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Total Synthesis of Cleomiscosin A, a Coumarinolignoid

HITOSHI TANAKA, ICHIRO KATO, and KAZUO ITO*

Faculty of Pharmacy, Meijo University, Tempaku-ku, Nagoya 468, Japan

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The coumarinolignoid cleomiscosin A (1), having antitumor activity, has been synthesized *via* the keto-ester (10), which was prepared from readily available starting materials (6 and 9). Removal of the methoxymethyl group of 10 followed by reduction with lithium borohydride gave a mixture of diols (12a, b). Treatment of 12a, b with 5% sulfuric acid in acetic acid provided cleomiscosin A monoacetate (14) and subsequent hydrolysis with sodium hydroxide afforded 1.

Keywords—coumarinolignoid; cleomiscosin A; benzodioxane; fraxetin; antitumor activity; coumarin

The coumarinolignoid cleomiscosin A (1) has been isolated¹⁾ from the defatted seeds of *Cleome viscosa* (Capparidaceae), and was recently found²⁾ in *Simaba multiflora* (Simaroubaceae), *Soulamea soulameoides* (Simaroubaceae), and *Matayba arborescens* (Sapindaceae). The structure of cleomiscosin A was first represented¹⁾ as formula 15, but recently the revised structure 1 was proposed³⁾ on the basis of studies of selective heterodecoupling in the carbon-13 nuclear magnetic resonance (¹³C-NMR) and proton nuclear magnetic resonance (¹H-NMR) spectra of cleomiscosin A diacetate (2).

Cleomiscosin A, having no optical activity (racemic compound), shows⁴⁾ antitumor activity *in vivo* in the P-388 lymphocytic leukemia test system. Further, it possesses a novel skeleton in which a phenylpropane unit is linked to a coumarin nucleus through a dioxane bridge. Recently, two groups have independently accomplished the synthesis of cleomiscosin A by an oxidative coupling reaction of fraxetin (5) and coniferyl alcohol. Namely, Merlini and co-workers⁵⁾ reported that the oxidative coupling of fraxetin (5) and coniferyl alcohol in the presence of silver oxide afforded cleomiscosin A in 8% yield together with a regioisomer cleomiscosin B (15) in 4% yield. On the other hand, Cordell and Lin⁶⁾ reported that treatment of the same reactants in buffered aqueous solution with horseradish peroxidase produced cleomiscosin A in 23% yield as the main product.

Herein, we wish to describe a large-scale synthesis of cleomiscosin A from readily available materials (6 and 9). The starting material, 8-acetyl-7-hydroxy-6-methoxycoumarin (3), prepared according to the literature, was converted to the corresponding methoxymethyl (MOM) ether (4) by treatment with chloromethyl methyl ether. The Baeyer-Villiger oxidation of 4 with alkaline hydrogen peroxide in dioxane gave the expected phenolic compound (6) in 58% yield. The latter compound was also easily prepared in 65% yield by treatment of commercially available fraxetin (5) with chloromethyl methyl ether. Compound 6 was condensed with ethyl 2-bromo-3-(4-benzyloxy-3-methoxyphenyl)-3-oxopropionate (9)⁸⁾ in the presence of potassium butoxide in N,N-dimethylformamide (DMF) to give an ether (10) in excellent yield. Next, we investigated simultaneous reduction of the keto and ethyl ester groups in 10 with various reagents (sodium borohydride, potassium borohydride, lithium borohydride, and L-selectride). However, all attempts were unsuccessful and gave an intractable tar. Since attack of reagents on the ethyl ester group is thought to be difficult because of steric hindrance by the MOM group, 10 was transformed to a less-hindered ethyl

