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Studies on the Constituents of Asclepiadaceae Plants. XXXIX.<sup>1)</sup> Component of *Marsdenia tomentosa* Decne: Structure of Deacetyldehydrotomentodin, 20-O-Acetylpenupogenin, Deacetylkidjoladinin, and Kidjoladinin

HIDEO SETO, TAMIHIKO SASAKI, KOJI HAYASHI, and HIROSHI MITSUHASHI

Faculty of Pharmaceutical Sciences, Hokkaido University<sup>2)</sup>

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Four new polyoxypregnane derivatives, deacetyldehydrotomentodin ( $12\beta$ -O-cinnamoylutendin) (III), 20-O-acetylpenupogenin ( $12\beta$ -O-cinnamoyl-20-O-acetylsarcostin) (V), deacetylkidjoladinin ( $12\beta$ -O-tigloylsarcostin) (VIII), and kidjoladinin ( $12\beta$ -O-tigloyl-20-O-acetylsarcostin) (IX), were isolated from the stem of Marsdenia tomentosa Decne. Deacetyldehydrotomentodin is the first example of a monoester possessing the utendin skeleton to be isolated from the Asclepiadaceae plants.

Our previous papers reported the isolation and characterization of tomentosin,<sup>3)</sup> tomentin,<sup>4)</sup> dehydrotomentin,<sup>5)</sup> tomentodin,<sup>5)</sup> dehydrotomentosin,<sup>5)</sup> deacetyltomentosin,<sup>6)</sup> and tomentidin,<sup>6)</sup> the new polyoxypregnane derivatives possessing tomentogenin or utendin skeleton from the stem of *Marsdenia tomentosa* Decne. In this paper, we report some later findings on the components of this plant.

The aglycone mixture, obtained by a mild acid hydrolysis of the crude glycoside, was chromatographed over silica gel and separated into several main fractions. Preparative thin-layer chromatography (TLC) of the most polar fraction yielded six crystalline substances, tentatively named compounds I, J, K, L, M, and N.

Among these, compounds I (I) and J (II) were identical with kidjolanin<sup>7a)</sup> and penupogenin,<sup>7a,8)</sup> respectively, from the comparison of spectral data and mixed mp with authentic samples. Compound K (III) mp 210—215°,  $[\alpha]_{50}^{20} + 7^{\circ}$  (c=0.8, CHCl<sub>3</sub>), with a molecular formula of  $C_{30}H_{40}O_{6}$  from its elemental analysis and mass spectrum of m/e 496 (M<sup>+</sup>) ,whose

Fig. 1

infrared (IR) spectrum showed absorptions for hydroxyl groups at 3450 and 1070 cm<sup>-1</sup>, and an  $\alpha,\beta$ -unsaturated ester at 1710, 1690, 1635, and 1150 cm<sup>-1</sup>. The nuclear magnetic resonance (NMR) spectrum of III showed signals for two tertiary methyl groups at  $\delta$  1.00 (s) and 1.32 (s), one secondary methyl group at 1.12 (d, J=6 Hz), three hydroxy-methines at 3.50 (m),

<sup>1)</sup> Part XXXVIII: H. Bando, T. Yamagishi, K. Hayashi, and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 24, 1842 (1976).

<sup>2)</sup> Location: Kita-12-jo, Nishi-5-chome, Kita-ku, Sapporo, 060, Japan.

<sup>3)</sup> H. Seto, K. Hayashi, and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 23, 1552 (1975).

<sup>4)</sup> H. Seto, K. Hayashi, and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 23, 2397 (1975).

<sup>5)</sup> H. Seto, K. Hayashi, and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 24, 443 (1976)

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<sup>7)</sup> a) H. Mitsuhashi, I. Takemori, Y. Shimizu, T. Nomura, and E. Yamada, Chem. Pharm. Bull. (Tokyo), 10, 804 (1962); b) H. Mitsuhashi, T. Sato, T. Nomura, and I. Takemori, ibid., 13, 267 (1965); c) M. Fukuoka, and H. Mitsuhashi, ibid., 16, 1634 (1968); d) T. Sasaki, K. Hayashi, and H. Mitsuhashi, ibid., 20, 628 (1972).

<sup>8)</sup> H. Mitsuhashi and Y. Shimizu, Chem. Pharm. Bull. (Tokyo), 8, 565 (1960).

