

Dehydro-*para*-asebotin, a New Chalcone Glucoside in the Flowers of *Gnaphalium affine* D. DON

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Besides luteolin 4'-D-glucoside (I) and luteolin (II), a chalcone glycoside (III), $C_{22}H_{24}O_{10} \cdot 1\frac{1}{2}H_2O$, mp 199—200°, $[\alpha]_D^{20} -85.2^\circ$ (methanol), ultraviolet spectrum (UV) λ_{max}^{EtOH} 368 nm, was isolated from the flowers of *Gnaphalium affine* D. DON. III was acid hydrolyzed to give glucose and a flavanone (V), mp 256°, UV λ_{max}^{EtOH} 228, 284 nm, which was identified as 4',7-dihydroxy-5-methoxyflavanone by chemical and spectral data and by direct comparison of the diethyl ether (VIII) with an authentic sample. The sample, 4',7-diethoxy-5-methoxyflavanone (VIII) was prepared, according to the result of a preliminary experiment (Table I), by stepwise alkylations of naringenin (IX) with an ether solution of diazoethane in absolute methanol for 3 hr and then with diazomethane in 70% methanol for 24 hr. Further examination of the methanolysis products of III permethylate (XV) and the spectral data of XV evidenced that III is 4'-O- β -D-glucopyranoside of 2',4,4'-trihydroxy-6'-methoxychalcone (dehydro-*para*-asebotin), from which V is yielded on acid treatment. This is the first isolation of III and V from natural sources.

In the previous paper,²⁾ the presence of luteolin 4'-D-glucoside (I) in the flowers of *Gnaphalium affine* D. DON was demonstrated. Now, in addition to I, luteolin (II) and a new chalcone glucoside (III) have been isolated from the same source, and this paper describes the identification of III as 2',4,4'-trihydroxy-6'-methoxychalcone 4'- β -D-glucopyranoside.

The methanol extracts of the fresh flowers were treated with lead acetate, and from the precipitates II was obtained, while the non-precipitable fraction gave I and III by polyamide column chromatography.

III was crystallized from dilute methanol as yellow needles, $C_{22}H_{24}O_{10} \cdot 1\frac{1}{2}H_2O$, mp 199—200°, $[\alpha]_D^{20} -85.2^\circ$ (methanol), ultraviolet spectrum (UV) λ_{max}^{EtOH} 368 nm. It gave, on acetylation with acetic anhydride and pyridine, a syrupy acetate (IV), UV λ_{max}^{EtOH} 306 nm, and, on acid hydrolysis, an aglycone (V), $C_{16}H_{14}O_5$, mp 256°, UV λ_{max}^{EtOH} 228, 284 nm, and glucose in respective yields 59—66% and 37% (by anthrone method³⁾).

V afforded a diacetate (VI), $C_{20}H_{18}O_7$, mp 174—175°, UV λ_{max}^{EtOH} 285 nm, a dimethyl ether (VII), $C_{18}H_{18}O_5$, mp 121°, UV λ_{max}^{EtOH} 228, 282 nm, and a diethyl ether (VIII), $C_{20}H_{22}O_5$, mp 121—122°, UV λ_{max}^{EtOH} 229, 282 nm, and was presumed to be 4',7-dihydroxy-5-methoxyflavanone on the basis of color reactions,⁴⁾ nuclear magnetic resonance (NMR)^{5,6)} and mass⁷⁾ spectral data and identification of VII with an authentic 4',5,7-trimethoxyflavanone.⁸⁾

- 1) Location: a) Kurokami, Kumamoto; b) Katakasu, Fukuoka.
- 2) M. Aritomi, M. Shimojo, and T. Mazaki, *Yakugaku Zasshi*, **84**, 895 (1964).
- 3) K. Anno and N. Seno, "Seibutsu Kagaku Jikkenho," Vol. 11, Kyoritsu Shuppan Kabushiki Gaisha, Tokyo, 1968, p. 13.
- 4) K. Venkataraman, "The Chemistry of Flavonoid Compounds," ed. by T.A. Geissman, Pergamon Press, Oxford, 1962, p. 70.
- 5) T.J. Batterham and R.J. Highet, *Aust. J. Chem.*, **17**, 428 (1964).
- 6) T.J. Mabry, K.R. Markham, and M.B. Thomas, "The Systematic Identification of Flavonoids," Springer-Verlag, Berlin, Heidelberg, New York, 1970, p. 252.
- 7) S. Sasaki, Y. Itagaki, T. Kurokawa, E. Watanabe, and T. Aoyama, *Shitsuryo Bunseki*, **14**, 82 (1966).
- 8) K. Nakazawa and S. Matsuura, *Yakugaku Zasshi*, **75**, 469 (1955).

