methyl)benzamidoxime, needles from EtOH, mp 148—150°. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3421, 3285 (NH), 1749, 1220 (COO), 1648, 1535 (CONH), 1618 (C=N). NMR (in DMSO- d_e) δ : 2.11 (3H, s, CH₃), 4.56 (2H, t, J=6 Hz, CH₂), 6.42 (1H, t, J=6 Hz, NH), 7.23—7.76 (11H, m, aromatic protons and NH). *Anal.* Calcd. for C₁₇H₁₇N₃O₃: C, 65.58; H, 5.50; N, 13.50. Found: C, 65.55; H, 5.50; N, 13.69. From the foregoing ethereal solution 0.14 g (6%) of N,N'-methylenebisbenzamide, mp 217—218°, was obtained.

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Lactams. XIV.¹⁾ cis-trans Isomerization in the 5-Ethyl-2-oxo-4-piperidine-acetic Acid System under Fischer-Speier Esterification Conditions²⁾

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It has been found that the reaction of the trans- (2a or 3a) or the cis-isomer (2b or 3b) of 5-ethyl-2-oxo-4-piperidineacetic acid (2) or its ethyl ester (3) with 10% EtOH-HCl at reflux for 20—27 hr gave an equilibrated 70: 30 mixture of the trans- (3a) and the cisisomer (3b) of the ester (3) in a good yield. Such a cis-trans isomerization did not occur at all when the trans- (2a) or the cis-lactam acid (2b) was similarly esterified but at 15° for 16 hr, whereas at 32° it did occur to a slight extent. In contrast to the N-unsubstituted lactam acids (2a, b), trans- (5a) and cis-1-benzyl-5-ethyl-2-oxo-4-piperidineacetic acid (5b) could be converted into the corresponding ethyl esters (7a, b) by similar esterifications even at 32° with complete retention of their original stereochemistry. The cis-trans isomerization of the methyl esters (6a, b) of 2a, b in 10% MeOH-HCl seemed somewhat easier than that of the ethyl esters (3a, b) in 10% EtOH-HCl. On the other hand, the N-benzylated methyl esters (8a, b) did not isomerize in 10% MeOH-HCl even after 5 hours' reflux.

Keywords—lactam acid; lactam ester; Fischer-Speier esterification; cis-trans isomerization; equilibrium; C-13 NMR spectroscopy; isomer ratio; quantitative analysis

Probably one of the most salient features in our recent synthetic incorporation of cincholoipon ethyl ester, derived from the cinchona alkaloid cinchonine, into some of the ipecac alkaloids⁴⁾ and the *Alangium* alkaloids ankorine⁵⁾ and alangicine⁶⁾ was the utilization of the potential molecular symmetry present in the title system (type 1) for $cis \rightarrow trans$ isomerization (1b \rightarrow 1a). We found that such an isomerization was feasible through the $cis \rightarrow trans$ equilibration (1b \rightarrow 1a) under acid hydrolytic conditions^{4,7)} or, more efficiently, on thermal treatment (e.g., 180°, without solvent).⁴⁻⁷⁾ The hydrolytic $cis \rightarrow trans$ isomerization was assumed⁷⁾ to proceed by a mechanism of ring opening followed by rotation and recyclization with another carboxyl group, apart from the mechanism⁷⁾ proposed for the thermal isomerization. In the

¹⁾ Paper XIII in this series, T. Fujii, S. Yoshifuji, and K. Yamada, Chem. Pharm. Bull. (Tokyo), 26, 2071 (1978).

²⁾ Presented in part at the 2nd Symposium on Progress in Chemical Reaction and Synthesis organized by Pharmaceutical Society of Japan, Osaka, Japan, November 5—6, 1975.

³⁾ Location: 13-1 Takara-machi, Kanazawa 920, Japan.

⁴⁾ T. Fujii and S. Yoshifuji, Tetrahedron Lett., 1975, 731.

⁵⁾ S. Yoshifuji and T. Fujii, Tetrahedron Lett., 1975, 1965.

⁶⁾ T. Fujii, S. Yoshifuji, S. Minami, S. C. Pakrashi, and E. Ali, Heterocycles, 8, 175 (1977).