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3.62 (q, J=6 Hz), and 4.70 (d.d, J=6, 11 Hz), and eight olefinic protons at 5.36 (1H, t, J=3.5 Hz), 6.40 (1H, d, J=16 Hz), 7.40 (5H, m), and 7.70 (1H, d, J=16 Hz). The mass spectrum of III showed the presence of a cinnamoyl group at m/e 348 (M<sup>+</sup>-C<sub>9</sub>H<sub>8</sub>O<sub>2</sub>), 148, 131, and 103 which was supported by ultraviolet (UV) absorptions at 217 (log  $\varepsilon$  4.30), 223 (4.28) and 279 nm (4.15).

Hydrolysis of III with 5% methanolic potassium hydroxide afforded utendin<sup>9)</sup> (IV) as a neutral product. These facts suggest that III is a monoester of utendin with cinnamic acid, and mass spectral peak of III at m/e 451 (M<sup>+</sup>—45)<sup>10)</sup> definitely indicated that cinnamate moiety was not at C-20 of utendin.

The NMR spin decoupling experiments were carried out to confirm the position of the ester linkage of III. Irradiation of 21-Me group protons at  $\delta$  1.12 collapsed the quartet at  $\delta$  3.62 to a singlet but the double-doublet at  $\delta$  4.70, while that of the hydroxy-methine at  $\delta$  3.62 collapsed 21-Me group protons to a singlet, so that the hydroxy-methines at 3.62 and 4.70 (d.d, J=6, 11 Hz) correspond to 20- and  $12\alpha$ -H,<sup>11)</sup> respectively. From these evidences, the structure of compound K (III) was determined as  $12\beta$ -O-cinnamoylutendin and was named deacetyldehydrotomentodin, which is the first example of a monoester possessing an utendin skeleton to be isolated from a plant of the Asclepiadaceae family. Compound L (V), mp

Fig. 2

142—145°, [α]<sub>D</sub><sup>18</sup> +52° (c=0.4,CHCl<sub>3</sub>), with a molecular fornula of C<sub>32</sub>H<sub>42</sub>O<sub>8</sub> from its elemental analysis, and mass spectrum of m/e 494 (M<sup>+</sup>—AcOH), whose IR spectrum showed absorptions for hydroxyl groups at 3400 and 1050 cm<sup>-1</sup>, a saturated ester at 1730 and 1260 cm<sup>-1</sup>, and an α,β-unsaturated ester at 1710, 1680, 1635, and 1170 cm<sup>-1</sup>. The NMR spectrum of V showed signals for two tertiary methyl groups at δ 1.14 (s) and 1.46 (s), one secondary methyl group at 1.20 (d, J=6 Hz), one acetyl group at 1.95 (s), three hydroxy-methines at 3.60 (m), 4.62 (q, J=6 Hz), and 4.68 (d.d, J=6, 11 Hz), and eight olefinic protons at 5.40 (1H, t, J=3.5), 6.34 (1H, d, J=16 Hz), 7.50 (5H, m), and 7.66 (1H, d, J=16 Hz). The mass spectral peaks of V at m/e (M<sup>+</sup>—C<sub>9</sub>H<sub>8</sub>O<sub>2</sub>), 131 (C<sub>9</sub>H<sub>7</sub>O), and 103 (C<sub>8</sub>H<sub>6</sub>), as well as m/e 494 (M<sup>+</sup>—AcOH) and 43 (COCH<sub>3</sub>), showed the presence of a cinnamoyl group and an acetyl group, respectively. The presence of cinnamoyl group was supported by UV absorptions at 217 (log  $\varepsilon$  4.29), 223 (4.22), and 278 nm (4.38).

Hydrolysis of V with 5% methanolic potassium hydroxide afforded sarcostin<sup>12)</sup> (VI) as a neutral product. These facts suggest that V is a diester of sarcostin (VI) with acetic acid and cinnamic acid. Acetylation of V with acetic anhydride-pyridine afforded a monoacetate (VII), which was identical with  $3\beta$ ,20-O,O-diacetylpenupogenin ( $3\beta$ ,20-O,O-diacetyl-12 $\beta$ -O-cinnamoylsarcostin). From these evidences, the structure of compound L (V) was determined as  $12\beta$ -O-cinnamoyl-20-O-acetylsarcostin and was named 20-O-acetylpenupogenin, which

<sup>9)</sup> E. Abisch, Ch. Tamm, and T. Reichstein, Helv. Chim. Acta, 42, 1014, (1959).

<sup>10)</sup> M. Fukuoka, K. Hayashi, and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 19, 1469 (1971).

<sup>11)</sup> N.S. Bhacca and D.H. Williams, "Application of NMR Spectroscopy in Organic Chemistry—Illustrations from Steroid Field," Holden-Day, Inc., San Francisco, 1964.