ester (11) by hydrolysis with hydrochloric acid. It was reported⁹⁾ that sodium borohydride is a good reagent for reduction of a β -ketoester to form prominently the *erythro* diastereomer of the corresponding diols. In the case of 11, however, reduction using sodium borohydride did not proceed satisfactorily in several solvents such as methanol, ethanol, and 2-propanol at $10-30\,^{\circ}$ C. Eventually, the successful reduction of 11 to the diols (12a, b) was accomplished with lithium borohydride¹⁰⁾ in tetrahydrofuran (THF) at room temperature. These diols were homogeneous on thin-layer chromatography (TLC) in a variety of solvent systems. The high resolution mass spectrum (MS) gave the molecular formula $C_{27}H_{26}O_9$ and the infrared (IR) spectrum lacked both of the carbonyl bands (1675 and 1745 cm⁻¹) which were found in the keto ester (11). In the ¹H-NMR spectrum, signals due to two methoxyl groups at δ 3.68 and 3.72, one methylene group at δ 4.92, and one aromatic proton of the coumarin ring at δ 6.48 were each observed as a singlet. Furthermore, the proton assigned to the C-7' position gave split signals at δ 5.08 (d, J=8 Hz) and δ 5.18 (d, J=4 Hz), and the protons at C-3 and C-4 similarly appeared as doublets at δ 6.04 and 6.05 (J=9.5 Hz, C_3 -H), and δ 7.39 and 7.40 (J=9.5 Hz, C_4 -H), respectively. Hence, these compounds (12a, b) were found to be a mixture of

the *erythro* and *threo* forms. The ratio of the two isomers was estimated to be about 2:1 by 1 H-NMR spectral analysis. Attempts to separate the diols by chromatography were unsuccessful under various conditions. Therefore, the diols were led to the corresponding triacetates (13a, b) by normal acetylation, and these were successfully separated into 13a and 13b in pure form by preparative TLC. In order to clarify the stereochemistry of the triacetates, the 1 H-NMR spectra were measured. The stereochemical assignaments of 13a, b were made on the basis of comparison of the J values for coupling between C_7 -H and C_8 -H. The 1 H-NMR spectrum of the major product (13a) exhibited a doublet at δ 6.07 (J=4 Hz) attributable to C_7 -H, whereas that of the minor product (13b) showed a doublet at δ 6.02 (J=8 Hz) ascribable to C_7 -H. It is known¹¹⁾ that the vicinal coupling constant (J_{AB}) of the *erythro* isomer is smaller than that of the *threo* isomer. Accordingly, the structure of the major reduction product was assigned as the *erythro* form (12a) and the minor product was assigned as the *threo* form (12b).

Finally, the mixture (12a, b) underwent¹²⁾ cyclization with 5% sulfuric acid to give cleomiscosin A monoacetate (14) in 50% yield. The MS showed the characteristic fragment peak at m/z 222 due to the retro Diels-Alder reaction¹⁾ of the benzodioxane moiety in addition to the parent peak at m/z 428. The coupling constant between the C_7 -H and C_8 -H signals in the ¹H-NMR spectrum was 8 Hz, demonstrating that the two hydrogens are *trans*-oriented. Subsequent treatment of 14 with 1% sodium hydroxide afforded the desired cleomiscosin A (1) in 96% yield.

Experimental

All melting points are uncorrected. Column chromatography was run on Merck Silica gel 60 (70—230 mesh) and TLC was performed on glass plates precoated with Kieselgel 60 F_{254} (Merck). MS were recorded on a Hitachi M-52 spectrometer, and high resolution MS and secondary ion mass spectrometry (SIMS) on a Hitachi M-80 spectrometer. IR spectra were obtained on a JASCO IRA-3 spectrophotometer. 1 H-NMR spectra were recorded on a JEOL JNM-PS-100 nuclear magnetic resonance spectrometer and 13 C-NMR spectra on a JEOL JNM-FX-100 Fourier transform spectrometer operating at 25.00 MHz, with tetramethylsilane as an internal standard. Chemical shifts are quoted in parts per million (s=singlet, d=doublet, dd=doublet of doublets, t=triplet, q=quartet, br=broad).

