(Chem. Pharm. Bull.) 30(8)2971—2979(1982)

A Proposed Method for the Prediction of Stability based on Actual Field Temperatures

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(Received January 25, 1982)

The present paper is concerned with an application of the annual atmospheric temperature distribution to the shelf life prediction of pharmaceutical preparations in distribution channels. The method attempts to correct for the relatively wide variation in storage temperature by utilizing actually recorded thermal histories in the respected market. The calculated storage breakdown rates obtained by this method have been found to correlate well with actual experience.

Keywords—stability study; shelf life prediction; stability of pharmaceutical preparation; atmospheric temperature distribution; stability prediction temperature

Reliable and practical methods for stability prediction are important for the rational formulation and the quality assurance of pharmaceutical preparations during extended storage in distribution channels. Kinetic methods were introduced in this field,¹⁻³⁾ and several reports on such methods have been presented.⁴⁾ Several authors, Tootill,⁵⁾ Rogers,⁶⁾ Kennon,⁷⁾ Lordi and Scott,⁸⁾ and Amirjahed,⁹⁾ among others, have reported practical kinetic approaches. The room temperature used in their predictions, however, was frequently only 25°C and the conditions of real distribution channels were not taken into account. Temperature consideration should not be limited to a fixed value such as 25°C. It is necessary to determine what room temperature variation actually is in order to increase the accuracy of prediction.

The ultimate solution to the problem of variation in ambient storage temperature is to use the real temperatures in predicting shelf life. For this purpose, the temperature used for prediction should not, of course, be merely the arithmetical average temperature.

The problem, then, is how to use a field temperature distribution adequately. Several

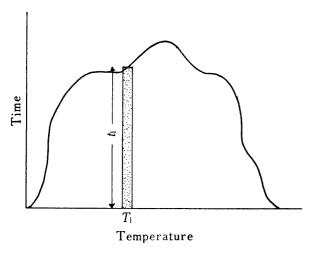


Fig. 1. A Model Atmospheric Temperature Distribution Chart

attempts have been made in this direction. Terao presented a method for stability prediction utilizing atmospheric temperature distribution and predicted the stability of preservatives^{10a)} and injections^{10b)} through this method. Egawa¹¹⁾ presented a method to obtain a kinetically average temperature from atmospheric tempera-In this method, however, the highest, the lowest and the annual arithmetical average values of the temperature were simply used for the calculation. Okusa¹²⁾ reported a novel prediction method of stability, "the multilevel nonisothermal method," and a graphical calculation was employed in it. Haynes¹³⁾

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proposed a method for calculating "the virtual temperature," which aids in the choice of a standard temperature for expiration dating. In these studies, the monthly arithmetical average temperatures were used for stability prediction.

In the present study, all of the temperatures and their durations in an actual field temperature distribution are exactly utilized. Therefore, the stability data predicted by this method are very accurate and compare favorably with the results of actual market storage.

Theory

Fig. 1 is a model chart of annual temperature distribution. If the concentration of a drug stored at a temperature T_i for a time t_i is represented by C_i (initial concentration $C_i=1$), and if the degradation rate of the drug is first order, then,

$$\log C_1 = -1/2.303 \, k_i t_1 \tag{1}$$

where k_i is the rate constant at the temperature T_i .

The concentration, C, after storage under all of the conditions (at every temperature and time of the temperature distribution) is given by Eq. 2:

$$\log C = \Sigma \log C_{\rm i} \tag{2}$$

Combination of Eqs. (1) and (2) gives Eq. (3):

$$\log C = -1/2.303 \ \Sigma \ k_i t_i \tag{3}$$

Therefore, when the annual temperature distribution at a market and the rate constant at each temperature of the temperature distribution are given, the concentration of a drug after storage for one year at the market can be estimated by means of Eq. (3). In this case, the rate order has been chosen to be first order, is since the majority of preparations in our stability studies showed first-order degradation.

