

Chemical Studies on Sweet Diterpene-Glycosides of *Stevia rebaudiana*: Conversion of Stevioside into Rebaudioside-A

By means of enzymatic and chemical procedures, stevioside (1), the major sweet glycoside of *Stevia rebaudiana* was converted efficiently into another constituent of this plant, rebaudioside-A (2) which tastes sweeter and more pleasant than 1.

Keywords—natural sweetener; *ent*-kaurene type diterpene glucoside; stevioside; rebaudioside-A; enzymatic hydrolysis; Takadiastase Y; glucosylation by *ortho*-ester; ¹³C NMR spectra; *Stevia rebaudiana*; Compositae

From leaves of *Stevia rebaudiana* BERTONI (Compositae), a wild herb of Paraguay, stevioside (1) has been isolated as the major sweet glycoside in a yield of 5–10%.¹⁾ In the search for new natural sweeteners, the present authors have investigated constituents of this plant, isolating other new sweet glycosides named rebaudiosides-A (2),^{2a)} -C, -D, and -E.^{2b)} Since 2 tastes sweeter (about 1.2–1.5 times as much as 1) and more pleasant than 1, much industrial interest has been shown recently in production of 2 rather than 1. In this regard, the present authors have investigated efficient conversion of 1 into 2.

Selective hydrolysis of the terminal glucosyl linkage of the β -sophorosyl moiety of 1 is the key step of the present study. This was achieved by the hydrolysis with Takadiastase Y, the crude preparation of amylase prepared from *Aspergillus oryzae*.³⁾ Incubation of 1 with this enzyme mixture at 37° in McIlvain buffer (pH 4.0) for 80 hr yielded a sweet desgluco-compound (3), colorless prisms, mp 178–182° (from MeOH), $[\alpha]_D^{20} -29.3^\circ$ ($c=0.3$, MeOH) in a quantitative yield. The structure of 3 was substantiated by ¹H and ¹³C nuclear magnetic resonance (NMR)^{2,4,5)} (see Table I). Saponification of 3 by refluxing with 5% NaOH in MeOH for 3 hr afforded steviolmonoside (4)⁴⁾ almost quantitatively, which was converted into 4',6'-benzylidene derivative (5) by the action of benzaldehyde in 98% HCOOH at room temperature for 15 min (yield 87%); colorless needles, mp 167–170° (from MeOH), $[\alpha]_D^{20} -54.4^\circ$ ($c=0.25$, MeOH). The ¹H and ¹³C NMR spectra of 5 indicated that no migration of the double bond took place during the process of this benzylidene formation in the acidic medium.

Previously, the ester glucoside was prepared from Ag-salt of *ent*-kaur-16-en-19-oic acid by the condensation of acetobromoglucose followed by mild deacetylation.⁴⁾ Whereas, treatment of Ag-salt of steviol (6, the common aglycone of 1 and 2) in the same way afforded only trace of the desired ester glucoside (7), unexpectedly. Preparation of 7 in a high yield was furnished by glucosylation with the orthoester; refluxing of a solution of 6 and 3,4,6-tri-O-acetyl- α -D-glucopyranose 1,2-(*tert*-butyl orthoacetate)⁶⁾ (8) in chlorobenzene followed by deacetylation with BaO in MeOH at 5° gave 7, colorless needles mp 185–187° (from MeOH-H₂O), $[\alpha]_D^{20} -31.0^\circ$ ($c=0.1$, MeOH) in a yield of 70%. The structure of 7 was confirmed by IR, ¹H and ¹³C NMR (see Table I) and mass spectrometry of its trimethylsilyl ether. The

- 1) E. Mosettig, U. Beglinger, F. Dolder, H. Lichiti, P. Quitt, and J.A. Waters, *J. Am. Chem. Soc.*, **85**, 2305 (1963) and the references cited therein.
- 2) a) H. Khoda, R. Kasai, K. Yamasaki, K. Murakami, and O. Tanaka, *Phytochemistry*, **15**, 981 (1976). Rebaudioside-B reported in this paper seems now to be an artifact formed from 2 by accidental partial hydrolysis during the process of the isolation; b) I. Sakamoto, K. Yamasaki, and O. Tanaka, *Chem. Pharm. Bull.* (Tokyo), **25**, 844 (1977).
- 3) The crude enzyme was kindly supplied by Dr. A. Endo, Institute of Sankyo Co. Ltd., to whom authors' thanks are due.
- 4) K. Yamasaki, H. Kohda, T. Kobayashi, N. Kaneda, R. Kasai, and O. Tanaka, *Chem. Pharm. Bull.* (Tokyo), accepted.
- 5) K. Yamasaki, H. Kohda, T. Kobayashi, R. Kasai, and O. Tanaka, *Tetrahedron Lett.*, **1976**, 1005.
- 6) N.K. Kochetkov, A.F. Bochkov, T.A. Sokolovskaya, and V. Snyatkova, *Carbohydrate Res.*, **16**, 17 (1971); K. Honma and A. Hamada, *Chem. Pharm. Bull.* (Tokyo), **24**, 1165 (1976).