In order to get the chemical evidence for the above presumption, it was hoped to prepare 4',7-diethoxy-5-methoxyflavanone as a reference compound. So far, it has been reported that methylations of 4',5,7-trihydroxyflavanone (naringenin) (IX) with an excess and an equimolar amount of diazomethane provide 4',7-diether and 7-monoether, respectively.⁹⁾ As a preliminary experiment, methylation of IX with an ether solution of diazomethane in some different conditions was then undertaken, and it was found that the kind of product was dependent on solvent and reaction time. Thus, the reaction for 3 hr in anhydrous dioxane solution gave mainly 7-monomethyl ether (X), in anhydrous methanol for 3 hr provided 4',7-dimethyl ether (XI) as an only isolable product, whereas in the reaction for 24 hr in 70% methanol even the chelated hydroxyl group was methylated to give 4',5,7-trimethyl ether (VII) (Table I).

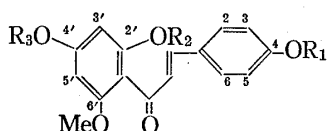
TABLE I. Yields (%) of Methyl Ethers on Methylation of Naringenin (IX) with Diazomethane-ether in Different Media and in Different Times

Products	Solvent and reaction time					
	Abs. dioxane		Abs. MeOH		70% MeOH	
	3 hr	24 hr	3 hr	24 hr	3 hr	24 hr
7-Mono-O-methylnaringenin (X)	70	47	+	—	—	—
4',7-Di-O-methylnaringenin (XI)	0.5	5	42	33	22	—
Tri-O-methylnaringenin (VII)	—	—	+	31	48	67

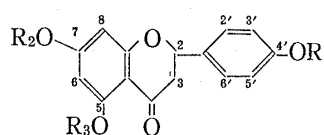
+ : detected on TLC as a minor product but could not be isolated in crystalline state
 — : could not be detected on TLC

Taking the above result into account, an analogous ethylation of IX in anhydrous methanol with an ether solution of diazoethane¹⁰⁾ for 3 hr was conducted and 4',7-di-O-ethyl-naringenin (XII), $C_{19}H_{20}O_5$, mp 97—98°, UV λ_{\max}^{EtOH} 226, 289 nm, was obtained, accompanied by 7-mono-O-ethyl-naringenin (XIII), $C_{17}H_{16}O_5$, mp 130—131°, UV λ_{\max}^{EtOH} 222, 288 nm. Their structures were determined by color reaction with ferric chloride⁴⁾ and NMR spectral data^{5,6)} of XII and NMR^{5,6)} as well as mass⁷⁾ spectra of mono-O-methylated XIII (XIV), $C_{18}H_{18}O_5$, mp 111°, UV λ_{\max}^{EtOH} 224, 289 nm. Subsequent methylation of XII in 70% methanol with diazomethane for 24 hr provided 4',7-diethoxy-5-methoxyflavanone, $C_{20}H_{22}O_5$, mp 121—122°, which was identified with VIII, furnishing the definite proof for the structure V.

Treatment of III with dimethyl sulfate and potassium carbonate in dry acetone followed by the Kuhn's methylation¹¹⁾ provided hexamethyl ether (XV), $C_{28}H_{36}O_{10}$, mp 112—113°,



- III: $R_1=R_2=H$, $R_3=\beta$ -D-glucopyranosyl
 IV: $R_1=R_2=OAc$, $R_3=\text{tetra-O-acetyl-}\beta$ -D-glucopyranosyl
 XV: $R_1=R_2=Me$, $R_3=\text{tetra-O-methyl-}\beta$ -D-glucopyranosyl
 XVI: $R_1=R_2=Me$, $R_3=H$



- V: $R_1=R_2=H$, $R_3=Me$
 VI: $R_1=R_2=OAc$, $R_3=Me$
 VII: $R_1=R_2=R_3=Me$
 VIII: $R_1=R_2=Et$, $R_3=Me$
 IX: $R_1=R_2=R_3=H$
 X: $R_1=R_3=H$, $R_2=Me$
 XI: $R_1=R_2=Me$, $R_3=H$
 XII: $R_1=R_2=Et$, $R_3=H$
 XIII: $R_1=R_3=H$, $R_2=Et$
 XIV: $R_1=Me$, $R_2=Et$, $R_3=H$

- 9) J. Shinoda and S. Sato, *Yakugaku Zasshi*, **48**, 933 (1928).
 10) K. Makino, A. Watanabe, and Y. Joh, *Seikagaku*, **32**, 788 (1960).
 11) R. Kuhn, H. Trischmann, and I. Löw, *Angew. Chem.*, **67**, 32 (1955).

