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### Steroid Saponins from *Paris polyphylla* Sm.—Supplement<sup>1)</sup>

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Previously, four steroid glycosides (Pa—d) were isolated from the rhizomes of *Paris polyphylla* Sm. and characterized. In a continuation of our studies on this plant, a further five new steroid glycosides (1, 2, 12, 14 and 16) together with three known compounds (13, 18 and 19; the latter two have been obtained synthetically as acetyl derivatives) have been isolated and their structures established as follows. 1 and 2: (22 $\xi$ ,25R)-22-methoxyfurostanol 3,26-O-bisglycosides (proto-type saponins) corresponding to Pa [diosgenin 3-O- $\alpha$ -L-ara·fur-(1 $\rightarrow$ 4)-[ $\alpha$ -L-rha·pyr-(1 $\rightarrow$ 2)]- $\beta$ -D-glc·pyranoside (3)] and Pb [diosgenin 3-O- $\alpha$ -L-rha·pyr-(1 $\rightarrow$ 4)- $\alpha$ -L-rha·pyr-(1 $\rightarrow$ 4)-[ $\alpha$ -L-rha·pyr-(1 $\rightarrow$ 2)]- $\beta$ -D-glc·pyranoside (6)], respectively; 12 and 13: pennogenin oligosides having the same sugar moieties as 3 and 6, respectively; 14 and 18: pennogenin derivatives carrying 3-O- $\alpha$ -L-ara·fur-(1 $\rightarrow$ 4)- $\beta$ -D-glc·pyranoside and hexaacetyl 3-O- $\alpha$ -L-rha·pyr-(1 $\rightarrow$ 2)- $\beta$ -D-glc·pyranoside, respectively; 16 and 19: diosgenin derivatives carrying the same sugar moieties as 14 and 18, respectively.

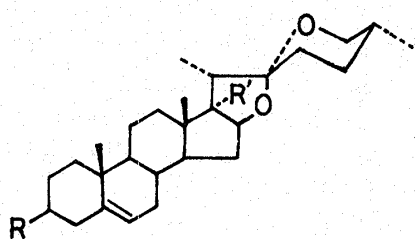
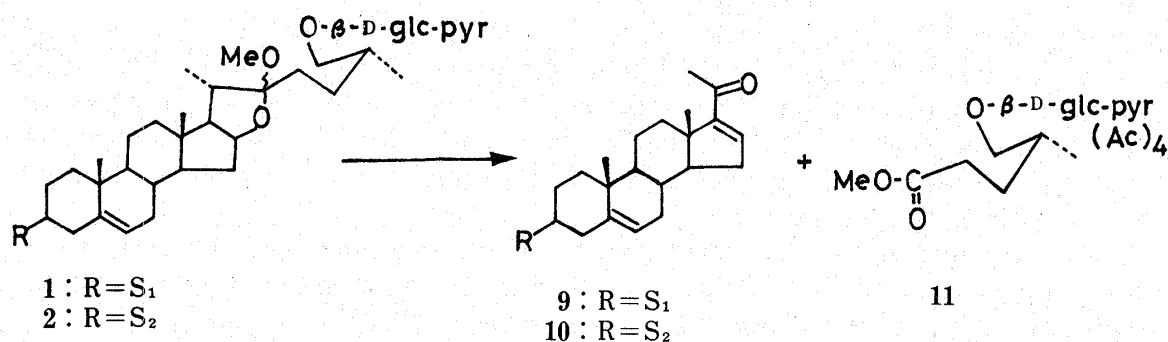
**Keywords**—*Paris polyphylla*; Liliaceae; furostanol glycoside; diosgenin glycoside; pennogenin glycoside; arabinofuranoside

In the previous paper,<sup>1)</sup> we reported four steroid glycosides (Pa through Pd) from the rhizomes of Himarayan *Paris polyphylla* Sm. (Liliaceae), which is used as a folk medicine in some parts of Nepal. In China, this plant has been used to treat chronic bronchitis. In a continuation of our studies on the ingredients of this plant, we have obtained a further eight steroid glycosides (1, 2, 12, 13, 14, 16, 18 and 19) together with Pa (3) and Pb (6). This paper deals with their structure determinations.

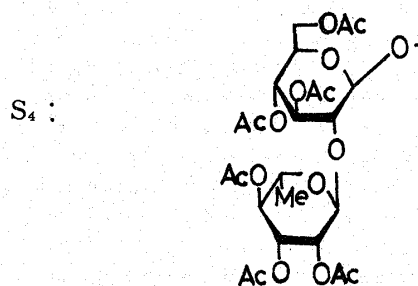
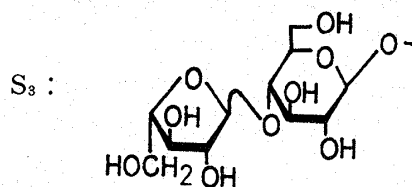
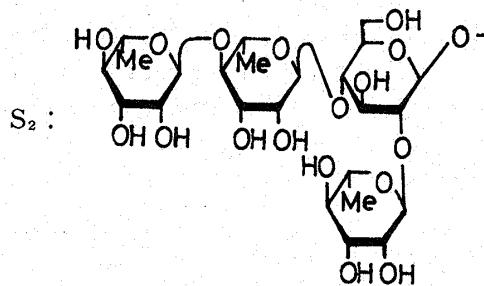
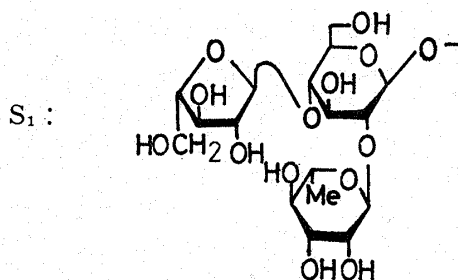
Separation of the MeOH extractive of the crude drug, commercial dried rhizomes of *Paris polyphylla* purchased in a market in China, was undertaken as described in Experimental, affording 1, 2, 12, 13, 14, 16, 18 and 19 in addition to Pa—d previously isolated.