<sup>12)</sup> J.W. Cornforth and J.C. Earl, J. Chem. Soc., 1939, 737.

corresponds to the acylgenin of amplexoside A isolated from Asclepias amplexicaulis by Piatak.<sup>13)</sup> Compound M (VIII), mp 205—209°, [ $\alpha$ ]<sub>D</sub><sup>19</sup> +9.5° (c=0.85, CHCl<sub>3</sub>), with molecular formula of C<sub>26</sub>H<sub>40</sub>O<sub>7</sub> from its elemental analysis and mass spectrum of m/e 464 (M<sup>+</sup>), whose IR spectrum of VIII showed absorptions for hydroxyl groups at 3400 and 1035 cm<sup>-1</sup>, and an  $\alpha$ , $\beta$ -unsaturated ester at 1710, 1690, 1640, and 1140 cm<sup>-1</sup>, which was supported by UV absorption at 215 nm (log  $\epsilon$  3.9). The NMR spectrum of VIII showed signals for two tertiary methyl groups at  $\delta$  1.12 (s) and 1.46 (s), one secondary methyl group at 1.20 (d, J=6 Hz), two vinyl methyl groups at 1.82 (d, J=6 Hz), and 1.83 (s), three hydroxy methines at 3.58 (q, J=6 Hz), 3.60 (m), 4.64 (d.d, J=6, 11 Hz), and two olefinic protons at 5.40 (t, J=3.5 Hz), and 6.86 (d, J=6 Hz).

Hydrolysis of VIII with methanolic potassium hydroxide afforded sarcostin (VI) as a neutral product. Prominent mass spectral peaks of VIII indicative of tiglate functional group were observed at m/e 83 (C<sub>5</sub>H<sub>7</sub>O) and 55 (C<sub>4</sub>H<sub>7</sub>). Further evidences were secured from the mass spectral peaks of VIII, since there were a faint parent ion at m/e 464 and other fragments at 446 (M<sup>+</sup>—H<sub>2</sub>O), 419 (M<sup>+</sup>—CHOH·Me),<sup>10)</sup> 374 (M<sup>+</sup>—5H<sub>2</sub>O), 364 (M<sup>+</sup>—tiglic acid), 346 (M<sup>+</sup>—tiglic acid—H<sub>2</sub>O), and 83 (tigloyl cation). These evidences indicated that VIII is a monoester of sarcostin (VI) with tiglic acid. The mass spectral peak at m/e 419, and the chemical shift and coupling constant<sup>11)</sup> of the NMR spectrum at  $\delta$  4.64 (d.d, J=6, 11 Hz) definitely indicated that tiglate moiety was not at C-20 but at C-12 $\beta$  of sarcostin.

In order to confirm the position of ester linkage of VIII, the NMR spin decoupling experiments were carried out. Irradiation of 21-Me group protons ( $\delta$  1.20, 3H, d, J=6 Hz) collapsed the quartet at  $\delta$  3.58 to a singlet but not the double-doublet at 4.64 and that of the hydroxymethine at  $\delta$  3.58 collapsed 21-Me to a singlet, so that the hydroxymethines at 3.58 and 4.64 correspond to 20- and  $12\alpha$ -H,<sup>11)</sup> respectively. From these results, the structure of compound M (VIII) was determined as  $12\beta$ -O-tigloyl sarcostin and was named deacetylkidjoladinin.

Compound N (IX), mp 250—255°,  $[\alpha]_D^{15}+43.4^\circ$  (c=1.0, CHCl<sub>3</sub>), with a molecular formula of  $C_{28}H_{42}O_8$  from its elemental analysis and mass spectrum of m/e 506 (M+), whose IR spectrum showed absorptions for hydroxyl groups at 3500, 3430, and 1070 cm<sup>-1</sup>, a saturated ester at 1710 and 1280 cm<sup>-1</sup>, and an  $\alpha,\beta$ -unsaturated ester at 1700, 1645 and 1145 cm<sup>-1</sup>. The NMR spectrum of IX showed signals for two tertiary methyl groups at  $\delta$  1.15 (s) and 1.44 (s), one secondary methyl group at 1.23 (d, J=6 Hz), two vinyl-methyl groups at 1.85 (s) and 1.86 (d, J=6 Hz), one acetyl group at 1.95 (s), three hydroxy-methines at 3.52 (m), 4.63 (d.d, J=6 Hz)

<sup>13)</sup> A.M. Ahsan, D.M. Piatak, and P.D. Sorensen, Experientia, 29, 788 (1973).

