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Chemical Studies on the Constituents of Polygonum nodosum

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A new cyclobutane derivative, which may be formed from dehydrokawain by [2+2] cycloaddition, named compound B (8a), mp 226— 227° C, $C_{28}H_{24}O_{6}$, and a new flavanonol, named compound C (10a), mp 142— 143° C, $C_{17}H_{14}O_{6}$, together with pinobanksin(1a), taxifolin(2a), quercetin- 3β -D-glucopyranoside 2''-gallate(3), pinosylvin(4a), methyl gallate(5a), dehydrokawain(6) and compound A(7a) were isolated from Polygonum nodosum (Polygonaceae). The structures of the new compounds, 8a and 10a, were established to be rel-1,trans-3-bis-(4-methoxy-2-oxopyran-6-yl)-cis-2,trans-4-diphenyl cyclobutane and (2R,3R)-3-hydroxy-5-methoxy-6,7-methylenedioxy-flavanone, respectively, on the basis of physicochemical evidence. The structure of 7a was also established to be rel-(1R,6S,7S,8S)-5-methoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)styryl-2-oxabicyclo-[4,2,0]-oct-4-en-3-one and this compound was found to be identical with aniba-dimer-A.

Keywords—*Polygonum nodosum*; Polygonaceae; flavanonol; dehydrokawain dimer; cyclobutane derivative; [2+2] cycloaddition; styrylpyrone derivative; 13 C-NMR

The genus *Polygonum* (polygonaceae) contains a large number of species, and several flavonoids and phenolic compounds have been isolated from some of these. However, few studies on the chemical constituents have been carried out. From a chemotaxonomical veiwpoint, we interested in the constituents of the plants of the genus. So far, quercetin, kaempferol, quercetin- 3β -D-glucopyranoside, kaempferol- 3β -D-glucopyranoside 2"-gallate and quercetin- 3β -D-glucopyranoside 2"-gallate (3) have been reported²⁾ as a constituents of P. nodosum P_{ERS} (Japanese name "Ohinutade"). The present paper describes a further characterization of the constituents of the plant.

The methanol (MeOH) extract of the aerial part of the plant was fractionated into three fractions, *i.e.*, those soluble in ethyl acetate (AcOEt), *n*-butanol (BuOH) and water. Silica gel column chromatography and preparative thin-layer chromatography (PLC) of the AcOEt fraction gave nine constituents, pinobanksin (1a).³⁾ taxifolin (2a),⁴⁾ 3, pinosylvin (4a),⁵⁾ methyl gallate (5a), dehydrokawain (6),⁶⁾ and compounds A (7a), B(8a) and C (10a).

The compounds 1a and 2a gave a tri-(1b) and a pentaacetate (2b), respectively. The proton nuclear magnetic resonance (1 H-NMR) spectra of 1a, 1b, 2a and 2b indicated that 1a is pinobanksin and 2a is taxifolin. The constituent 3 was identical with quercetin-3 β -D-glucopyranoside 2"-gallate isolated from the same source as a molluscidal principle.²⁾ The constituent 4a gave a diacetate (4b) and from the 1 H-NMR spectra of 4a and 4b, 4a was concluded to be pinosylvin. The constituent 5a gave a triacetate (5b) and from the spectral data of 5a and 5b, 5a was identified as methyl gallate. The constituent 6 gave the molecular formula $C_{14}H_{12}O_3$. The 1 H-NMR spectrum of 6 indicated the presence of a transstyryl group (δ 7.25—7.50 (5H, m), 6.50 (1H, d, J=16 Hz) and 7.42 (1H, J=16 Hz)) and a 4-methoxy-2-oxopyran-6-yl group (δ 3.78 (3H, s), 5.43 (1H, J=2.2 Hz) and 5.90 (1H, J=2.2 Hz)), and 6 was concluded to be dehydrokawain.