The concentration of a drug after m years storage can be estimated from Eq. (4):

$$\log C_m = -1/2.303 \, m\Sigma \, k_i t_i \tag{4}$$

If k_p is the rate constant at a certain constant temperature, T_p , which gives the same concentration, C, after time t_p ($t_p = \sum t_i$), then Eq. (5) can be derived:

$$-1/2.303 \Sigma k_i t_i = -1/2.303 k_p t_p \tag{5}$$

Eq. (5) is further simplified to Eq. (6):

$$\Sigma k_{\rm i} \gamma_{\rm i} = k_{\rm p} \tag{6}$$

where,

$$\gamma_{\rm i} = t_{\rm i}/t_{\rm p}$$

If the Arrhenius equation, $\log k = \log A - E_a/4.574T$, is written in exponential form, $k = A \cdot 10^{-ax} (X = 1/T \times 10^3, a = E_a(\text{kcal/mol})/4.574)$, Eq. (7) can be obtained from Eq. (6):

$$\Sigma(A \cdot 10^{aXi} \cdot \gamma_i) = A \cdot 10^{-aX} p \tag{7}$$

When the logarithmic value of Eq. (7) is taken and rearranged, Eq. (8) results:

$$X_{\rm p} = \frac{\log \Sigma \, 10^{-aXi} \cdot \gamma_{\rm i}}{-a} \tag{8}$$

Thus, the stability prediction temperature counterpart, activation energy, and time ratio can be expressed in terms of (X_p) , (a) and (γ_i) respectively. X_p corresponds to the room temperature counterpart generally used for the prediction of stability.

On the other hand, the accelerated test condition causing the same degradation as that under the atmospheric temperature distribution can also be estimated from Eq. (8).

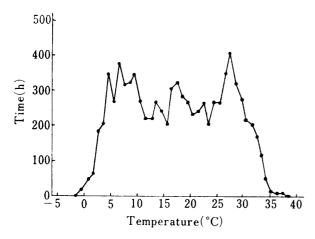


Fig. 2. Annual Atmospheric Temperature Distribution Chart measured at a Storehouse in Osaka

The time (t_a) at the accelerated temperature counterpart (X_a) which causes the same degradation as that under condition (X_p, t_p) is given by Eq. (9):

$$t_{\rm a} = 10^{a(X_{\rm a} - X_{\rm p})} \cdot t_{\rm p} \tag{9}$$

Experimental

Annual Atmospheric Temperature Distribution Chart—The annual atmospheric temperature distribution charts at a woody plane warehouse and a reinforced concrete workshop in Osaka are shown in Figs. 2 and 3, respectively. The temperature was measured by automatic temperature recorders (Yokogawa Electric Work and Ota Keiki). Table I shows the annual

TABLE I. Annual Atmospheric Temperature Distribution in Osaka

Temperature range (°C)	Mean of temperature range (°C)	Cumulative time per year (h)
-4.04.9	-4.5	3
-3.03.9	-4.5	9
-2.0 - 2.9	-2.5	18
-1.01.9	-1.5	48
-0.10.9	-0.5	81
0.00.9	0.5	99
1.0— 1.9	1.5	219
2.0-2.9	2.5	195
3.0— 3.9	3.5	208
4.0-4.9	4.5	195
5.0— 5.9	5.5	237
6.0 - 6.9	6.5	222
7.0— 7.9	7.5	213
8.0— 8.9	8.5	258
9.09.9	9.5	213
10.0— 10.9	10.5	252
11.0 11.9	11.5	276
12.0 - 12.9	12.5	309
13.0 13.9	13.5	291
14.0— 14.9	14.5	279
15.0— 15.9	15.5	360
16.0-16.9	16.5	282
17.0— 17.9	17.5	249
18.0— 18.9	18.5	342
19.0— 19.9	19.5	309
20.0— 20.9	20.5	330
21.0— 21.9	21.5	372
22.0-22.9	22.5	306
23.0— 23.9	23.5	321
24.0— 24.9	24.5	372
25.0 - 25.9	25.5	339
26.0 - 26.9	26.5	321
27.0— 27.9	27.5	321
28.0— 28.9	28.5	273
29.0— 29.9	29.5	180
30.0 - 30.9 $31.0 - 31.9$	$\frac{30.5}{31.5}$	159 111
32.0 32.9	32.5	96
33.0— 33.9	33.5	63
34.0 - 34.9	34.5	$\frac{33}{24}$
35.0 - 35.9	35.5	6

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atmospheric temperature distribution in Osaka (1977). Fig. 4 shows the relationship between the annual atmospheric temperature distribution chart (A, 1977, Osaka) which was determined at three-hour intervals, equivalent to Table I, and the daily average annual atmospheric temperature distribution chart (B, 1941—1970, Osaka). Table II is the annual atmospheric temperature distribution in Bangkok (1969) showing a typical tropical climate. Table I, ¹⁵⁾ Fig. 4(A), ¹⁵⁾ Fig. 4(B) ¹⁶⁾ and Table II ¹⁷⁾ were prepared from meteorological observatory data in each country.

