(Chem. Pharm. Bull.) 27(6)1383—1394(1979)

UDC 547.913.02.05:581.192

The Constituents of Schizandra chinensis Baill. I. Isolation and Structure Determination of Five New Lignans, Gomisin A, B, C, F and G, and the Absolute Structure of Schizandrin¹⁾

YUKINOBU IKEYA, HEIHACHIRO TAGUCHI, ITIRO YOSIOKA, ^{2a)} and HIROSHI KOBAYASHI^{2b)}

Tsumura Laboratory^{2a)} and Faculty of Pharmaceutical Sciences, University of Tokyo^{2b)}

(Received November 20, 1978)

Five new dibenzocyclooctadiene lignans, named gomisin A(2), B(3), C(4), F(5) and G(6), were isolated from the petroleum ether extract of fruits of *Schizandra chinensis* Ball. (Schizandraceae) and their absolute structures were elucidated by chemical and spectral techniques.

The absolute structure of schizandrin(1), the plane structure of which had already been elucidated by Kochetkov *et al.*,³⁾ was also elucidated on the basis of spectroscopic results.

Keywords—Schizandra chinensis Baill.; Schizandraceae; lignans; gomisin A; gomisin B; gomisin C; gomisin F; gomisin G; schizandrin; NOE

The fruits of *Schizandra chinensis* Balll. (Schizandraceae) are used as an antitussive and a tonic under the names "Hoku-(or Kita-)gomishi"(北五味子) in Japan and "Wu-wei-zi" in China.⁴⁾

Sesquiterpenoids (α - and β -chamigrene, chamigrenal and sesquicarene) were isolated from the essential oil of the fruits by Ohta *et al.*,⁵⁾ and several dibenzocyclooctadiene lignans (schizandrin, deoxyschizandrin and γ -schizandrin) have been isolated from the unhydrolyzed fraction of seed oil of this plant by Kochetkov *et al.*³⁾

This paper deals with the isolation and structure elucidation of five new lignans, named gomisin A (2), B (3), C (4), F (5) and G (6), and also describes the absolute structure of schizandrin (1), the plane structure of which has been elucidated by Kochetkov *et al.*^{3,6)} The isolation procedure for the compounds is summarized in Chart 1 and described in the experimental section.

Compound 1 was isolated as colorless prisms(ether–n-hexane) (yield, 0.26%), $C_{24}H_{32}O_7$, mp 128.5—130°, $[\alpha]_D^{22}$ +88.7° (in CHCl₃), and was suggested to be schizandrin on the basis of comparison of the physical data with those of schizandrin reported by Kochetkov *et al.*,³⁾ as well as proton nuclear magnetic resonance (PMR) (Table I) and mass spectral analyses (Chart 3). Furthermore, oxidation of 1 with potassium permanganate followed by methyla-

¹⁾ Preliminary communications, see: a) H. Taguchi and Y. Ikeya, Chem. Pharm. Bull. (Tokyo), 23, 3296 (1975); b) Idem., ibid., 25, 364 (1977); c) Y. Ikeya, H. Taguchi, and I. Yosioka, ibid., 26, 328 (1978); d) After this work had been completed, Liu et al. reported the isolation of gomisin B(3) and C(4) from Schisandra sphenanthera under the names of schisantherin B and A, respectively [J.S. Liu, S.D. Fang, M.F. Huang, Y.L. Gao, and J.S. Hsu, Hua Hsueh Hsueh Pao, 34, 229 (1976) [Chem. Abstr., 89, 24193d (1978)].

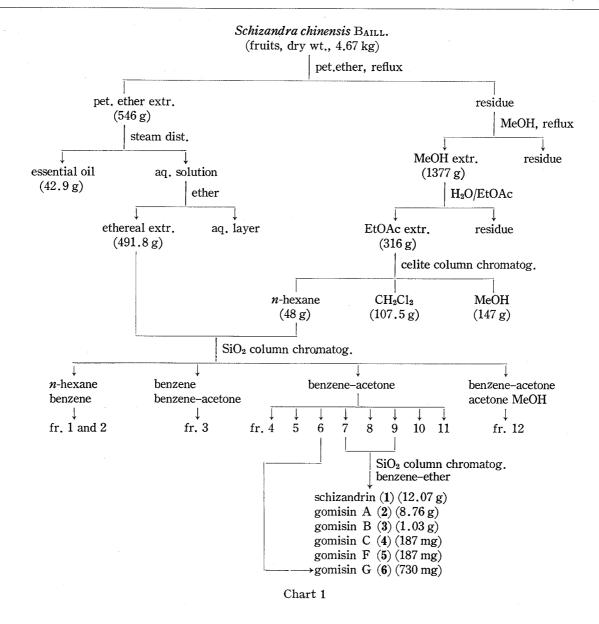
²⁾ Location: a) Izumi 1421, Komae-shi, Tokyo 201 Japan; b) Hongo, Bunkyo-ku, Tokyo 113 Japan.

³⁾ a) N.K. Kochetkov, A. Khorlin, and O.S. Chizhov, Zh. Obshch. Kim., 31, 3454 (1961); b) N.K. Kochetkov, A. Khorlin, O.S. Chizhov, and V.I. Sheichenko, Tetrahedron Lett., 1961, 730; c) N.K. Kochetkov, A. Khorlin, and O.S. Chizhov, Tetrahedron Lett., 1962, 361.

⁴⁾ H.-Y. Hsu and W.G. Peacher, "Chinese Herb Medicine and Therapy," Oriental Healing Arts Institute of U.S.A. and Aurora Publishers Incorporated., 1976, p. 176.

⁵⁾ Y. Ohta and H. Hirose, Tetrahedron Lett., 1968, 1251, 2483.

⁶⁾ The partial stereo-structure of schizandrin (presence of a cis-dimethyl) was elucidated by Chen et al. [Y.Y. Chen, Z.B. Shu, and L.L. Li, Scientia Sinica (Peking), 19, 276 (1976)].



tion with diazomethane gave dimethyl 4,4′,5,5′,6,6′-hexamethoxydiphenate (1a),7′ $C_{22}H_{26}O_{10}$, mp 90—91°, $[\alpha]_{D}^{24}$ +20.1° (in CHCl₃). On the basis of these results, 1 was identified as schizandrin.

