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### Carbon-13 Nuclear Magnetic Resonance Spectral Assignments of Grayanotoxin-I

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The <sup>13</sup>C-NMR signals of grayanotoxin-I, a tetracyclic diterpene with an A-nor-B-homo-*ent*-kaurane skeleton, were assigned by means of single-frequency off-resonance decoupling, selective proton decoupling and by comparison with spectra of derivatives.

**Keywords**—<sup>13</sup>C-NMR spectral assignments; grayanotoxins; *ent*-kaurane skeleton; lyoniol; *Leucothoe grayana*

Grayanotoxins, isolated from *Leucothoe grayana* Max., are tetracyclic diterpenoids with an A-nor-B-homo-*ent*-kaurane skeleton. As most of them have physiological and pharmaceutical activities,<sup>2)</sup> various workers have investigated their structure-activity relationships.<sup>3)</sup>

In previous papers,<sup>4)</sup> we reported the first isolation of two grayanotoxin glycosides. The <sup>13</sup>C-nuclear magnetic resonance (NMR) spectra of grayanotoxins are useful in determining the structures of the glycosides, and this report describes the assignment of the <sup>13</sup>C-NMR signals of all the carbons in grayanotoxin-I.

### Experimental

Grayanotoxin-I (4),<sup>5)</sup> -II (5),<sup>6)</sup> -III (2),<sup>5)</sup> -IV (6),<sup>7)</sup> and -XVIII (10)<sup>4)</sup> were isolated from *Leucothoe grayana* and lyoniol-B (12)<sup>8)</sup> from *Lyonia ovalifolia* var. *elliptica*. Other compounds were prepared from the natural

- 1) Location: a) *Tanabe-dori, Mizuho-ku, Nagoya 467, Japan*; b) *Hirosawa, Idogaya-cho, Kariya, Aichi 448, Japan*.
- 2) T. Narahashi and I. Seyama, *J. Physiol.*, **242**, 471 (1974); D.D. Ku, T. Akera, M. Frank, T.M. Brody, and J. Iwasa, *J. Pharmacol. Exp. Ther.*, **200**, 363 (1977); Y. Hotta, K. Takeya, J. Sakakibara, and N. Shirai, *J. Aichi Med. Univ. Ass.*, **6**, 8 (1978).
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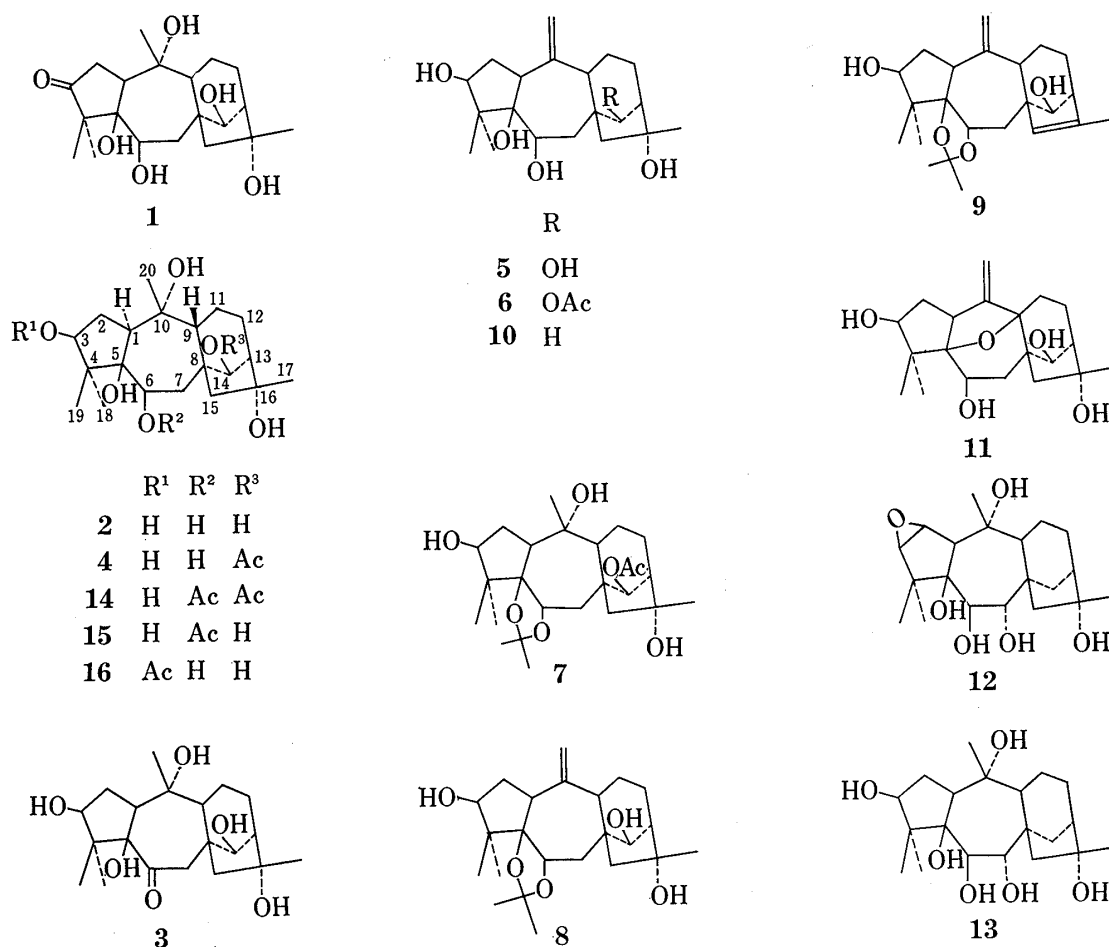


