phenolic substance which was shown up by its characteristic bluish violet FeCl₃ coloration. In this communication, we present the chemical and physicochemical evidence on the structure of the new substance (now named oxyphysodic acid) which leads to the formulation II.

Successive ether and acetone extraction followed by preparative silica gel TLC separation of the lichen *Parmelia enteromorpha* Ach. (syn. *Hypogymnia enteromorpha* (Ach.) Nyl.), which was collected at Mt. Ōdaigahara in Nara prefecture, furnished three known phenolic substances, i.e. physodic acid (I, 1.8%), physodalic acid (III, 1.5%), and atranorin (IV, 1.5%) in addition to oxyphysodic acid (II, 0.5%).

Oxyphysodic acid (II), $C_{26}H_{30}O_9$, m/e 486 (M⁺), mp 187° (decomp.) (colorless needles from aq. MeOH), IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3550, 3150 (br.) (OH), 1690, 1660 (depsidone, CO, COOH), 1620, 1605 (aryl C=C), showed a positive color reaction for FeCl₃ reagent (bluish violet) but was negative for NaOCl and p-phenylenediamine reagents.³⁾ On ethereal diazomethane treatment, II was converted to a methyl ester (IIa), $C_{25}H_{29}O_7 \cdot {\rm COOCH}_3$, m/e 500 (M⁺), mp

$$(b) \qquad \qquad R^1 \quad R^2 \quad R^8$$

$$CO-CH_9CH_9CH_9CH_9CH_3 \qquad \qquad I \quad : H \quad H \quad physodic acid$$

$$(a) \rightarrow CH_2 \qquad \qquad I \quad : H \quad H \quad Ac$$

$$I \quad : H \quad H \quad H \quad physodic acid$$

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1) M. Nuno, J. Japan. Botany, 39, 97 (1964).

2) C.F. Culverson, "Chemical and Botanical Guide to Lichen Products," The University of North Carolina Press, Chapel Hill, North Carolina, U.S.A., 1967, p. 361.

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104—106°, IR $v_{\text{max}}^{\text{CHCb}}$ cm⁻¹: 3250 (OH), 1725 (depsidone, CO), 1670 (COOCH₃), 1615 (aryl C=C), while acetylation of II with acetic anhydride and sulfuric acid afforded a triacetate (IIb), $C_{26}H_{27}O_6(\text{OCOCH}_3)_3$, m/e 612 (M+), mp 104° (decomp.), IR $v_{\text{max}}^{\text{CHCb}}$ cm⁻¹: 1780 (OAc), 1745 (br.), 1725 (sh.) (depsidone, CO, COOH), 1610 (aryl C=C).

The proton magnetic resonance (PMR) examinations of II, IIa, and IIb in comparison with those of physodic acid (I),⁴⁾ methyl physodate (Ia),⁴⁾ and diacetylphysodic acid (Ib) (Table I) indicate that II is a closely related compound with I. The significant difference observed between two is the presence of one more phenolic hydroxyl but one less aromatic proton in II as compared with I. Based on the biogenetic considerations keeping in mind the co-occurrence of both acids and based on lack of one of the *meta*-coupled protons in oxyphysodic acid, two alternative structures II and [A] have become rational for oxyphysodic acid.

TABLE I. δ Values at 90 MHz, Coupling Constants in Hz^a).

CH ₃	$\mathrm{CH_2}$	Ar-H	gree see est others
$I^{b)}$ 0.86 (t, $J=6$) 0.91 (t, $J=6$)		6.60 (1H, s) 6.62 (2H, s)	1.11—2.14 (12H, br, -CH ₂ -×6)
$ \Pi^{b} $ 0.88 (t, $J=6$) 0.92 (t, $J=6$)	(c) 2.81 (br) (a) 3.82 (s) (b) 2.42 (t, <i>J</i> =7 (c) 3.04 (br)		1.11—2.11 (12H, br, -CH ₂ -×6)
	(a) 3.80 (s) (b) 2.38 (t, J =7 (c) 3.20 (br)		3.96 (3H, s, COOCH₃)
IIa ^{c)} 0.90 (t, $J=6$) 0.92 (t, $J=6$)	(a) 3.80 (s) (b) 2.61 (t, J =7 (c) 3.26 (br)	6.40 (1H, s) 6.71 (1H, s)	1.11—1.84 (12H, br, $-CH_2-\times 6$) 3.94 (3H, s, COOCH ₃)
0.91 (t, /=6)	(b) 2.51 (t, $J=7$	7) 7.00 (1H, s)	2) 1.15—2.05 (12H, br, +CH ₂ -×6)
Ibc) 0.88 (t, J=6) 0.89 (t, J=6)	(c) 3.00 (br) (a) 4.05 (s) (b) 2.51 (t, <i>I</i> =7	7,03 (1H, d, J = 6.96 (1H, s) 7,05 (1H, s)	2) 2.24, 2.30 (3H each, s, OAc×2) 1.04—1.85 (12H, br, -CH ₂ -×6)
og desilan Mögrig. Omre med sedager	(c) 3.09 (br)	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	2.23, 2.27, 2.41