Compound A was obtained in two crystalline states, **7a**, mp 185—188°C, and **7b**, mp 207—209°C. The IR spectra (KBr disk) of both crystals were different, but their ¹H-NMR spectra were identical. The fragment pattern of the mass spectrum (MS) of compound A is almost superimposable on that of **6** (Fig. 1) and the elemental analysis data of compound A coincide

Chart 1

well with the molecular formula, $C_{14}H_{12}O_3$, of 6. However, from the ¹H-NMR and carbon nuclear magnetic resonance (13C-NMR) spectra the molecular formula of compound A was confirmed to be C₂₈H₂₄O₆. Compound A gave a dihydro derivative (7c) upon catalytic hydrogenation. From the ¹H-NMR and ¹³C-NMR spectra of compound A and 7c, the structure of compound A was elucidated. The unusual chemical shifts of a methoxy group (δ 3.30) and a methine proton (δ 3.60), which resonated at higher field than another methoxyl group and other methine protons, can be accounted for by anisotropy of the adjacent cis-oriented phenyl group on the cyclobutane ring, as observed in the case of truxinic acid and truxillic acid methyl esters,7) in which the methoxyl group having an adjacent trans-phenyl group resonated at normal field (δ 3.75) but the methoxyl group having an adjacent cis-phenyl group did not (δ 3.23) (Chart 2). These unusual chemical shifts and the coupling constants $(J_{7,8}=9.0, J_{6,7}=11.0 \text{ Hz})$ of methine protons on the cyclobutane ring indicated the relative configuration. Thus, the structure of compound A was concluded to be rel-(1R,6S,7S,8S)-5methoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-2-oxabicyclo[4,2,0]-oct-4-en-3-wethoxy-7-phenyl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-8-(4-methoxy-2-oxopyran-6-yl)-1-(E)-styryl-8-(EThis structure is identical with that of aniba-dimer-A, which has been isolated from Aniba species⁸⁾ and whose structure has been confirmed by X-ray analysis by Gottlieb et al.⁹⁾ They reported that a diastereomer of aniba-dimer-A was synthesized from dehydrokawain by photo-dimerization; its melting point and IR spectrum (KBr disk) were different from those of aniba-dimer-A, but the ¹H-NMR spectrum and MS were identical with those of anibadimer-A, so that the stereo structure of the synthetic dimer was rel-(1R, 6R, 7S, 8S)(7d). Crysatlline 7a and 7b were identical with aniba-dimer-A each other in chloroform solution as judged from the IR spectra. These results indicate that 7a and 7b are the same compound and that aniba-dimer-A and the synthetic dimer are also the same compound, differing only in crystalline state.

Compound B (8a) was obtained as colorless prisms, mp 226—227°C, $[\alpha]_D \pm 0^\circ$ (MeOH), showing the same MS fragment pattern (Fig. 1) and elemental analysis data as 6 and 7a. The ¹H-NMR spectrum of 8a showed the presence of phenyls (δ 7.22 (10H, br s)), a 4-methoxy-2-oxopyran-6-yl moiety (δ 3.68 (6H, s), 5.19 (2H, d, J=2.2 Hz) and 5.81 (2H, J=2.2 Hz)) and two kinds of methine groups (δ 4.28 (2H, m), 4.44 (2H, m)) instead of the *trans* olefinic

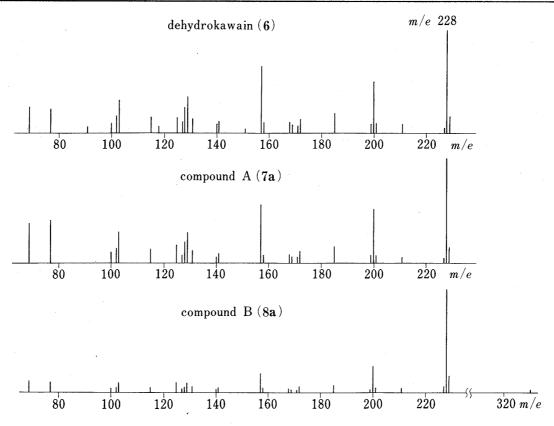
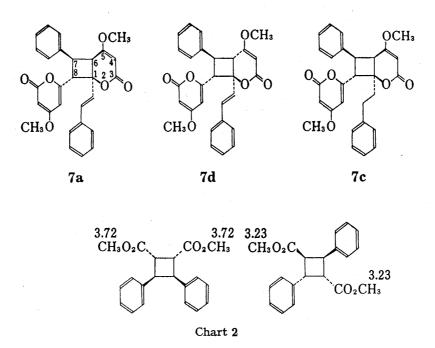