8-Acetyl-6-methoxy-7-methoxymethoxycoumarin (4)—A mixture of 3^{7} (1.869 g) and DMF (20 ml) was added to a suspension of NaH (60% in mineral oil) (480 mg) in THF (4 ml) under ice-cooling. The reaction mixture was stirred at room temperature for 1 h and then chloromethyl methyl ether (0.91 ml) was added gradually. The mixture was stirred at room temperature for 3 h, then poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to dryness. The residue was purified by column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v) to afford a solid (4). The solid was recrystallized from EtOH. Colorless needles (1.50 g) (68%). mp 108-109 °C. Anal. Calcd for $C_{14}H_{14}O_6$: C, 60.43; H, 5.07. Found: C, 60.26; H, 4.96. MS m/z: 278 (M⁺), 246, 233, 216, 204 (100%). IR $v_{max}^{CHCl_3}$ cm⁻¹: 1740, 1725, 1570. ¹H-NMR (CDCl₃) δ : 2.62 (3H, s, COCH₃), 3.49 (3H, s, OCH₂OCH₃), 3.88 (3H, s, OCH₃), 5.16 (2H, s, OCH₂OCH₃), 6.33 (1H, d, J= 9.5 Hz, C_3 -H), 6.95 (1H, s, C_5 -H), 7.62 (1H, d, J=9.5 Hz, C_4 -H).

8-Hydroxy-6-methoxy-7-methoxymethoxycoumarin (6)—a) A solution of 30% hydrogen peroxide (20 ml) was added at once to a solution of 4 (400 mg) in dioxane (5 ml) and 1 n sodium hydroxide (4 ml) under ice-cooling. The mixture was stirred at room temperature for 2 d and poured into water. The resulting mixture was acidified by the dropwise addition of 5% acetic acid and extracted with ether. The ether layer was washed with water, dried over Na₂SO₄, and evaporated to dryness. The residue was purified by column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v) to yield a solid (6). The solid was recrystallized from EtOH. Colorless needles (211 mg) (58%). mp 161—163 °C. Anal. Calcd for $C_{12}H_{12}O_6$: C, 57.14; H, 4.80. Found: C, 56.95; H, 4.77. MS m/z: 252 (M⁺), 222 (100%), 220, 207, 123. IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3550, 1720, 1570. ¹H-NMR (acetone- d_6) δ : 2.88 (1H, br s, OH), 3.56 (3H, s, OCH₂OCH₃), 3.88 (3H, s, OCH₃), 5.22 (2H, s, OCH₂OCH₃), 6.30 (1H, d, J=9.5 Hz, C₃-H), 6.80 (1H, s, C₅-H), 7.85 (1H, d, J=9.5 Hz, C₄-H).

b) A solution of fraxetin (5) (500 mg) in THF (0.5 ml) and DMF (0.5 ml) was added to a suspension of NaH (60% in mineral oil) (96 mg) in THF (1 ml) under ice-cooling. The reaction mixture was stirred at room temperature for 1 h and then chloromethyl methyl ether (0.185 ml) was added dropwise. The mixture was stirred at room temperature for 3 h, then poured into water and extracted with AcOEt. The organic layer was washed with water, dried over Na₂SO₄, and evaporated to yield a semisolid. The crude solid was crystallized from a small amount of

EtOH to yield unreacted fraxetin (5) (230 mg), and concentration of the mother liquor afforded a brown oil, which was subjected to column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v). The substance eluted first was the dimethoxymethyl ether (8) (56 mg) (15% based on consumed starting material) and the compound eluted second was the monomethoxymethyl ether (7) (24 mg) (7%). The last elution product was the desired monomethoxymethyl ether (6) (211 mg) (65%).