$[\alpha]_D^{24}$ —18.6° (chloroform), UV $\lambda_{\max}^{\text{EtOH}}$ 327 nm, which was methanolized to give methyl 2,3,4,6-tetra-O-methyl- α - and β -glucopyranosides and a trimethoxy-monohydroxychalcone (XVI), $\text{C}_{18}\text{H}_{18}\text{O}_5$, mp 204—205°, UV $\lambda_{\max}^{\text{EtOH}}$ 326 nm, identical in all respects with a synthetic specimen of 2',4,6'-trimethoxy-4'-hydroxychalcone,¹²⁾ mp 202—203°. Accordingly III should be the 4'-glucopyranoside of 2',4,4',6'-tetrahydroxychalcone or its methyl ether. Since III was acid hydrolyzed to give V,¹³⁾ the aglycone is regarded as 6'-(2'-) monomethyl ether.

Consequently, if the component glucose is assumed to be β -linked¹⁴⁾ D-enantiomer as is common in natural flavonoid glycosides, III is defined as 2',4,4'-trihydroxy-6'-methoxychalcone 4'- β -D-glucopyranoside (dehydro-*para*-asebotin).

III was reported in 1944 by Zemplen, *et al.*¹⁵⁾ as an intermediate in the synthetic studies of phlorizin and related dihydrochalcone glucosides, but as far as we are aware, natural existence of III and V has never been reported.¹⁶⁾

Experimental

Melting points were measured on Kofler block (Yazawa BY-1) and are uncorrected. Optical rotations, UV and mass spectra were respectively taken with JASCO automatic polarimeter DIP-SL, Hitachi spectrophotometer 181 and JMS-01-SG mass spectrometer at an ionizing voltage of 75 eV. NMR spectra were determined on a JEOL-JNM-C-60H spectrometer at 60 MHz using Me_4Si as internal reference. Chemical shifts are given in δ (ppm) values. Abbreviations used are s=singlet, d=doublet, t=triplet, q=quartet, m=multiplet. Unless otherwise indicated, thin-layer chromatographies (TLC) were carried out on "Kieselgel G nach Stahl" (Merck) and the spots were visualized by charring with 10% H_2SO_4 . For column chromatographies "Kieselgel 60" (Merck) (a mixture (5:1, w/w) of "0.063—0.200 mm" and "unter 0.08 mm") was employed. The solvent systems (v/v) were hexane-acetone (9:1) (solv. 1), hexane-acetone (8:2) (solv. 2), benzene-acetone (20:1) (solv. 3), benzene-acetone (9:1) (solv. 4), benzene-acetone (8:2) (solv. 5), benzene-EtOAc (8:2) (solv. 6) and CHCl_3 -MeOH (8:2) (solv. 7). The phenolic compounds and reducing sugars were examined on Avicel SF in the solvent systems of 40% (v/v) AcOH (visualizing reagent, 10% Na_2CO_3 (method 1) or diazotized *p*-sulfanilic acid (method 2)) and BuOH-pyridine- H_2O (3:2:1, v/v) (reagent, *p*-anisidine-HCl (method 3)). Gas-liquid chromatographies (GLC) were measured at column temperature 100° with Toyo GC-SD equipped with hydrogen flame detector and glass columns (4 mm \times 1.5 m) packed with butanediol succinate (5%) on Chromosorb W (80—100 mesh). Carrier gas was nitrogen (50 ml/min).

Extraction and Isolation of Constituents—The fresh flowers of *G. affine* D. Don were percolated twice with MeOH. The MeOH extract was evaporated *in vacuo* to give a dark brown liquid, which was repeatedly extracted with benzene and then with EtOAc. The EtOAc extracts were dissolved in MeOH and treated with a MeOH solution of $\text{Pb}(\text{OAc})_2$, giving precipitable (Fr. 1) and non-precipitable (Fr. 2) fractions. They were bubbled with H_2S , and the Pb salts were filtered off.