a) Abbreviations: br=broad signal, d=doublet, s=singlet, t=triplet

Irradiation at δ 4.03 (methylene (a) in Ib) or δ 4.05 (methylene (a) in IIb) resulted in the increase of aromatic proton signal intensity at δ 6.87 in Ib (12%) or at δ 6.96 in IIb (8%) due to the nuclear Overhauser effect. Therefore, the substituent disposition in the left aromatic ring in oxyphysodic acid has been corroborated as in II.

Formic acid treatment of oxyphysodic acid furnished an oxyphysodone (VI), $C_{25}H_{28}O_6$, m/e 424 (M⁺), mp 202—203°, IR ν_{\max}^{KBF} cm⁻¹: 3200 (OH), 1695 (pyrone), PMR: 0.86, 0.91 (3H each, t, J=6 Hz, -CH₃×2), 1.16—1.95 (12H, br, -CH₂-×6), 2.25, 2.72 (2H each, t, J=7, Ar-CH₂-×2), 6.13, 6.52 (1H each, s, Ar-H, pyrone-H), 6.26, 6.35 (1H each, d, J=3, Ar-H ×2). The latter was then subjected to nitric acid oxidation in acetic acid⁵ to furnish an orange yellow benzoquinone derivative, $C_{11}H_{14}O_3$, mp 172—173°, IR ν_{\max}^{CHO} cm⁻¹: 3300 (OH), 1705, 1660, 1619 (benzoquinone), which was proved identical in all respects (TLC, mixed

b) measured in d_6 -DMSO

c) measured in CDCl $_{\mathrm{8}}$

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mp, and infrared (IR)) with 2-hydroxy-6-pentyl-1,4-benzoquinone (VII), PMR: 0.88 (3H, t, J=6, -CH₃), 1.05—1.90 (6H, br., -CH₂-×3), 2.45 (2H, t-like, J=6, -C=C-CH₂-), 6.10, 6.46 (1H each, s, ring H×2), prepared analogously from I via physodone (V).60

Consequently, the structure of oxyphysodic acid has been established as II.

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Neridienone A, a C₂₁-Steroid in Nerium odorum¹⁾

Concerning the constituents of *Nerium odorum*, we have previously reported the isolation of pregnenolone glucosides²⁾ from its root bark. This communication deals with the isolation and the structure elucidation of neridienone A, a pregnane compound in the same source.

The methanol percolate of the root bark was extracted with benzene, and through successive column chromatography of the benzene extractives followed by crystallization from acetone-hexane, a substance, sensitive to 2,4-dinitrophenylhydrazine reagent on thin-layer chromatography, was obtained as prisms and named neridienone A (NA). NA, (mp 210-211°, $[\alpha]_D + 71.5^\circ$, Mass. Calcd. for $C_{21}H_{26}O_3$ m/e 326.1875; Found 326.1882), shows nuclear magnetic resonance (NMR) signals at δ (ppm) 0.98 (3H, s), 1.16 (3H, s), 2.43 (3H, s), 3.71 (1H, dd), 5.67 (1H, s), 5.81 (1H, s), 6.12 (2H, s) and 6.99 (1H, dd). Two oxygens in NA were found to belong to the unsaturated ketones, 4,6-dien-3-one and 16-en-20-one of steroid system, according to the ultraviolet absorptions at 283 nm (ε 24100) and 245 nm (ε 12900), infrared (IR) bands at 1665, 1640, 1612 and 1580 cm⁻¹, and NMR signals at 5.67 (C₄-H), 6.12 (C_{6,7}-H) and 6.99 ppm (C₁₆-H). The IR bands at 3420 and 3320 cm⁻¹ as well as the disappearance of the peak at δ 5.81 ppm with D₂O exchange indicated that the remaining oxygen function forms a hydroxyl group. NA, on reduction with NaBH₄, gave three products, dihydro-, tetrahydroand hexahydro-compounds, the first of which, (Calcd. for C₂₁H₂₈O₃ 328.2038; Found 328.2079) bears no 16-en-20-one system in its molecule, and afforded a diacetate (Calcd. for $C_{25}H_{32}O_5$ 412.2249; Found 412.2233) on acetylation, whereas NA was unreactive with Ac_2O and pyridine. The above results suggested that the hydroxyl group is oriented in the position under the influence of 16-en-20-one, and the location and the configuration were determined as C_{12} - β -OH

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