8: The substance was recrystallized from EtOH. Colorless needles. mp 125—126 °C. TLC (silica gel/CHCl₃–acetone (10:1, v/v), Rf=0.62). Anal. Calcd for $C_{14}H_{16}O_7$: C, 56.75; H, 5.44. Found: C, 56.81; H, 5.52. MS m/z: 296 (M⁺), 251, 235, 220 (100%), 205, 192. IR $v_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 1720, 1570. ¹H-NMR (acetone- d_6) δ : 3.56, 3.63 (6H, 2×s, 2×OCH₂OCH₃), 3.90 (3H, s, OCH₃), 5.20, 5.25 (2H, 2×s, 2×OCH₂OCH₃), 6.32 (1H, d, J=9.5 Hz, C_3 -H), 7.70 (1H, s, C_5 -H), 7.88 (1H, d, J=9.5 Hz, C_4 -H).

7: The compound was recrystallized from EtOH. Colorless needles. mp 134—135 °C. TLC (silica gel/CHCl₃-acetone (10:1, v/v), Rf = 0.43). Anal. Calcd for $C_{12}H_{12}O_6$: C, 57.14; H, 4.80. Found: C, 57.27; H, 4.84. MS m/z: 252 (M⁺), 222 (100%), 220, 207, 123. IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3550, 1720, 1570. ¹H-NMR (acetone- d_6) δ : 3.15 (1H, br s, OH), 3.64 (3H, s, OCH₂OCH₃), 3.91 (3H, s, OCH₃), 5.25 (2H, s, OCH₂OCH₃), 6.21 (1H, d, J = 9.5 Hz, C_3 -H), 7.03 (1H, s, C_5 -H), 7.86 (1H, d, J = 9.5 Hz, C_4 -H).

6: The recrystallization of 6 was carried out from EtOH. Colorless needles. mp 159—161 °C. TLC (silica gel/CHCl₃-acetone (10:1, v/v), Rf=0.40). This compound was identical with the product obtained in a) on the basis of a comparison of spectral (IR, ¹H-NMR, and MS) properties and mixed melting point determination.

Condensation of 6 with 9 (Formation of 10)—A solution of 6 (197 mg) in DMF (0.5 ml) was added to a suspension of tert-BuOK (96 mg) in DMF (0.5 ml) at room temperature. The mixture was stirred for 1 h at the same temperature, then a solution of 9^{8} (343 mg) in DMF (0.7 ml) was added under ice-cooling. Stirring was continued for a further 3 h at room temperature. The resulting mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to dryness. Purification of the residue by column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v) gave an oil (10). A colorless oil (444 mg) (98%). High MS m/z: 578.1786. Calcd for C₃₁H₃₀O₁₁ (M⁺). Found: 578.1818. MS m/z: 578 (M⁺), 328 (100%), 285, 252, 241. IR $v_{max}^{\text{CHCl}_3}$ cm⁻¹: 1750, 1720, 1675, 1565. ¹H-NMR (CDCl₃) δ : 1.20 (3H, t, J=7 Hz, CH₂CH₃), 3.49 (3H, s, OCH₂OCH₃), 3.86, 3.95 (6H, 2×s, 2×OCH₃), 4.24 (2H, q, J=7 Hz, OCH₂CH₃), 5.24 (4H, s, CH₂Ph and OCH₂OCH₃), 6.16 (1H, s, C₈-H), 6.29 (1H, d, J=9.5 Hz, C₃-H), 6.69 (1H, s, C₅-H), 6.97 (1H, d, J=8 Hz, C₅-H), 7.18—7.56 (5H, m, 5×aromatic protons), 7.57 (1H, d, J=9.5 Hz, C₄-H), 7.71 (1H, d, J=3 Hz, C₂-H), 7.88 (1H, dd, J=8, 3 Hz, C₆-H).