The MeOH solution of Fr. 1, which exhibited at least four spots at *Rf* 0.74 (brown), 0.58 (orange), 0.35 (yellow) and 0.26 (yellow) on TLC (method 1), was placed on a column of polyamide (C-200, Wako Pure Chemical Industries, Ltd.) and eluted with MeOH. The eluate was crystallized from MeOH, giving II, yellow needles, mp >230°, UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 255 (4.51), 352 (4.56). Its infrared (IR) spectrum was identical with that of authentic luteolin, and the acetate, colorless needles (from MeOH), showed mp 221—223°, either alone or on admixture with luteolin tetraacetate.

Fr. 2 revealed at least three spots (*Rf* 0.58 (orange), 0.35 (yellow) (tailing) and 0.22 (brown)) on TLC (method 1) and gave, by polyamide column chromatography, the compounds of *Rf* 0.58 and 0.35 respectively in TLC homogeneous state.

Crystallization of the compound of *Rf* 0.35 from H_2O furnished I as pale yellow needles (yield 0.3%), which melted at around 164—178°. Its IR spectrum was identical with that of I reported previously.²⁾

12) N. Narasimhachari and T.R. Seshadri, *Proc. Indian Acad. Sci.*, **27A**, 223 (1948) [*C.A.*, **44**, 1493 (1950)].

13) a) T.R. Seshadri, "The Chemistry of Flavonoid Compounds," ed. by T.A. Geissman, Pergamon Press, Oxford, 1962, p. 156; b) M. Shimokoriyama, *ibid.*, p. 286.

14) In harmony with the finding of Mabry, *et al.*,⁶⁾ that the anomeric proton of 7-glucosidoxyflavanoids appears as complex multiplet, XV exhibited on its NMR spectrum the anomeric proton signal at 5.07 ppm (4.84 ppm at 100 MHz) as broad doublet. The spacing between the major peaks of them measured 9 Hz (*cf.* Experimental).

15) G. Zemplen, R. Bognar, and K. Thiele, *Chem. Ber.*, **77**, 446 (1944).

16) Birch, *et al.* (A.J. Birch and M. Salahuddin, *Tetrahedron Letters*, **1964**, 2211) reported isolation of naringenin and 5-O-methylnaringenin from *Xanthorrhoea preissii*, but the details have not appeared yet.

The compound of *Rf* 0.58 was dissolved in hot 30% MeOH and then cooled. The yellow precipitates separated were removed by filtration and the filtrate was left stand for one week to give III as yellow needles. Yield 0.07%. It melted at 199–200° with the foregoing sintering at 140–160°, and showed $[\alpha]_D^{20} -85.2^\circ$ (*c* 0.71, MeOH), UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 368 (4.51) (lit.,¹⁵) changes to a glassy mass at 162° and melts at 199° (hydrated form) or 202–203° (nonhydrated form), $[\alpha]_D^{20} -38.6^\circ$ (pyridine). *Anal.* Calcd. for $\text{C}_{22}\text{H}_{24}\text{O}_{10} \cdot 1\frac{1}{2}\text{H}_2\text{O}$: C, 55.57; H, 5.72; H₂O, 5.7. Found: C, 55.57; H, 5.56; H₂O, 6.2. NMR ((CD₃)₂SO) δ ppm: 7.54 (2H, s, olefinic H), 7.56 and 6.82 (4H, two d, *J*=8 Hz, C₂-, C₆-, C₃- and C₅-H), 6.20 and 6.21 (2H, two d, *J*=2 Hz, C₃'- and C₅'-H), 3.89 (3H, s, OMe), 10.07 (H, s, C₄-OH), 13.07 (H, s, C₂'-OH). Dark brown with FeCl₃, negative to Mg-HCl and Zn-HCl tests.

III Hexaacetate (IV)—III (516 mg) was acetylated with Ac₂O (2 ml) and pyridine (2 ml) at room temperature overnight, and the product (*Rf* 0.18 (solv. 6)) was purified through column chromatography over silica gel (solv. 6) to give IV as colorless syrup (yield 74 mg), UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 306 (4.18). NMR (CDCl₃) δ ppm: 7.66 and 6.90 (2H, two d, *J*=17 Hz, olefinic H), 7.57 and 7.13 (4H, two d, *J*=8 Hz, C₂-, C₆-, C₃- and C₅-H), 6.54 and 6.44 (2H, two d, *J*=2 Hz, C₃'- and C₅'-H), 3.80 (3H, s, OMe), 2.30 and 2.17 (6H, two s, OAc \times 2 on aromatic ring), 2.08, 2.07 and 2.05 (12H, three s, OAc \times 4 on sugar moiety), 5.2, 4.2 and 3.8 (7H, m, protons on sugar moiety).