Fig. 1

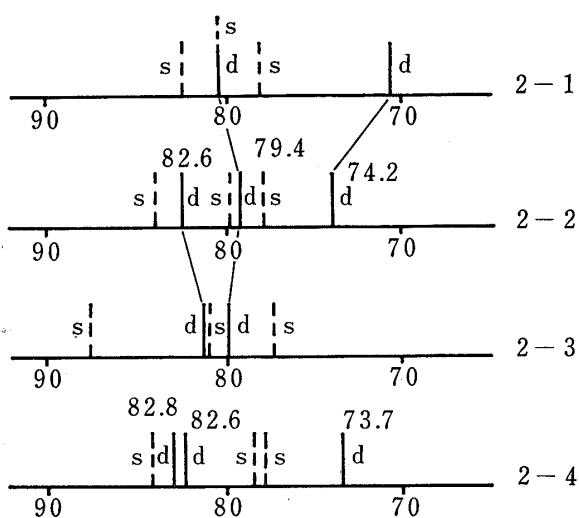


Fig. 2

compounds. Their structures were confirmed by  $^1\text{H}$ -NMR, infrared (IR), mass spectrum (MS), mp and/or elemental analysis.  $^{13}\text{C}$ -NMR spectra were measured using a 10 mm glass tube on a JEOL FX-60 spectrometer (15 MHz) with Fourier transform techniques, controlled by a JEC-98B 16K computer. The sample (about 100 mg) was dissolved in  $d_5$ -pyridine (about 1 ml) with  $\text{Me}_4\text{Si}$  as an internal reference.

### Results and Discussion

The structures of grayanotoxins to be discussed in this paper are shown in Fig. 1. The  $^{13}\text{C}$ -NMR signals were divided into 11 groups, as shown in Table I, based on the information obtained from chemical shifts and coupling patterns in off-resonance experiments. Abbreviations are s, singlet; d, doublet; t, triplet; and q, quartet. The sign 2—3 refers to the NMR spectrum of compound 3 in Fig. 2, and so on.