Both 1, an amorphous powder (mp 209—212°C),  $[\alpha]_D -135.6^\circ$ , and 2, an amorphous powder (mp 174—177°C),  $[\alpha]_D -87.0^\circ$ , showed strong absorptions (3600—3200 cm<sup>-1</sup>) due to hydroxyl groups, but not due to spiroketal functions<sup>2)</sup> in their infrared (IR) spectra and were positive to Ehrlich reagent, which is a convenient reagent<sup>3)</sup> to detect proto-type saponins such as proto-dioscin,<sup>4)</sup> for example. They also showed methoxy signals<sup>4)</sup> reminiscent of 22-methoxyfurostane derivatives in their <sup>1</sup>H nuclear magnetic resonance (<sup>1</sup>H NMR) spectra. On the assumption that 1 and 2 were proto-type saponins, the compounds were hydrolyzed with  $\beta$ -glucosidase (almond emulsin) to afford spirostanol derivatives, 3, colorless needles, mp 274—276°C,  $[\alpha]_D -133.0^\circ$ , identical (mp,  $[\alpha]_D$ , thin layer chromatography (TLC), <sup>13</sup>C nuclear magnetic resonance (<sup>13</sup>C NMR)) with Pa, and 6, colorless needles, mp 203—206°C,  $[\alpha]_D -128.0^\circ$ , identical (mp,  $[\alpha]_D$ , TLC, <sup>13</sup>C NMR) with Pb, respectively, both accompanied with D-glucose ( $[\alpha]_D$ , TLC). Furthermore, the identity of 3 and 6, obtained by enzymic hydrolysis of 1 and 2, with Pa and Pb, respectively, was substantiated by their transformation into the acetylated derivatives (4, colorless needles, mp 122—128°C,  $[\alpha]_D -100.0^\circ$ , and 7, colorless needles, mp 148—153°C,  $[\alpha]_D -74.0^\circ$ , respectively) and methylated derivatives (5, colorless needles, mp 160—162°C,  $[\alpha]_D -92.0^\circ$ , and 8, colorless needles, mp 135—139°C,  $[\alpha]_D -110.0^\circ$ , respectively). Therefore, 1 and 2 could be deduced to be proto-type saponins corresponding to Pa (3) and Pb (6), respec-

tively. To verify the structures of **1** and **2**, they were acetylated with  $\text{Ac}_2\text{O}$ -pyridine, refluxed with  $\text{Ac}_2\text{O}$  for pseudomerization into  $\Delta^{20(22)}$  and oxidized with  $\text{CrO}_3$  in  $\text{AcOH}$  (Marker's degradation<sup>5</sup>). After treatment of the oxidized products with alkali, **1** and **2** were finally decomposed respectively into  $3\beta$ -hydroxy-pregna-5,16-dien-20-one glycosides, **9**, an amorphous powder (mp 214—216°C),  $[\alpha]_{\text{D}} -42.0^\circ$ , and **10**, colorless needles, mp 234—237°C,  $[\alpha]_{\text{D}} -117.7^\circ$ , respectively, both accompanied with a product formed from the steroidal side chain moiety, which on acetylation followed by methylation with  $\text{CH}_2\text{N}_2$  afforded methyl 4(*R*)-methyl-5-hydroxy-



- 3** : R = S<sub>1</sub>, R' = H (=Pa)  
**4** : R = S<sub>1</sub>(Ac)<sub>8</sub>, R' = H  
**5** : R = S<sub>1</sub>(Me)<sub>8</sub>, R' = H  
**6** : R = S<sub>2</sub>, R' = H (=Pb)  
**7** : R = S<sub>2</sub>(Ac)<sub>10</sub>, R' = H  
**8** : R = S<sub>2</sub>(Me)<sub>10</sub>, R' = H  
**12** : R = S<sub>1</sub>, R' = OH  
**13** : R = S<sub>2</sub>, R' = OH  
**14** : R = S<sub>3</sub>, R' = OH  
**15** : R = S<sub>3</sub>(Ac)<sub>6</sub>, R' = OH  
**16** : R = S<sub>3</sub>, R' = H  
**17** : R = S<sub>3</sub>(Ac)<sub>6</sub>, R' = H  
**18** : R = S<sub>4</sub>, R' = OH  
**19** : R = S<sub>4</sub>, R' = H



pentanoate-D-glucopyranoside tetraacetate (**11**)<sup>4,5b</sup>) as a colorless oil. The structures of **9**, **10** and **11** were supported by their IR, mass (MS) and <sup>1</sup>H NMR spectra.