Acid Hydrolysis of III—III (284.3 mg) in H₂O (40 ml) and 1*N* HCl (10 ml) was heated on a water bath for 4 hr. After being cooled, the precipitates (V) (*Rf* 0.71 (yellow) (method 2)) were collected (yield 149.2 mg). Extraction of the filtrate with EtOAc gave further crop (18.5 mg) of V (*Rf* 0.71). Total yield 167.7 mg, 59%. In another hydrolysis III (57.7 mg) yielded V (38.0 mg, 66%).

The aqueous solution was neutralized by passing through a column of Amberlite IR 45 (OH⁻), evaporated *in vacuo* to a syrup and examined by TLC (method 3). Only one spot was detected at *Rf* 0.24, identical with that of *D*-glucose. Quantitative determination of glucose after anthrone method,⁹ 21.5 mg, 37%.

Aglycone (V)—V obtained above was recrystallized from dimethylsulfoxide-H₂O to give almost colorless needles, mp 256°, UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 228 (4.43), 284 (4.28) (lit.,¹⁵) mp 263° with foregoing browning at 248°. *Anal.* Calcd. for C₁₆H₁₄O₅: C, 67.12; H, 4.93. Found: C, 67.05; H, 4.90. Red with Mg-HCl, reddish violet with Zn-HCl and no color with FeCl₃. NMR ((CD₃)₂SO) δ ppm: 5.36, 3.05 and 2.52 (3H, ABX system due to C₂- and C₃-H, *J*=4, 16 and 12 Hz), 7.33 and 6.81 (4H, two d, *J*=8 Hz, C₂'-, C₆'-, C₃'- and C₅'-H), 6.10 and 5.99 (2H, two d, *J*=2 Hz, C₃- and C₆-H), 3.76 (3H, s, OMe), 9.6 and 10.55 (2H, two broad s, OH \times 2).

V Diacetate (VI)—V (76 mg) was acetylated with Ac₂O (2 ml) and pyridine (2 ml) at room temperature overnight to afford VI as colorless needles (from CHCl₃-hexane), mp 174–175°, UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 285 (3.30) (lit.,¹⁵) mp 176°. *Anal.* Calcd. for C₂₀H₁₈O₇: C, 64.86; H, 4.90. Found: C, 64.41; H, 4.81. NMR (CDCl₃) δ ppm: 5.31, 3.05 and 2.71 (3H, ABX system due to C₂- and C₃-H, *J*=5, 10 and 16 Hz), 7.24 and 6.91 (4H, two d, *J*=8 Hz, C₂'-, C₆'-, C₃'- and C₄'-H), 6.29 and 6.17 (2H, two d, *J*=2 Hz, C₃- and C₆-H), 3.81 (3H, s, OMe), 2.23 (6H, s, OAc \times 2).

V Dimethyl Ether (VII)—V (159 mg) in a mixture of dioxane (15 ml) and MeOH (5 ml) was methylated with ether solution of CH₂N₂ at room temperature overnight. The product (*Rf* 0.86 (violet), 0.82 (brown), 0.63 (red), 0.49 (yellow), 0.35 (dark violet) (solv. 5)) was chromatographed (solv. 3), and the fraction showing one spot at *Rf* 0.49 was crystallized from 70% MeOH to give VII as colorless needles (144 mg), mp 121°, UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 228 (4.24), 282 (4.02). *Anal.* Calcd. for C₁₈H₁₈O₅: C, 68.78; H, 5.77. Found: C, 68.58; H, 5.88. NMR (CDCl₃) δ ppm: 5.31, 3.06 and 2.67 (3H, ABX system due to C₂- and C₃-H, *J*=4, 12 and 17 Hz), 7.33 and 6.87 (4H, two d, *J*=8 Hz, C₂'-, C₆'-, C₃'- and C₅'-H), 6.03 and 6.09 (2H, two d, *J*=2 Hz, C₆- and C₈-H), 3.78 and 3.87 (9H, two s, OMe \times 3). Mixed melting point with authentic 4',5,7-trimethoxyflavanone,⁸ mp 121–122°, showed no depression and their IR spectra were superimposable.