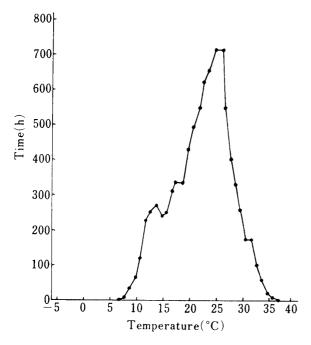


Fig. 3. Annual Atmospheric Temperature Distribution Chart measured at a Workshop in Osaka

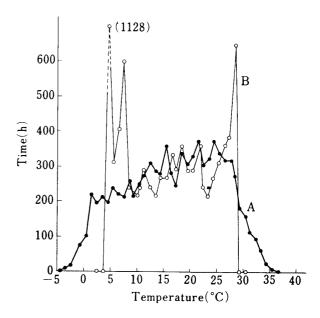


Fig. 4. Annual Atmospheric Temperature Distribution Chart derived from the Data of the Osaka Meteorological Observatory

A: cumulative curve of the annual temperature-time data measured at three-hour intervals (1977).

B: cumulative curve of the annual, daily averaged, temperature data (1941—1970).

Table II. Annual Atmospheric Temperature Distribution in Bangkok

Temperature range $(^{\circ}C)$	Mean of temperature range ($^{\circ}$ C)	Cumulative time per year (h)
15.0—15.9	15.5	3
16.0-16.9	16.5	13
17.0-17.9	17.5	22
18.0-18.9	18.5	56
19.0-19.9	19.5	63
20.0-20.9	20.5	85
21.0-21.9	21.5	100
22.0-22.9	22.5	128
23.0-23.9	23.5	267
24.0 - 24.9	24.5	451
25.0 - 25.9	25.5	940
26.0-26.9	26.5	1129
27.0-27.9	27.5	1097
28.0 - 28.9	28.5	1031
29.0-29.9	29.5	850
30.0-30.9	30.5	688
31.0—31.9	31.5	643
32.0-32.9	32.5	544
33.0-33.9	33.5	365
34.0-34.9	34.5	208
35.0-35.9	35.5	66
36.0-36.9	36.5	116

TABLE III. Distribution of the Activation Energy Counterpart of Common Drug Compounds