6, 11 Hz), 4.65 (q, J=6 Hz), and two olefinic protons at 5.35 (t, J=3.5 Hz) and 6.85 (q, J=6 Hz).

Hydrolysis of IX with methanolic potassium hydroxide afforded sarcostin (VI) as a neutral product. The mass spectrum of IX showed the presence of a tigloyl group at m/e 406 (M<sup>+</sup>—C<sub>5</sub>H<sub>8</sub>O<sub>2</sub>), 83 (C<sub>5</sub>H<sub>7</sub>O), and 55 (C<sub>4</sub>H<sub>7</sub>), and an acetyl group at m/e 446 (M<sup>+</sup>—AcOH) and 43 (COCH<sub>3</sub>). The presence of tigloyl group was supported by UV absorption at 214 nm (log  $\epsilon$  4.14). These facts indicate that IX is a diester of sarcostin (VI) with acetic acid and tiglic acid. Acetylation of IX with acetic anhydride–pyridine afforded a monoacetate (X), which was identical with deacetylkidjolanin diacetate. From these results, the structure of compound N (IX) was determined as  $12\beta$ -O-tigloyl-20-O-acetylsarcostin and was named kidjoladinin.

Fig. 4

## Experimental

Melting points were determined on a Kofler hot stage and are uncorrected. Optical rotations were measured in CHCl<sub>3</sub> solution on a Hitachi S115-4 polarimeter. NMR spectra were determined on a JEOL PS-100 spectrometer operating at 100 MHz with tetramethylsilane (TMS) as an internal standard. Mass spectra were determined on a Hitachi RMU-7 mass spectrometer, IR spectra in Nujol mull on a Hitachi 215 spectrometer, and UV spectra in EtOH solution on a Hitachi EPS-3T spectrometer. TLC was performed on silica gel HF<sub>254</sub> (Merck, Type 60), and silica gel 0.05-0.2 mm (Merck, 70-325 mesh ASTM) was used for column chromatography.

Deacetyldehydrotomentodin (III) — From 15 g of the ester-type aglycone mixture obtained by the same procedure as reported previously,<sup>3)</sup> 74 mg of deacetyldehydrotomentodin (III) was obtained by silica gel column chromatography and preparative TLC (CHCl<sub>3</sub>: MeOH=19: 1, cyclohexane: EtOAc=1: 1). III was recrystallized from acetone–hexane to prisms, mp 210—215°,  $[\alpha]_D^{so} + 7^\circ$  (c=0.8, CHCl<sub>3</sub>). Mass Spectrum m/e: 496 (M<sup>+</sup>), 478 (M<sup>+</sup>—H<sub>2</sub>O), 460 (M<sup>+</sup>—2H<sub>2</sub>O), 451 (M<sup>+</sup>—CHOH·Me), 442 (M<sup>+</sup>—3H<sub>2</sub>O), 433 (M<sup>+</sup>—CHOH·Me—H<sub>2</sub>O), 415 (M<sup>+</sup>—CHOH·Me—2H<sub>2</sub>O), 397 (M<sup>+</sup>—CHOH·Me—3H<sub>2</sub>O), 348 (M<sup>+</sup>—cinnamic acid), 330 (M<sup>+</sup>—cinnamic acid—H<sub>2</sub>O), 312 (M<sup>+</sup>—cinnamic acid—2H<sub>2</sub>O), 303 (M<sup>+</sup>—cinnamic acid—CHOH·Me), 294 (M<sup>+</sup>—cinnamic acid—3H<sub>2</sub>O), 285 (M<sup>+</sup>—cinnamic acid—CHOH·Me—H<sub>2</sub>O), 267 (M<sup>+</sup>—cinnamic acid—CHOH·Me—2H<sub>2</sub>O), 249 (M<sup>+</sup>—cinnamic acid—CHOH·Me—3H<sub>2</sub>O), 148, 147, 131 (base peak), 105, 103. IR  $r_{\text{max}}^{\text{Nujol}}$  cm<sup>-1</sup>: 3450, 1710, 1690, 1635, 1575, 1280, 1150, 1070, 1050. UV  $\lambda_{\text{max}}^{\text{EtOH}}$  nm (log  $\varepsilon$ ): 217 (4.30), 223 (4.28), 279 (4.15). NMR  $\delta_{\text{ppm}}^{\text{cDCl}_1}$ : 1.00 (3H, s, 19-Me), 1.12 (3H, d, J=6 Hz, 21-Me), 1.32 (3H, s, 18-Me), 3.50 (1H, m, 3α-H), 3.62 (1H, q, J=6 Hz, 20-H), 4.70 (1H, d.d, J=6, 11 Hz, 12α-H), 5.36 (1H, t, J=3.5 Hz,  $\Delta^{5}$ -olefinic proton), 6.40 (1H, d, J=16 Hz), 7.40 (5H, m, aromatic protons), 7.70 (1H, d, J=16 Hz). Anal. Calcd. for C<sub>30</sub>H<sub>40</sub>O<sub>6</sub>: C, 72.55; H, 8.12. Found: C, 72.63; H, 7.95.

