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New Derivatives of Saikosaponins

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In the course of studies on the metabolism of saikosaponins, which are the main constituents of Bupleurum falcatum L., five new compounds derived from saikosaponins a, c and d were isolated after incubation with acid and snail enzyme. On incubation of saikosaponins a and c at 60 °C in a 1 N sulfuric acid—dioxane solution, saikosaponins g and i possessing a homoannular diene moiety at C-9(11),12 were isolated, and their structures were elucidated as 3β ,16 β ,23,28-tetrahydroxyoleana-9(11),12-diene 3-O- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-fucopyranoside and 3β ,16 β ,28-trihydroxyoleana-9(11),12-diene 3-O- β -D-glucopyranosyl-(1 \rightarrow 6)-[α -L-rhamnopyranosyl-(1 \rightarrow 4)]- β -D-glucopyranoside, respectively. Furthermore, on incubation of saikosaponins b₁, g and b₂ with snail enzyme for 12 h at 37 °C, prosaikogenins A, H, and D were formed; they were identified as the 3- β -D-fucopyranosides of saikogenins A, H and D.

Keywords—saikosaponin; prosaikogenin; saikogenin; snail enzyme; intestinal flora

Studies on the triterpenoidal saponins of the roots of *Bupleurum falcatum* L. were originated by Shibata *et al.*^{1,2)} and Kubota *et al.*³⁾ who reported chemical studies on saikogenins A, C and D. Then, Kubota *et al.*^{4,5)} identified the genuine genins, saikogenins E, F and G, and glycosides, saikosaponins a, b, c and d. Later, Shimaoka *et al.*⁶⁾ reinvestigated the fraction of saikosaponins a and b, and showed that so called saikosaponins a and b were a mixture of saikosaponins a, b₁ and b₃ and a mixture of saikosaponins b₂ and b₄, respectively. Recently, Ishii *et al.*⁷⁾ reported the isolation of saikosaponins e and f, and five monoacetyl saikosaponins. He also reported comparative carbon-13 nuclear magnetic resonance (13 C-NMR) studies on 23 related compounds excluding saponins possessing a homoannular diene moiety. The structure of saikosaponin h was clarified by Kimata *et al.*, but the spectral data were not given. As to the structures of saikogenins possessing a homoannular diene, saikogenin B was reported by Kubota and Tonami⁹⁾ and saikogenin H by Takeda, but no detailed spectral data for saikogenin H were presented. As to the report on 13 C-NMR studies of triterpenoid possessing a homoannular diene moiety, only Diaz *et al.* reported 11 0 about 12 1 about 12 2 acetoxyoleana-9,12-diene.

This paper reports the isolation and structural identification of eighteen derivatives, including five new compounds, which were formed by pseudo-metabolic reactions, as well as comparative ¹³C-NMR studies of the compounds obtained, which include derivatives possessing a homoannular diene moiety.

Results and Discussion

Treatment of saikosaponin a (1) with 1 N sulfuric acid—dioxane (1:1) for 1 h at 60 °C, followed by separation by reversed-phase high performance liquid chromatography (HPLC) or on a Lobar column afforded known saikosaponin b_1 (2), identical with an authentic sample, and a new saikosaponin g (3) (resulting ratio, 3:1). Compound 3 was suggested to have a homoannular diene moiety at C-9(11),12 on the basis of the observation of a

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Chart 1

ultraviolet (UV) absorption maximum at 280 nm. This was supported by the singlet signal due to two olefinic protons at δ 5.62 in the proton nuclear magnetic resonance (1 H-NMR) spectrum of the peracetate (22), prepared by acidic hydrolysis of 3 and acetylation of the resulting saikogenin H (16). The 13 C-NMR signals due to the sugar moiety of 3 were identical with those of 1 (Chart 1 and Table I). Thus, the structure of 3 was determined to be 3β , 16β ,23,28-tetrahydroxyoleana-9(11),12-diene 3-O- β -D-glucopyranosyl-($1\rightarrow 3$)- β -D-fucopyranoside. Saikosaponin c (4) was also treated by the same procedure as described for 1 to give saikosaponin h (5) and a new compound, saikosaponin i (6). The structure of 6 was determined to be 3β , 16β ,28-trihydroxyoleana-9(11),12-diene 3-O- β -D-glucopyranosyl-($1\rightarrow 6$)-[α -L-rhamnopyranosyl-($1\rightarrow 4$)]- β -D-glucopyranoside mainly based on the 13 C-NMR spectral analysis (Chart 2 and Table I). On acidic hydrolysis of 5 and 6, saikogenin C (18) and saikogenin B (19) were obtained, respectively. Saikosaponin d (7) was treated with 1 N sulfuric acid-dioxane (1:1) for 30 min at 30 °C to give known saikosaponin b₂ (8) only, no saponin possessing a homoannular diene moiety being detected in this case (Chart 3).