The conformational structure of 1 was elucidated by measurements of intramolecular nuclear Overhauser effects (NOE) (in CDCl₃) as in the cases of kadsurin and kadsurarin, which are similar lignans isolated from the stem of *Kadsura japonica* Dunal (Schizandraceae),⁸⁾ and gomisin D.⁹⁾ As shown in Fig. 1 and Table II, irradiation of a methoxyl signal (δ 3.90) caused 14% and 13% increases in the integrated intensity of the C₍₄₎ proton (δ 6.60) and C₍₁₁₎ proton (δ 6.53), respectively. Irradiation of a secondary methyl signal (δ 0.82) caused an 11% increase in the integrated intensity of the C₍₁₁₎ proton, while the C₍₄₎ proton was unaffected, indicating that the C₍₁₁₎ proton and secondary methyl are close to each other. Irradiation at δ 2.33, corresponding to the higher field protons of two methylene groups (C₍₆₈₎-H and C_(9 α)-H),

⁷⁾ D.E. Harthway, J. Chem. Soc., 1957, 519.

⁸⁾ Y. Chen, R. Lin, H. Hsu, S. Yamamura, Y. Shizuri, and Y. Hirata, Tetrahedron Lett., 1973, 4257; idem, Bull. Chem. Soc. Japan, 50, 1824 (1977).

⁹⁾ Y. Ikeya, H. Taguchi, and Y. Iitaka, Tetrahedron Lett., 1976, 1359; Y. Ikeya, H. Taguchi, I. Yosioka, Y. Iitaka, and H. Kobayashi, Chem. Pharm. Bull. (Tokyo), 27, 1395 (1979).

Chart 2

caused 15% and 10% increases in the integrated intensity of the $C_{(4)}$ proton and $C_{(11)}$ proton, respectively. Irradiation of a tertiary methyl signal did not affect the aromatic protons.

On the basis of the above results and a Dreiding model examination, the conformational structure of schizandrin was deduced to be $1.^{10}$. The J values between the $C_{(8)}$ proton and $C_{(9)}$ methylene protons in the PMR spectrum were consistent with the structure $1(J_{8,9\alpha}=7 \text{ Hz}, \phi_{8,9\beta}=2 \text{ Hz}, \phi_{8,9\beta}=90^{\circ})$, as shown in Fig. 2.

Gomisin A (2) was isolated as colorless needles (from MeOH) (yield, 0.185%), $C_{23}H_{28}O_7$, mp 88—89°, $[\alpha]_D^{25}$ +67.9° (in CHCl₃). The ultraviolet (UV) spectrum of 2 showed absorption maxima at 218 (log ε , 4.88), 253 (4.30), 281 (sh 3.70) and 290 nm (sh 3.57), indicating that 2 is a dibenzocyclooctadiene lignan. The PMR spectrum (Table I) showed the presence of a secondary methyl shielded by the aromatic ring, a tertiary methyl attached to a carbon carrying a hydroxyl group, and two benzylic methylene groups, and also indicated the presence of a methylenedioxyl moiety and four methoxyls on the aromatic rings. On the other hand, one aromatic proton appeared at higher field than that of schizandrin, indicating that the methylenedioxyl moiety may be linked adjacent to the aromatic proton.¹¹⁾

 ^{(±)-}Schizandrin was recently synthesized by Ghera et al. (E. Ghera and Y. Ben-David, Chem. Commun., 1978, 480).

¹¹⁾ M. Mervic and E. Ghera, J. Am. Chem. Soc., 99, 7673 (1977).

Table I. PMR Spectral Data for 1—6, 1c, 2a, 3a, 3c, 5a and 5b (in CDCl₃, ô Value, 100 MHz)

Compound	4-H, s 11-H, s (.	6α -H $(J=Hz)$	$_{(J=Hz)}^{6eta-H}$	9α -H d,d $(J=Hz)$	9β -H d,d $(J=Hz)$	$ \begin{array}{c c} \downarrow \\ CH_3-\zeta-H^d)\\ d\\ m\\ (J=Hz) \end{array} $	Hz	CH3-C	CH3-¢-OH6)	S s	OCH_2O $(J=Hz)$
1	6.60	2.70 d (14)		2.33 (14/7)	2.68 (14/2)	0.82	1.80	1.25	1.86	$3.59(\times 2), 3.90(\times 2)$	-
10		2.67 d (14)		3.20 (14.5/8)	2.78 (14.5/2)	0.90	1.75	1.33	2.02	3.47, 3.60	
61	6.62 6.49	2.72d (13.5)		2.30 (14/7)	2.64 (14/2)	0.80	1.80	1.25	1.85	$3.52(\times 2)$, $3.91(\times 2)$ 3.50, 3.80 , $3.90(\times 2)$	5.96 s
2a]	2.69d (14)		3.15 (14.5/8)	2.72 (14.5/2)	0.90	1.73	1.33	2.03	3.43, 3.82, 3.92, 3.97	6.07 s
$3^a,c)$	6.83 6.48	5.70 s		2.00- n	-2.45 n	1.13	1.90	1.33	1.60	3.57, 3.75 3.85, 3.93	5.90 s
38	$6.62 \\ 6.51$	4.59 s		2.35 (12/8.5)	2.05 (12/1)	1.11	1.85	1.40	1.56	$3.61, 3.90$ $3.93(\times 2)$	5.95 s
$3c^{a)}$	6.70 6.40	1		2.53 (14/9)	2.20 (14/2)	1.10	1.70	1.37	1.92	3.70, 3.80	5.93 s
$4^a,c$	6.89 6.60	5.90 s		2.0- n	_2.5 n	1.16	1.65	1.36	1.65	$3.32, 3.60, 3.93(\times 2)$	d, 1
5 ⁶)	6.82 6.48	5.88 s		2.52 (14/9)	2.05 (14/1)	1.10	1.80	1.25	1.30	3.53, 3.67, 3.83, 3.85	5.30(1H, d, 1.5) 5.43(1H, d, 1.5)
$\mathbf{5a}^{a)}$	6.57 6.53	4.53 d f) (11)		2.43 (14/8)	2.08 (14/1)	1.13	1.78	1.39	1.70	$3.68, 3.85, 3.90(\times 2)$, s (s)
${f 5b}^a$	6.60 6.45	1		2.37 (cent	er, 2H, m)	1.13	1.75	1.37	1.83	3.60, 3.83, 3.88, 3.92	6.02s
6 a,c)	6.78	5.90 s	1	2.0- n	2.0—2.6 m	1.17 (7)	2.07	1.33	1.57	3.15, 3.43, 3.82, 3.98	6.00 s

a) These compounds were measured at 60 MHz. b) This compound was measured in C_6D_6 .

c)

8:1.40 (3H, q, J=1.5 Hz, α -Me), 1.86 (3H, q, J=7/1.5 Hz, β -Me), 6.00 (1H, q, q, J=7/1.5 Hz, β -H). 5:1.33 (3H, q, J=1Hz, α -Me), 1.82 (3H, d, q, J=7/1Hz, β -H). 4: 7.3—7.7 (5H, m, Ph-CO-). 6: 7.35 (center, 5H, m, Ph-CO-). Hydroxyl signals of all compounds were confirmed by addition of D₂O. display after addition of D₂O. decouplet, m=multiplet, s=singlet.