TABLE I. ^{13}C Chemical Shifts^{a)}

	3	4	7
C-1	40.7	41.0	40.7
2	19.3	19.7	19.4
3	38.2	38.4	38.2
4	44.0	43.8	44.0
5	57.3	56.9	57.4
6	22.0	22.5	22.1
7	41.6	41.6	41.7
8	42.4	42.1	41.7
9	53.9	54.1	54.2
10	39.8	39.7	39.7
11	20.7	20.6	20.7
12	37.1	38.4	40.7
13	85.8	86.4	79.7
14	44.4	44.6	47.2
15	47.8	48.2	48.0
16	154.3	153.7	157.5
17	104.5	104.9	102.8
18	28.2	29.2	28.4
19	176.9	180.0	176.8
20	15.5	15.7	15.6
G'-1 ^{b)}	95.7		95.7
2	73.8		73.9
3	78.9		79.1
4	70.9		71.0
5	78.9		79.1
6	61.9		62.0
G''-1 ^{c)}	99.5	99.4	
2	75.2	75.3	
3	78.5 ^{d)}	78.5 ^{e)}	
4	72.1	71.5	
5	77.8 ^{d)}	77.9 ^{e)}	
6	62.9	62.5	

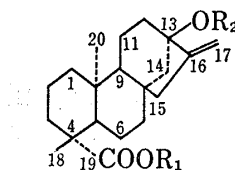
a) δ ppm from internal TMS in $\text{C}_6\text{D}_6\text{N}$. Taken at 25° with JEOL JNM-PFT 100 NMR spectrometer at 25.15 MHz, computer limited resolution: ± 0.1 ppm.

b) Corresponding to ester glucose (R_1).

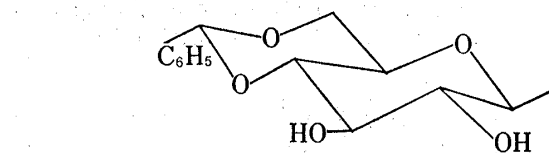
c) Corresponding to 13-O-glucose (R_2).

d, e) Values may be interchanged.

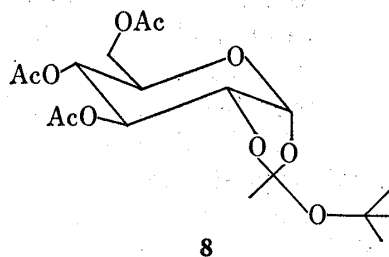
-Glc:-D-glucopyranosyl



	R_1	R_2
1	-Glc	-Glc $\xrightarrow{2-1}$ Glc
2	-Glc	-Glc $\begin{cases} \xrightarrow{2-1} \text{Glc} \\ \xrightarrow{3-1} \text{Glc} \end{cases}$
3	-Glc	-Glc
4	H	-Glc
5	H	



6	H	H
7	-Glc	H



8

benzylidene derivative (5) was subjected to glucosylation in the same way; a solution of 5 and excess of 8 in chlorobenzene was refluxed for 2 hr. The product was desbenzalated with 30% AcOH at 80° for 15 min and then deacetylated with 0.5 N BaO in MeOH at 0° for 2 hr, affording a tetraglucoside, colorless needles, mp 242–244° (from MeOH), $[\alpha]_D^{20} -19.5^\circ$ ($c=0.2$, MeOH) in a yield of 75%, which was proved to be identical with an authentic sample of natural 2 by comparison of thin-layer chromatogram, ^{13}C NMR,^{2,5)} and other physical constants.

Acknowledgement The authors are grateful to Dr. Y. Miyazaki and Mr. K. Nishi, the Experimental Station of Medicinal Plants, National Institute of Hygienic Sciences for their kind supply of leaves of cultivated *Stevia rebaudiana* and to Prof. A. Hamada and Ms. K. Honma, Showa University for their valuable information. This work was supported in part by Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture which is acknowledged.

Institute of Pharmaceutical Sciences
Hiroshima University School of Medicine
1-2-3 Kasumi, Hiroshima-shi, 734, Japan

NORITO KANEDA
RYOJI KASAI
KAZUO YAMASAKI
OSAMU TANAKA⁷⁾

Received June 27, 1977

7) Correspondence should be addressed to O. Tanaka.