Compounds, 1, 2, 3, 7, and 8, were incubated with snail enzyme for 12 h at 37 °C and prosaikogenins F (9), A (10), H (11), G (12) and D (13), respectively, were formed by the

Chart 3

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TABLE I. 13C-NMR Spectral Data for Saikosaponins

					TABLE 1.		C-14WIK Spectral Bata for Salkosaponina				
	1	9	14	7	12	20	4	17 ^{a)}	2 ^{a)}	10	
C-1	38.7	38.6	38.6	38.5	38.7	38.6	38.4	38.8	38.7	38.4	
C-2	$26.0^{b)}$	$26.0^{b)}$	27.5	26.0	26.1	27.6	26.5	27.9	25.8	26.0	
C-3	81.6	81.7	73.2	81.7	81.8	73.4	88.9	78.0	82.4	81.6	
C-4	43.7	43.7	43.1	43.7^{d}	43.8^{d}	43.1	39.6	39.5	43.7	43.6	
C-5	47.3	47.4	49.7	47.3	47.6	48.4	55.8	55.3	48.1	47.4	
C-6	17.6	17.4	18.0	17.6	17.7	18.0	18.4	18.2	18.8	18.9	
C-7	31.5	31.6	31.6	31.5	31.6	31.6	31.5	31.9	32.2	32.2^{h}	
C-8	42.2	42.2	42.2	41.9	42.0	41.9	42.1	42.2	40.8	40.5	
C-9	53.0	53.1	53.1	53.0	53.1	53.0	52.7	53.0	54.7	54.5	
C-10	36.2	36.3	36.6	36.3	36.4	36.6	36.2	36.7	36.9	36.6	
C-11	132.2	132.2	132.2	131.9	132.0	132.0	132.0	132.1	127.1	127.1	
C-12	131.1	131.2	131.2	131.9	132.0	132.0	131.0	131.2	125.7	125.7	
C-13	83.9	84.0	84.0	84.9	84.9	85.0	83.9	84.0	136.8	136.6	
C-14	45.6	45.7	45.7	43.6^{d}	43.7^{d}	43.6	45.6	45.6	44.6^{f})	44.3 ^f	
C-15	36.2	36.1	36.1	35.4	35.5	35.4	36.2	36.3	$35.1^{g)}$	34.9^{g}	
C-16	64.0	64.0	64.0	77.1	77.2	77.2	64.0	64.0	76.5	76.5	
C-17	46.9	47.0	47.0	45.3	45.4	45.4	46.9	47.0	44.8^{f})	44.8 ^f	
C-18	52.1	.52.1	52.1	51.3	51.4	51.4	52.0	52.2	133.6	133.6	
C-18 C-19	37.7	37.8	37.8	38.5	38.5	38.5	38.1	37.8	38.7	38.4	
C-19	31.5	31.6	31.6	31.9	32.0	31.9	31.5	31.6	32.8	32.7	
C-20 C-21	34.7	34.7	34.7	36.8	36.9	36.8	34.8	34.7	$35.4^{g)}$	35.19	
C-21 C-22	25.7^{b}	25.7^{b}	25.7	31.2	31.3	31.3	25.7	25.7	30.2	30.0	
C-22 C-23	64.0	64.3	67.5	64.1	64.5	67.8	27.8	28.4	65.2	64.3	
C-23 C-24	13.0	13.0	12.5	13.0	13.1	12.5	16.3	15.9	12.9	12.9	
C-24 C-25	18.7	18.8	18.6	18.8	18.9	18.7	18.1	18.2	18.5	18.2	
C-25	20.0	20.0	20.0	19.5	19.6	19.5	19.9	20.0	17.3	17.4	
C-20 C-27	20.8	20.0	20.9	18.1	18.2	18.0	20.9	20.9	22.1	21.9	
C-27	73.0	72.8	73.0	77.7	77.9	77.8	72.9	73.0	64.1	63.9	
C-28 C-29	33.6	33.6	33.6	33.7	33.8	33.8	33.6	33.7	25.0	24.8	
C-29 C-30	23.8	23.8	23.8	24.4	24.5	24.4	23.8	23.8	32.8	32.3	
C-30 C-1'		106.2	23.0	105.9	106.3	27.7	106.4	23.0	105.5	106.3	
C-1 C-2'	105.9	73.0		71.5^{e}	73.1		75.4		71.8	73.0	
C-2 C-3'	71.5	75.5		85.1	75.5		76.7		85.2	75.5	
	85.1			71.7 ^{e)}	73.3 72.9		79.7		72.1	72.8	
C-4′	71.7	72.8		71.7	72.9		75.4		71.0	71.3	
C-5'	70.9	71.3			17.5		68.9		17.0	17.1	
C-6′	17.2	17.3		17.2	17.3		102.7		106.0	17.1	
C-1′′	106.4			106.5	•		72.5		75.7		
C-2′′	75.7			75.7			72.5 72.5		78.3		
C-3''	78.6 ^{c)}			$78.6^{c)}$ $72.1^{e)}$			73.7		73.3		
C-4''	72.1						70.5		78.3		
C-5''	78.3°)			78.3°)			18.1		63.1		
C-6''	62.6			62.7			104.9		03.1		
C-1'''							104.9 74.9				
C-2'''											
C-3'''							78.2				
C-4'''						i.	71.3				
C-5'''							78.2				
C-6'''							62.4				

