Communications to the Editor

Chem. Pharm. Bull. 32(7)2882—2885(1984)

3,4-DIMETHOXYBENZYL GROUP: A NEW PROTECTING GROUP FOR THE GUANOSINE RESIDUE DURING OLIGONUCLEOTIDE SYNTHESIS

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The protection of the 0^6 -amide group of deoxyguanosine with the 3,4-methoxybenzyl group is described. This protecting group could be introduced effectively and was readily removed with DDQ. This was used to demonstrate the synthesis of the oligodeoxyribonucleotide.

KEYWORDS——protecting group; 3,4-dimethoxybenzyl group; deoxyguanosine; oligodeoxyguanylate synthesis

Various side reactions of guanine residue during the phosphorylation and condensation steps have been reported in recent years. Several methods for the 0⁶-protection of guanosine have been reported by Reese, Hata, Jones, and Pfleiderer Pfleiderer, however, unstable in alkali conditions.

In this paper, we report a useful protecting group, 3,4-dimethoxybenzyl group, for the protection of the 0^6 -amide group of deoxyguanosine. This is stable in acid and alkali, and removable by treatment with 2,3-dichloro-5,6-dicyanobenzoquinone (DDQ)⁶⁾ in $\text{CH}_3\text{CN-H}_2\text{O}$ (4:1, v/v).

We first investigated the effect of the 3,4-dimethoxybenzyl group of the 06amide of the deoxyguanosine derivative (1) by use of the procedure reported by Pfleiderer. 5) 3',5'-O-N²-Triisobutyryldeoxyguanosine (1) (2.68 g, 6.0 mmol) was treated with 3,4-dimethoxybenzyl alcohol (5.01 g, 30 mmol) in the presence of triphenylphosphine (4.33 g, 16.5 mmol) and diethyl azodicarboxylate (2.61 g, 15 mmol) in THF (60 ml) at room temperature. After 20 h, TLC analysis showed complete conversion of starting material 1 into a high Rf product. The reaction mixture was quenched with ice water (5 ml), extracted with $\mathrm{CH_2Cl_2}$ (60 ml X 2), and washed with 5% NaHCO $_3$ solution. The organic layer was dried with Na $_2$ SO $_4$ and evaporated in vacuo. The residue was dissolved in a mixture of EtOH (30 ml) and pyridine (10 ml) and the solution was treated with 2N NaOH (28 ml) at 0°C for 15 min. mixture was neutralized with Dowex 50W-X2 (pyridinium form). The resin was removed by filtration and the filtrate was evaporated in vacuo. The residue was dissolved in CH₂Cl₂ (50 ml), and washed with 5% NaHCO₃ solution and with water. layer was dried with $\mathrm{Na_2SO_4}$ and evaporated under reduced pressure to give an oil. The oil was dissolved in a small amount of CH₂Cl₂ and subjected to silica gel column The appropriate fractions (eluted with a stepwise gradient of MeOH (0-2%) in CH_2Cl_2) were evaporated to give the corresponding 0^6-3 ,4-dimethoxybenzyl- \tilde{N}^2 -isobutyryldeoxyguanosine (2), which was isolated (3.69 g, 78%) as a white soild by precipitation from hexane: mp 107-109°C; Rf 0.33 (CH2Cl2-MeOH, 9:1, v/v); UV λ max (MeOH) nm: 265, 230; ¹H-NMR(DMSO-d₆): 89.02(1H, s, NH), $8.21(1H, s, C_8-H)$,

6.64(3H, d, ArH), 6.01(1H, t, $J_{1',2'}$ =6.0Hz, $C_{1'}$ -H), 5.30-4.61(3H, m, $C_{3'}$ -H, $C_{3'}$ -OH, and $C_{5'}$ -OH), 4.21(2H, br s, ArCH₂), 3.70(1H, m, $C_{4'}$ -H), 3.51(6H, s, OCH₃), 3.39(2H, br s, $C_{5'}$ -H), Anal. Calcd for C_{23} H₂₉N₅O₇.2H₂O: C, 52.76; H, 6.35; N, 13.38. Found: C, 53.17; H, 6.45; N, 13.28.