Alkaline Hydrolysis of Deacetyldehydrotomentodin (III)——A solution of 20 mg of deacetyldehydrotomentodin (III) in 1 ml of 5% MeOH-KOH was refluxed for 30 min and the reaction mixture was purified directly by preparative TLC (MeOH: CHCl<sub>3</sub>=9:1). Recrystallization from acetone gave 10 mg of utendin

(IV) as prisms, mp 240—243°. Mass Spectrum m/e: 366 (M+), 348 (M+-H<sub>2</sub>O), 330 (M+-2H<sub>2</sub>O), 321 (M+-CHOH·Me), 312 (M+-3H<sub>2</sub>O), 303 (M+-CHOH·Me-H<sub>2</sub>O), 285 (M+-CHOH·Me-2H<sub>2</sub>O), 267 (M+-CHOH·Me-3H<sub>2</sub>O), 260, 234, 159, 145. IR  $v_{\rm max}^{\rm Nuloi}$  cm<sup>-1</sup>: 3400, 1070, 1050, 1035.

20-O-Acetylpenupogenin (V)—From the same column chromatographic fraction which contained III, 55 mg of 20-O-acetylpenupogenin (V) was obtained by preparative TLC (ether, CHCl<sub>3</sub>: MeOH=99: 1). V was recrystallized from acetone-hexane to fine needles, mp 142—145°, [α]<sub>1</sub><sup>18</sup> +52° ( $\alpha$ =0.4, CHCl<sub>3</sub>). Mass Spectrum m/e: 494 (M<sup>+</sup>—AcOH), 476 (M<sup>+</sup>—AcOH—H<sub>2</sub>O), 467 (M<sup>+</sup>—CHOAc·Me), 458 (M<sup>+</sup>—AcOH—2H<sub>2</sub>O), 449 (M<sup>+</sup>—CHOAc·Me—H<sub>2</sub>O), 440 (M<sup>+</sup>—AcOH—3H<sub>2</sub>O), 431 (M<sup>+</sup>—CHOAc·Me—2H<sub>2</sub>O), 406 (M<sup>+</sup>—cinnamic acid, 346 (M<sup>+</sup>—cinnamic acid—AcOH), 328 (M<sup>+</sup>—cinnamic acid—AcOH—H<sub>2</sub>O), 319 (M<sup>+</sup>—cinnamic acid—CHOAc·Me—2H<sub>2</sub>O), 265 (M<sup>+</sup>—cinnamic acid—CHOAc·Me—3H<sub>2</sub>O), 148, 147, 131 (base peak), 103, 43. IR  $\nu_{\text{max}}^{\text{Nujol}}$  cm<sup>-1</sup>: 3400, 1730, 1710, 1680, 1635, 1260, 1170, 1050. UV  $\lambda_{\text{max}}^{\text{EtoH}}$  nm (log  $\varepsilon$ ): 217 (4.29), 223 (4.22), 278 (4.38). NMR  $\delta_{\text{ppm}}^{\text{CDCl}_3}$ : 1.14 (3H, s, 19-Me), 1.20 (3H, d, J=6 Hz, 21-Me), 1.46 (3H, s, 18-Me), 1.95 (3H, s, OAc), 3.60 (1H, m, 3α-H), 4.62 (1H, q, J=6 Hz, 20-H), 4.68 (1H, d.d, J=6, 11 Hz, 12α-H), 5.40 (1H, t, J=3.5 Hz,  $\Delta$ <sup>5</sup>-olefinic proton), 6.34 (1H, d, J=16 Hz), 7.50 (5H, m, aromatic protons), 7.66 (1H, d, J=16 Hz), Anal. Calcd. for C<sub>32</sub>H<sub>42</sub>O<sub>8</sub>: C, 69.29; H, 7.63. Found: C, 69.07; H, 7.85.