 $\widehat{g}(\widehat{f})$

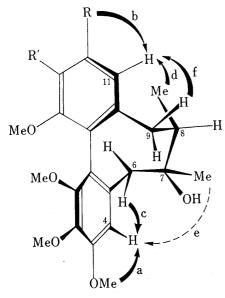


Fig. 1. NOE in 1 and 2

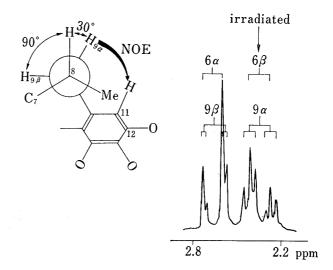


Fig. 2. Relationship between J Values and Dihedral Angle between the $C_{(8)}$ and $C_{(9)}$ Protons in 1 and 2

TABLE II. Results of NOE Measurements in 1 and 2

	1: R=R'=OMe	2: R, R'=OCH ₂ O
	NOE	NOE
a	14% (irrad. at δ 3.90)	14% (irrad. at δ 3.80)
b	14% (irrad. at δ 3.90)	0 (irrad. at each OMe)
c	15%	12%
d	11%	10%
e	0	0
f	10%	12%

$$\begin{array}{c} R \\ R' \\ MeO \\ MeO \\ MeO \\ OMe \\ OM$$

1:
$$R=R'=OMe$$
, m/e 432 (100%)
2: R , $R'=OCH_2O$, m/e 416.1869 (100%) R' H
 MeO CH_2 CH_2 $R=R'=OMe$, 330 (38%) R , $R'=OCH_2O$, 314.1159 (50%) $C_{18}H_{18}O_5$

R = R' = OMe 361 (15%)R, $R' = OCH_2O 345.1287 (17\%)$

Chart 3

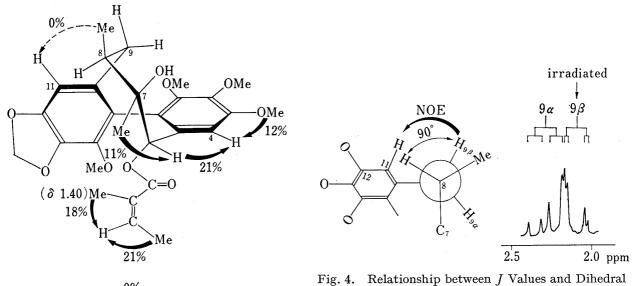
Treatment of 2 with bromine in carbon tetrachloride saturated with water gave the dibromoderivative (2a),¹²⁾ the PMR spectrum of which lacked two aromatic proton signals and showed downfield shifts of two methylene signals, as observed in the case of 4,11-dibromoschizandrin (1c). This suggested that the aromatic protons and methylene groups are close to each other.

On the basis of the above results, as well as mass spectral analysis (Chart 3), the plane structure of gomisin A was deduced. The structure of 2 was confirmed by measurements of the NOE (in CDCl₃) as in the case of 1. The results are shown in Fig. 1 and Table II (the $C_{(11)}$ proton was unaffected by irradiation of each methoxyl signal, indicating that the methylenedioxyl moiety is linked to the 12 and 13 positions). Thus, the structure of gomisin A was elucidated as 2.

Gomisin B (3) was isolated as colorless prisms (from ether-n-hexane), $C_{28}H_{34}O_{9}\cdot 1/2$ - $C_2H_5OC_2H_5$, mp 95—97°, $[\alpha]_D^{23}$ —26.6° (in CHCl₃) (yield 0.022%). The UV spectrum of 3, with absorption maxima at 218 (log ε 4.98), 257 (sh 4.24) and 292 nm (sh 3.65) and the infrared (IR) spectrum, with bands at 3500 (OH), 1715 (ester), 1640 (C=C), 1610 and 1590 (aromatic) cm⁻¹, indicated that 3 is a dibenzocyclooctadiene lignan possessing a hydroxyl and an ester linkage. The PMR spectrum showed two olefinic methyls δ 1.40 (q, J=1.5Hz) and 1.86 (d, q, J=7/1.5 Hz)], and an olefinic proton [δ 6.00(q, q, J=7/1.5 Hz)] due to an angeloyl group. The strong peaks at m/e 83 (base peak) and m/e 55 in the mass spectrum also suggested the presence of an angeloyl group (Chart 4 and Table III). On hydrolysis with 3% ethanolic potassium hydroxide, 3 gave a diol (3a), $C_{23}H_{28}O_8$, mp $209-210^\circ$, $[\alpha]_D^{23}$ -91.8° (in CHCl₃), IR: 3500, 3425 cm⁻¹ (OH) and no C=O band, and tiglic acid. Although angelic acid was not obtained in the above hydrolysis, it is clear that 3 possesses an angeloyl group from the chemical shift of the olefinic proton (δ 6.00, in CDCl₃) in the PMR spectrum, in comparison with those of angelic acid (δ 6.22) and tiglic acid (δ 7.00). The PMR spectrum of 3a showed the presence of a methylenedioxyl moiety and four methoxyls on the aromatic rings, as well as a secondary methyl and a tetriary methyl (CH_3 -C-OH). The signals around δ 2.00—2.40 (2H) appeared as an ABX octet, indicating the presence of an Ar-CH₂-CH-CH₃ moiety. The singlet at δ 4.59, which appeared at δ 5.70 in 3, was assigned to a benzylic methine (C₍₆₎-H). Oxidation of **3a** with chromium trioxide in acetic acid gave compound **3b**, $C_{23}H_{26}O_8(m/e~430)$, which showed carbonyl (1710 cm⁻¹) and aldehyde (1685 cm⁻¹) bands in the IR spectrum, and an aldehyde proton (δ 9.57) and methylketone (δ 1.83) signals in the PMR spectrum. Oxidation of 3a with chromium trioxide in pyridine gave compound 3c, $C_{23}H_{26}O_8$, mp 129—130°, $[\alpha]_p^{22}$ —88.5° (in CHCl₃), IR: 1690 cm⁻¹ (C=O). The structure of the diol (3a) was thus elucidated as 3a except for the positions of the functional groups on the aromatic rings.

