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The Constituents of Schizandra chinensis Baill. The Structures of Three New Lignans, Angeloylgomisin H, Tigloylgomisin H and Benzoylgomisin H, and the Absolute Structure of Schizandrin

Three new lignans named angeloylgomisin H(I), tigloylgomisin H(II) and benzoylgomisin H(III), which possess dibenzocyclooctadiene skeleton, have been isolated from the fruits of Schizandra chinensis BAILL (Schizandraceae). Their structures including the absolute configurations were established by chemical and spectral evidence.

The absolute structure of schizandrin was also established by spectral studies (nuclear Overhauser effects and circular dichroism spectrum).

Keywords—Schizandra chinensis; Schizandraceae; lignan; angeloylgomisin H; tigloylgomisin H; benzoylgomisin H; schizandrin; NOE; CD spectrum

In the preceding papers of this series, it was reported that the petroleum ether extract of the fruits of Schizandra chinensis Baill. (Schizandraceae) contains six new lignans possessing dibenzocyclooctadiene skeleton.¹⁾ This paper concerns with isolation and characterization of three additional lignans named angeloylgomisin H(I, yield, 0.031%), tigloylgomisin H(II, 0.014%) and benzoylgomisin H(III, 0.003%), and the absolute structure of schizandrin(V).2) The physical and spectral data of three new compounds are shown below and in Table I.

TABLE I. PMR Spectral Data of I—V, VIII and IX (in CDCl₃, & Value)

	C ₄ -H,s l C ₁₁ -H,s	$C_{6\alpha}$ -H $(J=Hz)$	$C_{6\beta}$ -H $(J=Hz)$	$C_{9\alpha}$ -H $(J=Hz)$	$C_{9\beta}$ -H $(J=Hz)$	$ \begin{array}{c} C\underline{\mathbf{H}}_{3}-\dot{\mathbf{C}}-\underline{\underline{\mathbf{H}}}\\ (\mathbf{d}, \mathbf{J}=\mathbf{H}\mathbf{z}) & \mathbf{m} \end{array} $	$ \begin{array}{ccc} C\underline{H}_3 - \dot{C} - O\underline{\underline{H}} \\ s & s \end{array} $	OCH ₃
Ia)	6.67 6.56	2.75, d (13.5)	2.30, d (13.5)	2.37, d, d (13.5/7)	2.75, d,d (13.5/2)	0.83(7) 1.83	1.23 1.83	3.53, 3.83(×2) 3.88, 3.90
][a)	6.68 6.57	2.75, d (13.5)	2.30, d (13.5)	2.35, d, d (13.5/7)	2.75, d, d (13.5/2)	0.84(7) 1.93	1.23 1.93	3.50, 3.83(×2) 3.89, 3.90
(a)	6.77 6.53	2.80, d (13.5)	2.33, d (13.5)	2.40, d, d (13.5/7)	2.78, d,d (13.5/2)	0.88(7) 1.87	1.27 1.87	3.53, 3.67, 3.80 3.87, 3.95
IV	6.63 6.35	2.70, d (13.5)	2.33, d (13.5)	2.31, d,d (13.5/7)	2.68, d,d (13.5/2)	0.83(7) 1.83	1.25 1.83	$3.63, 3.90(\times 2)$ $3.92(\times 2)$
T	6.62 6.56	2.70, d (14)	2.32, d (14)	2.33, d, d (14/7)	2.68, d, d $(14/2)$	0.82(7) 1.80	1.25 1.86	$3.59(\times 2) \\ 3.90(\times 4)$
VIII	6.62, s ^b , 6.72, s 6.75, s)	2.1—2.	9(4H, m)		0.87(7) 1.80	1.25 1.82	3.49, 3.86 3.91(×3)
IX	6.59, s (C ₄ -H)	2.78, d (15)	2.45, d (15)	2.97, d,d (15/9)	(15/2)	0.87(7) 1.87	1.27 1.68	3.78, 3.87 3.92, 4.05(×2)

α) Other signals: I, $\frac{\text{Me}}{\text{H}}$ $\overset{\alpha}{\text{CO}}$ $\overset{\alpha}{\text{CO}}$ 1.73 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta}{\text{CO}}$ 1.70 (6H, m, β and γ- Me), 5.88 (1H, m, γ -H); II, $\frac{\text{Me}}{\text{H}}$ $\overset{\beta}{\text{CO}}$ $\overset{\beta$

and γ -Me), 6.78(1H, m, γ -H); III, 7.38(3H, m), 7.98(2H, d,d)(C₆H₆CO-). b) in D_6 -acetone: 6.70, 6.77 and 6.85 (each singlet).

