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Electrochemical Oxidation of Chlorpromazine-Cyclodextrin Inclusion Complex

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The electrochemical oxidation of a chlorpromazine (CPZ)-cyclodextrin (CyD) inclusion complex was studied in phosphate buffers by means of cyclic voltammetry.

The voltammogram of the CPZ-CyD system exhibits three oxidation waves. The first wave, which is due to the oxidation of CPZ to form its cation radical, is catalytic in nature, and its peak current depends on pH and on the buffer concentration. The second wave is attributed to the oxidation of CPZ cation radical-phosphate adduct. The third wave appears to be the result of further oxidation of CPZ sulfoxide-CyD complex.

The formation of an inclusion complex of CPZ with CyD leads to a lowering of the peak current of the first wave and a shift in the peak potential to the positive side at the same time. The presence of CyD exerts no appreciable effect on the second wave, suggesting a minor interaction between the radical and CyD, and this is consistent with the absorption, circular dichroism, and ¹H-nuclear magnetic resonance (NMR) spectra.

Based on these results, an electrochemical reaction mechanism for CPZ-CyD system is proposed.

Keywords—chlorpromazine-cyclodextrin inclusion complex; electrochemical oxidation; electrode reaction; glassy carbon electrode; voltammetry

Phenothiazine-based major tranquilizers such as chlorpromazine (CPZ) are very widely used as antipsychotic drugs, and much attention has been directed to their undesirable side effects, such as drug-induced hemolysis.¹⁾ To avoid such effects as well as to protect the drugs from photodecomposition, combination with various components has been investigated.^{1,2)}

Recently, cyclodextrins (CyDs), cyclic- α -1,4-D-glucose oligomers which form host–guest inclusion complexes with CPZ, have been found promising for alleviating the adverse side effects of CPZ. Fortunately, CyDs do not lessen the therapeutic effects of CPZ on the central nervous system of the rat and are not toxic to the human body. However, a detailed explanation has not yet been presented for the effectiveness of CyDs in alleviating the defects of CPZ.

Chlorpromazine is easily oxidized electrochemically,^{5,6)} photochemically,⁷⁾ and enzymatically⁸⁾ to form CPZ sulfoxide (CPZO) via the CPZ cation radical (CPZ·⁺). It has often been pointed out that phenothiazine cation radicals may be active pharmacological entities⁸⁻¹²⁾ and their oxidation potentials are correlated to clinical activity.¹³⁾ Thus, we considered it worthwhile to observe the oxidation potentials of CPZ and its cation radical in the presence of CyDs. If the electrode surface is regarded as a simple model of the receptor surface, the electrode reaction mechanism of CPZ-CyD systems may be suggestive of the interaction of CPZ with CyDs at the receptor site.

In this paper, an attempt has been made to elucidate the roles of CyDs in the overall electrochemical oxidation processes of CPZ.

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Experimental

Apparatus—Cyclic voltammetry and controlled potential electrolysis were carried out using an HA-101 potentiostat (Hokuto Denko, Ltd.) and an HA-107A function generator (Hokuto Denko, Ltd.). A glassy carbon rod polished with alumina powder $(0.3 \,\mu\text{m})$ was used as a working electrode. Before each measurement, the electrolytic solution was deaerated with purified nitrogen gas. During measurement, the solutions were shielded from light and air, and the temperature was kept at $25 \pm 1\,^{\circ}\text{C}$.

Absorption and circular dichroism (CD) spectra were recorded on a Hitachi model 200-10 spectrometer and a JASCO J-500C spectropolarimeter, respectively. Proton nuclear magnetic resonance (¹H-NMR) spectra were obtained on a JEOL PS-100 machine.

Electroconductivity was measured with a model CM-2A instrument (Toa Electronics, Ltd.). The measurement was carried out in freshly distilled water at 25 ± 0.1 °C.