Alkaline Hydrolysis of 20-O-Acetylpenupogenin (V)——A solution of 15 mg of 20-O-acetylpenupogenin (V) in 1 ml of 5% MeOH-KOH was allowed to stand for 24 hr at room temperature and the reaction mixture was purified directly by preparative TLC (MeOH:  $CHCl_3=1:9$ ). Recrystallization from MeOH-acetone gave 6 mg of sarcostin (VI) as prisms, mp 245—250°. Mass Spectrum  $m/e:382~(M^+),364~(M^+-H_2O),346~(M^+-2H_2O),337~(M^+-CHOH\cdotMe),328~(M^+-3H_2O),319~(M^+-CHOH\cdotMe-H_2O),310~(M^+-4H_2O),301~(M^+-CHOH\cdotMe-2H_2O),283~(M^+-CHOH\cdotMe-3H_2O),265~(M^+-CHOH\cdotMe-4H_2O),244~(M^+-138),^{10,14})$  226 (M<sup>+</sup>-138-H<sub>2</sub>O), 161, 138, 114, 105, 43. IR  $\nu_{mis}^{naio}$  cm<sup>-1</sup>: 3470, 3380, 3250, 175, 1045, 1040.

Acetylation of 20-O-Acetylpenupogenin (V) — A solution of 20 mg of 20-O-acetylpenupogenin (V) in 1 ml of Ac<sub>2</sub>O and 1 ml of pyridine was allowed to stand for 24 hr at room temperature, and poured into ice water. A white powder that appeared was collected and recrystallized from acetone-hexane to afford 18 mg of an acetate (VII) as needles, mp 137—139°. Mass Spectrum m/e: 536 (M+—AcOH), 518 (M+—AcOH—H<sub>2</sub>O), 590 (M+—AcOH—2H<sub>2</sub>O), 476 (M+—2×AcOH), 448 (M+—cinnamic acid), 388 (M+—cinnamic acid—AcOH), 370 (M+—cinnamic acid—AcOH—H<sub>2</sub>O), 352 (M+—cinnamic acid—AcOH—2H<sub>2</sub>O), 328 (M+—cinnamic acid—2×AcOH), 310 (M+—cinnamic acid—2×AcOH—H<sub>2</sub>O), 292 (M+—cinnamic acid—2×AcOH—2H<sub>2</sub>O), 148, 147, 131 (base peak), 120, 103, 43. IR  $v_{\text{max}}^{\text{Nuloi}}$  cm<sup>-1</sup>: 3450, 1725, 1705, 1690, 1635, 1245, 1160, 1070, 1030. NMR  $\delta_{\text{ppm}}^{\text{cncl}}$ : 1.16 (3H, s, 19-Me), 1.20 (3H, d, J=6 Hz, 21-Me), 1.46 (3H, s, 18-Me), 2.02 (3H, s, OAc), 2.18 (3H, s, OAc), 4.60 (3H, m, 3\alpha-H+12\alpha-H+20-H), 5.36 (1H, t, J=3.5 Hz,  $\Delta^5$ -olefinic proton), 6.24 (1H, d, J=16 Hz), 7.40 (5H, m, aromatic protons), 7.56 (1H, d, J=16 Hz). Anal. Calcd. for  $C_{34}H_{44}O_9$ : C, 68.43; H, 7.43. Found: C, 68.28; H, 7.34.

Acetylation of Penupogenin (II)——A solution of 55 mg of penupogenin (II) in 1 ml of Ac<sub>2</sub>O and 1 ml of pyridine was allowed to stand for 19 hr at room temperature and worked up in the same manner as in the acetylation of V to produce 50 mg of an amorphous product, which was recrystallized from acetone-hexane to needles, mp 136—138.5°, and miexd mp with VII 135—139°. All spectral data were identical with those of VII.