Confirmed by decoupling experiments.

d) d=doublet, m=multiplet, s=singlet.

¹⁾ a) H. Taguchi and Y. Ikeya, Chem. Pharm. Bull. (Tokyo), 23, 3296 (1975); b) H. Taguchi and Y. Ikeya, ibid., 25, 364 (1977); c) Y. Ikeya, H. Taguchi and Y. Iitaka, Tetrahedron Lett., 1976, 1357.

N.K. Kochetkov, A. Khorlin, O.S. Chizov and V.I. Sheichenko, Tetrahedron Lett., 1961, 730; N.K. Kochetkov, A. Khorlin, O.S. Chizov, ibid., 1962, 361.

Angeloylgomisin H(I), $C_{28}H_{36}O_8(M^+, m/e, Calcd.: 500.240; Observed: 500.237)$, amorphous, $[\alpha]_D^{22} + 19.4^{\circ}(c=1.60, CHCl_3)$, Ultraviolet (UV) spectrum: λ_{max}^{EDOH} nm (log ε): 215 (4.72), 248 (sh, 4.18), 286 (sh, 3.14). Infrared (IR) spectrum: ν_{max}^{KDr} cm⁻¹: 3500 (OH), 1735 (ester), 1645 (>C=C<). Mass spectrum (MS), m/e (%): 500 (M+, 51), 482 (M+-H₂O, 5), 83[CH₃CH=C(CH₃)-CO, 100], 55[CH₃CH=C(CH₃), 54].

Tigloylgomisin H(II), $C_{28}H_{36}O_8(M^+, m/e, Calcd.: 500.240;$ Observed: 500.237), amorphous, $[\alpha]_D^{23} + 67.7^{\circ}(c=1.30, CHCl_3),$ UV λ_{\max}^{ECH} nm(log ε): 215 (4.75), 248 (sh, 4.21), 285 (sh, 3.47). IR ν_{\max}^{KE} cm⁻¹: 3500 (OH), 1725 (ester), 1645 (C=C), MS, m/e (%): 500 (M⁺, 57), 482 (M⁺-H₂O, 4), 418 (19), 83 [CH₃CH=C(CH₃)CO, 100], 55[CH₃CH=C(CH₃), 85].

Benzoylgomisin H(III), $C_{30}H_{34}O_8(M^+, m/e, Calcd.: 522.225, Observed: 522.224)$, amorphous, $[\alpha]_D^{24} + 96.8^{\circ}(c=1.25, CHCl_3)$, UV λ_{\max}^{EOH} nm (log ϵ): 217 (4.66), 250 (sh, 4.20), 282 (sh, 3.56). IR ν_{\max}^{KBr} cm⁻¹: 3450 (OH), 1735 (ester), 1595, 705 (aromatic), MS, m/e (%): 522 (M+, 47), 504 (M+-H₂O, 5), 105 (C₆H₅CO, 100), 77 (C₆H₅, 25).

These spectral data suggested that three compounds are lignans possessing dibenzocyclooctadiene skeleton and have five methoxyl groups attached to the aromatic rings, a tertiary methyl group attached to the carbon atom having the hydroxyl group and a secondary methyl group.¹⁾ Their proton magnetic resonance (PMR), infrared (IR) and mass spectra indicate that I, II and III possess angeloyl, tigloyl and benzoyl groups respectively. These compounds (I—III), on hydrolysis with 3% ethanolic potassium hydroxide, afforded the same phenolic