Chemicals—Hydrochlorides of CPZ and CPZO were kindly provided by Shionogi Co., Ltd. and were used without further purification. The perchlorate salt of CPZ^+ was synthesized electrochemically according to the method of McCreery et al. $^{14)}$ α -, β - and γ -Cyclodextrins were kindly provided by Nippon Shokuhin Kako Co., Ltd., and were recrystallized from water. Maltopentaose was obtained from Wako Pure Chemical Industries Ltd. Phosphate buffer was used as base electrolyte solution, and the ionic strength was adjusted to 0.2 using NaCl. In order to prevent its decomposition, CPZ^+ was dissolved in 3 N HCl for measurement of CD and absorption spectra and in 3 N DCl for the 1 H-NMR analysis. Under these conditions, CyDs and CPZ underwent almost no decomposition during measurement.

Results and Discussion

As shown in Fig. 1, the cyclic voltammogram of CPZ without CyD has three oxidation waves at $+0.65\,\mathrm{V}$, $+0.90\,\mathrm{V}$ and $+1.10\,\mathrm{V}$ vs. a saturated calomel electrode (SCE). These waves correspond to the oxidation of CPZ, that of CPZ·+-supporting electrolyte adduct and that of CPZO, respectively.¹⁵⁾ When α -, β - or γ -CyD was added to the CPZ solution, the peak current of the first oxidation wave (i_{p1}) decreased and the peak potential (E_{p1}) shifted to the positive side in the order of β -> γ -> α -CyD (Fig. 1, dotted lines A-C). This order is the same as that of the stability constants of the CPZ-CyD complexes.^{16,17)} In contrast to CyD, maltopentaose, adopted as a linear-structured analogue of CyD having no tendency to form an inclusion complex, hardly affects the voltammogram of CPZ (Fig. 1, curve D). This indicates that the formation of CPZ-CyD complexes results in both a shift of E_{p1} and a decrease in i_{p1} .

To examine the electrochemical reaction of CPZ with CyD, a controlled potential

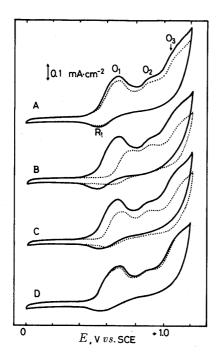


Fig. 1. Cyclic Voltammograms of 1.0×10^{-3} M CPZ with (-----) and without (-----) CyD in 0.1 M Phosphate Buffer

Concentration of CyD: 1.0×10^{-2} M α -CyD (A):

Concentration of CyD: $1.0\times 10^{-2}\,\text{M}$ α -CyD (A); $1.0\times 10^{-2}\,\text{M}$ β -CyD (B); $1.0\times 10^{-2}\,\text{M}$ γ -CyD (C); $1.0\times 10^{-2}\,\text{M}$ maltopentaose (D). pH = 3.

electrolysis was carried out and the oxidation products were detected by absorption spectroscopy and thin-layer chromatography (TLC). The final product of CPZ for each CyD used at the potential of either $E_{\rm p1}$ or $E_{\rm p2}$ was found to be CPZO-CyD by a comparison of the ultraviolet (UV) spectra and Rf values on TLC with those of pure CPZO-CyD mixture. During electrolysis, the solutions had a red tinge owing to a colored product identified as CPZ. on the basis of a comparison of its absorption spectrum with that of synthetic CPZ. From these results, it may be said that the electrochemical oxidation of CPZ in the presence of CyDs takes place by essentially the same pathway as that for CPZ alone. 15)

first peak

$$\begin{array}{c}
CPZ & \stackrel{-e}{\Longrightarrow} CPZ^{+} \\
 & + e \\
 & CPZ^{+} + H_{2}PO_{4}^{-} & = [CPZ(HPO_{4})]^{-} + H^{+} \\
 & [CPZ(HPO_{4})]^{-} + CPZ^{+} & = [CPZ(HPO_{4})] + CPZ \\
 & [CPZ(HPO_{4})] + H_{2}O & = CPZO + H_{2}PO_{4}^{-} + H^{+}
\end{array}$$
(1)

$$\begin{array}{c}
 & (1) \\
 & (2) \\
 & (3) \\
 & (4) \\
 & (4) \\
 & (4) \\
 & (4) \\
\end{array}$$