Demethoxymethylation of 10 (Formation of 11)——A mixture of 10 (100 mg) and 2 n HCl–MeOH (5 ml) was stirred at room temperature for 30 min. The mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to give a solid (11), which was recrystallized from AcOEt. Colorless needles (91 mg) (99%). mp 157—158 °C. Anal. Calcd for C₂₉H₂₆O₁₀: C, 65.16; H, 4.90. Found: C, 64.96; H, 4.86. SIMS m/z: 535 (M⁺+1), 517, 372, 329, 281, 241 (100%), 219. IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3630, 1745, 1730, 1675, 1580. ¹H-NMR (CDCl₃) δ: 1.24 (3H, t, J=7 Hz, OCH₂CH₃), 3.90, 3.94 (6H, 2×s, 2×OCH₃), 4.31 (2H, q, J=7 Hz, OCH₂CH₃), 5.24 (2H, s, CH₂Ph), 6.20 (1H, s, C₈-H), 6.26 (1H, d, J=9.5 Hz, C₃-H), 6.72 (1H, s, C₅-H), 7.02 (1H, d, J=8 Hz, C₅-H), 7.36 (5H, br s, 5×aromatic protons), 7.59 (1H, d, J=9.5 Hz, C₄-H), 7.70 (1H, d, J=3 Hz, C₂-H), 7.98 (1H, dd, J=8, 3 Hz, C₆-H).

Reduction of 11 with Lithium Borohydride (Formation of 12a, b)—A mixture of 11 (81 mg) and LiBH₄ (13 mg) in THF (5 ml) was stirred at room temperature for 1.5 h. The mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to dryness. Purification of the residue by column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v) afforded an oil (12a, b). A colorless oil (56 mg) (75%). High MS m/z: 494.1575 Calcd for C₂₇H₂₆O₉ (M⁺). Found: 494.1610. MS m/z: 494 (M⁺), 476, 385, 286, 270, 234 (100%), 208. IR $v_{max}^{CHCl_3}$ cm⁻¹: 3350, 1720, 1615, 1575. ¹H-NMR (CDCl₃) δ: 3.40—4.16 (3H, m, C₉.-H and C₈.-H), 3.68, 3.72 (6H, 2×s, 2×OCH₃), 4.92 (2H, s, CH₂Ph), 5.08 (1/3H, d, J=8 Hz, C₇.-H), 5.18 (2/3H, d, J=4 Hz, C₇.-H), 6.04 (1/3H, d, J=9.5 Hz, C₃-H), 6.05 (2/3H, d, J=9.5 Hz, C₃-H), 6.48 (1H, s, C₅-H), 6.56—6.92 (3H, m, C₂.-H, C₅.-H, and C₆.-H), 7.19 (5H, br s, 5×aromatic protons), 7.39 (1/3H, d, J=9.5 Hz, C₄-H), 7.40 (2/3H, d, J=9.5 Hz, C₄-H).

Acetylation of 12a, b—A mixture of 12a, b (50 mg), acetic anhydride (0.3 ml), and pyridine (0.5 ml) was stirred at room temperature for 2 h, then poured into water and extracted with AcOEt. The extract was washed with water, dried over Na_2SO_4 , and evaporated to give a crude oil (13a, b). The oil was subjected to preparative TLC with a mixture of CHCl₃ and acetone (50:1, v/v) as a developing solvent. Elution of the adsorbent corresponding to Rf 0.46 with the same solvent gave 13a (30 mg) (48%) and the band at Rf 0.42 afforded 13b (9 mg) (14%).

13a: A colorless oil. TLC (silica gel/CHCl₃-acetone (50:1, v/v), Rf=0.46). High MS m/z: 620.1891 Calcd for $C_{33}H_{32}O_{12}$ (M⁺). Found: 620.1870. MS m/z: 620 (M⁺), 518, 458, 427, 367 (100%), 335, 312, 293, 251, 208. IR $v_{\max}^{\text{CHCl}_3}$ cm⁻¹: 1765, 1740, 1575. ¹H-NMR (CDCl₃) δ : 1.90 (3H, s, COCH₃), 2.16 (6H, s, 2×COCH₃), 3.78, 3.86 (6H, 2×s, 2×OCH₃), 4.07 (1H, dd, J=13, 4 Hz, C_9 -H), 4.45 (1H, dd, J=13, 7 Hz, C_9 -H), 4.99 (1H, m, C_8 -H), 5.07 (2H, s, CH₂Ph), 6.07 (1H, d, J=4 Hz, C_7 -H), 6.30 (1H, d, J=9.5 Hz, C_3 -H), 6.62 (1H, s, C_5 -H), 6.67—7.70 (3H, m, 3×aromatic protons), 7.29 (5H, br s, 5×aromatic protons), 7.50 (1H, d, J=9.5 Hz, C_4 -H).