Consequently **1** and **2** can be defined as (22ξ, 25R)-22-methoxyfurost-5-ene-3,26-diol 26-O-β-D-glucopyranosides of 3-O-α-L-arabinofuranosyl-(1→4)-[α-L-rhamnopyranosyl-(1→2)]-β-D-glucopyranosyl and 3-O-α-L-rhamnopyranosyl-(1→4)-α-L-rhamnopyranosyl-(1→4)-[α-L-rhamnopyranosyl-(1→2)]-β-D-glucopyranosyl, respectively.

Next, **12**, colorless needles, mp 274—278°C, [α]<sub>D</sub> -108.9°, showed absorptions due to hydroxyl (3600—3200 cm<sup>-1</sup>) and spiroketal side chain (980, 920, 900 and 890) moieties. The mass spectrum of **12** exhibited characteristic peaks,<sup>6)</sup> *m/z* 412 (C<sub>27</sub>H<sub>40</sub>O<sub>3</sub><sup>+</sup>), 155 (C<sub>9</sub>H<sub>15</sub>O<sub>2</sub><sup>+</sup>), 153 (C<sub>9</sub>H<sub>13</sub>O<sub>2</sub><sup>+</sup>) and 126 (C<sub>8</sub>H<sub>14</sub>O<sup>+</sup>), derived from pennogenin. Moreover, the <sup>13</sup>C NMR spectrum of **12** revealed signals attributable to the sugar moiety superimposed on those of Pa (**3**), as described in Experimental. Accordingly, **12** was deduced to be pennogenin 3-O-α-L-arabinofuranosyl-(1→4)-[α-L-rhamnopyranosyl-(1→2)]-β-D-glucopyranoside, whose sugar moiety is identical with that of Pa (**3**).

**13**, colorless needles, mp 224—228°C (dec.), was identical (mp, TLC, IR) with Tg,<sup>7)</sup> pennogenin tetraglycoside, having the same sugar residue as Pb (**6**) previously isolated from the fresh underground parts of *Trillium kamtschaticum* PALL.

**14**, an amorphous powder, [α]<sub>D</sub> -98.0°, and **16**, colorless needles, mp 239—242°C, were derived into the corresponding acetates, **15**, colorless needles, mp 172.5—176°C, [α]<sub>D</sub> -44.6°, and **17**, colorless needles, mp 174—176°C, [α]<sub>D</sub> -37.4°, respectively. Both acetates, **15** and **17**, showed peaks derived from the terminal peracetylated-pentosyl hexosyl cation (*m/z* 547)<sup>6a)</sup> and -pentosyl cation (*m/z* 259)<sup>6a)</sup> in the mass spectra. Furthermore, a comparison of the signals ascribable to the sugar moieties in the <sup>13</sup>C NMR spectra of **15** and **17** indicated that the moieties were almost identical. The hexamethyl ether derived from **14**, on acid hydrolysis with 2 N HCl-MeOH, gave two methylglycosides of 2,3,5-tri-O-methyl arabinofuranose and 2,3,6-tri-O-methyl glucopyranose besides aglycone derivatives (bethogenin and kryptogenin).<sup>8)</sup> Since the mass spectra of **14** and **16** showed peaks at *m/z* [412, 155, 153, 126]<sup>6)</sup> and [396(C<sub>27</sub>H<sub>40</sub>O<sub>2</sub><sup>+</sup>), 282 (C<sub>21</sub>H<sub>30</sub><sup>+</sup>), 139 (C<sub>9</sub>H<sub>15</sub>O<sup>+</sup>)]<sup>9)</sup> characteristic of pennogenin and diosgenin, respectively, and the chemical shifts of the signals due to the aglycones in the <sup>13</sup>C NMR spectra of **15** and **17** were in good agreement with those of pennogenin and diosgenin, respectively, **14** and **16** are pennogenin- and diosgenin-glycosides, respectively, having the same sugar moiety, 3-O-α-L-arabinofuranosyl-(1→4)-β-D-glucopyranoside, and correspond to prosapogenins of **12** and **3**, respectively.

**18**, colorless needles, mp 193—195°C, [α]<sub>D</sub> -52.0°, and **19**, colorless needles, mp 203—207°C, [α]<sub>D</sub> -60.0°, isolated after acetylation, were identical (mp, [α]<sub>D</sub>, TLC, MS) with Tb hexaacetate<sup>7)</sup> and Ta hexaacetate,<sup>7)</sup> respectively, previously obtained from the fresh underground parts of *T. kamtschaticum* PALL.

### Experimental

All melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. Optical rotations were measured with a Union Giken PM-201 automatic digital polarimeter at 20—25°C. IR and UV spectra were obtained with Hitachi IR-215 and Hitachi UV-124 machines, respectively. <sup>13</sup>C NMR and <sup>1</sup>H NMR spectra were taken with JEOL JNM-FX-90Q (22.5 MHz) and JEOL JNM-PS-100 (100 MHz) spectrometers, respectively. Chemical shifts are given on a δ (ppm) scale with tetramethylsilane as an internal standard. Mass spectra were taken with a JEOL JMS D-300 machine. Gas liquid chromatography (GLC) was run on a Shimadzu GC-6A unit with a flame ionization detector using a glass column (3 mm × 2.0 m) packed with neopentylglycol succinate. Paper partition chromatography (PPC) for sugar was conducted on Toyo Roshi No. 50 paper using the upper layer of *n*-BuOH-pyridine-water (6:2:3) + pyridine (1) as a solvent and aniline phthalate as a staining agent. TLC was performed on precoated silica gel plates 0.25 mm thick (Kieselgel 60, Merck) and detection was achieved by spraying 10% H<sub>2</sub>SO<sub>4</sub> followed by heating. Column chromatography was performed on Kieselgel 60 (Merck, 70—230 mesh), Sephadex LH-20 (Pharmacia Fine Chem.) and Amberlite XAD-2 (Organo).