TABLE I.  $^{13}\text{C}$ -NMR Data for Grayanotoxins and Related Compounds

Compounds	1	2	3	4	4'	5	6	7	8	9	10	11	12	13	
-CH <sub>3</sub>	18.1	19.8	17.9	19.8	19.0	18.7	18.5	20.7	20.3	15.3	19.2	19.2	20.6	20.2	
	22.3	23.3	23.9	21.2	19.0	23.8	21.1	21.4	23.5	20.2	24.1	23.3	21.3	23.3	
	24.0	23.9	26.9	23.4	22.7	24.6	23.8	24.2	27.0	27.1	25.4	24.2	23.9	23.9	
	27.9	28.3	26.9	24.0	23.9		25.6	25.9	27.4	27.2			30.9	28.5	
				28.3	27.4				25.9	27.8	27.7				
									27.2						
								27.7							
-CH <sub>2</sub> -	22.3	22.6	20.7	22.5	21.2	24.6	23.8	20.7	22.1	21.8	24.1	23.3	23.1	23.3	
	27.0	27.1	26.9	27.3	26.2	24.8	24.8	27.2	24.2	23.3	25.8	29.5	26.9	27.1	
	38.8	35.8	36.0	35.8	34.7	39.4	39.6	35.3	35.8	31.1	36.5	37.3	29.3	29.5	
	44.3	44.4	50.9#	44.1	42.5	42.2	40.8	37.5	38.3	38.2	39.4	42.6	52.8	35.8	
	61.0	60.4	59.7	61.1	59.8	60.7	61.4	63.6	61.5		46.7	50.0		53.4	
											62.5				
>CH-	47.6	51.7	51.8#	51.4	49.5	45.9	45.3	49.5	44.2	38.6	44.5	51.2	50.7	50.8	
	54.3	55.2	51.8#	55.0	53.5	50.8	51.7	50.3	45.8	45.8	47.9	52.6	51.4	52 <sup>a)</sup>	
	55.7	56.4	54.3	55.6	54.5	54.3	52.8	53.1	55.0	54.7	52.3		54.5	52 <sup>a)</sup>	
-C-	52.2	51.8	50.4	51.0	49.5	50.6	49.0	48.6	48.1	48.0	44.7	48.5	48.1	52 <sup>a)</sup>	
	57.1	52.6	53.7	51.8	50.4	50.6	50.3	50.9	51.3	54.5	50.6	50.8	51.7	52 <sup>a)</sup>	
$\begin{array}{c} \text{H} \quad \text{H} \\   \quad   \\ -\text{C} \quad \text{C}- \\ \diagdown \quad / \\ \text{O} \end{array}$														60.0	
															64.4
-CH-O-	70.8	74.2	79.9	73.7	72.3	69.5	69.3	76.1	77.1	77.3	70.6	67.9	76.0	76.8	
	80.4	79.4	81.2	82.6	80.8	81.0	80.8	79.0	78.3	78.4	81.2	82.1	79.1	80.3	
		82.6		82.8	81.0	81.2	84.0	86.2	82.8	84.2		85.3		82.8	
-C-O-	78.0	78.1	77.5	77.9	76.5	81.2	79.5	77.3	81.0	93.5	79.5	81.2	77.1	77.8	
	80.4	79.7	81.0	78.6	77.0	83.9	83.4	80.4	93.5	107.3	83.5	89.1	78.0	78.0	
	82.5	84.6	87.6	84.4	83.0			93.5	107.4			95.6	79.0	83.7	
									108.0						
=CH <sub>2</sub>						111.9	112.5		111.2	111.5	112.2	107.4			
=CH-										133.5					
=C<						153.0	151.8		149.9	137.4	153.0	161.8			
										149.5					
>C=O	221.3		216.1	170.2	169.7		170.3	170.6							

Proton off-resonance spectra were obtained for all compounds.

*d*<sub>5</sub>-Pyridine solvent, except in the case of 4' (*d*<sub>6</sub>-DMSO).