13b: A colorless oil. TLC (silica gel/CHCl₃-acetone (50:1, v/v), Rf=0.42). High MS m/z: 620.1891 Calcd for $C_{33}H_{32}O_{12}$ (M⁺). Found: 620.1853. MS m/z: 620 (M⁺), 518, 458, 427, 367 (100%), 335, 312, 293, 251, 208. IR $v_{max}^{CHCl_3}$ cm⁻¹: 1765, 1740, 1575. ¹H-NMR (CDCl₃) δ : 1.95, 1.97, 2.28 (9H, 3×s, 3×COCH₃), 3.82, 3.85 (6H, 2×s, 2×OCH₃), 3.89 (1H, dd, J=11, 4Hz, C_9 -H), 4.29 (1H, dd, J=11, 3 Hz, C_9 -H), 4.95 (1H, m, C_8 -H), 5.07 (2H, s, CH₂Ph), 6.02 (1H, d, J=8 Hz, C_7 -H), 6.32 (1H, d, J=9.5 Hz, C_3 -H), 6.64 (1H, s, C_5 -H), 6.53—6.93 (3H, m, 3×aromatic protons), 7.29 (5H, br s, 5×aromatic protons), 7.52 (1H, d, J=9.5 Hz, C_4 -H).

Cleomiscosin A Monoacetate (14)——A mixture of 12a, b (62 mg) and acetic acid (10 ml) was heated in the presence of 5% sulfuric acid (1.5 ml) under stirring at 60 °C for 10 min. After cooling, the reaction mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated. The resulting oil was purified by column chromatography on silica gel with a mixture of CHCl₃ and acetone (10:1, v/v) to yield an oil (14). A colorless oil (27 mg) (50%). High MS m/z: 428.1106 Calcd for C₂₂H₂₀O₉ (M⁺). Found 428.1107. MS m/z: 428 (M⁺), 368 (100%), 222, 179, 162. IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3550, 1740, 1720, 1570. ¹H-NMR (CDCl₃) δ: 2.06 (3H, s, COCH₃), 3.91 (6H, s, 2 × OCH₃), 3.93—4.49 (3H, m, C₈-H and C₉-H), 4.91 (1H, d, J=8 Hz, C₇-H), 5.81 (1H, br s, OH), 6.33 (1H, d, J=9.5 Hz, C₃-H), 6.55 (1H, s, C₅-H), 6.77—7.01 (3H, m, 3 × aromatic protons), 7.61 (1H, d, J=9.5 Hz, C₄-H).