**Extraction and Isolation of Steroid Saponins**—Commercial dried rhizomes (480 g) purchased in a market

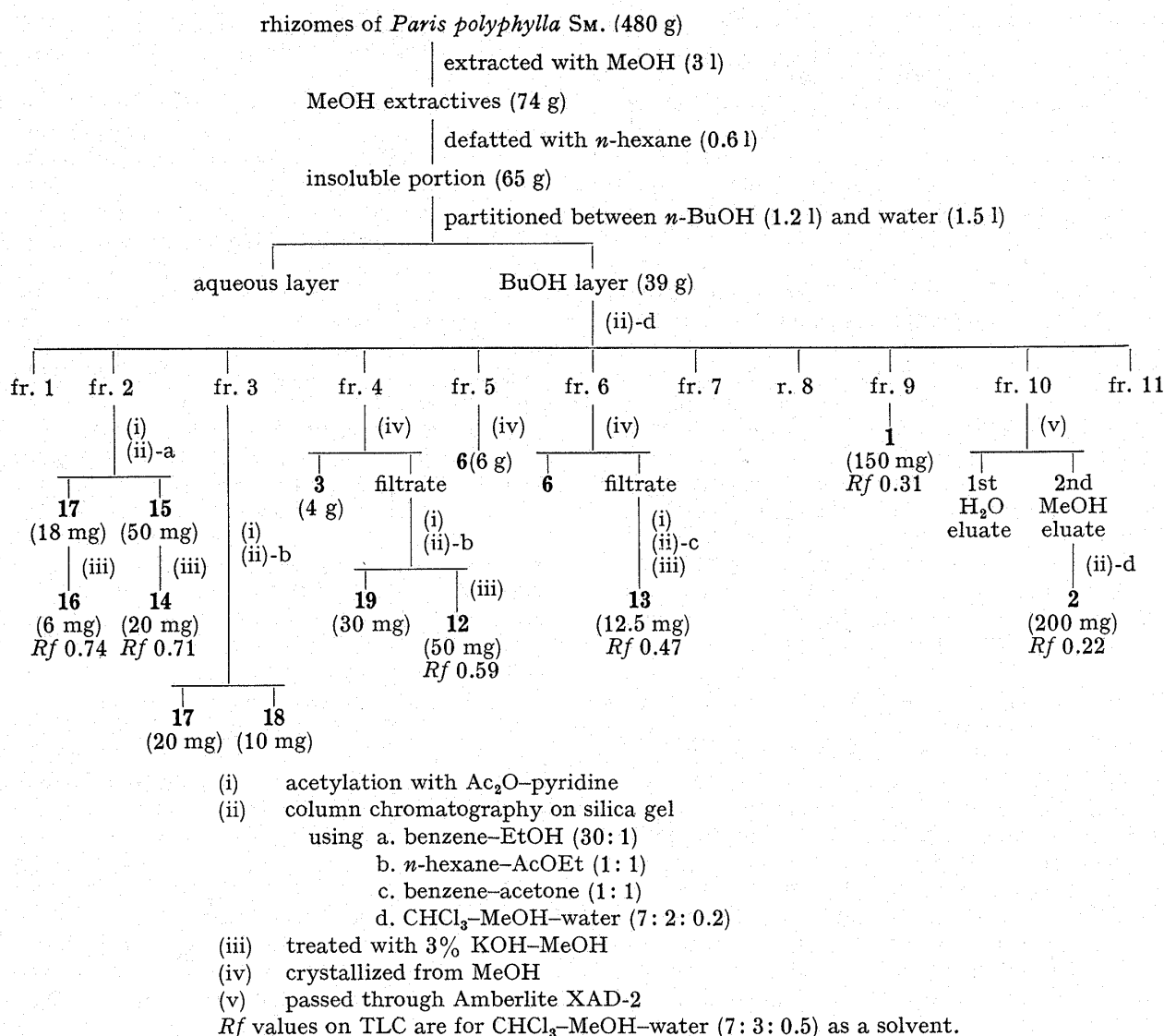


Chart 1

in China were extracted with refluxing MeOH (3 l), and the extract was concentrated under reduced pressure to give a residue (74 g), which was treated as shown in Chart 1.

1—An amorphous powder (mp 209–212°C),  $[\alpha]_D -135.6^\circ$  ( $c=0.45$ , MeOH). IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>-1</sup>: 3600–3300 (OH). Ehrlich reagent: positive. Anal. Calcd for C<sub>50</sub>H<sub>84</sub>O<sub>22</sub>·H<sub>2</sub>O: C, 56.91; H, 8.11. Found: C, 56.51; H, 7.92. <sup>1</sup>H NMR (*d*<sub>5</sub>-pyridine)  $\delta$  (ppm): 3.24 (OMe).