Finally, the structure of 3 was confirmed by measurements of the NOE in 3 and 3a as shown in Fig. 3. In the case of 3, irradiation of methoxyl (δ 3.93) and methine signals (δ 5.70, $C_{(6)}$ –H) caused 12% and 21% increases in the integrated intensity of the aromatic proton signal at δ 6.83 ($C_{(4)}$ –H), respectively, while no enhancement of the signal intensity of the aromatic proton at δ 6.48 ($C_{(11)}$ –H) was detected upon irradiation of each methoxyl signal. These findings indicate that the methylenedioxyl moiety must be located adjacent to the $C_{(11)}$ proton. Irradiation of the tertiary methyl signal (δ 1.33) caused an 11% increase in the integrated intensity of the $C_{(6)}$ proton (δ 5.70), indicating that the tertiary methyl and $C_{(6)}$ proton must be close to each other. The aromatic protons were unaffected by irradiation of the two methyl signals. The presence of an angeloyl group in 3 was also confirmed by the NOE measurements. On the other hand, the interaction between the higher field proton (δ 2.05, $C_{(9\beta)}$ –H) of the methylene group and the $C_{(11)}$ proton was detected in the NOE measurements in 3a. On the basis of the above results, the relative structure of gomisin

¹²⁾ R.M. Keefer, J.H. Blake III and L.J. Andrews, J. Am. Chem. Soc., 76, 3062 (1954).



Angles between the $C_{(8)}$ and $C_{(9)}$ Protons in 3a

Fig. 3. NOE in 3 and 3a (in CDCl₃)

Chart 4. Mass Fragments of 3, 3a, 4, and 5

TABLE III. Mass Spectral Data of 3,3a,4 and 5

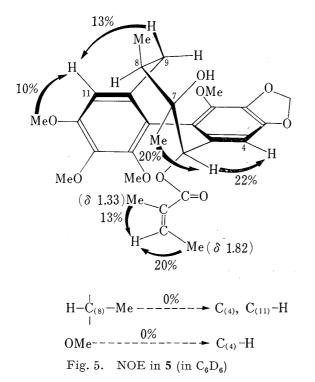
C- 1	M+			Fragment	t, m/e (%)		
Compound	m/e (%)	Í	Ι	П	IV	V	VI
3	514(56)	414(16)	343 (34)	83 (100)	55 (98)		
3a 4 5	432 (91) 536 (58) 514 (62)	414 (42) 414 (66) 414 (48)	343 (100) 343 (100) 343 (96)	— 83 (91)	 55 (100)	105 (90)	77 (49) —

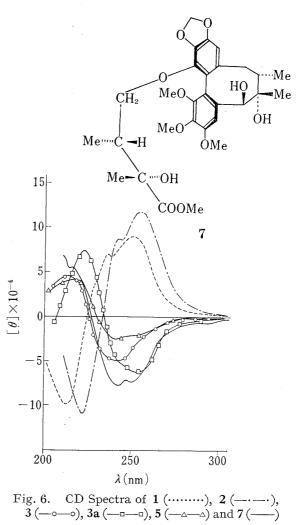
B was elucidated as 3. The J values between the $C_{(8)}$ proton and $C_{(9)}$ -methylene protons of 3a were consistent with this structure, as shown in Fig. 4 ($J_{8,9a}=8.5 \text{ Hz}$, $\phi_{8,9a}=150^{\circ}$; $J_{8,98}=1 \text{ Hz}$, $\phi_{8,9a}=90^{\circ}$). In addition, the IR spectrum of 3c showed an unconjugated carbonyl band (1690 cm⁻¹) and the PMR spectrum showed no downfield shift of the aromatic proton, 11,13) supporting the view that 3 has a twist-boat-chair conformation of the cyclooctadiene ring. 14)

Gomisin C (4) was isolated as colorless prisms (from ether–n-hexane), $C_{30}H_{32}O_{9}\cdot 1/2$ - $C_{2}H_{5}OC_{2}H_{5}$, mp 115—116°, $[\alpha]_{D}^{25}$ —186° (in CHCl₃) (yield 0.004%). The IR, PMR and mass spectra of 4 indicated that 4 possesses a benzoyl group. On hydrolysis with 3% ethanolic potassium hydroxide, 4 gave compound 3a and benzoic acid. The structure of gomisin C was thus elucidated as 4.

Gomisin F (5) was isolated as colorless needles (from ether–n-hexane) (yield 0.004%), $C_{28}H_{34}O_9$, mp 119.5—120.5°, $[\alpha]_D^{23}\simeq 0^\circ$ (in CHCl₃) and gomisin G (6) was isolated as colorless prisms (from ether–acetone) (yield 0.016%), $C_{30}H_{32}O_9\cdot 1/2C_2H_5OC_2H_5$, mp 97—98°, $[\alpha]_D^{25}=126^\circ$ (in CHCl₃). The UV and IR spectra of 5 and 6 indicated that the two compounds are cyclo-octadiene lignans possessing a hydroxyl and an ester linkage. The PMR spectral data of 5

and 6 (Table I) closely resembled those of 3 and 4, respectively, and showed that 5 possesses an angeloyl group [PMR (in C_6D_6): δ 1.33 (3H, q, J=1 Hz), 1.82 (3H, d, q, J=7/1 Hz), 5.63 (1H, q, q, J=7/1 Hz); mass spectrum: m/e, 83 (91%) and 55 (base peak)] and that 6 possesses a benzoyl group [PMR (in CDCl₃): δ 7.35 (5H, m); IR (in KBr): 722 cm⁻¹, isolated phenyl].





¹³⁾ a) A.S. Kende, L.S. Liebeskind, C. Kubiak, and R. Eisenberg, J. Am. Chem. Soc., 98, 6389 (1976);
b) Our unpublished data show that the C-6 carbonyl groups of lignans of the same type, which have a boat conformation of the cyclooctadiene ring, appear as a conjugated carbonyl band around 1660 cm⁻¹ in the IR spectra and the C-4 protons appear at around δ 7.50 in the PMR spectra (in CDCl₃).

14) F.A.L Anet and I. Yavari, Tetrahedron Lett., 1975, 1567.

On hydrolysis with 3% ethanolic potassium hydroxide, 5 and 6 gave the same diol (5a), $C_{23}H_{28}O_8$, mp 222.5—224°, $[\alpha]_D^{25}$ -73.1° (in CHCl₃) and the corresponding acids (during the hydrolysis of 5, angelic acid was isomerized to tiglic acid). The PMR spectrum of 5a also closely resembled that of 3a obtained from 3 and 4 by the same procedure, suggesting that 5a possesses two equatorial methyls (a trans-dimethyl) and has the same conformational structure as 3a. The highfield shift of the $C_{(6)}$ proton of 5a, compared with those of 5 and 6, indicated that the acyl groups in 5 and 6 are linked to the C₍₆₎-hydroxyl group of 5a. On oxidation with chromium trioxide in pyridine, 5a gave compound 5b, C₂₃H₂₆O₈, mp 227.5— 229.5°, $[\alpha]_{D}^{23}$ -93.6° (in CHCl₃), IR (in KBr): 3460 (OH), 1695 (C=O) cm^{-1.13}) The spectral data (IR, PMR and mass spectra) for 5b were very similar to those of 3b, but the physical constants (mp and $[\alpha]_p$) of the two compounds were quite different. Thus, it appears that 5a has the same structure as 3a except for the positions of the functional groups on the aromatic rings. The structures of 5 and 6 were confirmed by measurements of the NOE in 5 (in C_6D_6) as in the case of 3. The results are shown in Fig. 5. In conclusion, 5 appears to possess a methylenedioxyl moiety at C-(2-3), four methoxyls at the C-1, -12, -13 and -14 positions, and the same cyclooctadiene ring moiety as 3. The relative structures of gomisin F and G were thus elucidated as 5 and 6, respectively.