⁷⁾ T. Fujii, S. Yoshifuji, and M. Tai, Chem. Pharm. Bull. (Tokyo), 23, 2094 (1975).

above alkaloid syntheses,^{4–6)} protection of the carboxyl group in **1a** was required for the operations in further steps and we decided to embody it by the Fischer–Speier esterification method.⁸⁾ However, the mechanistic similarity between the acidic hydrolysis and the acid-catalyzed alcoholysis of the lactam carbonyl–nitrogen bond could not exclude the possibility that the *trans*—*cis* isomerization might

occur under these particular esterification conditions. We, therefore, scrutinized the reactions of some model compounds with ethanolic or methanolic hydrogen chloride in order to learn about reaction conditions under which the stereochemical purity of the lactam esters of type 1 could be secured.

Stereochemically pure samples of the trans- (a-series) and the cis-isomers (b-series) of 5-ethyl-2-oxo-4-piperidineacetic acid (2) and of its N-benzyl analog (5) were separately allowed to react with 10% (w/w) ethanolic hydrogen chloride (EtOH-HCl) under various conditions. All the esterifications proceeded smoothly to furnish the stereochemically pure or impure lactam esters (3, 7) in good yields. The quantitative analysis of the cis- and the trans-isomers in the ester fractions was then carried out according to the previously reported carbon-13 nuclear magnetic resonance (NMR) spectroscopic method, which was found to be most satisfactory (accurate to $\pm 1\%$) and convenient among those tested. It consisted of the measurement of relative heights of the methylene carbon signals of the $C_{(5)}$ -ethyl groups appeared in the noise-decoupled C-13 NMR spectra of the lactam ester fractions (see Table I). It may be seen from Table II that the esterifications of the N-unsubstituted lactam acids (2a,b) at 32° for 16 hr were accompanied by the cis-trans isomerization to a slight extent. The isomerization became faster at reflux, attaining to an equilibrium (3a: 3b=70: 30) in ca. 27 hr. At 15°, however, no isomerization occurred for at least 16 hr. On the other hand, the esterifications of the trans- (5a) and the cis-isomer (5b) of the N-benzyl analog did not accompany any isomerized product even at 32° for 16 hr. As shown in Chart 2, the cis-trans isomerization may proceed through the proton-catalyzed ethanolysis of one lactam ester isomer to the ring-opened intermediate (type 4), which may rotate and recyclize to the other isomer.

Table I. $C_{(5)}$ -Ethyl Carbon Shieldings of Esters of 5-Ethyl-2-oxo-4-piperidineacetic Acid and Its N-Benzyl Analog

Compound	0 10 1 1	Chemical shifts for $C_{(5)}$ - C_2H_5 carbons in $CDCl_3^{\alpha}$			
	$C_{(4)}/C_{(5)}$ stereochemistry	CH ₃ carbon	CH ₂ carbon		
3a	trans	11.0	23.5		
3b	cis	11.9	20.5		
6a	trans	11.0^{b}	23.5^{b}		
6b	cis	$11.9^{b)}$	20.5^{b}		
7 a	trans	10.9	23.6		
7b	cis	11.6	20.4		
8a	trans	10.9^{b}	23.6^{b})		
8 b	cis	11.6^{b}	20.4^{b}		

a) In ppm downfield from internal tetramethylsilane. See ref. 7 for details of instrumentation and measurement.

b) From ref. 7.

⁸⁾ E. Fischer and A. Speier, Bev. Dtsch. Chem. Ges., 28, 3252 (1895).

Table II. Esterification of Lactam Acids by the Method of Fischer and Speier

Lactam acid		Reaction conditions		Lactam ester					
Compound	C ₍₄₎ /C ₍₅₎ sterechemistry	Reagent	Temp. (°C)	Time (hr)	Yield (%)	% trans-Isom	er % cis-Isome		
2a	trans	EtOH-HCl	15	16	94	100 (3a)	0 (3b)		
			32	16	95	98 ` ´	2		
			Reflux	0.83	92	95	5		
			Reflux	5	96	82	18		
2b	cis	EtOH-HCl	15	16	97	0 (3a)	100 (3b)		
			32	16	92	3	97		
			Reflux	0.83	87	8	92		
			Reflux	2.5	93	20	80		
			Reflux	5	90	41	59		
			Reflux	27	92	70	30		
5a	trans	EtOH-HCl	15	16	96	100 (7a)	0 (7b)		
			32	16	98	100	0 ` '		
5b	cis	EtOH-HC1	32	16	95	0 (7a)	100 (7b)		

Table III. Reaction of Lactam Esters under Fischer-Speier Esterification Conditions