**Enzymic Hydrolysis of 1**—A solution of 1 (60 mg) in dist. water (5 ml) was incubated with almond emulsin (Sigma Chem. Co.) (20 mg) at 37°C for 8 h, then the reaction mixture was evaporated to dryness *in vacuo* to give a residue, to which MeOH was added. The mixture was filtered. The MeOH solution was evaporated to dryness to give a residue, which was subjected to Sephadex LH-20 column chromatography using MeOH as an eluent to give a glycoside (3), colorless needles (from MeOH), mp 274–276°C,  $[\alpha]_D -133.0^\circ$  ( $c=0.56$ , MeOH), identical with Pa (30 mg), and a sugar identical with D-glucose (12 mg), Rf 0.46 (on PPC),  $[\alpha]_D +59.5^\circ$  ( $c=0.51$ , water). 3: Anal. Calcd for C<sub>44</sub>H<sub>72</sub>O<sub>16</sub>·H<sub>2</sub>O: C, 60.39; H, 8.52. Found: C, 60.37; H, 8.45. <sup>13</sup>C NMR (in *d*<sub>5</sub>-pyridine)  $\delta$  (ppm): aglycone (diosgenin);<sup>10</sup> 37.4, 30.0, 77.6, 40.4, 140.8, 121.6, 32.2, 31.6, 50.3, 37.0, 21.0, 39.8, 40.4, 56.6, 32.1, 81.1, 62.4, 16.3, 19.3, 41.9, 14.9, 109.3, 31.6, 29.1, 30.4, 66.8, 17.2 (C<sub>1-27</sub>), glucose moiety; 100.0, 78.2, 77.2, 76.2, 77.6, 62.7 (C<sub>1'-6'</sub>), rhamnose moiety; 101.6, 72.4, 71.9, 73.6, 69.2, 18.3 (C<sub>1''-6''</sub>), arabinofuranose moiety; 109.4, 82.5, 77.6, 86.1, 61.2 (C<sub>1'''-5'''</sub>).

**Octaacetate (4) of 3**—3 (85 mg) was acetylated with Ac<sub>2</sub>O-pyridine (each 4 ml) at 80°C for 1 h to give a peracetate (4), colorless needles (from MeOH, 68 mg), mp 122–128°C,  $[\alpha]_D -100.0^\circ$  ( $c=0.50$ , CHCl<sub>3</sub>). MS  $m/z$ : 777, 396 (C<sub>27</sub>H<sub>40</sub>O<sub>2</sub><sup>+</sup>), 282 (C<sub>21</sub>H<sub>30</sub><sup>+</sup>), 273 (C<sub>12</sub>H<sub>17</sub>O<sub>7</sub><sup>+</sup>), 259 (C<sub>11</sub>H<sub>15</sub>O<sub>7</sub><sup>+</sup>), 253, 139 (C<sub>9</sub>H<sub>15</sub>O<sub>2</sub><sup>+</sup>), 115 (C<sub>6</sub>H<sub>11</sub>O<sub>2</sub><sup>+</sup>).

**Octamethylether (5) of 3**—3 (50 mg) was methylated by Hakomori's method<sup>11</sup> to give a permethylate (5), colorless needles (from MeOH, 26 mg), mp 160–162°C,  $[\alpha]_D -92.0^\circ$  ( $c=0.50$ , CHCl<sub>3</sub>). MS  $m/z$ : 553, 396, 282, 253, 189 (C<sub>9</sub>H<sub>17</sub>O<sub>4</sub><sup>+</sup>), 175 (C<sub>8</sub>H<sub>15</sub>O<sub>4</sub><sup>+</sup>), 139, 115. <sup>1</sup>H NMR (in CDCl<sub>3</sub>)  $\delta$  (ppm): 0.80 (3H, s, 18-CH<sub>3</sub>),

0.85 (3H, d,  $J=6$  Hz, 27-CH<sub>3</sub>), 0.98 (3H, d,  $J=6$  Hz, 21-CH<sub>3</sub>), 1.02 (3H, s, 19-CH<sub>3</sub>), 3.53–3.70 (OMe), 4.33 (1H, d,  $J=7$  Hz, glucosyl anomeric proton), 5.17 (1H, s, rhamnosyl anomeric proton), 5.22 (1H, broad, s, arabinofuranosyl anomeric proton).