Deacetylkidjoladinin (VIII) — From the same column chromatographic and thin-layer chromatographic fraction which contained V, 37 mg of deacetylkidjoladinin (VIII) was obtained by preparative TLC (ether, CHCl<sub>3</sub>: MeOH=99: 1). VIII was recrystallized from acetone-hexane to prisms, mp 205—209°, [α]<sub>D</sub><sup>19</sup> +9.5° (c=0.85, CHCl<sub>3</sub>). Mass Spectrum m/e: 464 (M<sup>+</sup>), 446 (M<sup>+</sup>—H<sub>2</sub>O), 428 (M<sup>+</sup>—2H<sub>2</sub>O), 419 (M<sup>+</sup>—CHOH·Me), 413 (M<sup>+</sup>—2H<sub>2</sub>O—Me), 410 (M<sup>+</sup>—3H<sub>2</sub>O), 401 (M<sup>+</sup>—CHOH·Me—H<sub>2</sub>O), 392 (M<sup>+</sup>—4H<sub>2</sub>O), 383 (M<sup>+</sup>—CHOH·Me—2H<sub>2</sub>O), 374 (M<sup>+</sup>—5H<sub>2</sub>O), 365 (M<sup>+</sup>—CHOH·Me—3H<sub>2</sub>O), 364 (M<sup>+</sup>—tiglic acid), 346 (M<sup>+</sup>—tiglic acid—H<sub>2</sub>O), 328 (M<sup>+</sup>—tiglic acid—2H<sub>2</sub>O), 310 (M<sup>+</sup>—tiglic acid—3H<sub>2</sub>O), 301 (M<sup>+</sup>—tiglic acid—CHOH·Me—H<sub>2</sub>O), 277, 240, 208, 83 (base peak), 55. IR  $v_{\text{max}}^{\text{Nulol}}$  cm<sup>-1</sup>: 3400, 1710, 1690, 1640, 1280, 1140, 1070, 1035. UV  $\lambda_{\text{max}}^{\text{EtoH}}$  215 nm (log  $\varepsilon$  3.93). NMR  $\delta_{\text{ppm}}^{\text{CDCH}}$ : 1.12 (3H, s, 19-Me), 1.20 (3H, d, J=6 Hz, 21-Me), 1.46 (3H, s, 18-Me), 1.82 (3H, d, J=6 Hz, vinyl-Me), 1.83 (3H, s, vinyl-Me), 3.58 (1H, q, J=6 Hz, 20-H), 3.60 (1H, m, 3α-H), 4.64 (1H, d.d, J=6, 11 Hz, 12α-H), 5.40 (1H, d, J=3.5 Hz,  $\Delta$ 5-olefinic proton), 6.86 (1H, d, J=6 Hz). Anal. Calcd. for C<sub>26</sub>H<sub>40</sub>O<sub>7</sub>: C, 67.21; H, 8.68. Found: C, 67.09; H, 8.75.

Alkaline Hydrolysis of Deacetylkidjoladinin (VIII)——A solution of 7 mg of deacetylkidjoladinin (VIII) in 0.5 ml of 5% MeOH–KOH was allowed to stand for 18 hr at room temperature and worked up in the same manner as in the hydrolysis of V to afford 3 mg of sarcostin (VI) as prisms.

Acetylation of Deacetylkidjoladinin (VIII) ——A solution of 20 mg of deacetylkidjoladinin (VIII) in 1 ml of Ac<sub>2</sub>O and 1 ml of pyridine was allowed to stand for 23 hr at room temperature and worked up in the same manner as in the acetylation of V to afford 20 mg of a diacetate (X), which was recrystallized from acetone—hexane to plates, mp 231—236°. Mass Spectrum  $m/\varepsilon$ ; 488 (M<sup>+</sup>—AcOH), 470 (M<sup>+</sup>—AcOH—H<sub>2</sub>O), 452 (M<sup>+</sup>—AcOH—2H<sub>2</sub>O), 448 (M<sup>+</sup>—tiglic acid), 434 (M<sup>+</sup>—AcOH—3H<sub>2</sub>O), 430 (M<sup>+</sup>—tiglic acid—H<sub>2</sub>O), 428 (M<sup>+</sup>—2×

<sup>14)</sup> M. Fukuoka and H. Mitsuhashi, Chem. Pharm. Bull. (Tokyo), 17, 2448 (1969).

AcOH), 410 (M<sup>+</sup>-2×AcOH-H<sub>2</sub>O), 392 (M<sup>+</sup>-2×AcOH-2H<sub>2</sub>O), 388 (M<sup>+</sup>-tiglic acid-AcOH), 370(M<sup>+</sup>-tiglic acid-AcOH-H<sub>2</sub>O), 352 (M<sup>+</sup>-tiglic acid-AcOH-2H<sub>2</sub>O), 328 (M<sup>+</sup>-tiglic acid-2×AcOH), 310 (M<sup>+</sup>-tiglic acid-2×AcOH-H<sub>2</sub>O), 292 (M<sup>+</sup>-tiglic acid-2×AcOH-2H<sub>2</sub>O), 274 (M<sup>+</sup>-tiglic acid-2×AcOH-3H<sub>2</sub>O), 120, 105, 83 (base peak), 55, 43. IR  $\nu_{\rm max}^{\rm Nulol}$  cm<sup>-1</sup>: 3450, 1730, 1705, 1690, 1640, 1260, 1240, 1150, 1070, 1030. NMR  $\delta_{\rm ppm}^{\rm ODCl_3}$ : 1.14 (3H, s, 19-Me), 1.20 (3H, d, J=6 Hz, 21-Me), 1.42 (3H, s, 18-Me), 1.82 (3H, d, J=6 Hz, vinyl-Me), 1.83 (3H, s, vinyl-Me), 1.94 (3H, s, OAc), 2.01 (3H, s, OAc), 4.60 (1H, m, 3\alpha-H), 4.64 (1H, d.d, J=6, 11 Hz, 12\alpha-H), 4.66 (1H, q, J=6 Hz, 20-H), 5.38 (1H, t, J=3.5 Hz,  $\Delta$ <sup>5</sup>-olefinic proton), 6.82 (1H, d, J=6 Hz).