In the following discussion, β -CyD is preferred to other CyDs, unless otherwise noted, because of its ability to form the most stable complex with CPZ. In our previous paper, we proposed the following electrochemical oxidation mechanism for the first wave of CPZ.¹⁵⁾ The cation radical CPZ.⁺ produced through a one-electron oxidation of CPZ reacts with a phosphate anion, one of the supporting electrolyte components, to form the CPZ.⁺-phosphate adduct. The adduct is then further oxidized by a neighboring CPZ.⁺ molecule through a disproportionation step.

The first oxidation peak current i_{p1} may be represented by Eq. 5.¹⁸⁾

$$i_{\rm pl} = nFAC_0^* \sqrt{Dk_{\rm f}} \tag{5}$$

where D is the diffusion coefficient, C_0^* , the bulk concentration of CPZ, A, the area of the

electrode, F, the Faraday constant, n, the number of electrons, and k_f , the rate constant of the degradation of CPZ^{+} in the bulk solution given by Eq. $6.^{13}$)

$$k_{\rm f} = k'[{\rm H_2PO_4^-}]/[{\rm H^+}]$$
 (6)

On the basis of Eq. 5 and 6, linear relations should be obtainable for both i_{p1} vs. $[H_2PO_4^-]^{1/2}$ and $\log i_{p1}$ vs. pH. This was actually found to be the case in the previous investigation.¹⁵⁾

In the presence of β -CyD, similar relations were also obtained for the first peak height (Fig. 2), indicating that a similar reaction scheme holds for the first oxidation wave of the CPZ- β -CyD system.

As mentioned above, the addition of β -CyD to the CPZ solution leads to a decrease in i_{p1} . The reason for this should be the lower diffusion coefficient of the resulting CPZ- β -CyD complex. The diffusion coefficient and Stokes' diameter of CPZ- β -CyD were estimated by CPZ electroconductivity measurement with and without β -CyD. The straight lines in Fig. 3, in which the equivalent conductivity was plotted vs. square root of the concentration of CPZ, show that CPZH+Cl⁻ is completely dissociated even in the presence of β -CyD. The peak current i_{p1} is directly proportional to the square root of the diffusion coefficient D, and D is

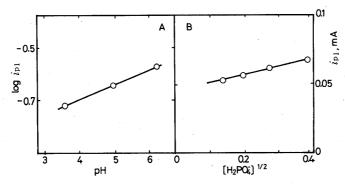


Fig. 2. A. Logarithm of Peak Current i_{p1} vs. pH of Solution B. Peak Current i_{p1} vs. Square Root of Buffer Concentration $[CPZ] = 1.0 \times 10^{-3} \text{ M}$; $[\beta - CyD] = 1.0 \times 10^{-2} \text{ M}$.

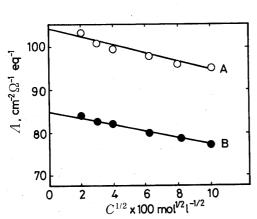


Fig. 3. Electroconductivity of CPZ with (B) and without (A) β -CyD in Distilled Water at 25 °C

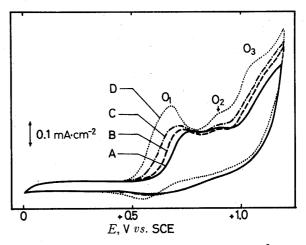


Fig. 4. Cyclic Voltammogram of 1.0×10^{-3} M CPZ with (A—C) and without (D) CyD in 0.1 M Phosphate Buffer, pH=3.8

Concentration of CyD: 1.0×10^{-2} M β -CyD (A); 5.0×10^{-2} M γ -CyD (B); 5.0×10^{-2} M α -CyD (C).

proportional to the equivalent conductivity at infinite dilution Λ_0 . That Λ_0 of CPZH⁺Cl⁻ $-\beta$ -CyD is lower than that of CPZH⁺Cl⁻ is obvious from Fig. 3, and thus it is apparent that the lowering of D results in a decrease in $i_{\rm p1}$.