Compound	Activation energy counterpart	
Acetaminophen ²⁰⁾	3.7(17)	
Actinospectacin ²¹⁾	4.4(20)	
Ampicillin ²²⁾	3.6(16), 4.0(18), 4.9(22)	
Adenine monophosphate ²³⁾	6.4(20)	
Ascorbic acid ²⁴⁾	4.2(19), 5.5(25), 5.3(24), 5.0(23), 3.5(16) 5.0(23), 4.6(21)	
Aspirin ²⁵⁾	3.7(17), 316(16), 3.9(18), 2.7(13), 3.3(15)	
Atropine ²⁶⁾	2.8(13), 3.0(14), 5.5(25), 3.8(17), 2.8(13) 4.6(21), 2.8(13)	
Barbital ²⁷⁾	5.1(24), 2.4(11), 3.3(15), 3.5(16)	
Benzocaine ²⁸⁾	2.8(13), 4.0(19), 4.2(19)	
Benzyl penicillin ²⁹⁾	3.9(18), 4.6(21), 5.0(23)	
Cephalothin ³⁰⁾	5.0(24), 3.4(15)	
Chloramphenicol ³¹⁾	5.3(24), 4.4(20), 4.6(21), 5.0(23), 5.3(24) 7.7(35)	
Chlordiazepoxide ³²⁾		
Chlorobutanol ³³⁾	5.0—5.5(23—25), 6.2(28)	
Clindamycin ³⁴)	4.2(19), 5.6(26) 8.3(38), 6.4(20), 7.2(22)	
Diazepam ³⁵⁾	8.3(38), 6.4(29), 7.2(33)	
	4.3(20), 4.0(18), 5.0(23)	
Epinephrine ³⁶⁾	5.0(23)	
Ethylparaben ³⁷⁾	4.1(19)	
Filipin 8)	2.0(9)	
5-Fluorouracil ³⁹⁾	5.3(24)	
Folic acid ²⁴⁾	3.7(17), 5.9(27), 5.1(23)	
Fumagillin ²⁴⁾	2.0(9), 3.7(17)	
Glucose ⁴⁰⁾	6.8(31), 7.0(32)	
Homatropine ^{26,41)}	2.4(11), 2.6(12), 2.8(13)	
Hydrochlorothiazide ⁴²⁾	5.5(25), 6.7(31)	
Hydroco tisone ^{24,43)}	1.5(7), 3.7(17), 4.4(20)	
5-Iodo-2'-deoxyuridine44)	6.7(30), $6.8(31)$, $5.9(27)$, $3.9(18)$	
Isoamylnitrate ⁴⁵⁾	4.4(20)	
Lincomycin ⁴⁶⁾	7.0(32), 8.3(38)	
Methicillin ⁴⁷⁾	3.8(17), 4.0(18)	
Methylparaben ³⁶⁾	4.5(20), 4.4(20), 3.3(15), 5.3(24), 4.6(21), 5.6(26)	
Methylphenidate ⁴⁸⁾	3.3(15), 5.5(25)	
Methylprednisolone ⁴⁹⁾	3.1(14)	
Morphine ⁵⁰⁾	5.0(23)	
Naphazoline ⁵¹⁾	1.1(5), 1.3(6)	
2-PAM ⁵²⁾	3.7(17), 6.3(29)	
Pantothenyl alcohol ²⁴⁾	4.4(20), 4.6(21)	
Phenethicillin ⁵³⁾	3.9(18)	
Phenobarbital ⁵⁴⁾	5.6(26), 5.4(25)	
Pilocarpine ⁵⁵⁾	5.5(25)	
Prednisolone ⁵⁶⁾	2.4(11)	
Procainamide ²⁸⁾	2.8(13)	
Procaine ^{28,57)}	3.7(17), 2.6(12), 3.0(14)	
Propylparaben ⁵⁸⁾	4.3(20), 4.4(20), 2.8(13), 4.2(19)	
Riboflavin ⁵⁹⁾	4.4(20)	
Salicylalcohol ⁶⁰⁾	5.3(24)	
Streptozotocin ²⁴⁾	4.6(21), 5.9(27)	
Sulfacetamide ⁶¹⁾	5.0(23)	
Thiamine ^{2,24,62,63)}	2.8(13), 4.6(21), 5.3(24), 5.5(25), 5.7(26),	
	6.3(29), 6.8(31), 6.3(29), 6.0(27), 6.7(31)	
Thiamine propyl disulfide ⁶⁴⁾	5.0(23)	
Vitamin A ²⁴⁾	3.3(15), 5.0(23)	
	\ // \ \ =/	

Note: Numbers in parenthesis are approximate values of activation energy, kcal/mol.

Calculation of Stability Prediction Temperature Counterpart (X_p) ——An IBM 370 computer was used for the computation of Eq. (8).

Activation Energy Counterpart (a)——The activation energy counterpart of drug compounds and some adjuvants under various conditions relevant to pharmaceutical preparations were surveyed and the results are shown in Table III. $^{20-64}$)

Results and Discussion

Stability Prediction Temperature

As shown in Table III, the values of activation energy counterpart (a) of the compounds related to pharmaceutical preparations mostly fall in the range of 2—8 (ca. 10—35 kcal/mol as activation energy). Moreover, the majority of compounds represented in Table III can be

practically treated as having first-order degradation rates. Therefore, Eq. (8) is applicable in almost all cases and the calculation of Eq. (8) can be performed in the range of a=2-8. The values of prediction temperature counterpart, $X_{\rm p}$, solved by computer for the atmospheric temperature distribution charts of Figs. 2 and 3 are shown by curves A and B in Fig. 5, respectively.

The values of prediction temperature change depending on the nature of preparations, *i.e.*, the values of activation energy: the lower the values of a, the higher the values of X_p . It has been made clear that the room temperature used for the prediction of stability should not be limited to one temperature only (say 25° C), but instead it should be corrected for the values of activation energy and the shape of the temperature distribution charts.

From Fig. 5 it is readily seen that the wider the range of atmospheric temperature distribution, the wider the range of prediction temperature.