 $^{^{13}}$ C-NMR Spectra were observed at 25 °C. a) Taken from ref. 6. b—l) Assignments may be reversed in each column.

removal of the terminal glucose from the starting saponins (Charts 1 and 3). Compounds 10 and 13 were shown to be the 3-O- β -D-fucopyranosides of saikogenin A (15) and saikogenin D (21), respectively, based on observation of a UV absorption maximum at 254 nm, indicative of

15 ^{a)}	8 ^{a)}	13	21	5	18 ^{a)}	3	11	16	6	19
38.4	38.7	38.4	38.4	38.6	38.8	37.7	37.6	37.6	37.4	37.8
27.6	25.9	26.0	27.6	26.5	28.1	26.9	26.8	28.4	27.3	28.8
73.1	82.6	81.8	73.3	89.0	78.5	81.6	81.7	73.0	88.8	77.7
43.1	43.7	43.7	43.0	39.6	39.6	43.8	43.7	43.2	39.7	39.6
48.2	48.2	47.5	48.4	55.3	55.7	43.8	43.7	44.6	51.8	51.8
18.6	18.8	18.9	18.7	18.5	19.0	18.0	18.1	18.5	18.5	18.7
32.4^{h}	32.5	32.4	31.9	32.7	33.1	32.1	32.2	32.2	32.4	32.5
40.5	41.4	41.1	41.0	40.4	40.7	43.2^{k}	43.2^{k}	43.2	43.1^{k}	43.1
54.5	54.2	54.0	54.0	54.2	54.6	155.0	155.0	155.0	154.7	154.9
36.8	36.9	36.6	36.8	36.5	37.3	38.7	38.7	39.0	38.7	39.1
127.1	126.3	126.2	126.2	127.0	127.1	116.1	116.1	116.1	116.1	116.1
125.7	126.3	126.2	126.2	125.7	125.8	121.2	121.2	121.2	121.2	121.2
136.4	137.1	136.1	136.0	135.8	136.7	145.3	145.3	145.4	145.3	145.4
44.4	42.2	41.9	41.9	44.3^{f})	44.6^{f})	$43.3^{k)}$	$43.3^{k)}$	43.2	43.2^{k}	43.3
34.8^{g}	32.8	32.6	32.8	34.8^{g}	$35.1^{g)}$	36.2	36.2	36.2	36.1	36.1
76.5	68.8	67.7	67.6	76.6	76.5	66.8	66.8	66.8	66.8	66.8
44.4	45.4	45.3	45.3	44.4^{f})	44.8^{f})	40.6	40.6	40.6	40.6	40.6
33.3	133.0	133.1	133.0	133.4	133.7	42.7	42.7	42.7	42.7	42.7
38.4	39.2	39.0	39.0	38.5	38.7	47.0	47.0	47.0	47.1	47.1
32.7	32.8	32.6	32.6	32.7	32.8	31.0	31.0	31.0	31.0	31.0
$35.1^{g)}$	35.8	35.4	35.4	$35.1^{g)}$	$35.4^{g)}$	34.1	34.1	34.2	34.1	34.1
29.9	24.9	24.4	24.4	30.0	30.2	26.2	26.2	26.1	26.1	26.1
67.4	65.3	64.7^{i}	$64.7^{i)}$	28.0	28.5	64.4	64.7	67.7	28.4	28.9
12.6	12.9	12.6	12.6	16.4	15.8	13.7	13.6	13.2	17.1	16.6
18.6	18.6	18.7	18.7	18.5	18.3	$21.1^{(l)}$	$21.1^{l)}$	$21.1^{(l)}$	21.1^{l}	21.0
17.1	17.5	17.3	17.3	17.0	17.2	$21.3^{(l)}$	$21.3^{l)}$	$21.3^{l)}$	$21.2^{l)}$	21.3
22.0	22.3	21.9	21.9	22.0	22.1	26.2	26.2	26.1	25.4	25.6
64.0	65.6	$64.4^{i)}$	$64.4^{i)}$	63.9	64.1	69.5	69.4	69.4	69.3	69.4
24.8	25.3	25.1	25.1	24.8	25.0	33.2	33.2	33.2	33.2	33.2