Fig. 1. Mass Spectra of 6, 7a and 8a



group of 6. The presence of the two kinds of methin groups was also supported by the 13 C-NMR spectrum (δ 43.6 (d) and 45.0 (d)) of 8a. Thus, 8a was considered to be a cyclobutane derivative formed by dimerization of dehydrokawain at the styryl olefinic moiety, and so the molecular formular was considered to be $C_{28}H_{24}O_6$, the same as that of 7a. This was confirmed by the fact that the molecular ion peak of 8a could be detected at m/e 456 (below 0.5%) in the enlarged spectrum. Similar cyclobutane derivatives of natural products, truxinic and truxillic acid derivatives, are well known. In such compounds, head-to-head and head-to-tail dimers

may be formed. In the case of the head-to-head dimer, which has been synthesized from 6 by photo dimerization, ¹⁰⁾ three typical fragments at m/e 228, 180 and 276 due to the ions 9a, 9b and 9c, respectively (Fig. 2), appeared. The appearance of a base peak corresponding to the ion 9a and disappearance of fragments 9b and 9c in 8a indicated 8a to be a head-to-tail dimer (Fig. 2). Five possible structures, 8a, 8b. 8c, 8d, and 8e, were considered. The coupling pattern of the methine protons on the cyclobutane ring was expected to be AA'BB' type¹²⁾ in the case of 8a, A_2B_2 type in the cases of 8b and 8c, and a more complex type in the cases of 8d and 8e. In the ¹H-NMR spectrum of compound B (measured at 200 MHz) four methine protons appeared as an AA'BB' type signal consisting of at least sixteen symmetrical peaks. These results indicated that the structure of compound B is rel-, trans-3-bis[6-(4-methoxy-2-pyronyl)]-cis-2, trans-4-diphenyl cyclobutane (8a).

Fig. 2. The Predicted Mass Fragments of Head-to-Tail and Head-to-Head Dimers

Chart 3

Compound C (10a) was obtained as colorless needles, mp 142—143°C, $C_{17}H_{14}O_6$, $[\alpha]_D$ $+29.0^{\circ}$ (MeOH), and gave a monoacetate (10b), mp 98–100°C, $C_{19}H_{16}O_7$. The ¹H-NMR spectrum of 10a showed the presence of a phenyl (δ 7.2—7.6 (5H, m)), a methoxyl (δ 4.07 (3H, s)), a methylenedioxy (δ 5.90 (2H, s)), an alcoholic hydroxyl (δ 3.92 (1H, d, J=2.8 Hz)), two methine groups (δ 4.34 (1H, dd, J = 12.0, 2.8 Hz) and 4.97 (1H, d, J = 12.0 Hz)) and an aromatic proton (δ 6.17 (1H, s)). The ¹H-NMR spectrum of 10b showed a new acetyl signal, and the hydroxy proton was lost. One of the two methine protons was also shifted to lower field (δ 5.65). These observations indicated that 10a is a flavanonol derivative having a methoxyl group and a methylenedioxy group on ring A. It has been reported that the carbon chemical shift of a methoxyl group situated between substituents such as hydroxyl, alkoxyl, acyloxyl acyl and other groups on a benzene ring is shifted to lower field (to near 60 ppm). 13) Since the methoxyl group of 10a resonated at δ 60.4 ppm, the substitution pattern on ring A was considered to be 6,7-methylenedioxy-5-methoxy (10a) or 6,7-methylenedioxy-8-methoxy (10c). In the ¹³C-NMR spectrum of 10a the $s\dot{\rho}^2$ carbon appearing at the highest field (δ 93.1 ppm) showed a doublet peak under off-resonance conditions and was assigned to carbon having O-functional groups on both ortho positions. Thus the structure of compound C was

Chart 4

concluded to be 10a, not 10c. The streochemistry at C-2 and C-3 was determined to be 2R, 3R from the coupling constant (J=12.0 Hz) of the methine protons and the positive optical rotation.¹⁴⁾

Truxinic acid and truxillic acid derivatives are well known as natural dimers having a cyclobutane ring, considered to be formed through [2+2] cycloaddition, and recently other such natural compounds have been reported. These natural compounds are optically inactive in general, and 7a and 8a are also optically inactive. This is the first report of isolation of such dehydrokawain derivatives from Polygonaceae plants.