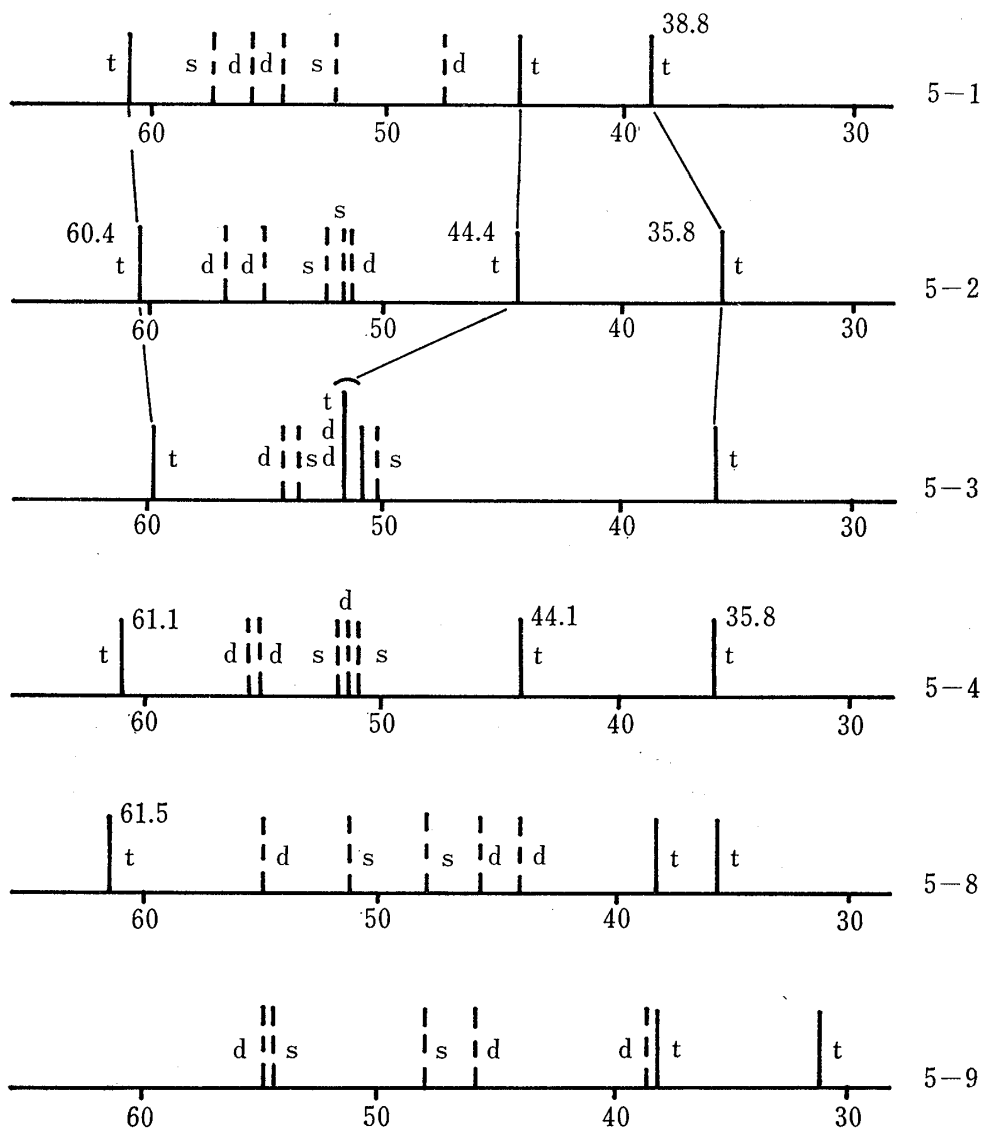
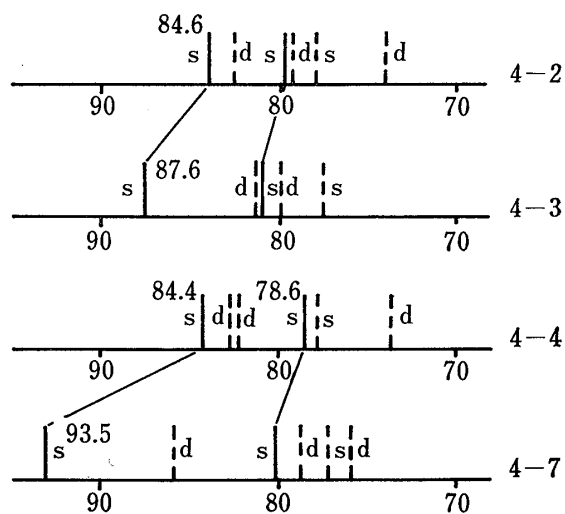
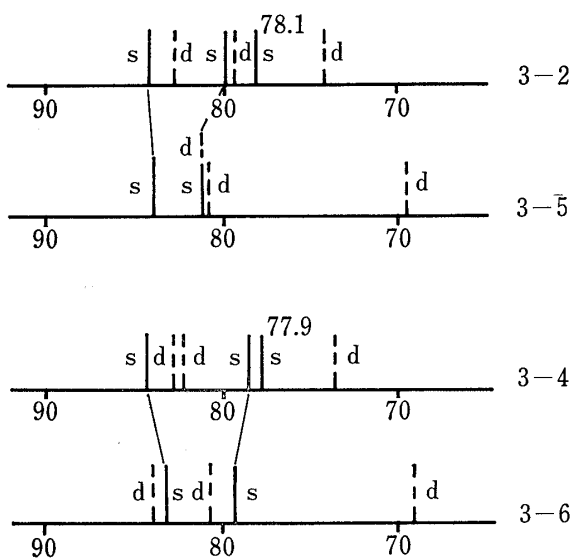
Owing to the complex proton off-resonance spectrum, signals marked # may be reversed.

a) These signals appear in the region of 51.9—52.2 ppm.