**Oxidation Products (9 and 11) of 1**—The peracetate (72 mg) of 1 in Ac<sub>2</sub>O (10 ml) was refluxed for 45 min, then the solution was cooled and EtOH was added. The whole was evaporated to dryness *in vacuo*, yielding a pale brown residue (60 mg) which included the acetyl  $\Delta^{20(22)}$  compound of 1. AcOH (1.6 ml) and AcONa (32 mg) were added to the residue, and the resulting solution was cooled to 12°C. CrO<sub>3</sub> solution (65 mg in 0.5 ml of 40% AcOH) was added with stirring, and the mixture was allowed to stand for 1 h at room temperature. The reaction mixture was diluted with water (50 ml) and shaken with CHCl<sub>3</sub> to extract oxidation products. Evaporation yielded a syrup (78 mg), which was mixed with an aqueous solution (1.2 ml) containing KOH (165 mg). After further addition of *t*-BuOH (4 ml), the mixture was incubated at 30°C for 3.5 h. The *t*-BuOH was distilled off, and the reaction mixture was extracted with *n*-BuOH. The extract was evaporated to dryness *in vacuo* to give a residue, which was chromatographed on silica gel using CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (7:3:0.2) to give a pregnenolone glycoside (9, sugar moiety=S<sub>1</sub>), a white powder (mp 214–216°, 25 mg),  $[\alpha]_D -42.0^\circ$  ( $c=0.50$ , MeOH). IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600–3200 (OH), 1650, 1580 ( $\alpha,\beta$ -unsaturated ketone). UV  $\lambda_{\max}^{\text{EtOH}}$  nm: 239 ( $\epsilon=9800$ ). <sup>1</sup>H NMR (in *d*<sub>5</sub>-pyridine)  $\delta$  (ppm): 0.95 (3H, s, 19-CH<sub>3</sub>), 1.05 (3H, s, 18-CH<sub>3</sub>), 1.75 (3H, d,  $J=6$  Hz, rhamnosyl 5-CH<sub>3</sub>), 2.25 (3H, s, 21-CH<sub>3</sub>), 5.80 (1H, d,  $J=9$  Hz, glucosyl anomeric proton), 6.18 (1H, s, arabinofuranosyl anomeric proton), 6.63 (1H, br s, rhamnosyl anomeric proton). The aqueous layer, after being adjusted to pH 3.0 with 3N HCl, was shaken in turn with *n*-BuOH and CHCl<sub>3</sub>, neutralized with 2N NaOH, and concentrated to afford a salt mixture. The salt were acetylated in a usual way and its acetate was taken up in MeOH. This solution was treated with diazomethane in ether. After removal of the solvent, the residue was chromatographed on a silica gel column with *n*-hexane–acetone (1:1) to give methyl 4(*R*)-methyl-5-hydroxypentanoate  $\beta$ -D-glucopyranoside tetraacetate (11), a colorless oil (11 mg),  $[\alpha]_D -16.8^\circ$  ( $c=0.42$ , CHCl<sub>3</sub>). MS  $m/z$ : 331 (C<sub>14</sub>H<sub>19</sub>O<sub>9</sub><sup>+</sup>), 243, 242, 200, 169, 157, 145, 140, 129 (C<sub>7</sub>H<sub>13</sub>O<sub>2</sub><sup>+</sup>), 115, 109, 103, 98, 97 (C<sub>8</sub>H<sub>9</sub>O<sup>+</sup>). <sup>1</sup>H NMR (in CDCl<sub>3</sub>)  $\delta$  (ppm): 0.90 (3H, d,  $J=7$  Hz, *sec.* CH<sub>3</sub>), 2.00–2.08 (12H, 4 × OAc), 3.62 (3H, s, 1 × OMe), 4.45 (1H, d,  $J=7$  Hz, glucosyl anomeric proton).


**2**—An amorphous powder (mp 174–177°C),  $[\alpha]_D -87.0^\circ$  ( $c=1.00$ , MeOH). IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600–3200 (OH). Ehrlich reagent: positive. Anal. Calcd for C<sub>57</sub>H<sub>90</sub>O<sub>25</sub>·3H<sub>2</sub>O: C, 55.40; H, 7.99. Found: C, 55.35; H, 8.10. <sup>1</sup>H NMR (in *d*<sub>5</sub>-pyridine)  $\delta$  (ppm): 3.21 (OMe). Enzymic hydrolysis of 2 (42 mg) with almond emulsin (15 mg) in the manner described for 1 and 2 yielded a glycoside (6), colorless needles (12 mg), mp 203–206°C,  $[\alpha]_D -128.0^\circ$  ( $c=0.50$ , MeOH), identical with Pb, and a sugar (11 mg) identical with D-glucose, *Rf* 0.45 (on PPC),  $[\alpha]_D +60.0^\circ$  ( $c=0.30$ , water). **6**: Anal. Calcd for C<sub>51</sub>H<sub>82</sub>O<sub>20</sub>·3H<sub>2</sub>O: C, 57.29; H, 8.30. Found: C, 57.49; H, 8.00. <sup>13</sup>C NMR (in *d*<sub>5</sub>-pyridine)  $\delta$  (ppm): aglycone; 37.4, 30.0, 77.2, 38.8, 140.7, 121.6, 32.2, 31.6, 50.2, 37.0, 21.0, 39.8, 40.4, 56.6, 32.0, 81.0, 62.6, 16.3, 19.3, 41.8, 14.9, 109.3, 31.6, 29.0, 30.4, 66.8, 17.2 (C<sub>1–27</sub>), glucose moiety; 100.1, 80.0, 76.4, 77.6, 78.1, 61.1 (C<sub>1'–6'</sub>), rhamnose moiety attached to C-2 of glucose; 102.0, 72.4, 71.9, 73.6, 69.3, 18.4 (C<sub>1''–6''</sub>), rhamnose moiety attached to C-4 of glucose; 102.7, 72.4, 70.0, 78.1, 68.2, 18.3 (C<sub>1'''–6'''</sub>), rhamnose moiety attached to C-4 of rhamnose; 102.0, 72.4, 71.9, 73.6, 69.3, 18.1 (C<sub>1''''–6''''</sub>).