The coexistence of flavanonols 1a and 10a, stilbene derivative 4a, and styrylpyrone derivatives 6, 7a and 8a, which contain phenyl groups is interesting from a biogenetic standpoint. These compounds may be biosynthesized through a common $C_6H_5-C_3$ unit which is then elongated by the addition of acetate units.

Experimental

All melting points were determined on a Yanagimoto melting point apparatus and are uncorrected. IR spectra were recorded on a JASCO IRA-2 machine, and UV spectra were recorded on a Hitachi model 200-10 spectrometer. Optical rotations were determined on a JASCO DIP-180 automatic polarimeter. ¹H-NMR spectra were recorded on Hitachi R-24B (60 MHz), Hitachi R-22 (90 MHz) and JEOL FX-200 (200 MHz) machines with tetramethylsilane (TMS) as an internal standard (δ value). ¹³C-NMR spectra were recorded on a JEOL FX-100 machine with TMS as an internal standard (δ value). MS were recorded on a JMS-01SG-2 mass spectrometer. Thin layer chromatography (TLC) was carried out on Kieselgel GF₂₅₄ (Merck) and precoated Kieselgel 60F₂₅₄ (Merck), and PLC was carried out on Kieselgel PF₂₅₄ (Merck, 200 × 200 × 0.75 mm). Column chromatography was carried out on Kieselgel type 60 (Merck).

Isolation of Constituents—Aerial parts (fresh 10 kg) of Polygonum nodosum, collected at Shizuoka city, Japan, on August 1980, were extracted with boiling MeOH. The MeOH extract was concentrated under reduced pressure. The residue was suspended in water, and extracted with AcOEt to give 250 g of AcOEt extract. The water layer was extracted with BuOH to give the BuOH extract (100 g) and water layer. The AcOEt extract was chromatographed on a silica gel column with a benzene-AcOEt gradient system as the developer. The resultant eluates were recombined on the basis of their TLC pattern to give seventeen fractions, fr. 1—17. Fr. 10 (21 g) was chromatographed repeatedly on a silica gel column and/or subjected to PLC to give pinobanksin (1a) (70 mg), pinosylvin (4a) (270 mg), dehydrokawain (6) (100 mg), compound A (550 mg) as crystals (7b) and compound C (10a) (2.2 g). Fr. 11 (23 g) was also chromatographed on a silica gel column and/or subjected to PLC to give taxifolin (2a) (120 mg), methyl gallate (5a) (250 mg), compound A (1.2 g) as crystals (7a) and compound B (8a) (700 mg). Fr. 15 gave quercetin-3β-p-glucopyranoside 2″-gallate (3) (4.6 g) upon filtration.

Pinobanksin (1a)—Colorless needles from CHCl₃, mp 171—174°C. [α]_D +6.1 (c=0.45, MeOH). IR ν_{\max}^{KBr} cm⁻¹: 3460, 3120, 1650, 1620, 1480, 1285,1178. ¹H-NMR (δ in CD₃OD): 4.48 (1H, d, J=12.0 Hz, at C-2), 5.03 (1H, d, J=12.0 Hz, at C-3), 5.90 (2H, s, at C-6 and 7), 7.25—7.52 (5H, m, phenyl). ¹³C-NMR (δ in CDCl₃): 72.5 (d), 83.5 (d), 96.0 (d) 96.9 (d), 100.5 (s), 127.6 (d), 128.6 (d), 129.2 (d), 136.5 (s), 163.6 (s), 167.5 (s), 196.0 (s).

Pinobanksin Triacetate (1b)—An amorphous solid. ¹H-NMR (δ in CDCl₃): 1.95 (3H, s, Ac at C-3), 2.25 (3H, s, Ac at C-7), 2.34 (3H, s, Ac at C-5), 5.35 (1H, d, J=12.0 Hz, at C-2), 5.75 (1H, d, J=12.0 Hz, at C-3), 6.54 (1H, d, J=2.0 Hz, at C-8), 6.73 (1H, d, J=2.0 Hz, at C-6), 7.26—7.43 (5H, m, phenyl).