V Diethyl Ether (VIII)—V (132 mg) in EtOH (100 ml) was treated with an ether solution of CH₃CHN₂¹⁰ at room temperature for 4 hr. The reaction mixture (*Rf* 0.59 (major) and 0.47 (solv. 5)) was passed through a silica gel column (solv. 4), and the major product of *Rf* 0.59 was crystallized from benzene-hexane (1:4, v/v) to provide VIII as pale yellow needles, mp 121–122°, UV $\lambda_{\max}^{\text{EtOH}}$ nm(log ϵ): 229 (4.58), 282 (4.38). *Anal.* Calcd. for C₂₀H₂₂O₅: C, 70.16; H, 6.48. Found: C, 70.24; H, 6.56. Mass Spectrum *m/e* (%): 342 (M⁺) (25), 221 (M⁺-C₆H₄(OEt)) (9), 194 (C₆H₂(OEt)(OMe)(=O)(=C=O)⁺) (18), 149 (*m/e* 194-OEt) (100), 148 (C₆H₄(OEt)(-CH=CH₂)⁺) (26).

Methylation of IX (Table I)—To a solution of IX (500 mg) in 20 ml of a solvent (anhyd. dioxane (purified after the procedure of Hess, *et al.*¹⁷), anhyd. MeOH and 70% MeOH), and ether solution (50 ml) of CH₂N₂ prepared from 5 g of nitrosomethylurea was added at 0°. The solution in 70% MeOH quickly consumed CH₂N₂ and, hence, further amount (50 ml) of the reagent was added. Each mixture was left stand for 3 and 24 hr at 0°, the excess of CH₂N₂ decomposed with AcOH, the solvent evaporated *in vacuo*, and the residue was chromatographed (solv. 4) over silica gel (20 g).

In anhyd. dioxane: Both products (reaction times 3 and 24 hr) exhibited six spots at *Rf* 0.85 (yellow) (XI), 0.82 (pink, trace), 0.73 (pink, trace), 0.57 (yellow) (X), 0.28 (black, trace) and 0.15 (black, trace) on

17) K. Hess and H. Frahm, *Chem. Ber.*, **71**, 2627 (1938).

TLC (solv. 4), and gave X (70% (3 hr) and 47% (24 hr)) and XI (0.5% (3 hr) and 5% (24 hr)). X was crystallized from benzene as colorless needles, mp 148—149° (lit.,⁹ mp 150°), UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 287 (4.28). Brown with FeCl₃. Anal. Calcd. for C₁₆H₁₄O₅: C, 67.12; H, 4.93. Found: C, 67.03; H, 4.85. NMR ((CD₃)₂CO) δ ppm: 5.49, 3.26 and 2.74 (3H, ABX system due to C₂- and C₃-H, $J=4, 12$ and 17 Hz), 7.41 and 6.90 (4H, two d, $J=9$ Hz, C_{2'}-, C_{6'}-, C_{3'}- and C_{5'}-H), 6.05 (2H, s, C₆- and C₈-H), 3.87 (3H, s, OMe), 8.52 (H, s, C_{4'}-OH), 12.13 (H, s, C₅-OH). XI was obtained as TLC homogeneous colorless needles, mp 106—110° (lit.,⁹ mp 118°), identified with an authentic sample described later by cochromatography and mixed melting point determination.

In anhyd. MeOH: The intensities of the spot at R_f 0.85 (XI), 0.82, 0.73, 0.28 and 0.15 were enhanced and three additional spots were newly detected at R_f 0.65 (pink), 0.51 (black) and 0.38 (VII) (yellow).

From the reaction mixture for 3 hr, only XI (yield 42%) was isolated as colorless needles (from acetone-H₂O), mp 113—114° (lit.,⁹ mp 118°), UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 226 (4.45), 288 (4.27). Anal. Calcd. for C₁₇H₁₆O₅: C, 67.99; H, 5.37. Found: C, 68.08; H, 5.40. NMR (CDCl₃) δ ppm: 5.34, 3.15 and 2.72 (3H, ABX system due to C₂- and C₃-H, $J=4, 12$ and 17 Hz), 7.39 and 6.94 (4H, two d, $J=9$ Hz, C_{2'}-, C_{6'}-, C_{3'}- and C_{5'}-H), 6.06 (2H, s, C₆- and C₈-H), 3.82 and 3.84 (6H, two s, OMe \times 2), 12.03 (H, s, C₅-OH). The compounds of R_f 0.57 and 0.38 were respectively identified with X and VII by cochromatography on TLC.