Lactam ester		Reaction conditions		Lactam ester recovered					
Compound	C ₍₄₎ /C ₍₅₎ sterechemistry	Reagent Te	emp. $(^{\circ}C)^{a}$	Time (hr)	Recovery (%)	% tran	s-Isomer	% cis-	-Isome _r
3a	trans	EtOH-HCl	15	16	98	100	(3a)	0	(3b)
			Reflux	2.5	97	88	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	12	()
			Reflux	5	94	80		20	
			Reflux	8.3	95	74		26	
			Reflux	20	94	70		30	
3 b	cis	EtOH-HCl	15	16	96	0	(3a)		(3b)
			Reflux	2.5	95	22	(/	78	()
			Reflux	5	94	41		59	
			Reflux	8.3	90	54		46	
			Reflux	27	91	70		30	
6a	trans	MeOH-HCl	r.t.	16	92	100	(6a)		(6b)
			Reflux	5	92	80	()	20	(0.0)
6b	cis	MeOH-HCl	r.t.	16	94	14	(6a)		(6b)
			Reflux	5	90	50	()	50	(0.0)
8 a	trans	MeOH-HCl	r.t.	16	99	100	(8a)	0	(8b)
			Reflux	5	98	100	· ·/	0	\-·-/
8b	cis	MeOH-HCl	r.t.	16	96	0	(8a)	100	(8b)
			Reflux	5	99	0	\ <i>y</i>	100	(/

a) The abbreviation r.t. stands for room temperature (14° \pm 5°).

If such a mechanism is operative in the above isomerizations, lactam esters of that type should also undergo cis-trans isomerization under similar Fischer-Speier esterification conditions. Thus, the ethyl esters (3a,b) were separately treated with 10% (w/w) EtOH-HCl at 15° for 16 hr or at reflux; the methyl esters (6a,b, 8a,b), with 10% (w/w) methanolic hydrogen chloride (MeOH-HCl) at room temperature for 16 hr or at reflux for 5 hr. The lactam ester fractions were recovered from these reaction mixtures and isomer ratios were determined C-13 NMR spectroscopically as described above (see Table I). Table III summarizes the results. It may be seen that in the reaction with EtOH-HCl 3a,b did not isomerize at all at 15° for 16 hr, whereas at reflux each of them isomerized rapidly to give an equilibrated mixture (3a: 3b=70: 30) within 20-27 hr, paralleling the results (Table II) of the esterifications of the corresponding lactam acids (2a,b). The cis-trans isomerization of the methyl esters (6a,b) in MeOH-HCl seemed somewhat easier than that of the ethyl esters (3a,b) in EtOH-HCl, and it suggested that the esterification of 2b with MeOH-HCl is not safe stereochemically even at 9-19°. On the other hand, the methyl esters (8a,b) of the N-benzyl analog seemed much more stable: no isomerization was observed even after 5 hours' reflux.

In conclusion, the present results have provided a guide to stereochemically safe preparation of the ethyl and methyl esters of the 5-ethyl-2-oxo-4-piperidineacetic acid system by the Fischer-Speier esterification method. It is hoped that the guide will encourage further stereoselective syntheses using such a system as a key synthon.

Experimental9)

Materials—Among the stereochemically pure samples used in the present work were those of trans-5-ethyl-2-oxo-4-piperidineacetic acid (2a), 7,10,11) cis-5-ethyl-2-oxo-4-piperidineacetic acid (2b), 7,10) ethyl trans-5-ethyl-2-oxo-4-piperidineacetate (3a), 7,10,11) trans-1-benzyl-5-ethyl-2-oxo-4-piperidineacetic acid (5a), 7,10,11) cis-1-benzyl-5-ethyl-2-oxo-4-piperidineacetate (6b), 7) methyl trans-5-ethyl-2-oxo-4-piperidineacetate (6b), 7) methyl trans-1-benzyl-5-ethyl-2-oxo-4-piperidineacetate (8b), 7) and methyl cis-1-benzyl-5-ethyl-2-oxo-4-piperidineacetate (8b), 7) They were prepared according to the procedure reported in the literature as indicated, whereas the other samples were obtained in the manner described below.