The range of prediction temperature in Osaka for various preparations is regarded as 20--25°C, which varies with the nature of the preparation. It should be reasonable for stability

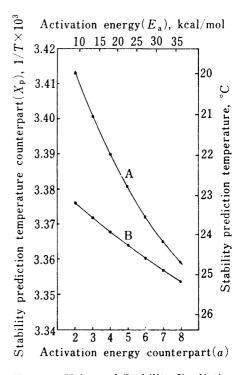


Fig. 5. Values of Stability Prediction Temperature Counterpart (X_p) for the Temperature Distribution Chart of Fig. 2(A) and 3(B)

prediction in the domestic market to utilize the atmospheric temperature of Osaka, because the annual average temperature of Osaka (15.6°C, 1941—1970)¹⁷⁾ is the highest among the major cities in Japan (Tokyo, 15.0°C; Yokohama, 14.8°C; Nagoya, 14.4°C; Kyoto, 14.8°C; Kobe, 15.5°C, 1941—1970)¹⁷⁾ where large amounts of medical preparations are consumed.

As a basic parameter for stability prediction, the atmospheric temperature in some building in which the pharmaceutical preparations are handled or stored may be suitable. The atmospheric temperature data measured at meteorological observatories may be most feasible. In this case, though, the average temperature, which has an apparently biased distribution, e.g., the curve B of Fig. 4, compared to the actual distribution (curve A), should not be applied to our purpose.

Prediction of Stability

(1) Prediction of Retention Percent after One Year of Storage—The method of prediction is illustrated by an example of thiamine in a B-complex with C injection. The Arrhenius

equation obtained from the accelerated stability test was as follows:

log
$$k(h^{-1}) = -5.212X + 12.95$$

 $(N=4,^{19}) X=1/T \times 10^3, \sigma_{\log k} = 1.01 \times 10^{-2})$

The prediction of stability was conducted on the basis that the injection was stored in a storehouse (Fig. 2) for one year.

From Fig. 6, the value of X_p for a=5.212 could be read as 3.364. $X_p=3.364$ was inserted into X of the above Arrhenius equation to obtain $k=2.2624\times 10^{-5}\ h^{-1}$.

The concentration, C, remaining after storage for one year could now be caculated from Eq. (3) (t=8760 h, C_o =1), i.e., $\log C = -1/2.303 \sum k_i t_i$, C=0.813. The retention rate of thiamine in the injection when stored for one year at the storehouse of Fig. 2 was therefore predicted to be 81.3%. The 95% confidence interval of the retention rate was in the range of 79.3–82.9%. The actual retention rate of thiamine in the injection observed after one

TABLE IV. Predicted Stability of Epinephrine Injection^{a)} at Domestic (Osaka) and Tropical (Bangkok) Atmospheric Temperatures

Storage period (Year)	Calculated retention percent of epinephrine ^{b)}	
	Osaka	Bangkok
1	90.8	78.0
2	82.6	60.8
3	75.0	47.3

a) Epinephrine injection: L-epinephrine hydrochloride 0.1%, sodium bisulfite 0.1%, sodium chloride 0.9%, pH=5.0 (upper limit of pH specified in JP IX).

$$\log k = 11.75 - \frac{22450}{4.574 T}$$

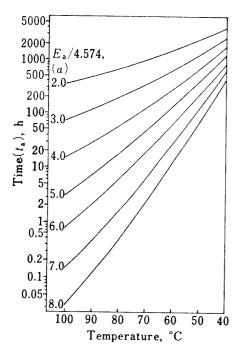


Fig. 6. Accelerated Stability Test Conditions determined by the Use of Eq. (9) and the Temperature Distribution Chart of Fig. 3

year in the storehouse was shown to be 81.0% (average of three lots.) Thus the calculated results obtained by our method compare satisfactorily with the data obtained from actual storage.

Monthly or seasonal stability prediction could also be possible by using the corresponding temperature distribution.

(2) Comparison of Stability in Different Territories—The stability of epinephrine injection containing sodium bisulfite as an antioxidant was estimated. It has already been found in an accelerated stability study that bisulfite has a destructive effect upon epinephrine⁶⁵⁾ but the long-term stability at ambient temperature of the injection is not clear.

The stability of the epinephrine injection was predicted by this method employing the temperature distributions of Tables I and II for the domestic market and tropical market, respectively. As shown in Table IV, the shelf life of the injection was estimated to be about one year in the domestic market, but it was not suitable for storage at room temperature in the

b) Arrhenius equation for the degradation of epinephrine in the injection obtained by accelerated stability testing:

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tropical market.

(3) Design of Accelerated Stability Test Corresponding to the Actual Storage Conditions—

The accelerated test conditions could be determined by the use of Eq. (9) and the temperature distribution of Fig. 3. The accelerated test conditions equivalent to the actual storage conditions are shown in Fig. 6, and appropriate test conditions could thus be selected.

