

## Communications to the Editor

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## RUGOSIN D, E, F AND G, DIMERIC AND TRIMERIC HYDROLYZABLE TANNINS

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Four new hydrolyzable tannins, rugosin D (**1**), E (**2**), F (**3**) and G (**4**), were isolated from *Rosa rugosa*, and their dimeric and trimeric structures, linked *via* a valoneoyl group, were elucidated.

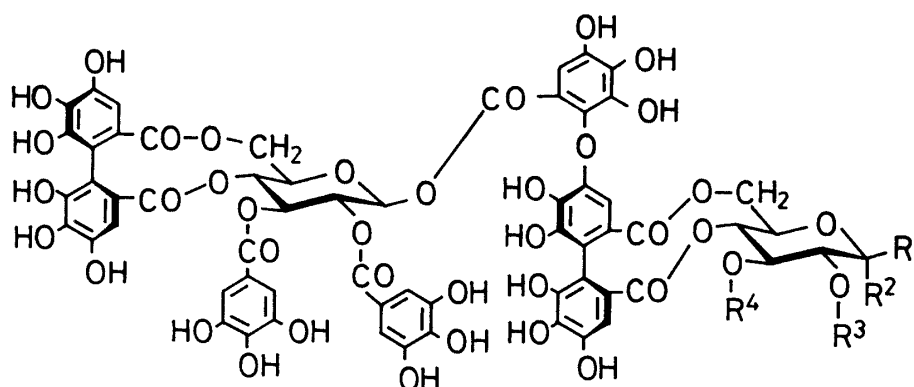
KEYWORDS — *Rosa rugosa*; Rosaceae; tannin; dimeric hydrolyzable tannin; trimeric hydrolyzable tannin; valoneoyl group; rugosin D; rugosin E; rugosin F; rugosin G

Recently, some dimeric hydrolyzable tannins have been found.<sup>1,2)</sup> We now report the isolation and the structures of dimeric tannins of a new type, rugosin D (**1**), E (**2**) and F (**3**), having two monomeric units linked through a valoneoyl group. We report likewise on rugosin G (**4**), the first example of a trimeric hydrolyzable tannin.

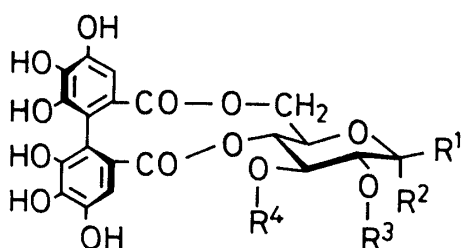
These four tannins, **1**~**4**, have been isolated from the ethyl acetate extract obtained from the crude extract of the flower petal of *Rosa rugosa* Thunb. by column chromatography on Sephadex LH-20, collecting the fractions eluted after monomeric tannins.<sup>3)</sup>

Rugosin D (**1**),  $C_{82}H_{58}O_{52} \cdot 9H_2O$ ,  $[\alpha]_D +118^\circ$  ( $c=1$ , acetone), UV  $\lambda_{max}^{MeOH}$  nm ( $\log \epsilon$ ) 219 (5.17) and 277 (4.84), was obtained as an off-white amorphous powder. Its  $^1H$ -NMR spectrum (200 MHz, in acetone- $d_6$ ) shows fifteen aromatic protons [ $\delta$ 7.149, 7.032, 7.025, 7.02, 6.99 (s, 2H each, galloyl), 7.146, 6.67, 6.49, 6.47 and 6.26 (s, 1H each, hexahydroxydiphenoyl and valoneoyl)] and fourteen glucose protons [ $\delta$ 6.20 (d,  $J=8$  Hz, H-1), 6.15 (d,  $J=8$  Hz, H-1), 5.85 (t,  $J=10$  Hz, H-3), 5.81 (t,  $J=10$  Hz, H-3), 5.61 (dd,  $J=8, 10$  Hz, H-2), 5.54 (dd,  $J=8, 10$  Hz, H-2), 5.33 (dd,  $J=6, 14$  Hz, 2H, H-6), 5.16 (t,  $J=10$  Hz, 2H, H-4), 4.54 (dd,  $J=6, 10$  Hz, H-5), 4.48 (dd,  $J=6, 10$  Hz, H-5), 3.83 (d,  $J=14$  Hz, H-6') and 3.79 (d,  $J=14$  Hz, H-6')], which indicate that **1** possesses five galloyl, a hexahydroxydiphenoyl and a valoneoyl group, and two glucose cores. The formation of *C1* conformation and  $\beta$ -anomer of the two glucose cores is shown by the coupling constants of the sugar protons.