Ethyl cis-5-Ethyl-2-oxo-4-piperidineacetate (3b) ——A solution of $2b^{7,10}$ (1.11 g, 6 mmol) in 10% (w/w) ethanolic hydrogen chloride (EtOH–HCl) (25 ml) was kept in a refrigerator (1—2°) for 24 hr. The solution was then evaporated in vacuo, and sat. aq. NaCl (20 ml) was added to the crystalline residue. The resulting mixture was made basic with anhyd. K_2CO_3 under cooling and extractad with CHCl₃ (1×40 ml, 2×20 ml). The CHCl₃ extracts were washed with sat. aq. NaCl (2×15 ml), dried over anhyd. Na₂SO₄, and evaporated in vacuo to leave 3b (1.24 g, 97%) as a colorless oil, bp 146° (10^{-3} mmHg); MS m/e: 213 (M+); IR v_{max}^{film} cm⁻¹: 3220 (b, NH), 1728 (ester CO), 1667 (lactam CO); IR $v_{max}^{\text{CHCl}_3}$ cm⁻¹: 3405, 3200 (NH), 1729 (ester CO), 1659 (lactam CO); NMR (CDCl₃) δ : 0.95 (3H, t, J=6.5 Hz, CCH₂Me), 1.26 (3H, t, J=7.0 Hz, OCH₂Me), 1.10—1.90 (4H, m, H₍₄₎, H₍₅₎, and CCH₂Me), 2.13—2.62 (4H, m, H₍₃₎'s and CH₂CO₂Et), 2.91—3.44 (2H, m, H₍₆₎'s), 4.14 (2H, q, J=7.0 Hz, OCH₂Me), 7.22—7.50 (1H, b, NH). The C-13 nuclear magnetic resonance (NMR) spectroscopic quantitative analysis as described below and infrared (IR) spectroscopy failed in detecting the presence of the trans-isomer (3a) in the crude or the distilled sample of 3b.¹²)

Ethyl trans-1-Benzyl-5-ethyl-2-oxo-4-piperidineacetate (7a)—A solution of $5a^{7,10,11}$) (826 mg, 3 mmol) in 10% (w/w) EtOH-HCl (30 ml) was allowed to stand in a refrigerator (1—2°) for 20 hr. The solution was evaporated in vacuo, and H_2O (10 ml) was added to the residue. The resulting mixture was made basic

⁹⁾ All melting points are corrected; boiling points, uncorrected. IR spectra were measured in Nujol mulls, in liquid films, or in CHCl₃ solutions at 0.2 m concentration. See also ref. 1 for details of instrumentation and measurement. The following abbreviations are used: b=broad, d=doublet, m=multiplet, q=quartet, s=singlet, t=triplet.

¹⁰⁾ T. Fujii, Chem. Pharm. Bull. (Tokyo), 6, 591 (1958).

¹¹⁾ T. Fujii, S. Yoshifuji, and M. Ohba, Chem. Pharm. Bull. (Tokyo), 26, 645 (1978).

¹²⁾ The esterifications of 2a and 2b were previously effected with 12.5% EtOH-HCl at reflux for 3.5 hr, giving 3a (83.2% yield; mp 93—94°) and 3b [82.6% yield; bp 165—166° (0.02 mmHg)], respectively. Although the highest stereochemical purity of the crystalline sample of 3a prepared by that method was secured by recrystallization, 7,10) it is most likely in the light of the results in Table II that the previous oily sample of 3b was contaminated with 3a. This was also supported by a separate experiment. 7)

with 10% aq. Na₂CO₃ under cooling and extracted with benzene (1 × 20 ml, 2 × 10 ml). The benzene extracts were washed with H₂O (10 ml), dried over anhyd. Na₂SO₄, and evaporated *in vacuo* to leave 7a (890 mg, 98%) as a colorless oil, MS m/e: 303 (M⁺); IR $v_{\rm max}^{\rm film}$ cm⁻¹: 1730 (ester CO), 1644 (lactam CO); IR $v_{\rm max}^{\rm cHOl_3}$ cm⁻¹: 1728 (ester CO), 1633 (lactam CO); NMR (CDCl₃) δ : 0.80 (3H, t, J=7.0 Hz, CCH₂Me), 1.25 (3H, t, J=7.0 Hz, OCH₂Me), 1.0—3.40 (10H, m, H₍₃₎'s, H₍₄₎, H₍₅₎, H₍₆₎'s, CCH₂Me, and CH₂CO₂Et), 4.12 (2H, q, J=7.0 Hz, OCH₂Me), 4.52 and 4.62 (1H each, AB type d's, J=14 Hz, CH₂Ph), 7.26 (5H, s, Ph). This sample was C-13 NMR (see below) and IR spectroscopically shown to be free from the contamination with the *cis*-isomer (7b).

