

Isolation of 23-Methylcholesta-5,22-dien-3 β -ol from the Soft Coral *Sarcophyton glaucum*

A new marine C₂₈ sterol was isolated from the soft coral, *Sarcophyton glaucum*, and its structure was proposed as (22*E*)-23-methylcholesta-5,22-dien-3 β -ol on spectral evidences, especially by the comparison of its ¹³C-NMR chemical shifts with those of 23,24-dimethylcholesta-5,22-dien-3 β -ol, which is present in the same soft coral.

Keywords—23-methylcholesta-5,22-dien-3 β -ol; soft coral; *Sarcophyton glaucum*; dinoflagellate; marine sterol; ¹³C-NMR

Previously we reported the occurrence of a minor unidentified sterol (I) which shows the slightly longer retention time than cholesterol on gas chromatography (GC), along with the biogenetically interesting 23,24-dimethylcholesta-5,22-dien-3 β -ol (II) and 23,24-dimethylcholesta-5,17(20)-dien-3 β -ol (III), in the sterol mixture of the soft coral *Sarcophyton glaucum*, collected at Ishigaki Island.¹⁾ The major components were (24*S*)-24-methylcholesterol and gorgosterol, and compound I represented less than 0.5% of total sterols. Repeated column chromatography of the sterol mixture (30 g) over silver nitrate-impregnated silica gel (benzene-hexane) and partition chromatography using Lipidex 5000 (hexane-methanol)²⁾ afforded 11 mg of a gas chromatographically pure specimen as needles from methanol.

Compound I showed mp 140.5—142.5°, [α]_D -39.2° (*c*=0.98, CHCl₃). The mass spectrum of I showed a molecular ion (M⁺) at *m/e* 398 and other ions at 380 (M⁺-H₂O), 383 (M⁺-CH₃), 365 (M⁺-CH₃ and H₂O), and 355 (M⁺-C₃H₇) indicating that it is a diunsaturated C₂₈ sterol. A series of fragments, *m/e* 271 (M⁺-side chain and 2H), 255 (M⁺-side chain and H₂O) and 213 (M⁺-side chain, H₂O, and ring D cleavage) are those normally observed with sterols having an unsaturated nucleus and unsaturated side chain.³⁾ Furthermore, an intense peak at *m/e* 125 (C₉H₁₇) seemed to match the entire side chain, excluding the possibility of an extra methyl group's presence on the nucleus. The proton magnetic resonance (PMR) signals for 18-Me (δ 0.71), 19-Me (1.01), hydroxymethine (3.3—3.76, 1H, m) and olefinic proton (5.36, 1H, m) suggested that I belongs to the conventional 3 β -hydroxy- Δ^5 sterols⁴⁾ as was the case with all of the other ten sterol components in the same organism.¹⁾ Assuming that I bears a methylated cholestane-type side chain, three doublets at δ 0.93 (3H, *J*=6.5 Hz), 0.83 (3H, *J*=6 Hz), and at 0.81 (3H, *J*=6 Hz) are assignable to 21-, 26- and 27-methyl signals respectively. Also the olefinic signals at δ 1.56 (3H, d, *J*=0.5 Hz) and 4.88 (1H, broad d, *J*=10 Hz) indicate the presence of a trisubstituted double bond bearing one methyl group in the side chain. The appearance of the doublet at δ 4.88 confined the location of the double bond to a position adjacent to a tertiary carbon (*i.e.* Δ^{22} or Δ^{23}), and thus the location of the olefinic methyl group is set at C-23. An intense ion at *m/e* 300 (cleavage at C-20 and C-22 with 1H transfer) in the mass spectrum is characteristic of Δ^{22} -sterols with a nuclear double bond while that of Δ^{23} -sterols is *m/e* 301.^{3b)} From these results the most plausible structure of I would be 23-methylcholesta-5,22-dien-3 β -ol. The previously determined homologous sterol (II) showed a similar PMR spectrum and mass spectral fragmentation pattern.¹⁾

- 1) M. Kobayashi, A. Tomioka, and H. Mitsuhashi, Steroids, in press. The GC was carried out using 1.5% OV-17 on 80—100 mesh Shimalite W at 260°. The relative retention time (*Rt_r*) of I to cholesterol was 1.06.
- 2) P.M. Hyde and W.H. Elliott, *J. Chromatogr.*, **67**, 170 (1972).
- 3) a) B.A. Knights, *J. Gas Chromatogr.*, **5**, 273 (1967); b) S.G. Wyllie and C. Djerassi, *J. Org. Chem.*, **33**, 305 (1968).
- 4) R.F. Zürcher, *Helv. Chim. Acta*, **40**, 2054 (1963).