The configurations of the biphenyl groups in 1, 2, 3, 4, 5 and 6 were decided by comparison of the circular dichroism (CD) spectra with that of compound 7 derived from gomisin D, the absolute structure of which has been elucidated by X-ray analysis and shown to have an S-biphenyl configuration.^{9,15)}

As shown in Fig. 6, the CD spectrum of 7 showed two negative Cotton effects at 249 ([θ] -72400, sh) and 240 nm ([θ] -78500), and a positive effect at 216 nm ([θ] +54000). Similarly, gomisin B (3), F (5) and 3a showed a negative Cotton effect around 235—255 nm and a positive one around 215—225 nm, while gomisin A (2) and schizandrin (1) showed two positive Cotton effects around 235—255 nm and a negative one around 215—225 nm, indicating that gomisin B, C, F and G possess an S-biphenyl configuration, while gomisin A and schizandrin possess an R-biphenyl configuration.

Consequently, the absolute structures of schizandrin and gomisin A, B, C, F and G are expressed by the formulae 1, 2, 3, 4, 5 and 6, respectively.

Experimental

All melting points were determined on a Yanagimoto micro-melting point apparatus (hot-stage type) and are uncorrected. The UV spectra were recorded with a Hitachi 624 digital spectrophotometer and the IR spectra with a Hitachi EPI-G2 unit. The PMR spectra were recorded with Varian T-60 and JEOL PS-100 spectrometers. Mass spectra (MS) were measured with Hitachi double-focusing and JEOL JMS-01SG-2 mass spectrometers. The specific rotations were measured with a JASCO DIP-SL machine. TLC plates were made with silica gel (Kieselgel GF $_{254}$, Merck), and silica gel (Kieselgel 70—325 mesh, Merck) was used for column chromatography.

Extraction—The dried fruits of Schizandra chinensis were pulverized and extracted with pet. ether (bp 37—39°) under reflux $(4 \times 12 \text{ l}, 8 \text{ hr} \text{ each})$ and the residue was extracted with hot MeOH $(4 \times 24 \text{ l}, 8 \text{ hr} \text{ each})$. The pet. ethereal extracts were concentrated to give a brown mass (yield 546 g, 11.6%), which was steam-distilled to remove the essential oil (essential oil, yield 42.9 g, 0.92%). The undistilled part was extracted with ether and the ethereal extracts were concentrated to give a yellowish-brown residue (491.8 g). The methanolic extracts were concentrated to give a brown mass (1377 g, 29%), which was dissolved in H_2O (6 l) and extracted with EtOAc (3×6 l). The EtOAc extract (316 g), after concentration, was dissolved in MeOH and mixed with Celite (No. 535, Wako Pure Chemical Industries Ltd., 1200 g). The mixture was dried at room temperature and chromatographed, developing with n-hexane (8 l), CH_2Cl_2 (5 l) and then MeOH (8 l). The n-hexane eluate was concentrated to give a residue (48 g), which was combined with the undistilled part of the pet. ethereal extract. This mixture (total 539 g) was chromatographed on silica gel

¹⁵⁾ K. Mislow, M.A.W. Glass, R.E. O'Brien, P. Rutkin, D.H. Steinberg, J. Weiss, and C. Djerassi, J. Am, Chem. Soc., 84, 1455 (1962); J.P. Ferris, C.B. Boyce, R.C. Briner, U. Weiss, I.H. Qureshi, and N.E. Sharpless, ibid., 93, 2963 (1971); B. Feringa and H. Wynberg, ibid., 98, 3372 (1976).

(5 kg), developing with n-hexane-benzene, benzene and then benzene-acetone solvent systems. The details of this chromatography are given in Table IV.

Table IV.	Silica Gel Column	Chromatography o	f the Pet. Et	ther Extract ((undistilled part)
and n -	Hexane Eluate from	n Celite Column Ch	iromatograph	hy of the MeC)H Extract

Fraction No.	Solvent	Volume (1)	Yield (g)
1	n-Hexane-benzene (1:1)	4.0	10.78
2	n-Hexane-benzene (9:1)	3.0	204.44
3	Benzene	6.5	
	Benzene-acetone (98: 2)	5.0	114.0
	Benzene-acetone (96:4)	5.0	
4	Benzene-acetone (94:6)	1.5	12.85
5	Benzene-acetone (92:8)	2.5	64.89
6	Benzene-acetone (90: 10)	9.5	10.97
7	Benzene-acetone (88: 12)	5.0	24.45
8	Benzene-acetone (85:15)	6.5	12.85
9	Benzene-acetone (80: 20)	9.5	31.11
10	Benzene-acetone (75:25)	6.0	6.07
11	Benzene-acetone (70:30)	1.5	6.91
	(60:40)	2.0	
	(50: 50)	1.0	
	(30:70)	1.5	
12	Benzene-acetone (10:90)	1.5	20.50
	Acetone	2.5	
	Acetone-methanol (70:30)	1.0	

Isolation of Schizandrin(1) and Gomisin A(2), B(3), C(4), F(5) and G(6) ——Fraction (fr.) 6 (Table IV) was rechromatographed on silica gel (240 g) using n-hexane—EtOAc. The fractions eluted with n-hexane—EtOAc (7:3) were combined and concentrated to give a residue (401 mg), which was purified by preparative TLC (PLC) [n-hexane—acetone (7:3)] to give 6 (275 mg). Fr. 7 was crystallized from MeOH to give 2 as colorless needles (3.97 g). Fr. 9 was crystallized from ether—n-hexane to give 1 as colorless prisms (4.21 g). The mother liquors of fr. 7 and fr. 9 were combined with fr. 8 and the mixture was concentrated to give a residue (70.47 g), which was rechromatographed on silica gel (700 g), developing with benzene containing increasing amounts of ether (6% ether—benzene—ether, total 10.2 l). The fractions eluted with 14% ether—benzene were combined (total 2.4 l) and concentrated to give a residue (5.11 g). Repeated column chromatography (SiO₂, 120 g, acetone—n-hexane) and PLC [acetone—n-hexane (3:7)] of this residue gave 5 (187 mg, yield 0.004%) and 6 (455 mg, total yield 730 mg, 0.016%). The fractions eluted with 16% ether—benzene were combined (total 2.6 l) and concentrated to give a residue (4.64 g). Repeated column chromatography (SiO₂, 100 g, benzene—ether) and PLC [n-hexane—EtOAc (1:1)] of this residue gave 3 (1.03 g, 0.022%) and 4 (187 mg, 0.004%). The fractions eluted with benzene containing from 18% to 50% ether gave 1 (7.86 g, total yield 12.07 g, 0.26%) and 2 (4.77 g, total yield 8.76 g, 0.19%).