Acknowledgement The authors are very grateful to Dr. T. Higuchi for his critical review of this manuscript.

References and Notes

- 1) A. Watanabe, Yakugaku Zassi, 59, 218 (1939).
- 2) E.R. Garrett, J. Am. Pharm. Assoc., Sci. Ed., 45, 470 (1956).
- 3) E.R. Garrett, J. Pharm. Sci., 51, 811 (1962).
- 4) J.A. Mollica, S. Ahuja, and J. Chohen, J. Pharm. Sci., 67, 443 (1978).
- 5) J.P.R. Tootill, J. Pharm. Pharmacol. 13, 75T (1961).
- 6) A.R. Rogers, J. Pharm. Pharmacol., 15, 101T (1963).
- 7) L. Kennon, J. Pharm. Sci., 53, 815 (1964).
- 8) G. Lordi and M.W. Scott, J. Pharm. Sci., 54, 531 (1965).
- 9) A.K. Amirjahed, J. Pharm. Sci., 66, 785 (1977).
- 10) M. Terao, a) the 12th Annual Meeting of the Pharmaceutical Society of Japan, Nagoya, April 1958; b) the 19th Annual Meeting of the Pharmaceutical Society of Japan, Tokyo, April 1964.
- 11) S. Egawa, the 11th Meeting of the Kinki Branch, Pharmaceutical Society of Japan, Kobe, October 1961.
- 12) N. Okusa, Chem. Pharm. Bull., 23, 794, 803 (1975).
- 13) J.D. Haynes, J. Pharm. Sci., 60, 927 (1971).
- 14) In cases where the rate order is zero or second order, $\log C = \sum \log (1 k_i t_i)$ or $\log C = \sum \log 1/k_i t_i + 1$, respectively.
- 15) Osaka Meteorological Observatory, unpublished data, 1977.
- 16) The Japan Meteorological Agency (ed.), "Climate Table of Japan," part 2, 1971.
- 17) Meteorological Department, Thailand, Monthly Meteorological Bulletin, 34, part 1, 1969.
- 18) The composition was as follows: thiamine hydrochloride 0.5%, riboflavin sodium phosphate 0.025%, niacinamide 1.0%, pyridoxine hydrochloride 0.1%, ascorbic acid 2.5% and benzyl alcohol 1.0%. This was chosen as an example because thiamine in this solution (pH=5) is rather unstable.
- 19) It is convenient for calculation of the Arrhenius equation to equalize the intervals of 1/T as 40.0, 49.7, 60.0 and 71.0°C.
- 20) K.T. Kosy and J.L. Lach, J. Pharm. Sci., 50, 113 (1961).
- 21) E.R. Garrett and G.R. Umbreit, J. Pharm. Sci., 51, 436 (1962).
- 22) J.P. Hou and J.W. Poole, J. Pharm. Sci., 58, 1510 (1969).
- 23) Y. Matsukura, unpublished observation.
- 24) E.R. Garrett, J. Pharm. Sci., 43, 539 (1954); 45, 171, 470 (1956); 49, 767 (1960); 51, 445 (1962).
- 25) E.R. Garrett, J. Am. Chem. Soc., 79, 3401 (1957); L.J. Leeson and A.M. Mattocks, J. Pharm. Sci., 47, 329 (1958); S.M. Blsug and J.W. Wesolowski, J. Pharm. Sci., 48, 691 (1959).
- P. Zvirblis, I. Socholitrky, and A.A. Kondritzer, J. Pharm. Sci., 45, 450 (1956); J. L. Patel and A.P. Lemberger, J. Pharm. Sci., 48, 106 (1956).
- 27) J.E. Goyan, Z.I. Shaikh, and J. Autian, J. Pharm. Sci., 49, 627 (1960).
- 28) A.D. Marcus and S. Baron, J. Pharm. Sci., 48, 85 (1959).
- R. Broderson, Trans. Faraday Soc., 43, 351 (1947); Acta Chem. Scand., 1, 403 (1947); Acta Pharmacol.,
 3, 345 (1947); J.M. Blaha, A.M. Knevel, D.P. Kessler, J.W. Mincy and S.L. Hem, J. Pharm. Sci., 65, 1165 (1976).
- 30) T. Yamana and A. Tsuji, J. Pharm. Sci., 65, 1563 (1976).
- 31) T. Higuchi and C.D. Bias, J. Pharm. Sci., 42, 707 (1953); T. Higuchi and A.D. Marcus, ibid., 43, 129, 530 (1954); A.S. Marcus and A.J. Taraszka, ibid., 48, 77 (1959).
- 32) A.D. Nair and J.L. Lach, J. Pharm. Sci., 48, 390 (1959); W.W. Han, G.J. Yakatan and D.D. Maness, ibid., 65, 1198 (1976).
- 33) K. Aoki, unpublished observation.
- 34) T.O. Oesterling and E.L. Rowe, J. Pharm. Sci., 59, 63, 75 (1970).
- 35) W. Mayer, S. Erbe, G. Wolf, and R. Voight, Pharmazie, 29, 700 (1974).
- 36) L.C. Schroeter and T. Higuchi, J. Pharm. Sci., 47, 426 (1958).
- 37) S.M. Blaug and D.E. Grant, J. Soc. Cosmet. Chem., 25, 495 (1974).
- 38) J.E. Tingstad and E.R. Garrett, J. Pharm. Sci., 49, 352 (1960).
- 39) E.R. Garrett, H.J. Nestler, and A. Somodi, J. Org. Chem., 33, 3460 (1968).