Chart 2

compound (IV, named gomisin H), $C_{23}H_{30}O_7(M^+, m/e, 418)$, colourless plates (ether-*n*-hexane), mp 145—146.5°, $[\alpha]_D^{24}$ +44.5° (c=0.97, CHCl₃), UV $\lambda_{\max}^{\text{EOH}}$ nm (log ε): 219 (4.81), 247—249 (sh, 4.32), 276 (3.72), 285 (sh, 3.65). IR ν_{\max}^{KBS} cm⁻¹: 3530, 3350 (OH), 1630, 1600 (aromatic) and corresponding acids.³⁾ Methylation of IV with dimethyl sulfate and potassium carbonate in acetone gave monomethyl ether (V), colourless needles, mp 129—131°, $[\alpha]_D^{\text{21}}$ +86.9°(c=0.35, CHCl₃), which was identified with schizandrin obtained from the same source by the direct comparison (mixed mp, IR and TLC), indicating that IV corresponds to norschizandrin.

The following experiments were thus carried out to confirm the position of the phenolic hydroxyl group in IV. At first, the intramolecular nuclear Overhauser effects (NOE) in benzyl ether of IV(VI) were measured. As shown in chart 2, the both aromatic protons were effected by irradiation of two methoxyl signals, indicating that two methoxyl groups are attached to C_3 and C_{12} positions.

Next, treatment of IV with 2,4-dinitrofluorobenzene in benzene-dimethylformamide mixture in the presence of sodium hydride as a catalyst afforded 2,4-dinitrophenyl ether (VII) (yield, 71.5%), $C_{29}H_{32}N_2O_{11}(M^+, m/e, 584)$, amorphous, $\lceil \alpha \rceil_D^{26} + 49.9^{\circ}$ (c=1.02, CHCl₃), UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm (log ε): 212 (4.70), 250 (4.31), 284 (4.06), 300 (sh, 3.98). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450 (OH), 1602 (aromatic), 1532, 1343 (NO₂). Catalytic hydrogenation of VII over platinum oxide in tetrahydrofuran-methanol mixture followed by cleavage with sodium in liquid ammonia $(-60-65^{\circ})$ afforded compound VIII (yield, 52.5%), $C_{23}H_{30}O_6(M^+, m/e, 402)$, colourless plates, mp 121—122.5°, $[\alpha]_{D}^{24}$ +86.0° (c=0.43, CHCl₃), UV λ_{max}^{EtOH} nm (log ε): 213 (4.63), 254 (4.14), 282 (3.76), 293 (sh, 3.67). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3480 (OH), 1600 (aromatic).⁴ spectrum of VIII shows three singlet signals assignable to the aromatic protons in the region δ 6.60—7.00, indicating that the hydroxyl group in IV is located at the para-position (C₁ or C_{14}) to the aromatic proton. On the other hand, oxidation of IV with Fremy salts $[(SO_3K)_3]$ NO]⁵⁾ afforded p-quinoid compound (IX) (yield, 97%), $C_{23}H_{28}O_8(M^+, m/e, 432)$, reddish needles, mp 65—66°, $[\alpha]_D^{23}$ +85.0° (c=0.64, CHCl₃), UV $\lambda_{\text{max}}^{\text{EiOH}}$ nm (log ε): 216 (4.59), 241 (sh, 4.16), 274 (4.01), 365 (3.44). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450 (OH), 1655, 1645 (C=O), indicating that the hydroxyl group in IV is located at C₁₄ position. The structure of IX was confirmed by the

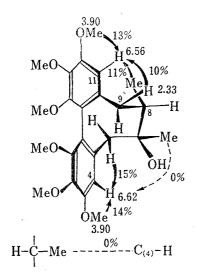


Fig. 1. NOE in V (in CDCl₃, δ value)

measurements of NOE(in CDCl₃) as shown in chart Irradiation of a methoxyl signal (δ 3.78) and a methylene proton signal ($\delta 2.45$) caused 16% and 19% increase in the integrated intensity of the aromatic proton signal, respectively. Irradiation of the tertiary methyl signal caused 7% increase of the integrated intensity of a methylene proton signal at δ 2.45 (d, J=15), while the aromatic proton showed no effect by irradiation of the both methyl Thus, the relative structure of IX including conformation is expressed by the formula IX. These facts agree with I value between C₈-proton and C₉-methylene protons, which show extreme down field (α -proton) and high field (β -proton) shifts by influence of the carbonyl group, in the PMR spectrum ($J_{8,9\alpha}$ =9 Hz, $\phi_{8,9\alpha}$ =30°; $J_{8,9\beta}$ =2 Hz, $\phi_{8,9\beta}$ = 90°).