Schizandrin (1)—Colorless prisms (ether—n-hexane), mp 128.5—130°, $[\alpha]_D^{22}$ +88.7° (c=0.767, CHCl₃), UV $\lambda_{\max}^{\text{Etoff}}$ nm (log ε): 217 (4.90), 246 (4.29), 251 (4.30), 256 (sh 4.23), 285 (sh 3.44). IR ν_{\max}^{KBr} cm⁻¹: 3495 (OH), 1593 (aromatic). CD (c=0.0232, MeOH), $[\theta]^{23}$ (nm): -104000 (213), +69000, sh (236), +92000 (250). The PMR and mass spectral data are given in Table I and Chart 3, respectively. Anal. Calcd. for C₂₄H₃₂O₇: C, 66.65; H, 7.46. Found: C, 66.62; H, 7.47.

Oxidation of 1 with Potassium Permanganate—KMnO₄ (1.5 g) was added to a solution of 1 (600 mg) in a mixture of pyridine (5 ml) and 2% NaOH, and the reaction mixture was kept at 60° for 14 hr. The reaction mixture was then acidified with 10% $\rm H_2SO_4$ and $\rm Na_2S_2O_3$ was added to dissolve $\rm MnO_2$. The mixture was extracted with ether (3×15 ml). The combined ethereal extract was extracted with 0.5 n NaOH (3×10 ml), washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated to recover 1 (297 mg). The combined ethereal extract was washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and extracted with ether (3×15 ml). The combined ethereal extract was washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated to give a residue, which was purified by PLC [benzene-ether-AcOH (10:10:2), Rf 0.6] to give an acid (38 mg). This compound was methylated with diazomethane in ether and the product was purified by PLC [benzene-ether (3:2), Rf 0.7] to give 1a as colorless prisms (ether-n-hexane), 24 mg, mp 90—91°, $[\alpha]_p^{24}$ +20.1° (c=0.686, CHCl₃), UV $\lambda_{\rm max}^{\rm Etoh}$ nm (log ε): 221 (4.66), 254 (4.22), 301 (3.71). PMR (δ in CDCl₃): 3.63 (12H, s, 4×OCH₃), 3.97 (12H, s, 4×OCH₃), 7.38 (2H, s, arom.-H). Anal. Calcd. for $\rm C_{22}H_{26}O_{10}$: C, 58.66; H, 5.82. Found: C, 58.86; H, 5.86. Reduction of 1a with Lithium Aluminium Hydride—LiAlH₄ (13 mg) was added to a solution of 1a (13

Reduction of 1a with Lithium Aluminium Hydride—LiAlH₄ (13 mg) was added to a solution of 1a (13 mg) in dry ether (4 ml). The mixture was tirred at room temperature for 1 hr, treated with wet ether and

filtered. The filtrate was concentrated to give R-2,2'-dihydroxymethyl-4,4',5,5',6,6'-hexamethoxybiphenyl (1b) as colorless needles (ether-n-hexane), mp 106—108°, $[\alpha]_{5}^{24}$ —38.2° (c=0.510, CHCl₃). PMR (δ in CCl₄): 3.33 (2H, br s, 2×OH, D₂O exchangeable), 3.60 (6H, s), 3.82 (6H, s), 3.87 (6H, s) (6×OCH₃), 3.98 (4H, s, 2×-CH₂OH), 6.75 (2H, s, arom.-H). MS m/e (%): 394 (M+, 41), 376 (M+-H₂O, 100), 314 (10), 302 (15). CD (c=0.0258, MeOH), $[\theta]^{23}$ (nm): -23000 (216), +13000 (233), -5700 (247).

Bromination of 1 giving 1c—Br₂ (4 drops) was added to a solution of 1 (33 mg) in CCl₄ (30 ml) saturated with H₂O. The reaction mixture was stirred at room temperature for 1 hr in the dark, then treated with 10% Na₂S₂O₃ until no color of Br₂ could be detected, and extracted with CHCl₃. The CHCl₃ extract was washed with H₂O, dried over Na₂SO₄ and concentrated to give a residue, which was purified by PLC [benzeneether (2:1)] to give 1c as an amorphous powder, $[\alpha]_D^{23}$ -64.2° (c=0.483, CHCl₃), IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450 (OH), 1570 (aromatic). MS m/e (%): 592 (C₂₄H₃₀⁸¹Br₂O₇, 40), 588 (C₂₄H₃₀⁷⁹Br₂O₇, 40). PMR spectral data are given in Table I.

Gomisin A(2)—Colorless needles (MeOH), mp 88—89°, $[\alpha]_D^{25}$ +67.9° (c=0.571, CHCl₃), gallic acid-conc-H₂SO₄: greenish-blue. UV $\lambda_{\max}^{\text{EtOH}}$ nm (log ε): 218 (4.88), 253 (4.30), 281 (sh 3.70), 290 (sh 3.57). IR ν_{\max}^{KBr} cm⁻¹: 3500 (OH), 1610, 1590 (aromatic). CD (c=0.0124, MeOH) $[\theta]^{23}$ (nm): -110000 (222), +88000 sh (242), +120000 (253). PMR and mass spectral data are given in Table I and Chart 3, respectively. *Anal.* Calcd. for C₂₃H₂₈O₇: C, 66.33; H, 6.78. Found: C, 66.27; H, 6.83.

Bromination of 2 giving 2a—Gomisin A(2) (43 mg) was treated with Br₂ as described for 1 to give 2a as an amorphous powder, 38 mg, $[\alpha]_D^{23}$ – 57.1° (c=0.42, CHCl₃), IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3550 (OH), 1610, 1570 (aromatic). MS m/e (%): 576 (C₂₃H₂₆⁸¹Br₂O₇,54), 575 (27), 574 (100), 573 (14), 572 (C₂₃H₂₆⁷⁹Br₂O₇, 52), 495 (48), 493 (48) (M⁺-Br), 414 (M⁺-2Br).