- N.E. Webb, G.J. Sperandio, and A.N. Martin, J. Pharm. Sci., 47, 101 (1958); K.R. Heimlich and A.N. Martin, ibid., 49, 592 (1960).
- 41) J.L. Patel and A.P. Lemberger, J. Pharm., 47, 878 (1958).
- 42) J.A. Mollica, C.R. Rehm, J.B. Smith and H.K. Govan, J. Pharm. Sci., 60, 1380 (1971).
- 43) A.D. Marcus, J. Pharm. Sci., 49, 383 (1960).
- 44) E.R. Garrett, P.B. Chemburkar, and T. Suzuki, Chem. Pharm. Bull., 13, 1113 (1965).
- 45) M.H. Tunker, D. Szulczewski, and T. Higuchi, J. Pharm. Sci., 47, 613 (1958).
- 46) T.O. Oesterling, J. Pharm. Sci., 59, 63 (1970).
- 47) M.A. Schwartz, E. Bara, I. Rabyez and A.P. Granatek, J. Pharm. Sci., 54, 149 (1965).
- 48) S. Siegel, L. Lachman, and L. Malspeis, J. Pharm. Sci., 48, 431 (1959).
- 49) E.R. Garrett and M.E. Royer, J. Pharm. Sci., 51, 451 (1962).
- 50) S. Yeh, J.L. Lach, J. Pharm. Sci., 50, 35 (1960).
- 51) M.J. Stern, L.D. King, and A.D. Marcus, J. Pharm. Sci., 48, 641 (1959).
- 52) R.I. Ellin, J.S. Carlese, and A.A. Kondritzer, J. Pharm. Sci., 51, 141 (1962).
- 53) M.A. Schwartz, A.P. Granatek and F.H. Buckwalter, J. Pharm. Sci., 51, 523 (1962).
- 54) J. Hasegawa, K. Ikeda, and T. Matsuzawa, Chem. Pharm. Bull., 6, 36 (1958).
- 55) M.A. Nunes and E. Brochmann-Hanssen, J. Pharm. Sci., 63, 717 (1974).
- 56) E.R. Garrett and M.E. Royer, J. Pharm. Sci., 51, 451 (1962).
- 57) T. Higuchi, A. Havinga, and L.W. Busse, J. Pharm. Sci., 39, 405 (1950).
- 58) A. Kamada, N. Yata, K. Kubo, and M. Arakawa, Chem. Pharm. Bull., 21, 2073 (1973).
- 59) D.E. Guttman, J. Pharm. Sci., 51, 1162 (1962).
- 60) L.C. Schroeter, J. Pharm. Sci., 51, 258 (1962).
- 61) B.J. Meakin, I.P. Tansey, and D.J.G. Davies, J. Pharm. Pharmacol., 23, 252 (1971).
- 62) J.J. Windheuser and T. Higuchi, J. Pharm. Sci., 51, 354 (1962).
- 63) K. Aoki, unpublished observation.
- 64) M. Terao, unpublished observation.
- 65) L.C. Schroeter, T. Higuchi, and E. Schuler, J. Am. Pharm. Assoc., Sci. Ed., 47, 723 (1958).