Ethyl cis-1-Benzyl-5-ethyl-2-oxo-4-piperidineacetate (7b)—The cis-lactam acid (5b)^{7,10} was esterified as described above for the esterification of 5a, affording 7b (95% yield) as a colorless oil, MS m/e: 303 (M⁺); IR $v_{\rm max}^{\rm flim}$ cm⁻¹: 1730 (ester CO), 1644 (lactam CO); IR $v_{\rm max}^{\rm cHCl_3}$ cm⁻¹: 1728 (ester CO), 1633 (lactam CO); NMR (CDCl₃) δ : 0.79 (3H, t, J=7.0 Hz, CCH₂Me), 1.25 (3H, t, J=7.0 Hz, OCH₂Me), 1.07—3.22 (10H, m, H₍₃₎'s, H₍₄₎, H₍₅₎, H₍₆₎'s, CCH₂Me, and CH₂CO₂Et), 4.12 (2H, q, J=7.0 Hz, OCH₂Me), 4.45 and 4.69 (1H each, AB type d's, J=15 Hz, CH₂Ph), 7.27 (5H, s, Ph). No contamination of this sample with the trans-ester (7a) was indicated by means of C-13 NMR (see below) and IR spectroscopy.

Quantitative Analysis of the cis- and trans-Lactam Esters (3a,b, 6a,b, 7a,b, or 8a,b) in an Isomeric Mixture—Performed C-13 NMR spectroscopically in a manner similar to that described previously⁷⁾ for analysis of 2a,b, 5a,b, 6a,b, or 8a,b (see Table I).

Isomerization Study of the Lactam Acids (2a,b,5a,b) under Fischer-Speier Esterification Conditions—The lactam acids (2a,2b,5a, and 5b) (0.6 mmol) were separately dissolved in 10% (w/w) EtOH-HCl (6 ml). The resulting solutions were refluxed for the time specified in Table II or sealed in small ampoules and placed in a thermoregulated constant temperature bath kept at $15^{\circ}\pm0.1^{\circ}$ or at $32^{\circ}\pm0.1^{\circ}$ for 16 hr. The reaction mixtures were evaporated in vacuo and sat. solutions (5 ml) each) of NaHCO3 in H2O were added to the residues. The resulting mixtures were separately extracted with CHCl3 $(3\times10 \text{ ml})$. The CHCl3 solutions were washed with sat. aq. NaCl (10 ml), dried over anhyd. Na2SO4, and evaporated to dryness in vacuo, leaving crude lactam esters. After having been weighed, the ester fractions were submitted to the C-13 NMR spectroscopic analysis (see above) for determination of isomer ratios. The results are summarized in Table II.^{12,13})

Stereochemical Stability of the Lactam Esters (3a,b, 6a,b, 8a,b) under Fischer-Speier Esterification Conditions—The stereochemical stability of the ethyl esters (3a, b) (0.6 mmol) in 10% (w/w) EtOH-HCl (6 ml) was examined in a similar manner to that described above for the lactam acids (2a,b, 5a,b). In the cases of the methyl esters (6a,b, 8a,b), they were dissolved in 10% (w/w) methanolic hydrogen chloride (MeOH-HCl) (3 ml) at 0.14 m concentration and the resulting solutions were kept at room temp. (14°±5°) for 16 hr or refluxed for 5 hr. For isolation and isomer analysis of ester fractions, the reaction mixtures were worked up as described above for the esterification of the lactam acids (2a,b, 5a,b). Table III summarizes the results thus obtained. Small portions of the ester fractions from the reactions of 6a (room temp., 16 hr, or refluxing, 5 hr) and of 6b (refluxing, 5 hr) with MeOH-HCl were recrystallized from isopropyl ether to give colorless needles, mp 92—93°, identical (by IR spectrum and mixed melting-point test) with authentic 6a.7) The crude ester fractions obtained from the reactions of 8a and of 8b with MeOH-HCl, respectively, showed IR spectra superimposable on those of authentic 8a7) and 8b.7)

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¹³⁾ For stereochemically safe esterification procedure for preparation of 3a and 3b, see ref. 11 (3a) and the present paper (Experimental section, under the name of 3b).