The Stokes' diameters of $CPZH^+$ and the $CPZH^+-\beta$ -CyD complex were calculated according to Nakagaki and Okada.¹⁹⁾ The diameters r were obtained from the following Eq. 7.

$$r = 0.82 \cdot |Z|/((\Lambda_{\text{CPZH}^+}^0 - \lambda_{\text{Cl}^-}^0)\eta^0)(\gamma_{\text{C}}/\gamma_{\text{S}})$$
(7)

In this equation, |Z| signifies the charge number, $\gamma_{\rm C}/\gamma_{\rm S}$ the correction factor, η^0 the viscosity of the solvent, $\Lambda^0_{\rm CPZH^+}$ the equivalent conductivity at infinite dilution of CPZH⁺, and $\lambda^0_{\rm Cl^-}$ the equivalent conductivity of Cl⁻. The charge of CPZ and viscosity of the solution were unchanged irrespective of the presence of β -CyD. The obtained size of CPZH⁺ and that of its β -CyD complex were about 5 and 14 Å, respectively. These values seem reasonable, taking into account the sizes of molecular models of CPZ and CPZ- β -CyD. Thus, enlargement of the molecular size of the diffusing entity is responsible for the lowering of D.

The cyclic voltammograms of CPZ with α -, β - or γ -CyD were obtained by keeping the complex concentrations constant at 9×10^{-4} M, and the results are shown in Fig. 4. In contrast with the first oxidation wave, the peak potential of the second wave $E_{\rm p2}$ was not affected by

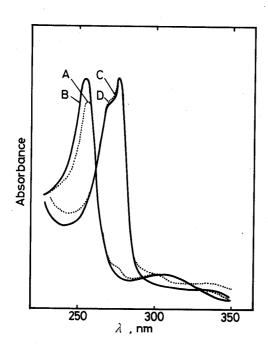


Fig. 5. Absorption Spectra of 1.9×10^{-5} M CPZ with (A) and without (B) 1.0×10^{-3} M β -CyD and 1.5×10^{-5} M CPZ $^+$ with (C) and without (D) 1.0×10^{-3} M β -CyD in 3 N HCl

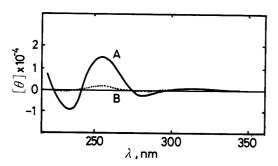


Fig. 6. CD Spectra of 2.5×10^{-5} M CPZ with 5.0×10^{-4} M β -CyD (A), 2.3×10^{-5} M CPZ·+ with 5.0×10^{-4} M β -CyD (B) in 3 N HCl

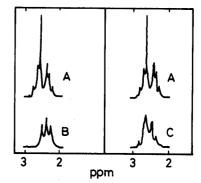


Fig. 7. ¹H-NMR Spectra of β-CyD in 3 N DCl Components: 1.0×10^{-2} M β-CyD (A); 5.0×10^{-3} M β-CyD with 5.8×10^{-3} M CPZ (B); 5.0×10^{-3} M β-CyD with 5.8×10^{-3} M CPZ·+ (C).

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the existence of CyD. The fact that α -, β - and γ -CyD had no effect on the oxidation potential of CPZ $^+$ may suggest that no complexation of CPZ $^+$ with CyDs took place.

To determine whether β -CyD can form a complex with CPZ· $^+$ or not, absorption, CD and 1 H-NMR spectra of CPZ· $^+$ with β -CyD were examined. In Fig. 5, which shows the UV absorption spectra of CPZ and CPZ· $^+$ with β -CyD, the absorptivity of CPZ with β -CyD differs from that of CPZ alone. On the other hand, the presence of β -CyD causes no appreciable change in absorption of CPZ· $^+$. No CD band was observed for any solution of CPZ, CPZ· $^+$ or β -CyD, but the CPZ– β -CyD mixture showed marked induced CD bands (Fig. 6, curve A) arising from the inclusion complex formed. In contrast, the CPZ· $^+$ - β -CyD system hardly showed any induced CD band, giving only a small band (curve B) derived from CPZ reproduced through reaction (3). These findings indicate that β -CyD is not capable of forming an inclusion complex with CPZ· $^+$.