Taxifolin (2a)—Colorless needles from benzene-AcOEt, mp 234—237°C. [α]_D +28.5° (c=0.35, MeOH). IR ν_{\max}^{KBr} cm⁻¹: 3540, 3380, 1630, 1585, 1450, 1362, 1255, 1061. ¹H-NMR (δ in CD₃OD): 4.42 (1H, d, J=12.0 Hz, at C-2), 4.90 (1H, d, J=12.0 Hz, at C-3), 5.84 (2H, s, at C-6 and 8), 6.6—7.0 (3H, m, at C-2', 5')

and 6'). Anal. Calcd for $C_{15}H_{12}O_7$: C, 59.2; H, 3.98. Found: C, 59.17; H, 4.23. MS m/e: 304 (M+) for $C_{15}H_{12}O_7$.

Taxifolin Pentaacetate (2b)——An amorphous solid, ¹H-NMR (δ in CDCl₃): 2.04 (3H, s, Ac at C-3), 2.30 (9H, s, Ac at C-3', 4' and 7), 2.38 (3H, s, Ac at C-5), 5.45 (1H, d, J=12.0 Hz, at C-3), 5.65 (1H, d, J=12.0 Hz, at C-3), 6.55 (1H, d, J=2.0 Hz, at C-8), 6.72 (1H, d, J=2.0 Hz, at C-6), 7.27—7.35 (3H, m, at C-2', 5' and 6').

Quercetin-3 β -n-glucopyranoside 2"-Gallate (3)—Yellow needles from CHCl₃-MeOH, mp 208—210°C, IR ν_{\max}^{NBT} cm⁻¹: 3200, 1710, 1660, 1605, 1235, 1200. ¹H-NMR (δ in CD₃OD): 5.70 (1H, d, J=8.0 Hz, at C-1"), 6.10 (1H, d, J=2.0 Hz, at C-8), 6.25 (1H, d, J=2.0 Hz, at C-6), 6.75 (1H, d, J=9.5 Hz, at C-5'), 7.08 (2H, s, on gallate), 7.42 (1H, dd, J=9.5, 2.0 Hz at C-6'), 7.50 (1H, d, J=2.0 Hz at C-2'). Identical with an authentic sample.

Pinosylvin (4a)—Colorless needles from CHCl₃, mp 156°C, IR $\nu_{\max}^{\text{RB}r}$ cm⁻¹: 3400, 3300, 1610, 1590, 1155. MS m/e: 212 (M+) for C₁₄H₁₂O₂. ¹H-NMR (δ in CDCl₃): 6.21 (1H, t, J=2.0 Hz at C-4), 6.48 (2H, d, J=2.0 Hz, at C-2 and 6), 6.95 (2H, s, at C-7 and 8), 7.10—7.50 (5H, m, phenyl).

Pinosylvin Diacetate (4b)—Colorless needles from MeOH, mp 86—87°C. IR $\nu_{\text{max}}^{\text{RB}}$ cm⁻¹: 1758, 1602, 1575, 1210, 1188. ¹H-NMR (δ in CDCl₃): 2.20 (6H, s, Ac), 6.65 (1H, t, J=2.0 Hz, at C-4), 6.85 (2H, s, at C-7 and 8), 6.94 (2H, d, J=2.0 Hz, at C-2 and 6), 7.05—7.35 (5H, m, phenyl).

Methyl Gallate (5a)—Colorless needles from CHCl₃, mp 200—203°C. IR $\nu_{\text{max}}^{\text{RBr}}$ cm⁻¹: 3480, 3300, 1692, 1618, 1318, 1200, 1140. ¹H-NMR (δ in CD₃OD): 3.78 (3H, s, CH₃O-CO-), 7.00 (2H, s, at C-2 and 3). Identical with an synthetic sample.

Triacetate of 5a (5b)—Colorless needles from MeOH, mp 121°C. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1762, 1710, 1608, 1420, 1320, 1200, 1180, 1160. ¹H-NMR (δ in CDCl₃): 2.25 (9H, s, Ac), 3.83 (3H, s, CH₃O-CO-), 7.65 (2H, s, at C-2 and 6).