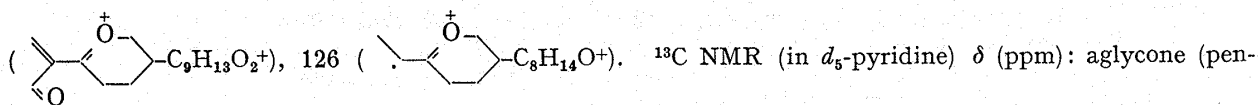
**Decaacetate (7) of 6**—6 (30 mg) was acetylated with Ac<sub>2</sub>O–pyridine (each 2 ml) at 80°C for 1 h to give 7 (30 mg) as colorless needles (from MeOH, 21 mg), mp 148–153°C,  $[\alpha]_D -74.0^\circ$  ( $c=1.00$ , CHCl<sub>3</sub>). MS  $m/z$ : 503 (C<sub>22</sub>H<sub>31</sub>O<sub>13</sub><sup>+</sup>), 414, 396, 282, 273, 253.

**Decamethyl Ether (8) of 6**—6 (50 mg) was methylated by Hakomori's method to give 8 (10 mg) as colorless needles (from MeOH, 12 mg), mp 135–139°C,  $[\alpha]_D -110.0^\circ$  ( $c=0.50$ , CHCl<sub>3</sub>). MS  $m/z$ : 1154 (M<sup>+</sup>), 414, 396, 363 (C<sub>17</sub>H<sub>31</sub>O<sub>8</sub><sup>+</sup>), 282, 253, 189 (C<sub>9</sub>H<sub>17</sub>O<sub>4</sub><sup>+</sup>). <sup>1</sup>H NMR (in CDCl<sub>3</sub>)  $\delta$  (ppm): 0.79 (3H, s, 18-CH<sub>3</sub>), 0.85 (3H, d,  $J=7$  Hz, 27-CH<sub>3</sub>), 0.97 (3H, d,  $J=6$  Hz, 21-CH<sub>3</sub>), 1.03 (3H, s, 19-CH<sub>3</sub>), 1.20, 1.22, 1.24 (each 3H, d,  $J=6$  Hz, rhamnosyl 5-CH<sub>3</sub>), 3.35–3.75 (OMe), 4.38 (1H, d,  $J=7$  Hz, glucosyl anomeric proton), 5.02 (1H, s, rhamnosyl anomeric proton), 5.20 (2H, s, 2 × rhamnosyl anomeric proton).

**Oxidation Products (10 and 11) of 2**—The peracetate (150 mg) of 2 was oxidized in the manner described for 1 to give a pregnenolone glycoside (10, sugar moiety=S<sub>2</sub>), colorless needles (from MeOH, 62 mg) mp 234–237°C,  $[\alpha]_D -117.7^\circ$  ( $c=0.51$ , MeOH) and methyl 4(*R*)-methyl-5-hydroxypentanoate  $\beta$ -D-glucopyranoside tetraacetate (11), a colorless oil (30 mg),  $[\alpha]_D -18.2^\circ$  ( $c=0.92$ , CHCl<sub>3</sub>). **10**: IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600–3000 (OH), 1650, 1580 ( $\alpha,\beta$ -unsaturated ketone). UV  $\lambda_{\max}^{\text{EtOH}}$  nm: 235 ( $\epsilon=10800$ ). <sup>1</sup>H NMR (in *d*<sub>5</sub>-pyridine)  $\delta$  (ppm): 0.95 (3H, s, 19-CH<sub>3</sub>), 1.06 (3H, s, 18-CH<sub>3</sub>), 1.60 (6H, d,  $J=6$  Hz, 2 × rhamnosyl 5-CH<sub>3</sub>), 1.77 (3H, d,  $J=6$  Hz, rhamnosyl 5-CH<sub>3</sub>), 2.24 (3H, s, 21-CH<sub>3</sub>), 5.75 (1H, d,  $J=7$  Hz, glucosyl anomeric proton), 6.22, 6.35, 6.56 (each 1H, s, rhamnosyl anomeric proton). **11**: Mass and <sup>1</sup>H NMR (in CDCl<sub>3</sub>) spectra were identical with those of the product of oxidation of 1.

**12**—Colorless needles (from MeOH), mp 274–278°C (dec.),  $[\alpha]_D -108.9^\circ$  ( $c=0.73$ , MeOH).  $[M]_D -949.6^\circ$ ,  $\Delta[M]_D(12-Tb) -80.2^\circ$ ,  $\Delta[M]_D(12-14) -257.7^\circ$ . IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600–3200 (OH), 980, 920, 900, 890 (intensity 900 > 920, 25*R*-spiroketal side chain). Anal. Calcd for C<sub>44</sub>H<sub>72</sub>O<sub>17</sub>·H<sub>2</sub>O: C, 59.31; H, 8.37.

Found. C, 58.96; H, 8.27. MS  $m/z$ : 430 (C<sub>27</sub>H<sub>42</sub>O<sub>4</sub><sup>+</sup>), 412 (C<sub>27</sub>H<sub>40</sub>O<sub>3</sub><sup>+</sup>), 155 (  C<sub>9</sub>H<sub>15</sub>O<sub>2</sub><sup>+</sup>), 153



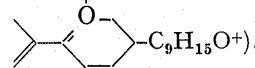
13—Colorless needles (from MeOH), mp 224—228°C (dec.),  $[\alpha]_D -132.0^\circ$  ( $c=0.50$ , MeOH). IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600—3200 (OH), 980, 920, 900, 890 (intensity 900—920, 25*R*-spiroketal side chain). Anal. Calcd for C<sub>51</sub>H<sub>82</sub>O<sub>21</sub>·3H<sub>2</sub>O: C, 56.44; H, 8.17. Found: C, 56.21; H, 8.08. Identical with Tg in terms of mp, *Rf* value on TLC and IR.