$^{13}\text{C}$ -NMR spectra of 14, 15 and 16 were not measured.

## A. Signals in the Region of 65—90 ppm

1. **Secondary Carbinyl Carbons (C-3, C-6 and C-14: Fig. 2)**—The signals arising from C-3 and C-6 can readily be assigned by comparison with the corresponding spectra of keto-derivatives as follows: a signal at 82.6 ppm(d) in 2—2 disappeared in 2—1 (3-keto derivative), so it was assigned to C-3 of 2. A signal at 74.2 ppm(d) in 2—2 was assigned to C-6 of 2 because of the absence of one doublet signal in 2—3 (6-keto derivative). The remaining signal at 79.4 ppm(d) was assigned to C-14 of 2. These assignments were confirmed by selective proton decoupling techniques with 2. Namely, a doublet at 82.6 ppm in the  $^{13}\text{C}$ -NMR of 2 changed to a singlet on irradiating a signal at 3.84 ppm (C-3-H, br-s,  $W_{1/2}=6$  Hz) in the  $^1\text{H}$ -NMR of 2 and a doublet at 79.4 ppm in the  $^{13}\text{C}$ -NMR of 2 changed to a singlet on irradiating a signal at 4.95 ppm (C-14-H, s) in the  $^1\text{H}$ -NMR of 2. A doublet at 82.8 ppm in the  $^{13}\text{C}$ -NMR of 4 changed to a singlet on irradiating a signal at 6.22 ppm (C-14-H, s) in the  $^1\text{H}$ -NMR of 4. Summarizing these assignments in 2—4, peaks at 73.7, 82.6 and 82.8 ppm were assigned to C-6, -3 and -14 of 4, respectively.



2. **Tertiary Carbinyl Carbons (C-5, C-10 and C-16: Fig. 3 and 4)**—Singlet signals at the highest field of 3—2 and 3—4 (78.1 and 77.9 ppm) were assigned to C-10, since these signals were absent in 3—5 and 3—6 ( $\Delta$  10(20) compounds).

It is well known that the  $\alpha$ -carbon of a carbonyl group is more deshielded than the  $\beta$ -carbon of a carbinyl carbon,<sup>9)</sup> and a signal at 84.6 ppm(s) in 4—2 shifted 3.0 ppm downfield in 4—3 (6-keto derivative). The signal at 84.6 ppm was assigned to C-5 of 2. A signal at 84.4 ppm(s) in 4—4 was assigned to C-5 of 4 in a similar way. The signal at 84.4 ppm in 4—4 showed a downfield shift to 93.5 ppm(s) in 4—7 (5,6-O-isopropylidene derivative). The remaining singlet at 78.6 ppm(s) in 4—4 was assigned to C-16 of 4.

## B. Signals in the Region of 30—65 ppm

1. **Methylene Carbons (C-2, C-7 and C-15: Fig. 5)**—The signal at 38.8 ppm in 5—1 is assigned to C-2, since the signal was absent in the spectrum of a deuterium-labelled derivative,<sup>10)</sup> 2,2- $d_2$ -3-keto-grayanotoxin-III. Only one triplet at 35.8 ppm in 5—2 moved downfield to 38.8 ppm(t) in 5—1 (3-keto derivative). Therefore, the signal at 35.8 ppm was assigned to C-2 of 2. A signal at 60.4 ppm(t) in 5—2 remained essentially unchanged in 5—3, whereas the triplet at 44.4 ppm in 5—2 showed a downfield shift to 51—52 ppm in 5—3 (6-keto derivative). Consequently, the latter triplet was assigned to C-7 of 2. In 5—4, a signal at 35.8 ppm was assigned to C-2 and a signal at 44.1 ppm to C-7.

In order to assign the signal of C-15, the acetonide and dehydrated derivatives (8 and 9) were prepared. In comparison with 5—8 and 5—9, a signal at 61.5 ppm(t) in 5—8 was absent in 5—9, so this signal was assigned to C-15 of 8. Therefore, a signal at 61.1 ppm(t) in 5—4 was assigned to C-15 of 4. In the spectra of 1—8 and 10, the triplet signals of C-15 were observed in the region of 58—63 ppm. Carbon-15 is not a carbinyl carbon but carbinyl carbon chemical shifts generally appear in this region (for example, 61.7 ppm in 1-butanol<sup>11)</sup>). On the other hand, the triplet signals of 11—13 were seen between 50 and 54 ppm. The triplet signal of 4 measured in  $d_6$ -DMSO shifted only slightly upfield to 59.8 ppm. This shows that the remarkable chemical shift of C-15 is independent of the solvent. The reason for this phenomenon is not clear.