5,6-Dehydrokawain (6)—Colorless needles from ethanol, mp 136—138°C. UV $\lambda_{\text{max}}^{\text{MoOH}}$ nm (log ε): 266 (3.84), 320 (3.93), 380 (3.83). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1720, 1628, 1600, 1544, 1400, 1252, 1145. ¹H-NMR (δ in CDCl₃): 3.78 (3H, s, CH₃O), 5.43 (1H, d, J=2.2 Hz, at C-3), 5.93 (1H, d, J=2.2 Hz, at C-5), 6.50 (1H, d, J=16 Hz, at C-8), 7.42 (1H, d, J=16 Hz, at C-7), 7.25—7.50 (5H, m, phenyl).

Compound A (7a)—Colorless needles from AcOEt, mp 185—188°C. [α]_D ±0° (c=0.21, MeOH₃). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 206 (4.77), 251 (4.54), 283 (3.95), 292 (3.88). IR $\lambda_{\max}^{\text{EBT}}$ cm⁻¹: 1720, 1710, 1645, 1625, 1569, 1460, 1445, 1400, 1262, 1250, 1245, 1145. IR $\nu_{\max}^{\text{CRCI}_3}$ cm⁻¹: 1720, 1710, 1643, 1625, 1568, 1455, 1415, 1396, 1256, 1242, 1142. ¹H-NMR (δ in CDCl₃): 3.30 (3H, s, CH₃O), 3.60 (1H, d, J=11.0 Hz, at C-6), 3.70 (3H, s, CH₃O), 4.20 (1H, d, J=9.0 Hz, at C-8), 4.40 (1H, dd, J=11.0 and 9.0 Hz, at C-7), 5.32 (1H, s, at C-4), 5.35 (1H, d, J=2.2 Hz, at C-3'), 5.94 (1H, d, J=2.2 Hz, at C-5'), 6.60 (1H, d, J=16.0 Hz, at C-7"), 7.00 (1H, d, J=16.0 Hz, at C-8"), 7.20—7.50 (10H, m, phenyl×2). ¹³C-NMR (δ in CDCl₃): 39.2 (d), 45.7 (d), 54.4 (d), 55.4 (q), 55.9 (q), 79.4 (s), 88.6 (d), 91.7 (d), 102.6 (d), 124.3 (d), 126.8 (d), 127.5 (d), 127.8 (d), 128.2 (d), 128.4 (d), 128.6 (d), 131.4 (d), 135.5 (s), 135.8 (s), 158.6 (s), 163.8 (s), 164.5 (s), 169.8 (s), 170.4 (s). MS m/e: 228 (M+×1/2). Anal. Calcd for C₂₈H₂₄O₆: C, 73.67; H, 5.30. Found: C, 73.52; H, 5.30. The crystals 7b, mp 207—209°C, IR ν_{\max}^{RBT} cm⁻¹: 1720, 1640, 1570, 1455, 1395, 1263, 1248, 1150, 1102. The crystals, 7a and 7b, gave identical IR spectra in CHCl₃ solution.

Catalytic Hydrogenation of 7a—A solution of 7a (60 mg) in MeOH (12 ml) was shaken with Pd-C (5%) (20 mg) in an H₂ atmosphere for 10 h, then the reaction mixture was filtered. The filtrate was concentrated to give a crystalline product, which was recrystallized from AcOEt to give colorless needles, 7c (35 mg), mp 169—171°C. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1720, 1640, 1570, 1454, 1397, 1261, 1246, 1150. ¹H-NMR (δ in CDCl₃): 2.23 (2H, m, CH₂ at C-8"), 2.76 (2H, m, CH₂ at C-7"), 3.24 (3H, s, CH₃O), 3.28 (1H, d, J=9.0 Hz, at C-6), 3.76 (3H, s, CH₃O), 4.02 (1H, d, J=11.0 Hz, at C-8), 4.13 (1H, dd, J=11.0, 9.0 Hz, at C-7), 5.27 (1H, s at C-4), 5.40 (1H, d, J=2.2 Hz, at C-3'), 5.95 (1H, d, J=2.2 Hz, at C-5'), 7.10—7.45 (10H, m, phenyl×2). ¹³C-NMR (δ in CDCl₃): 29.4 (t), 36.2 (t), 39.2 (d), 43.9 (d), 53.0 (d), 55.2 (q), 55.9 (q), 79.9 (s), 88.7 (d), 91.1 (d), 102.8 (d), 126.2 (d), 127.4 (d), 127.7 (d), 128.3 (d), 128.5 (d), 135.5 (d), 140.5 (s), 158.6 (s), 163.8 (s), 165.1 (s), 170.0 (s), 170.4 (s). Anal. Calcd for C₂₈H₂₆O₆: C, 73.35; H, 5.72. Found: C, 73.70; H, 5.72.