Gomisin B(3)—Colorless prisms (ether-*n*-hexane), mp 95—97°, $[\alpha]_D^{23}$ —26.6° (c=0.376, CHCl₃), gallic acid-conc-H₂SO₄: greenish-blue. UV $\lambda_{\max}^{\text{BioH}}$ nm (log ε): 218 (4.98), 257 (sh 4.24), 292 (sh 3.65). IR ν_{\max}^{KBr} cm⁻¹: 3500 (OH), 1715 (C=O), 1640 (C=C), 1610, 1590 (aromatic). CD (c=0.0119, MeOH) [θ]²³ (nm): +46000 (223), —52000 (252). PMR and mass spectral data are given in Table I, and Chart 4 and Table III, respectively. *Anal.* Calcd. for C₂₈H₃₄O₉·1/2C₂H₅CC₂H₅: C, 65.32; H, 7.13. Found: C, 65.41; H, 7.32; C, 65.37; H, 7.22.

Hydrolysis of 3——A solution of 3 (100 mg) in 3% KOH–EtOH (4 ml) was kept at 75° for 8 hr, then diluted with $\rm H_2O$ and extracted with ether (3×15 ml). The combined ethereal extract was washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated. The residue was purified by PLC [benzene–ether (1:1), Rf 0.35] to give 3a as colorless needles (ether–n-hexane), 42 mg, mp 209—210°, [α] $_{\rm b}^{23}$ —91.8° (c=0.305, CHCl $_{\rm 3}$). UV $\lambda_{\rm max}^{\rm Biot}$ nm (log ε): 217 (4.85), 257 (sh 4.12), 284 (sh 3.59). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3500, 3425 (OH), 1615, 1590 (aromatic). CD (c=0.0203, MeOH), [θ] $_{\rm 23}^{23}$ (nm): +74000 (222), -52000 sh (241), -64000 (253). PMR and mass spectral data are given in Table I, and Chart 4 and Table III, respectively. *Anal.* Calcd. for $\rm C_{23}H_{28}O_8$: C, 63.88; H, 6.53. Found: C, 64.05; H, 6.61.

The aqueous solution was acidified with 10% H₂SO₄ and extracted with ether (3×15 ml). The ethereal extracts were combined and washed with H₂O, then dried over Na₂SO₄ and concentrated. The residue was sublimed at 70° (15 mmHg) to give tiglic acid as colorless needles, mp 59— 61° (4.6 mg), which were identical with an authentic sample on direct comparison (IR, mixed mp and PMR).

Oxidation of 3a with Chromium Trioxide in Acetic Acid—CrO₃ (15 mg) was added to a solution of 3a (25 mg) in acetic acid (1.5 ml). The reaction mixture was stirred at room temperature for 5 hr, then diluted with H₂O and extracted with ether (3 × 15 ml). The combined ethereal extract was washed with 5% NaHCO₃, then H₂O, dried over Na₂SO₄ and concentrated. The residue was purified by PLC [*n*-hexane–EtOAc (3: 2), Rf 0.33] to give 3b as a colorless oil, $[\alpha]_{\rm D}^{26} \simeq 0^{\circ}$ (c=0.277, CHCl₃) (6 mg). UV $\lambda_{\rm max}^{\rm EiOH}$ nm (log ε): 218 (sh 4.59), 280 (4.01), 320 (sh 3.57). IR $\nu_{\rm max}^{\rm KBT}$ cm⁻¹: 1710 (C=O), 1685 (CHO), 1610, 1583 (aromatic). PMR (δ in CDCl₃): 0.89 (3H, d, J=7 Hz, CH₃–CH–), 1.83 (3H, s, CH₃–CO), 2.53 (1H, m, –CH), 2.0—2.93 (2H, m, ArCH₂–), 3.68 (3H, s), 3.85 (3H, s), 3.98 (6H, s) (4×OCH₃), 5.97 (2H, s, –OCH₂O–), 6.48 (1H, s, arom.-H), 7.37 (1H, s, arom.-H), 9.57 (1H, s, CHO). MS m/e (%): 430 (M⁺, C₂₃H₂₆O₈, 39), 55 (100).

3b (4.5 mg) was treated with 1% 2,4-dinitrophenylhydrazine in 2 N HCl–EtOH (2:1) to give the di-2,4-dinitrophenylhydrazone as reddish needles (acetone–EtOH), mp 129—132°. PMR (δ in acetone- d_{δ}): 1.13 (3H, d, J=6.5 Hz, CH₃–CH), 1.69 (3H, s, CH₃–C=N–), 3.66, 3.80, 3.94, 4.03 (each 3H, s, 4×OCH₃), 6.02, 6.12 (each 1H, d, J=1 Hz, –OCH₂O–), 6.75 (1H, s, arom.-H), 7.57—8.17 (5H, m, arom.-H), 8.67 (1H, d, J=2.5 Hz, arom.-H), 8.85 (1H, d, J=2.5 Hz, arom.-H). Anal. Calcd. for C₃₅H₃₄N₈O₁₅: C, 53.16; H, 4.33; N, 14.17. Found: C, 53.53; H, 4.34; N, 13.61.

Oxidation of 3a with Chromium Trioxide in Pyridine—CrO₃ (50 mg) was added to a solution of 3a (35 mg) in pyridine (1 ml). The reaction mixture was kept at room temperature overnight, then diluted with $\rm H_2O$ and extracted with ether (3×15 ml). The combined ethereal extract was washed with 5% HCl, then $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated to give 3c as colorless needles (ether-n-hexane), 23 mg, mp 129—130°, [α] $_{\rm D}^{22}$ -88.5° (c=0.26, CHCl $_{\rm 3}$). UV $\lambda_{\rm max}^{\rm EtOH}$ (log ε): 217 (4.56), 249 (sh 4.18), 256 (sh 4.11), 291—295 (sh 3.48). IR $\nu_{\rm max}^{\rm HBr}$ cm⁻¹: 3415 (OH), 1690 (C=O), 1615, 1595 (aromatic). MS m/e (%): 430 (M+, 100), 360 (6). CD (c=0.0123, MeOH) [θ] 23 (nm): +84000 (225), -98000 (252). PMR spectral data are given in Table I. Anal. Calcd. for $\rm C_{23}H_{26}O_8$: C, 64.17; H, 6.09. Found: C, 64.30; H, 6.20.

Gomisin C(4)—Colorless prisms (ether-*n*-hexane), mp 115—116°, $[\alpha]_D^{25}$ —186° (c=0.328, CHCl₃), gallic acid—conc. H₂SO₄: greenish-blue. UV $\lambda_{\max}^{\text{EtoH}}$ nm (log ε): 220 (4.76), 253 (sh 4.19), 284 (sh 3.58). IR

 $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3340 (OH), 1720 (C=O), 1610, 1590, 715 (aromatic). CD (c=0.0135, MeOH) [θ]²³ (nm): +19000 (223), -895000 (242), -60000 sh (255). PMR and mass spectral data are given in Table I, and Chart 4 and Table III, respectively. Anal. Calcd. for $C_{30}H_{32}O_9 \cdot 1/2C_2H_5OC_2H_5$: C, 67.00; H, 6.50. Found: C, 66.98; H, 6.44.