The reaction mixture for 24 hr showed no spot of R_f 0.57 (X) on TLC, and XI (yield 33%), mp 113—114°, and VII (yield 31%), mp 119—121°, were provided.

In 70% MeOH: A significant enhancement of the intensities of the spots, R_f 0.82, 0.73, 0.65, 0.51, 0.28 and 0.15, was observed. The reaction for 3 hr provided XI (yield 22%) and VII (yield 48%). The reaction mixture for 24 hr revealed no spot of XI on TLC and gave only VII (yield 67%).

Ethylation of IX with Diazoethane to XII and XIII—To a solution of IX (500 mg) in anhyd. MeOH (20 ml) was added at 0° an ether solution of CH₃CHN₂ prepared from 5 g of N-ethyl-N-nitroso-N'-nitroguanidine.¹⁰ CH₃CHN₂ was consumed completely in 3 hr at 0°. The reaction mixture (R_f 0.70 and 0.23 (solv. 2)) was chromatographed (solv. 4). The fraction of R_f 0.70 was crystallized from acetone-H₂O to give XII as colorless needles (yield 155 mg), mp 97—98°, UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 226 (4.47), 289 (4.29). Anal. Calcd. for C₁₉H₂₀O₅: C, 69.50; H, 6.14. Found: C, 69.58; H, 6.20. Brown with FeCl₃. NMR (CDCl₃) δ ppm: 5.34, 3.14 and 2.71 (3H, ABX system due to C₂- and C₃-H, $J=5, 12$ and 17 Hz), 7.36 and 6.92 (4H, two d, $J=8$ Hz, C_{2'}-, C_{6'}-, C_{3'}- and C_{5'}-H), 6.04 (2H, s, C₆- and C₈-H), 1.36 and 1.42 (3H \times 2, two t, $J=6$ Hz, OCH₂CH₃ \times 2), 4.03 and 4.05 (2H \times 2, two q, $J=6$ Hz, OCH₂CH₃ \times 2), 12.03 (H, s, C₅-OH). The fraction of R_f 0.23 was crystallized from benzene to afford XIII as colorless needles (yield 198 mg), mp 130—131°, UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 222 (4.45), 288 (4.30). Anal. Calcd. for C₁₇H₁₆O₅: C, 67.99; H, 5.37. Found: C, 67.73; H, 5.30. Brown with FeCl₃. NMR (CDCl₃) δ ppm: 5.27, 3.07 and 2.66 (3H, ABX due to C₂- and C₃-H, $J=5, 12$ and 16 Hz), 7.19 and 6.75 (4H, two d, $J=9$ Hz, C_{2'}-, C_{6'}-, C_{3'}- and C_{5'}-H), 5.94 (2H, s, C₆- and C₈-H), 1.36 (3H, t, $J=6$ Hz, OCH₂CH₃), 3.97 (2H, q, $J=6$ Hz, OCH₂CH₃), 11.76 (H, s, C₅-OH). Methylation of XIII (0.31 g) in anhyd. MeOH (20 ml) with CH₂N₂ for 3 hr and subsequent crystallization of the product from MeOH provided XIV as colorless needles (yield 0.14 g), mp 111°, UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 224 (4.52), 289 (4.35). Anal. Calcd. for C₁₈H₁₈O₅: C, 68.78; H, 5.77. Found: C, 68.92; H, 5.73. NMR (CDCl₃) δ ppm: 5.29, 3.10 and 2.67 (3H, ABX due to C₂- and C₃-H, $J=5, 12$ and 18 Hz), 7.26 and 6.84 (4H, two d, $J=8$ Hz, C_{2'}-, C_{6'}-, C_{3'}- and C_{5'}-H), 5.95 (2H, s, C₆- and C₈-H), 1.38 (3H, t, $J=6$ Hz, OCH₂CH₃), 3.96 (2H, q, $J=6$ Hz, OCH₂CH₃), 3.78 (3H, s, OMe), 11.81 (H, s, C₅-OH). Mass Spectrum m/e (%): 314 (M⁺) (74), 207 (M⁺-C₆H₄(OMe)) (25), 180 (C₆H₂(OEt)(OH)(=O)(=C=O)⁺) (19), 135 (m/e 180 - OEt) (19), 134 (C₆H₄(OMe)(-CH=CH₂)⁺) (100).