Compound B (8a)—Colorless needles from AcOEt, mp 226—227°C. [α]_D ±0° (C=0.39,MeOH). UV $\lambda_{\max}^{\text{MOOH}}$ nm (log ϵ): 207 (4.74), 287 (4.13). IR ν_{\max}^{RBr} cm⁻¹: 1720, 1640, 1560, 1447, 1400, 1253, 1132. ¹H-NMR (δ in CDCl₃): 3.67 (6H, s, CH₃O×2), 4.28 (2H, m, CH×2 on cyclobutane ring), 4.44 (2H, m, CH×2 on cyclobutane), 5.19 (2H, d, J=2.2 Hz, at C-3′ and C-3″), 5.74 (2H, d, J=2.2 Hz, at C-5′ and C-5″), 7.20—7.40 (10H, m, phenyl×2). ¹³C-NMR (δ in CDCl₃): 43.6 (d), 45.0 (d), 55.7 (q), 87.7 (d), 101.4 (d), 127.1 (d), 127.3 (d), 128.5 (d), 137.3 (s), 162.6 (s), 163.8 (s), 170.4 (s). MS m/ϵ : 456 (M+) (0.5% below), 228 (M+×1/2) (100%). Anal. Calcd for $C_{28}H_{24}O_6$: C, 73.67; H, 5.30. Found: C, 73.41; H, 5.30.

Compound C (10a)—Colorless needles from AcOEt, mp 142—143°C. [α]_D+29.0° (c=0.5, MeOH). UV $\lambda_{\max}^{\text{Nujol}}$ nm (log ε): 208 (4.28), 243 (4.16), 283 (3.95), 341 (3.50). IR ν_{\max}^{EBT} cm⁻¹: 3520, 3480, 1668, 1620, 1610, 1498, 1475, 1440, 1243, 1162, 1115, 1092. ¹H-NMR (δ in CDCl₃): 3.96 (1H, d, J=2.0 Hz, OH), 4.34 (1H, dd, J=12.0 and 2.0 Hz, at C-3), 4.97 (1H, d, J=12.0 Hz, at C-2), 4.09 (3H, s, CH₃O), 5.90 (2H, s, O-CH₂-O), 6.17 (1H, s, at C-8), 7.20—7.60 (5H, m, phenyl). ¹³C-NMR (δ in CDCl₃): 60.4 (q), 72.8 (d), 83.4 (d), 93.1 (d), 101.7 (t), 105.3 (s), 127.4 (d), 128.6 (d), 129.1 (d), 131.3 (s), 136.3 (s), 142.7 (s), 155.6 (s), 160.3 (s), 191.2 (s). MS m/e: 314 (M⁺). Anal. Calcd for C₁₇H₁₄O₆: C, 64.77; H, 4.52. Found: C, 64.96; H, 4.49.

Acetate (10b) of 10a—10a (400 mg) was acetylated with Ac₂O-Pyr at room temperature and the reaction mixture was poured into ice-water. The resulting precipitates were filtered off and recrystallized from MeOH to give colorless needles (10b) (300 mg), mp 98—100°C. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 1750, 1685, 1620, 1480, 1225. ¹H-NMR (δ in CDCl₃): 2.00 (3H, s, CH₃CO), 4.05 (3H, s, CH₃O), 5.25 (1H, d, J=12.0 Hz, at C-2), 5.63 (1H, d, J=12.0 Hz, at C-3), 5.89 (2H, s, O-CH₂-O), 6.18 (1H, s, at C-8), 7.20—7.50 (5H, m, phenyl). MS m/e: 356 (M+). Anal. Calcd for C₁₉H₁₆O₆: C, 64.04; H, 4.53. Found: C, 63.87; H, 4.65.

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