The 3,4-dimethoxybenzyl group was found to be quite stable for 1 day in 80% AcOH and also in conc. ammonia. Recently, Okikawa et al. reported that the 4-methoxybenzyl group of alcohol is readily removed with DDQ in $\rm CH_2Cl_2-H_2O$ (18:1, v/v) under neutral conditions. Therefore, 2 was treated with DDQ (2 mol eq) in $\rm CH_2Cl_2-H_2O$ (18:1, v/v) at room temperature. However, the reaction did not proceed effectively. Thus, after 6 h, N²-isobutyryldeoxyguanosine was obtained in 84% yield, and 12% of 2 was recovered. It is now found that $\rm CH_3CN-H_2O$ (4:1, v/v) is much more effective as solvent for the DDQ dehydrogenation. The use of $\rm CH_3CN-H_2O$ shortened dramatically the time for removal of 3,4-dimethoxybenzyl group as compared with $\rm CH_2Cl_2-H_2O$.

The compound 2 (789 g, 1 mmol) was tritylated with dimethoxytrityl chloride (441 mg, 1.3 mmol) in dry pyridine for 3 h. After the usual workup, chromatography on silica gel afforded the tritylated product 3^{8} (744 mg, 82%). The tritylated compound 3 (789 mg, 1.0 mmol) was allowed to react with 2,6-dichlorophenyl 5-chloro-8-quinolyl phosphorochloridate (4)⁹ prepared from 2,6-dichlorophenyl phosphodichloridate (558 mg, 2.0 mmol) and 5-chloro-8-hydroxyquinoline (293 mg, 2.2 mmol) in the presence of 1-methylimidazole (0.31 ml, 4.0 mmol) in dry THF (10 ml) at room temperature for 1 h. The reaction mixture was quenched

with ice-water (1 ml), extracted with $\mathrm{CH_2Cl_2}$ (25 ml X 2), and washed with water. The organic layer was dried with $\mathrm{Na_2SO_4}$ and evaporated in vacuo. The residue was chromatographed on silica gel to give the corresponding mononucleotide unit (5) 10) (1.06 g, 90%).

The utility of this new protecting group, 3,4-dimethoxybenzyl group, can be demonstrated in the following synthesis of d-GpGpGp (10). According to the previous paper, 10) the unit 5 (708 mg, 0.6 mmol) was treated with 3% Cl₃CCOOH in CH₃CN-MeOH (95:5, v/v) at room temperature for 3 min to give the corresponding 5'-hydroxyl component 6 (483 mg, 92%). On the other hand, removal of the 2,6-dichlorophenyl group from 5 (841 mg, 0.75 mmol) was performed by treatment with pyridine- ${\rm H_2O-t-BuNH_2}$ (8:1:1, v/v, 10 ml) at room temperature for 1.5 h. The mixture was extracted with CH2Cl2 (15 ml), washed with 5% NaHCO3, dried over Na2SO4, and evaporated in vacuo. The phosphodiester 7 thus obtained was treated with 6 (439 mg, 0.5 mmol) in the presence of QS-Cl¹¹) (273 mg, 1.50 mmol) and 1-methylimidazole (0.12 ml, 1.55 mmol) in dry pyridine. The reaction was completed in 1 h and the usual workup gave the dimer (8) (821 mg, 91%). The yield was very increased and the undesirble side reactions in the condensation reaction were not Similarly, treatment of 8 (820 mg, 0.43 mmol) with 3% Cl₃CCOOH afforded the detritylated product (9) (624 mg, 92%), whereas mild treatment of 5(580 mg, 0.5 mmol) with pyridine- H_2 O-t-BuNH₂ (8:1:1, v/v) gave the phosphodiester A solution of both compounds 9 (401 mg, 0.4 mmol) and 7 in dry pyridine was treated with QS-C1 (228 mg, 1.0 mmol) and 1-methylimidazole (0.16 ml, 1.0 mmol) The fully protected trinucleotide (10) was obtained in 93% (941 mg) yield after separation by silica gel column chromatography.