Cleomiscosin A (1)——A mixture of 14 (23 mg), MeOH (2 ml), and 1% NaOH (20 drops) was stirred at 0 °C for 30 min. The mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to afford a solid (1), which was recrystallized from MeOH. Colorless needles (20 mg) (96%). mp 252—253 °C (lit.,¹) mp 247—249 °C; lit.,²) mp 250—252 °C; lit.,⁵) mp 257 °C). *Anal.* Calcd for C₂₀H₁₈O₈: C, 62.17; H, 4.70. Found: C, 61.90; H, 4.69. MS m/z: 386 (M +), 368, 356, 249, 219, 208, 181, 180, 162, 161, 152, 151, 150, 138, 137, (100%), 124. IR $v_{\text{max}}^{\text{KBr}}$ cm -¹: 3460, 1710, 1610, 1575. ¹H-NMR (pyridine- d_5) δ: 3.74, 3.82 (6H, 2×s, 2×OCH₃), 3.94 (1H, dd, J = 13, 3 Hz, C₉·-H), 4.35 (1H, dd, J = 13, 2 Hz, C₉·-H), 4.51 (1H, m, C₈·-H), 5.01 (1H, br s, OH), 5.62 (1H, d, J = 8 Hz, C₇·-H), 6.49 (1H, d, J = 9.5 Hz, C₃-H), 6.78 (1H, s, C₅-H), 7.22—7.56 (3H, m, C₂·-H, C₅·-H, and C₆·-H), 7.32 (1H, br s, OH), 7.81 (1H, d, J = 9.5 Hz, C₄-H). ¹³C-NMR (pyridine- d_5) δ: 160.9 (s, C-2), 149.8 (s, C-3′), 149.1 (s, C-4′), 146.5 (s, C-6), 144.6 (s, C-4), 139.4 (s, C-9), 138.5 (s, C-7), 133.1 (s, C-8), 127.6 (s, C-1′), 121.8 (d, C-6′), 116.7 (d, C-5′), 113.9 (d, C-3), 112. 4 (d, C-2′), 112.0 (s, C-10), 101.2 (d, C-5), 80.0 (d, C-8′), 77.6 (d, C-7′), 60.8 (t, C-9′), 56.3 (q, OCH₃), 55.9 (q, OCH₃).

This compound was identical with an authentic sample¹⁾ on the basis of mixed melting point determination and direct comparison of IR spectra (KBr).

Cleomiscosin A Diacetate (2)—A mixture of 1 (50 mg), acetic anhydride (0.42 ml) and pyridine (0.50 ml) was heated at 100 °C for 1 h. After cooling, the reaction mixture was poured into water and extracted with AcOEt. The extract was washed with water, dried over Na₂SO₄, and evaporated to afford a solid (2), which was recrystallized from EtOH. Colorless needles (59 mg) (97%). mp 180—181 °C (lit.,²) mp 178—180 °C). *Anal.* Calcd for $C_{24}H_{22}O_{10}$: C, 61.27; H, 4.72. Found: C, 61.52; H, 4.63. MS m/z: 470 (M⁺), 428, 368 (100%), 291, 223, 222, 219, 180, 179, 162, 161, 147. IR v_{max}^{KBr} cm⁻¹: 1755, 1725, 1715, 1610, 1560. ¹H-NMR (CDCl₃) δ: 2.01, 2.27 (6H, 2×s, 2×COCH₃), 3.80, 3.85 (6H, 2×s, 2×OCH₃), 4.06 (1H, m, $C_{8'}$ -H), 4.38 (2H, m, $C_{9'}$ -H), 4.99 (1H, d, J=7 Hz, $C_{7'}$ -H), 6.24 (1H, d, J=9.5 Hz, C_{3} -H), 6.49 (1H, s, C_{5} -H), 6.82—7.10 (3H, m, $C_{2'}$ -H, $C_{5'}$ -H, and $C_{6'}$ -H), 7.52 (1H, d, J=9.5 Hz, C_{4} -H). ¹³C-NMR (CDCl₃) δ: 170.2 (s, COCH₃), 168.5 (s, COCH₃), 160.5 (s, C-2), 151.7 (s, C-3'), 145.8 (s, C-6), 143.6 (d, C-4), 140.8 (s, C-9), 138.8 (s, C-4'), 137.0 (s, C-7), 133.6 (s, C-1'), 131.7 (s, C-8), 123.4 (d, C-5'), 120.0 (d, C-6'), 114.4 (d, C-3), 111.9 (s, C-10), 111.5 (d, C-2'), 100.6 (d, C-5), 76.7 (d, C-7'), 75.1 (d, C-8'), 62.5 (t, C-9'), 56.4 (q, OCH₃), 56.0 (q, OCH₃), 20.6 (q, COCH₃).

Spectral (IR, ¹H-NMR, ¹³C-NMR, and MS) properties of this substance were identical with those of the product prepared by Professor G. A. Cordell.²⁾

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