Methylation of XII to VIII—Because of the poor solubility in 70% MeOH, XII (288 mg) was dissolved in a mixture of ether (10 ml) and 70% MeOH (10 ml), and methylated with an ether solution of CH₂N₂ prepared from 5 g of nitrosomethylurea at 0° for 24 hr. The product (R_f 0.70 (pink), 0.65 (pink), 0.53 (brown), 0.48 (yellow), 0.40 (pink), solv. 4) was chromatographed (solv. 4). The fraction of R_f 0.48 (252 mg) was crystallized from 70% MeOH to give VIII as colorless needles, mp 121—122°. No coloration with FeCl₃. Anal. Calcd. for C₂₀H₂₂O₅: C, 70.16; H, 6.48. Found: C, 70.16; H, 6.46. On admixture with VIII derived from V, it did not depress the melting point and their IR spectra were identical.

III Hexamethyl Ether (XV)—A mixture of III (978 mg), Me₂SO₄ (3.38 g) and K₂CO₃ (12.0 g) in anhydrous acetone (50 ml) was refluxed for 24 hr. The reaction mixture was filtered and evaporated to dryness. The residue (1.63 g) was stirred in dimethylformamide (20 ml) with MeI (2 g) and Ag₂O (3.2 g) for 24 hr. After removal of Ag salt, the reaction mixture was evaporated *in vacuo* and the residue (R_f 0.63, 0.45, 0.39, 0.32 (major), 0.23—0.09 (solv. 4)) was chromatographed (solv. 4). Crystallization of the fraction of R_f 0.32 from 70% MeOH gave XV (yield 329 mg) as pale yellow needles, mp 112—113°, $[\alpha]_{\text{D}}^{24} -18.6^\circ$ (c 0.98, CHCl₃), UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 327 (4.41). Anal. Calcd. for C₂₈H₃₆O₁₀: C, 63.14; H, 6.81. Found: C, 63.27; H, 6.80. NMR (CDCl₃) δ ppm: 7.42 and 6.88 (2H, two d, $J=17$ Hz, olefinic H), 7.59 and 6.98 (4H, two d, $J=9$ Hz, C₂-, C₆-, C₃- and C₅-H), 6.47 (2H, s, C_{3'}- and C_{5'}-H), 3.46, 3.63, 3.72, 3.77, 3.83 and 3.92 (3H \times 6, six s, OMe \times 6), 5.07 (H, broad d, distance between the major peaks, 9 Hz, anomeric H). On a spectrum taken at 100 MHz (at 23° and 50°), the anomeric proton was detected as broad doublet (J =about 9 Hz) centering at 4.84 ppm.

Methanolysis of XV—A solution of XV (102 mg) in anhyd. MeOH containing 1% HCl was refluxed for 2 hr. The reaction mixture was made alkaline with 10% NaOH, concentrated to half volume, diluted with H₂O, and extracted with CHCl₃. The organic layer gave a violet syrup (46 mg, neutral fraction). The aqueous layer was acidified with 10% HCl and extracted with CHCl₃ to give acidic fraction as an orange syrup (42 mg). The neutral fraction showed on TLC (solv. 2) two spots at *R_f* 0.23 (black) and 0.13 (black, major), and on GLC two peaks at *t_R* 1.7 and 2.4 (major), identical respectively with the *R_f* and *t_R* values of methyl 2,3,4,6-tetra-O-methyl- β - and α -D-glucopyranosides.

The acidic fraction consisted of a compound (*R_f* 0.26 (solv. 4)) accompanied by a trace amount of methyl 2,3,4,6-tetra-O-methyl- α -glucopyranoside (*R_f* 0.36). Column chromatographic purification gave the homogeneous compound of *R_f* 0.26, which was crystallized from acetone-hexane to yield XVI as yellow plates, mp 204–205°, UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(log ϵ): 326 (4.31). *Anal.* Calcd. for C₁₈H₁₈O₅: C, 68.78; H, 5.77. Found: C, 68.41; H, 5.71. NMR ((CD₃)₂SO) δ ppm: 7.20 and 6.76 (2H, two d, *J*=18 Hz, olefinic H), 7.60 and 6.96 (4H, two d, *J*=8 Hz, C₂-, C₆-, C₃- and C₅-H), 6.13 (2H, s, C_{3'}- and C_{5'}-H), 3.67 and 3.81 (9H, two s, OMe \times 3). On admixture with an authentic specimen, mp 202–203°, prepared by methylation of naringin with Me₂SO₄ and NaOH followed by acid hydrolysis of the product,¹²⁾ it did not depress the melting point. IR, UV and NMR spectra were identical with those of the authentic specimen.

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