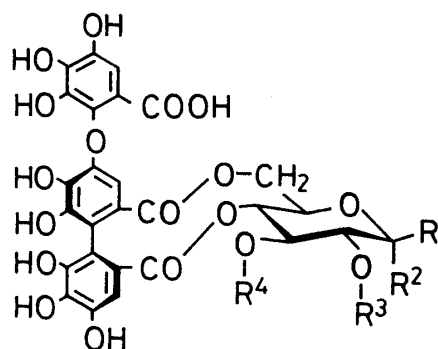
Methylation of **1** with dimethyl sulfate and potassium carbonate in dry acetone afforded nonacosa-*O*-methylrugosin D,  $C_{111}H_{116}O_{52} \cdot H_2O$ ,  $[\alpha]_D +52^\circ$  ( $c=1$ , acetone) [mass spectrum:  $m/z$  646 and 614 (methylated valoneoyl); 422, 404 and 360 (hexamethoxydiphenoyl); 212, 197 and 195 (trimethylgalloyl)].



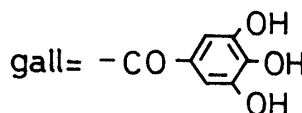
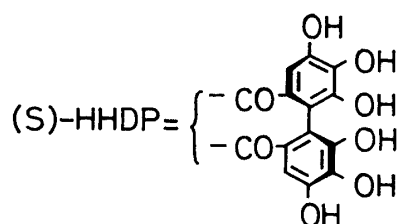
|   | R <sup>1</sup> | R <sup>2</sup> | R <sup>3</sup> | R <sup>4</sup> |
|---|----------------|----------------|----------------|----------------|
| 1 | O-gall         | H              | gall           | gall           |
| 2 | H, OH          |                | gall           | gall           |
| 3 | O-gall         | H              | (S)-HHDP       |                |



|    | R <sup>1</sup> | R <sup>2</sup> | R <sup>3</sup> | R <sup>4</sup> |
|----|----------------|----------------|----------------|----------------|
| 5  | H, OH          |                | gall           | gall           |
| 9  | O-gall         | H              | gall           | gall           |
| 10 | O-gall         | H              | (S)-HHDP       |                |



|   | R <sup>1</sup> | R <sup>2</sup> | R <sup>3</sup> | R <sup>4</sup> |
|---|----------------|----------------|----------------|----------------|
| 6 | O-gall         | H              | gall           | gall           |
| 7 | H, OH          |                | gall           | gall           |
| 8 | O-gall         | H              | (S)-HHDP       |                |



Hydrolysis of **1** in hot water gave two products, which were isolated by column chromatography on Sephadex LH-20, and were identified as tellimagrandin I (**5**)<sup>4,5</sup> and rugosin A (**6**)<sup>3</sup> respectively. The structure **1** in which two glucose cores are linked *via* the valoneoyl group is therefore assigned to rugosin D.

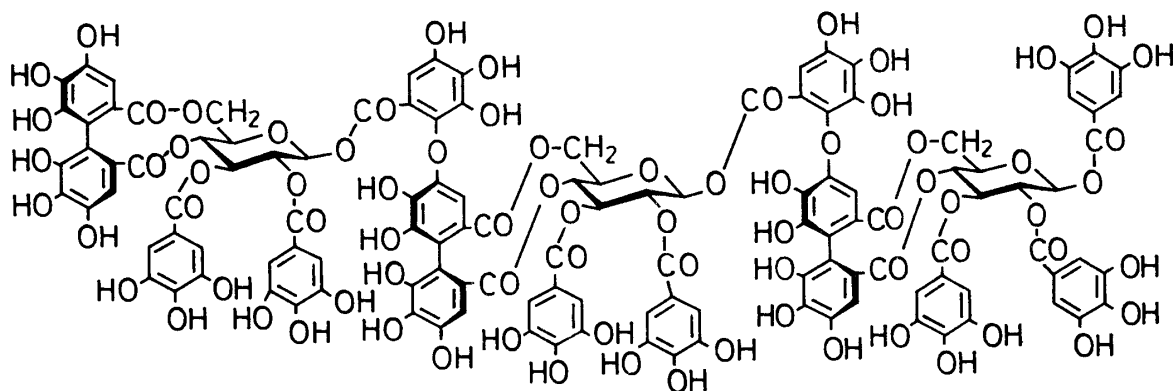
Rugosin E (**2**),  $C_{75}H_{54}O_{48} \cdot 5H_2O$ ,  $[\alpha]_D +140^\circ$  ( $c=1$ , acetone), was obtained as a light tan amorphous powder, UV  $\lambda_{max}^{MeOH}$  nm ( $\log \epsilon$ ) 220 (5.19) and 275 (4.87), which forms an anomer mixture as indicated by its  $^1H$ -NMR spectrum (200 MHz, in acetone- $d_6$ ) [ $\delta$ 7.09 (6/5H), 7.08 (4/5H), 7.034 (6/5H), 7.028 (14/5H) and 6.99 (2H) (four galloyl groups);  $\delta$ 7.16 (1H), 6.67 (1H), 6.49 (3/5H), 6.47 (7/5H), 6.251 (3/5H) and 6.245 (2/5H) (a hexahydroxydiphenoyl group and a valoneoyl group); protons of two glucose cores other than H-1 proton ( $\delta$ 6.17, d,  $J=8$  Hz) of a glucose core were complicated]. Partial hydrolysis of **2** in hot water afforded **5** and rugosin B (**7**).<sup>3)</sup> These results indicate that the structure of rugosin E is **2**.