Kidjoladinin (IX)—From the same column chromatographic and thin-layer chromatographic fractions which contained V, 52 mg of kidjoladinin (IX) was obtained by preparative TLC (ether, CHCl<sub>3</sub>: MeOH=99: 1). Recrystallization of IX from acetone-hexane gave needles, mp 250—252°, [α]<sub>0</sub><sup>15</sup> +43.4° (c=1.0, CHCl<sub>3</sub>). Mass Spectrum m/e: 506 (M+), 488 (M+-H<sub>2</sub>O). 470 (M+-2H<sub>2</sub>O), 452 (M+-3H<sub>2</sub>O), 446 (M+-AcOH), 428 (M+-AcOH-H<sub>2</sub>O), 413 (M+-AcOH-H<sub>2</sub>O-Me), 410 (M+-AcOH-2H<sub>2</sub>O), 406 (M+-tiglic acid), 395 (M+-AcOH-2H<sub>2</sub>O-Me), 388 (M+-tiglic acid-H<sub>2</sub>O), 370 (M+-tiglic acid-2H<sub>2</sub>O), 368 (M+-138), 352 (M+-tiglic acid-3H<sub>2</sub>O), 350 (M+-138-H<sub>2</sub>O), 346 (M+-tiglic acid-AcOH), 332 (M+-138-2H<sub>2</sub>O), 328 (M+-tiglic acid-AcOH-H<sub>2</sub>O), 314 (M+-138-3H<sub>2</sub>O), 310 (M+-tiglic acid-AcOH-2H<sub>2</sub>O), 190 (M+-tiglic acid-138-2H<sub>2</sub>O), 120, 105, 83 (base peak), 55, 43. IR  $v_{\text{max}}^{\text{Nujoi}}$ cm<sup>-1</sup>: 3500, 3430, 1710, 1700, 1650, 1645, 1280, 1145, 1070, 1050. UV  $\lambda_{\text{max}}^{\text{EtoH}}$ : 214 nm (log  $\varepsilon$  4.14). NMR  $\delta_{\text{ppm}}^{\text{puoi}}$ : 1.15 (3H, s, 19-Me), 1.23 (3H, d, J=6 Hz, 21-Me), 1.44 (3H, s, 18-Me), 1.85 (3H, s, vinyl-Me), 1.86 (3H, d, J=6 Hz, vinyl-Me), 1.95 (3H, s, OAc), 3.52 (1H, m, 3α-H), 4.63 (1H, d.d, J=6, 11 Hz, 12α-H), 4.65 (1H, q, J=6 Hz, 20-H), 5.35 (1H, t, J=3.5 Hz, J<sup>5</sup>-olefinic proton), 6.85 (1H, d, J=6 Hz). Anal. Calcd. for C<sub>28</sub>H<sub>42</sub>O<sub>8</sub>: C, 66.38; H, 8.36. Found: C, 66.64; H, 8.44.

Alkaline Hydrolysis of Kidjoladinin (IX)——A solution of 10 mg of kidjoladinin (IX) in 0.5 ml of 5%. MeOH-KOH was allowed to stand for 24 hr at room temperature and worked up in the same manner as in the hydrolysis of V to afford 4 mg of sarcostin (VI) as prisms.

Acetylation of Kidjoladinin (IX)——A solution of 25 mg of kidjoladinin (IX) in 1 ml of Ac<sub>2</sub>O and 1 ml of pyridine was allowed to stand for 24 hr at room temperature and worked up in the same manner as in the acetylation of V to afford 23 mg of an amorphous product, which was recrystallized from acetone-hexane to plates, mp 230—233° and mixed mp with X, 230—235°. All spectral data were identical with those of X.

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