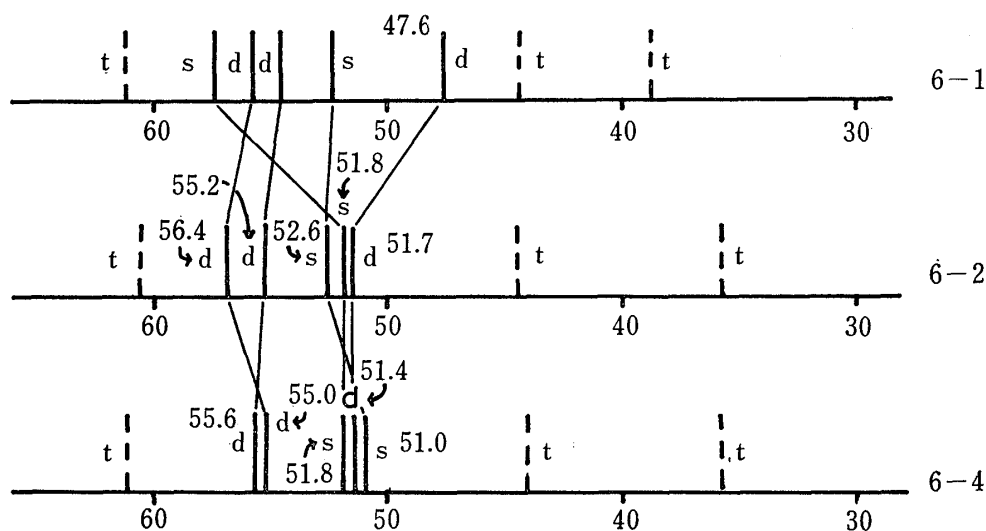


Fig. 6

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 10) H.J. Reich, M. Jautelat, M.T. Messe, F.J. Weigert, and J.D. Roberts, *J. Am. Chem. Soc.*, **91**, 7445 (1969).  
 11) J.B. Stothers, "Carbon-13 NMR Spectroscopy," Academic Press, New York, 1972, p. 140.

**2. Tertiary Carbons (C-1, C-9 and C-13: Fig. 6)**—A signal at 51.7 ppm(d) in 6—2 which moved upfield to 47.6 ppm(d) in 6—1 (3-keto derivative) was assigned to C-1 of 2, because C-1 is in the  $\gamma$  position relative to C-3.

When comparing 6—2 with 6—4 (14-acetate of 2), slight shifts of one singlet and one doublet (52.6— $\rightarrow$ 51.0 and 56.4— $\rightarrow$ 55.0) were observed. Since this was due to acetylation of the 14-hydroxyl group, the singlet and doublet peaks were assigned to C-8 and C-13, which are  $\beta$  to C-14. The remaining doublet at 55.2 ppm in 6—2 and 55.6 ppm in 6—4 were assigned to C-9. However, the assignments of C-9 and C-13 may be reversed.

**3. Quaternary Carbons (C-4 and C-8: Fig. 6)**—In the above discussion, the signal at 51.0 ppm in 6—4 was assigned to C-8 of 4. The remaining singlet at 51.8 ppm in 6—4 was assigned to C-4 of 4.

### C. Signals in the Region of 15—30 ppm

**1. Methyl Signals (C-17, C-18, C-19 and C-20: Fig. 7)**—Initially, we examined the  $^1\text{H-NMR}$  spectra. As shown in Fig. 7, a methyl signal which appears about 1.8 ppm in 7—2, 7—4, 7—14, 7—15 and 7—16 is absent in 7—6 ( $\Delta 10(20)$  compound). Therefore, the signal at 1.84 ppm in 7—4 was assigned to the 20-methyl group of 4.