Rugosin F (**3**),  $C_{82}H_{56}O_{52} \cdot 12H_2O$ ,  $[\alpha]_D +88^\circ$  ( $c=1$ , acetone), UV  $\lambda_{max}^{MeOH}$  nm ( $\log \epsilon$ ) 218 (5.18) and 273 (4.85), was obtained as a light tan amorphous powder. The  $^1H$ -NMR spectrum (200 MHz, in acetone- $d_6$ ) shows the presence of three galloyl ( $\delta$ 7.17, 7.00 and 6.97; 2H each), a valoneoyl and two hexahydroxydiphenoyl ( $\delta$ 7.12, 6.65, 6.54, 6.47, 6.46, 6.42 and 6.26; 1H each) groups and two glucose cores [ $\delta$ 6.19 (d,  $J=8.5$  Hz, H-1), 6.13 (d,  $J=8$  Hz, H-1), 5.80 (t,  $J=10$  Hz, H-3), 5.51 (dd,  $J=8, 10$  Hz, H-2), 5.44 (dd,  $J=9, 10$  Hz, H-3), 5.32 (dd,  $J=6, 14$  Hz, H-6), 5.29 (dd,  $J=6, 13$  Hz, H-6), 5.16 (t,  $J=10$  Hz, H-4), 5.15 (dd,  $J=8.5, 9$  Hz, H-2), 5.09 (t,  $J=10$  Hz, H-4), 4.48 (dd,  $J=6, 10$  Hz, H-5), 4.47 (dd,  $J=6, 10$  Hz, H-5), 3.83 (d,  $J=14$  Hz, H-6') and 3.78 (d,  $J=13$  Hz, H-6')] forming  $C1$  conformation and the  $\beta$ -anomer, as shown by the coupling constants.

Methylation of **3** afforded nonacosa-*O*-methylrugosin F,  $C_{111}H_{114}O_{52} \cdot 3/2H_2O$ ,  $[\alpha]_D +26^\circ$  ( $c=1$ , acetone) [mass spectrum:  $m/z$  646 and 614 (methylated valoneoyl); 422, 404 and 360 (hexamethoxydiphenoyl); 212, 197 and 195 (trimethylgalloyl)].

Hydrolysis of **3** afforded **5** and rugosin C (**8**)<sup>3)</sup> in an analogous way to the partial hydrolysis of **1** and **2**. Thus structure **3** was assigned to rugosin F.

Rugosin G (**4**),  $C_{123}H_{86}O_{78} \cdot 18H_2O$ ,  $[\alpha]_D +109^\circ$  ( $c=1$ , acetone), UV  $\lambda_{max}^{MeOH}$  nm ( $\log \epsilon$ ) 218 (5.37) and 276 (5.05), was obtained as a light tan amorphous powder. This tannin was pure and homogeneous on TLC, HPLC, and in the  $^1H$ -NMR spectrum. The retention volume on gel permeation chromatography on a co-polymer of styrene and divinylbenzene is consistent with the molecular weight of the trimeric version of the other rugosins. The  $^1H$ -NMR spectrum (200 MHz, in acetone- $d_6$ ) shows that



it has twenty-two aromatic protons [ $\delta$ 7.15 (3H), 7.14 (1H), 7.03 (2H), 7.02 (2H), 7.009 (2H), 7.006 (2H), 7.00 (2H), 6.98 (2H), 6.68 (1H), 6.48 (1H), 6.47 (2H), 6.23 (1H) and 6.22 (1H)] corresponding to seven galloyl, a hexahydroxydiphenoyl and two valoneoyl groups, and twenty-one glucose protons [ $\delta$ 6.19, 6.11 and 6.08 (d,  $J=8$  Hz, H-1); 5.85, 5.78 and 5.77 (t,  $J=10$  Hz, H-3); 5.62, 5.58 and 5.54 (dd,  $J=8, 10$  Hz, H-2); 5.32, 5.31 and 5.27 (dd,  $J=6, 14$  Hz, H-6); 5.17, 5.16 and 5.10 (t,  $J=10$  Hz, H-4); 4.53, 4.47 and 4.43 (dd,  $J=6, 10$  Hz, H-5); 3.79, 3.76 and 3.70 (d,  $J=14$  Hz, H-6')]. The coupling constants of glucose protons indicate three glucose cores form *C1* conformation and  $\beta$ -anomer.

Partial hydrolysis of **4** gave **5**, **6** and **7** in approximately the same amount. Therefore, rugosin G has the structure **4** having three tellimagrandin II (**9**)<sup>4~6</sup> units.

Among the tannins of this flower, **9** is the most abundant component (ca. 15% of the extract of fresh flower petal). The dimeric tannins (and also the trimer) may have been produced by ether linkage formation between the galloyl group of **9** and the hexahydroxydiphenoyl group of **5**, casuarictin (**10**),<sup>4</sup> or another molecule of **9**, although another biosynthetic route, esterification of the valoneoyl group of **6**, **7** or **8** with anomeric hydroxyl group of **5**, is also conceivable.

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