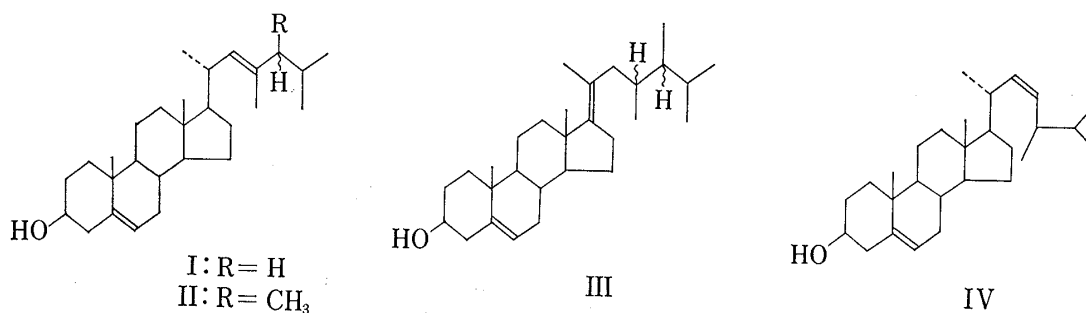


Fig. 1

The ¹³C-NMR spectrum of I and its comparison with that of II (Table I) provided strong evidence to support the structure of I. The both compounds showed almost the same chemical shifts for carbons 1 to 22. The difference is that compound I showed the signal of C-25 higher by 5.9 ppm and those of C-26 and C-27 slightly lower than the corresponding signals in II. These discrepancies can be well explained by the β- and γ-effects of the 24-methyl group.⁵⁾ The geometry of Δ²² double bond of I and II was deduced as *E* from the chemical shifts of C-24 and olefinic methyl. The chemical shifts of C-24 (I, 49.6 ppm; II, 50.1 ppm) are reasonably close to the calculated value when the β-methyl substituent effect (+8 ppm)⁶⁾ was taken into account on those of (22*E*)-cholesta-5,22-dien-3β-ol (42.0 ppm) and (22*E*)-24-methylcholesta-5,22-dien-3β-ol (43.1 ppm).⁷⁾ The chemical shifts of olefinic methyls (I, 16.2 ppm; II, 16.9 ppm) show the influence of the shielding effect by vicinal *Z* carbon (C-20) which is similar to that of (*E*)-polyisoprene (16.0 ppm) in contrast to (*Z*)-polyisoprene (23.6 ppm).⁸⁾ The geometry of the similar 23,24-dimethyl system in dinosterol, a 4α-methyl-5α,6-dihydro analog of II, from the toxic dinoflagellate *Gonyaulax tamarensis* by Shimizu *et al.* was also shown to have the *E* geometry by X-ray crystallography.⁹⁾

The proposed structure, (22*E*)-23-methylcholesta-5,22-dien-3β-ol, represents the second member of a class of 23-methylcholestane-type sterols found in marine organisms. The first example, 4α,23-dimethyl-5α-cholest-22-en-3β-ol, had been found in the cultured dinoflagellate, *Gonyaulax diagenesis* by Alam, *et al.*¹⁰⁾ Compound I may also be derived from the symbiotic dinoflagellate of *S. glaucum*.

Interestingly, I shows the spectroscopic and chromatographic data very similar to those reported for a C₂₈ sterol from a clam *Tapes philippinarum* by Teshima *et al.*¹¹⁾ Although we were unable to do the direct comparison, the published PMR and mass spectra are almost identical with those of I except for a minor difference. The sterol from the clam was assigned a structure of (22*Z*)-24-methylcholesta-5,22-dien-3β-ol (IV) on the basis of PMR (δ 4.79—4.88 and 4.15—4.22) and very weak infrared absorption (680 cm⁻¹). One of the signals (δ 4.15—4.22) attributed to the olefinic proton, however, is evidently too high for such an isolated *Z*-Δ²² disubstituted double bond which usually occurs at δ 4.9—5.1.¹²⁾

- 5) L.P. Lindeman and J.Q. Adam, *Anal. Chem.*, **43**, 1245 (1971).
- 6) F.W. Wehrli and T. Wirthlin, "Interpretation of Carbon-13 NMR Spectra," Heyden and Sons Ltd., London, Philadelphia, Rheine, 1978, p. 37.
- 7) J.L.C. Wright, A.G. McInnes, S. Shimizu, D.G. Smight, J.A. Walter, D.R. Idler, and W. Khalil, *Can. J. Chem.*, **56**, 1898 (1978).
- 8) M.W. Duch and D.M. Grant, *Macromolecules*, **3**, 165 (1970).
- 9) Y. Shimizu, M. Alam, and A. Kobayashi, *J. Am. Chem. Soc.*, **98**, 1059 (1976); J. Finer, J. Clardy, Y. Shimizu, A. Kobayashi, and M. Alam, *J. Org. Chem.*, **43**, 1990 (1978).
- 10) M. Alam, K.H. Schram, and S.M. Ray, *Tetrahedron Lett.*, **1978**, 3517.
- 11) S. Teshima, A. Kanazawa, and T. Ando, *Comp. Biochem. Physiol.* **47B**, 507 (1974). The reported *R_t* of IV to cholesterol is 1.05 on the similar condition with ref. 1.
- 12) A. Metayer and M. Barbier, *Chem. Commun.*, **424** (1973); M. Kobayashi, K. Todo, and H. Mitsuhashi, *Chem. Pharm. Bull.* (Tokyo), **22**, 236 (1974).

TABLE I. ^{13}C -NMR Chemical Shifts in Compounds I and II (in ppm)

Carbons	I	II	Carbons	I	II
1	37.2	37.7	15	24.3	24.3
2	31.9	31.9	16	28.2	27.8
3	71.7	71.7	17 ^{a)}	56.7	56.7
4	42.2	42.2	18	12.1	12.1
5	140.4	140.4	19	19.4	19.4
6	121.4	121.4	20	34.9	34.5
7	31.9	31.6	21	20.7	20.6
8	31.6	30.7	22	132.9	131.4
9	50.1	50.1	23	130.1	134.8
10	36.5	36.4	24	49.6	50.1
11	21.1	21.0	25	26.0	31.9
12	39.7	39.7	26 ^{b)}	22.3	20.0
13	42.1	42.1	27 ^{b)}	22.6	21.7
14 ^{a)}	56.6	56.7	28	—	13.2
			29 ^{c)}	16.2	16.9

a, b) Chemical shifts may be interchanged.

c) Methyl at C-23.

*Faculty of Pharmaceutical Sciences,
Hokkaido University
Sapporo 060, Japan*

MASARU KOBAYASHI
ATSUSHI TOMIOKA
TAKAAKI HAYASHI
HIROSHI MITSUHASHI

Received June 21, 1979