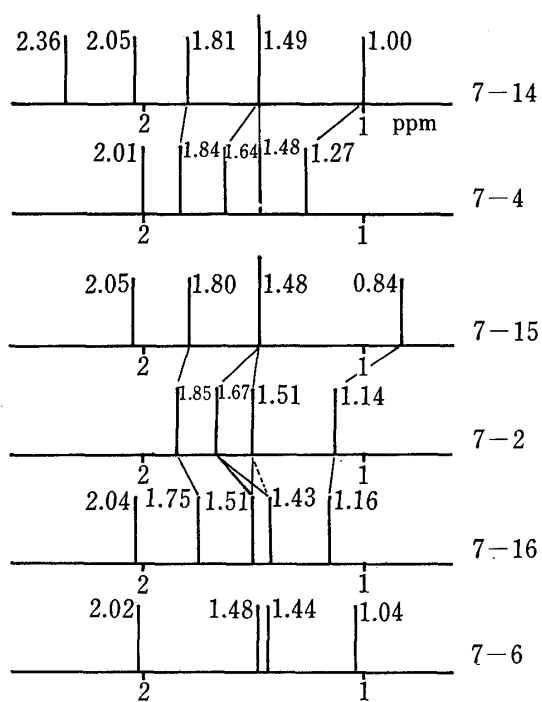


Fig. 7.  $^1\text{H-NMR}(\text{C}_5\text{D}_5\text{N})$

Kawazoe *et al.*<sup>12)</sup> reported that methyl proton resonances can shift to lower fields due to steric interaction with a neighboring hydroxyl group, and that such a methyl group can be identified by acetylation of the hydroxyl group; the resonance then appears at markedly higher field than that of the original hydroxyl compound. X-ray crystallographic analysis of 4<sup>13)</sup> and examination of its Dreiding model showed that the 18- and 19-methyl groups are close to the 6-hydroxyl group. Two signals at 1.27 and 1.64 ppm in 7—4 move to 1.00 and 1.49 ppm in 7—14 (6-acetate of 4). Comparing 7—2 with 7—15 (6-acetate of 2), similar shifts were observed for the two methyl groups. Therefore, these signals were assigned to the 18- and 19-methyl groups. Comparing 7—2 with 7—16 (3-acetate of 2), a signal at 1.14 ppm in 7—2 scarcely moved in 7—16, but a signal at 1.67 ppm in 7—2 shifted to 1.43 or 1.51 ppm in 7—16. Therefore, this signal was assigned to the 19-methyl group, because it is nearer to the 3-hydroxyl group than the 18-methyl is.

Consequently, five methyl proton signals, 1.27, 1.48, 1.64, 1.84 and 2.01 ppm, in the  $^1\text{H-NMR}$  spectrum of 4 were assigned to the 18-, 17-, 19-, 20- and acetyl methyl groups, respectively.

Using the single frequency selective decoupling technique, five methyl carbon signals in the  $^{13}\text{C-NMR}$  spectrum of 4 were then assigned. The signals at 19.8, 21.2, 23.4, 24.0 and 28.3 ppm were assigned to C-19, acetyl methyl, C-18, C-17 and C-20, respectively.

12) Y. Kawazoe, Y. Sato, T. Okamoto, and K. Tsuda, *Chem. Pharm. Bull.*, **11**, 328 (1963); T. Okamoto and Y. Kawazoe, *ibid.*, **11**, 643 (1963).

13) P. Narayanan, M. Rohrl, K. Zechmeister, and W. Hoppe, *Tetrahedron Lett.*, **1970**, 3943.

**2. Methylene Carbons (C-11 and C-12)**—Two signals at 22.5 and 27.3 ppm in the spectrum of **4** corresponded to C-11 and C-12, but an unambiguous assignment could not be made.

### Conclusion

The <sup>13</sup>-NMR spectral assignments of grayanotoxin-I (**4**) are as follows—C-1, 51.4 ppm; C-2, 35.8 ppm; C-3, 82.6 ppm; C-4, 51.8 ppm; C-5, 84.4 ppm; C-6, 73.7 ppm; C-7, 44.1 ppm; C-8, 51.0 ppm; C-9, 55.6 ppm; C-10, 77.9 ppm; C-11 and C-12, 22.5 and 27.3 ppm; C-13, 55.0 ppm; C-14, 82.8 ppm; C-15, 61.1 ppm; C-16, 78.6 ppm; C-17, 24.0 ppm; C-18, 23.4 ppm; C-19, 19.8 ppm; C-20, 28.3 ppm; acetyl methyl, 21.2 ppm; and acetyl carbonyl, 170.2 ppm.

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