³⁾ Tiglic acid and benzoic acid were identified by mixed mp, gas chromatography (GLC) and IR spectra. Angelic acid was identified by GLC.

⁴⁾ W.H. Pirkle and J.L. Zabriskie, J. Org. Chem., 29, 3124 (1964); W.H. Pirkle and M. Gates, ibid., 30, 1769 (1965).

⁵⁾ W. Moser and R.A. Howie, J. Chem. Soc. A, 1968, 3039.

Next, the absolute structure of schizandrin $(V)^{2,6}$ was determined by the measurements of NOE(in CDCl₃) and circular dichroism (CD) spectrum. As shown in Fig. 1, the results of NOE indicated that schizandrin has the same conformational structure as IX. On the other hand, the CD spectrum of V(in MeOH) showed two positive Cotton effects at 250 ([θ] +95000) and 235 nm ([θ] +75000), and a negative at 214 nm ([θ] -100000), indicating that V possesses R-biphenyl configuration. ^{1b,c)}

On the basis of the above observations, the absolute structures of angeloylgomisin H, tigloylgomisin H, benzoylgomisin H and schizandrin are expressed by the formulae I, II, III and V, respectively.

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6) Quite recently, Y.P. Chen, et al. suggested that schizandrin has trans-dimethyl groups at C₇ and C₈(Yuh-Pan Chen, Rolan Liu, Hon-Yen Hsu, S. Yamamura, Y. Shizuri and Y. Hirata, Bull. Chem. Soc. Japan, 50, 1824 (1977)). However, we believe that our assumption should be correct considering the results of spectral analyses as described above together with the results of C. Yan-Yang, et al. supporting that schizandrin has cis-dimethyl groups (Chen Yan-Yang, Shu Zeng-Bao and Li Lianf-Nianf, Scientia Scinica (Peking), 19, 276 (1976)).

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Structure of Gilvanol, a New Triterpene isolated from Quercus gilva Blume

A new triterpene named Gilvanol (Ia), $C_{30}H_{50}O_4$, mp 212—215°, which have been elucidated to be hopan-3 β -ol-17(21)-ozonide by chemical evidence and X-ray analysis, was isolated from *Quercus gilva* Blume.

Keywords—triterpene; hopane; gilvanol; ozonide; X-ray analysis; Quercus gilva Blume.

Previously, we reported about nine triterpenes isolated from *Quercus gilva* Blume.^{1,2)} One of them named as D-20 was not yet elucidated structurally. Now, we wish to report the structure of D-20 named as Gilvanol.

Gilvanol (Ia), $C_{30}H_{50}O_4$, mp 212—215°, possessed one hydroxy function (3410 cm⁻¹) by its infrared spectrum (IR), six tertiary methyls, two secondary methyls by its proton magnetic resonance (PMR) using Eu(dpm)₃ as a shift reagent, and showed the signal at δ 3.30 assignable to 3α -H. By mass spectrum of (Ia), the fragment ion peaks were observed at m/e 474 (M⁺), 456 (M⁺-H₂O), 207, 189 and 135 which had been characterized as the hopane skeleton.³⁾ On the other hand, the nature of three other oxygen atoms, was presumed to be an ozonide functional group, since no proton neighbouring to those oxygen atoms was observed by PMR and characteristic signals appeared at δ 112.4 ppm and 111.8 ppm on ¹³C-nuclear magnetic resonance (¹³C-NMR) spectrum.

¹⁾ Y. Kamano, Y. Tachi, T. Otake, J. Sawada, and I. Tanaka Yakugaku Zasshi, 96, 1202 (1976).

²⁾ Y. Tachi, Y. Kamano, J. Sawada, I. Tanaka, and H. Itokawa, Yakugaku ZAsshi, 96, 1213 (1976).

³⁾ G.R. Waller, "Biochemical Applications of Mass Spectrometry," Wiley-Interscience, 1972.