Deprotection of all the protecting groups from 10 was performed as follows: 1) To a solution of 10 (12.9 mg, 5 μ mol) in CH₃CN-H₂O (4:1, v/v, 0.5 ml) was added DDQ (4.5 mg, 20 μ mol). The mixture was stirred at room temperature for 1 h and

1. DDQ

2. pyridine- H_2O-t -BuN $H_2(8:1;!)$ d G

 $\label{eq:double_gradient} \text{d} \; \text{GpGpGp} \quad \begin{array}{c} \text{i=3\% Cl}_{3}\text{CCOOH } \; \text{(CH}_{3}\text{NO}_{2}\text{-MeOH,} \\ \text{95\%5}\text{)} \\ \text{ii=pyridine-H}_{2}\text{O-t-BuNH}_{2} \text{(8:1:1)} \end{array}$

3. zinc acetate (aqu. pyridine)

DMPM=3,4-dimethoxybenzyl

4. conc. ammonia

5. 80% AcOH

The solution was concentrated in vacuo. At this stage the 3,4-dimethoxybenzyl group was removed. 2) The residue was treated with pyridine-H₂O-t-BuNH₂ (8:1:1, v/v) at room temperature for 2 h to remove the 2,6-dichlorophenyl group. 3) The solution was concentrated and the residue was treated with zinc acetate (107 mg, 500 µmol) in aqueous pyridine at room temperature for 24 h.¹³⁾ The solution was treated with Dowex 50W-X2 (pyridinium form), and the resin was removed by filtration and washed with aqueous pyridine. 4) The filtrate was concentrated in vacuo and the residue was treated with conc. ammonia at 60°C for 6 h. 5) The solution was concentrated and 80% AcOH was added. After 15 min, the solution was coevaporated with water. The deblocked trimer, d-GpGpGp was isolated in 85% yield after chromatographic separation using Whatman 3MM paper with n-PrOH-conc. ammonia-water (55:35:10, v/v). The deblocked trimer was completly degraded by spleen phosphodiesterase to give a single spot of d-Gp.

REFERENCES AND NOTES

- P. K. Bridson, W. T. Markiewicz, and C. B. Reese, J. Chem. Soc., Chem. Commun., 1977, 447 and 791; C. B. Reese and A. Ubasawa, Tetrahedron Lett., 21, 2265 (1980); H. P. Daskalov, M. Sekine, and T. Hata, ibid., 21, 3899 (1980); H. Takaku, K. Kamaike, and K. Kasuga, Chem. Lett., 1982, 197; E. Ohtsuka, A. Yamane, and M. Ikehara, Nucleic Acids Res., 11, 1325 (1983).
- 2) S. S. Jones, C. B. Reese, S. Shibands, and A. Ubasawa, Tetrahedron Lett., $\underline{21}$, 4755 (1981).
- 3) M. Sekine, J. Matsuzaki, and T. Hata, Tetrahedron Lett., 23, 5287 (1982); T. Kamimura, M. Tsuchiya, K. Koura, M. Sekine, and T. Hata, ibid., 24, 2275 (1983).
- 4) B. L. Gaffeey and R. A. Jones, Tetrahedron Lett., 23, 2257 (1982).
- 5) T. Trichtinger, R. Charubala, and W. Pfleiderer, Tetrahedron lett., 24, 711 (1983).
- 6) Y. Okikawa and O. Yonemitsu, Heterocycles, 5, 233 (1976).
- 7) Y. Okikawa, T. Yoshioka, and O. Yonemitsu, Tetrahedron Lett., 23, 885 (1982).
- 8) 1 H-NMR(CDCl₃): § 8.21(1H,s, C₈-H), 7.68-7.52(10H, m, ArH), 6.45(6H, m, ArH), 6.35(1H, t, C₁,-H), 5.30(1H, m, C₃,-H), 4.20(2H, br s, ArCH₂), 4.00(1H, m, C₄,-H), 3.67(6H, s, OCH₃), 3.40(2H, m, C₅,-H).
- 9) H. Takaku, S. Ueda, H. Tsuchiya, and H. Kobayashi, submitted to Tetrahedron Lett.
- 10) mp 107-109°C; UV λ max (MeOH) nm: 282(sh), 270, 236; Rf 0.72 (CH₂Cl₂-MeOH, 9:1, v/v). Anal. Calcd for C₅₈H₅₄N₆O₁₂PCl₃: C, 59.82; H, 4.67; N, 7.23. Found: C, 60.09; H, 4.74; N, 7.38.
- 11) H. Takaku, K. Morita, and T. Sumiuchi, Chem. Lett., 1983, 1661.
- 12) H. Takaku, M. Yoshida, M. Kato, and T. Hata, Chem. Lett., 1979, 811,
- 13) K. Kamaike, S. Ueda, H. Tsuchiya, and H. Takaku, Chem. Pharm. Bull., <u>31</u>, 2928 (1983).

(Received May 21, 1984)