In the ¹H-NMR spectra of CPZ with β -CyD, as shown in Fig. 7, the peaks of β -CyD are shifted upfield by the anisotropic shielding of CPZ included in the cavity of β -CyD (curve B). ¹⁶⁾ In the case of CPZ· ⁺- β -CyD, only a broadening of the peaks was observed (curve C), implying that the aromatic portion of CPZ· ⁺ was not in the cavity of β -CyD.

The weak affinity of β -CyD for CPZ· + seems likely to be due to the polarity of the phenothiazine ring of CPZ· +. Recently, the relationship between the partition coefficients and stability constants of β -CyD-phenothiazine derivative complexes was discussed by Otagiri *et al.*¹⁶⁾ It is considered that hydrophilic derivatives having low partition coefficients cannot form stable complexes, because of the hydrophobic property of the β -CyD cavity. Thus, a mutual attraction cannot be expected between hydrophilic CPZ· + and the hydrophobic β -CyD cavity. Therefore, it may be reasonable to assume that a dissociation of CPZ· + from β -CyD takes place on the electrode surface during the electrolysis of the CPZ- β -CyD system.

On the basis of all the findings presented so far, the electrochemical oxidation mechanism of $CPZ-\beta$ -CyD system shown in Chart 3 is proposed.

first peak

$$\begin{array}{c} \text{CPZ} + \text{CyD} & \longrightarrow \text{CPZ-CyD} \\ \hline \begin{array}{c} -e \\ \text{CPZ-CyD} & \xrightarrow{-e} \text{CPZ} \cdot ^+ + \text{CyD} \\ \hline \end{array} & \begin{array}{c} (9) \\ \text{CPZ} \cdot ^+ + \text{H}_2 \text{PO}_4^- & \longrightarrow \text{[CPZ(HPO_4)]} \cdot ^- + \text{H}^+ \\ \hline \end{array} & \begin{array}{c} (10) \\ \text{[CPZ(HPO_4)]} \cdot ^- + \text{CPZ} \cdot ^+ & \longrightarrow \text{[CPZ(HPO_4)]} + \text{CPZ} \\ \hline \end{array} & \begin{array}{c} (11) \\ \end{array} & \end{array}$$

second peak

$$CPZ^{+} + H_{2}PO_{4}^{-} \Longrightarrow [CPZ(HPO_{4})]^{-} + H^{+}$$

$$[CPZ(HPO_{4})]^{-} \stackrel{-}{-} e \qquad [CPZ(HPO_{4})]$$

$$[CPZ(HPO_{4})] + H_{2}O \longrightarrow CPZO + H_{2}PO_{4}^{-} + H^{+}$$

$$CPZO + CyD \Longrightarrow CPZO - CyD$$

$$Chart 3$$

$$(13)$$

$$(14)$$

$$(15)$$

In this mechanism, β -CyD retards the first oxidation step of CPZ, but the subsequent reactions take place through the same pathway as that proposed for the oxidation of CPZ without β -CyD.¹⁵⁾ Thus it seems most likely that CyD serves as a stabilizer of CPZ by

preventing its photodecomposition, since β -CyD provides good protection against the oxidation of CPZ. The cation radical CPZ \cdot is an active pharmacological entity *in vivo*. The fact that the electrochemical oxidation potential of CPZ \cdot remains unchanged even when β -CyD is present in the solution probably indicates that the pharmacological activity of CPZ is not affected by β -CyD at the receptor site of CPZ. Therefore, CyD holds promise as a carrier of CPZ.

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