14—A white powder (mp 220—223°C),  $[\alpha]_D -98.0^\circ$  ( $c=0.50$ , MeOH),  $[M]_D -691.9^\circ$ . IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600—3100 (OH), 980, 915, 898, 890 (intensity 898>915, 25*R*-spiroketal side chain). MS *m/z*: 430, 412, 394, 155, 153, 127, 126. Anal. Calcd for C<sub>38</sub>H<sub>60</sub>O<sub>12</sub>: C, 64.38; H, 8.53. Found: C, 64.12; H, 8.41.

**Hexaacetate (15) of 14**—Colorless needles (from MeOH), mp 172.5—176°C,  $[\alpha]_D -44.6^\circ$  ( $c=0.92$ , CHCl<sub>3</sub>). MS (*m/z*): 684, 547, 412, 394, 282, 259, 157, 139, 115, 109. <sup>13</sup>C NMR (in CDCl<sub>3</sub>) δ (ppm): aglycone (pennogenin); 37.2, 29.5, 76.6, 39.0, 140.5, 121.7, 32.0, 31.6, 49.8, 36.8, 20.6, 36.8, 43.8, 52.9, 31.6, 91.0, 90.1, 17.1, 19.3, 44.6, 8.0, 110.0, 31.2, 28.1, 30.1, 66.8, 17.2 (C<sub>1-27</sub>), sugar moiety; 107.4, 99.5, 81.6, 81.1, 79.9, 76.6, 73.5, 72.7, 72.1, 63.1, 62.4.

**Methanolysis of Hexamethyl Ether of 14**—14 (10 mg) was methylated by Hakomori's method to give a permethyl ether (6 mg) of 14, a white solid, *Rf* 0.41 (on TLC, solv. *n*-hexane-acetone (2:1)), MS *m/z*: 412, 394, 376, 175, 101. The hexamethyl ether (4 mg) of 14 was subjected to methanolysis under reflux with 1*N* HCl-MeOH (5 ml) for 2 h to give bethogenin and kryptogenin (*Rf* 0.62 and 0.12, respectively, on TLC, solv. *n*-hexane-AcOEt (1:1) and methylglycosides, which were examined by GLC comparison with authentic samples. GLC, column temp. 108°C, N<sub>2</sub> 1.25 kg/cm<sup>2</sup>; *t*<sub>R</sub>, 5.5, 7.4 min (methyl 2,3,5-tri-*O*-methyl α- and β-arabinofuranoside): column temp. 180°C, *t*<sub>R</sub>, 1.60, 2.05 min (methyl 2,3,6-tri-*O*-methyl α- and β-glucopyranoside).

16—Colorless needles (from MeOH), mp 239—242°C. IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 3600—3100 (OH), 980, 918, 8.98, 860 (intensity 898>918, 25*R*-spiroketal side chain), Anal. Calcd for C<sub>38</sub>H<sub>60</sub>O<sub>13</sub>·H<sub>2</sub>O: C, 61.43; H, 8.41.

Found: C, 61.56; H, 8.38. MS *m/z*: 396 (C<sub>27</sub>H<sub>40</sub>O<sub>2</sub><sup>+</sup>), 282 (C<sub>21</sub>H<sub>30</sub><sup>+</sup>), 253, 139 ().

**Hexaacetate (17) of 16**—Colorless needles (from MeOH), mp 174—176°C,  $[\alpha]_D -37.4^\circ$  ( $c=0.54$ , CHCl<sub>3</sub>), MS (*m/z*): 684, 547, 396, 282, 259, 253, 139, 115. <sup>13</sup>C NMR (in CDCl<sub>3</sub>) δ (ppm): aglycone (diosgenin); 37.2, 30.7, 76.6, 39.0, 140.5, 121.8, 32.1, 31.5, 50.2, 36.9, 21.0, 39.8, 40.3, 56.6, 32.0, 80.8, 62.3, 16.3, 19.4, 41.7, 14.5, 109.2, 31.5, 28.9, 29.7, 66.9, 17.1 (C<sub>1-27</sub>), sugar moiety; 107.4, 99.5, 81.7, 81.2, 79.8, 76.0, 73.5, 72.8, 72.1, 63.2, 62.3.

18—Colorless needles (from MeOH), mp 193—195°C,  $[\alpha]_D -52.0^\circ$  ( $c=0.50$ , CHCl<sub>3</sub>), MS (*m/z*): 561, 412, 394, 273, 153, 139, 126, 111.

19—Colorless needles (from MeOH), mp 203—207°C,  $[\alpha]_D -60.0^\circ$  ( $c=1.00$ , CHCl<sub>3</sub>), IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 1750 (OAc), 980, 920, 900, 860 (intensity 900>920, 25*R*-spiroketal side chain), MS (*m/z*): 684, 561, 414, 396, 282, 273, 253, 139. Anal. Calcd for C<sub>51</sub>H<sub>74</sub>O<sub>18</sub>: C, 62.81; H, 7.65. Found: C, 62.50; H, 7.60.

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