Hydrolysis of 4—A solution of 4 (40 mg) in 3% KOH–EtOH (2 ml) was kept at 75° for 6 hr, then the reaction mixture was treated as described for the hydrolysis of 3 to give a neutral compound and an acid. The former was isolated as colorless needles (from ether–n-hexane), 18 mg, mp 208—209°, $[\alpha]_D^{13}$ –88.5° (c= 0.300, CHCl₃). Anal. Calcd. for C₂₃H₂₈O₈: C, 63.88; H, 6.53. Found: C, 64.03; H, 6.63. This compound was identified with 3a by direct comparison with an authentic sample (IR, mixed mp and PMR). The latter was purified by sublimation at 110° (15 mmHg) to give colorless needles, mp 115—116°, 4.7 mg. This compound was identified with benzoic acid by direct comparison (IR and mixed mp).

Gomisin F(5)—Colorless needles (ether-*n*-hexane), mp 119.5—120.5°, $[\alpha]_D^{23} \simeq 0^\circ$ (c=0.403, CHCl₃). Gallic acid-conc. H₂SO₄: greenish-blue. UV $\lambda_{\max}^{\text{EtoH}}$ nm (log ε): 220 (4.77), 256 (sh 4.13), 288 (sh 3.42). IR ν_{\max}^{KBr} cm⁻¹: 3450 (OH), 1715 (ester), 1640 (C=C). CD (c=0.0150, MeOH), $[\theta]^{23}$ (nm): +41000 (217), -25000 (240). PMR and mass spectral data are given in Table I, and Chart 4 and Table III, respectively. *Anal.* Calcd. for C₂₈H₃₄O₉: C, 65.36; H, 6.66. Found: C, 65.47; H, 6.86.

Gomisin G(6)—Colorless prisms (acetone–ether), mp 97—98°, $[\alpha]_D^{25}$ —126° (c=0.427, CHCl₃). Gallic acid–conc. H₂SO₄: greenish-blue. UV $\lambda_{\max}^{\text{EtoH}}$ nm (log ε): 224 (4.72), 255 (sh 4.08), 280 (sh 3.61), 290 (sh 3.35). IR ν_{\max}^{KBr} cm⁻¹: 3430 (OH), 1715 (ester), 927, 722 (isolated phenyl). CD (c=0.0454, MeOH) [θ]²³ (nm): +38000 (222), —77000 (239), —44000 sh (255), —12000 (285). PMR spectral data are given in Table I. *Anal.* Calcd. for C₃₀H₃₂O₉·1/2C₂H₅OC₂H₅: C, 67.00; H, 6.50. Found: C, 66.89; H, 6.46; C, 66.87; H, 6.48.

Hydrolysis of 5—A solution of 5 (42 mg) in 3% KOH–EtOH was kept at 70° for 17 hr, then diluted with $\rm H_2O$ and extracted with ether (3×15 ml). The combined ethereal extract was washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated to give a residue, which was purified by PLC [benzene-ether (1:1), Rf 0.56] to give 5a as colorless prisms (ether–n-hexane), 22 mg, mp 222.5—224°, [α] $_{\rm bos}^{\rm 15}$ —73.1° (c=0.219, CHCl $_{\rm 3}$). UV $\lambda_{\rm max}^{\rm Hom}$ nm (log ε): 220 (4.76), 253 (sh 4.19), 284 (sh 3.58). IR $\nu_{\rm max}^{\rm KBF}$ cm $^{-1}$: 3560 (OH), 3460 (OH), 1625, 1597 (aromatic). PMR spectral data are given in Table I. Anal. Calcd. for $\rm C_{23}H_{28}O_8$: C, 63.88; H, 6.53. Found: C, 64.03; H, 6.58. The aqueous solution was acidified with 5% $\rm H_2SO_4$ and extracted with ether (2×15 ml). The combined ethereal extract was washed with $\rm H_2O$, dried over $\rm Na_2SO_4$ and concentrated. The residue was sublimed at 70° (15 mmHg) to give tiglic acid as colorless prisms, 2 mg, mp 58—60°, which were identical with an authentic sample on direct comparison (IR and mixed mp).

Hydrolysis of 6—A solution of 6 (60 mg) in 3% KOH–EtOH was kept at 70° for 6 hr, then the reaction mixture was treated as described for the hydrolysis of 5 to give a neutral compound and an acid. The former was isolated as colorless needles (ether), 30 mg, mp $223.5-225^{\circ}$, $[\alpha]_{D}^{25}$ -72.1° (c=0.375, CHCl₃), and was identical with 5a on direct comparison with an authentic sample (IR, mixed mp and PMR). The latter was purified by sublimation at 110° (15 mmHg) to give colorless plates, 5 mg, mp $122-123^{\circ}$. This compound was identified as benzoic acid by direct comparison (IR and mixed mp).

Oxidation of 5a with Chromium Trioxide——CrO₃ (50 mg) was added to a solution of 5a (11.5 mg) in pyridine (0.5 ml). The reaction mixture was stirred at room temperature for 2 hr, then diluted with H₂O and extracted with ether (3 × 15 ml). The combined ethereal extract was washed with 1 n HCl, then with H₂O, dried over Na₂SO₄ and concentrated. The residue was purified by PLC [benzene–ether (1: 1), Rf 0.46] to give 5b as colorless needles (ether), mp 227.5—229.5°, [α]²³ —93.6° (c=0.235, CHCl₃), 10 mg. UV λ ^{Enormousless} nm (log ε): 218 (4.56), 247 (sh 4.23). IR ν ^{RBr}_{max} cm⁻¹: 3460 (OH), 1695 (C=O). MS m/e (%): 430 (M+, 100), 412 (M+-H₂O, 6.3), 358 (M+-CH₃-CH=C(OH)CH₃, 39). PMR spectral data are given in Table I. Anal. Calcd. for C₂₃H₂₆O₃: C, 64.17; H, 6.09. Found: C, 64.19; H, 6.11.

Acknowledgement The authors wish to express their gratitude to Prof. K. Hata, Osaka College of Pharmacy, for providing angelic acid. We are grateful to Prof. H. Ageta and Dr. R. Kamaya, Showa College of Pharmacy, for measurements of CD spectra, and also to Mr. Y. Shida, Tokyo College of Pharmacy, and to Dr. M. Uchida, Shizuoka College of Pharmacy, for measurements of mass spectra.

Thanks are also due to Prof. S. Shibata, Meiji College of Pharmacy, to Prof. U. Sankawa, Faculty of Pharmaceutical Sciences, University of Tokyo, and to Dr. K. Kuriyama, Shionogi Research Laboratory, for valuable discussions.