32.3 th)	32.5	32.4	32.4	32.3	32.3	24.1	24.1	24.1	24.0	24.0
	105.5	106.3		106.7		106.0	106.3		106.7	
	71.8	73.0		75.2		71.6	73.0		75.1	
	85.2	75.5		76.8		85.3	75.5		76.8	
	72.1	72.9		79.9		71.8	72.8		79.8	
	71.0	71.3		75.5		71.0	71.3		75.5	
	17.0	17.3		69.1		17.3	17.4		69.0	
	105.9			102.9		106.7			102.9	
	75.6			$72.5^{j)}$		75.8			72.5	
	78.2			$72.6^{j)}$		78.8			72.6	
	72.1			73.8		72.1			73.8	
	78.1			70.6		78.4			70.6	
	63.1			18.2		62.7			18.5	
				105.1					105.1	
				74.8					74.8	
				78.4					78.5	
				71.4					71.4	
				78.4					78.4	
				62.6					62.5	

the existence of a heteroannular diene moiety at C-11,13(18), as well as on the 13 C-NMR spectrum which exihibited signals due to carbons of the fucose moiety. Compound 11 was also characterized as the 3 -O- β -D-fucopyranoside of 16 based on analysis of the 13 C-NMR data.

When treated by the same procedure as mentioned above, 4, 5 and 6 remained unchanged and the starting materials were recovered.

Treatment of 1 with 2 N sulfuric acid—dioxane (1:1) for 2 h at 100 °C gave a mixture of 15 and 16, which were separated on a Lobar column. Treatment of 7 by the same procedure as mentioned above gave saikogenin D (21) only. Saikogenin C (18) and 19 were obtained by the treatment of 4 with 2 N sulfuric acid—dioxane (1:1) for 6 h at 100 °C, followed by purification on a Lobar column.

Saikogenins F (14), E (17) and G (20) were derived from 1, 4 and 7, respectively, by incubation with intestinal flora of mouse for 4h at 37 °C (Charts 1, 2 and 3).

As described above, we isolated 18 kinds of derivatives from saikosaponins a (1), c (4) and d (7) under pseudo-physiological and chemical conditions. The detection of these compounds in feces, urine and blood after the oral administration of saikosaponins is being attempted.

Experimental

All melting points were measured on a Yanagimoto microscope hot plate and are uncorrected. Infrared (IR) spectra were determined on a JASCO IR spectrometer. Optical rotations were measured with a DIP-140 degital polarimeter. UV spectra were taken with a Shimadzu UV-240 spectrometer. 1 H-NMR spectra were measured on a JEOL JNM-MH-100 spectrometer and 13 C-NMR spectra were measured on a JEOL FX-100 spectrometer with tetramethylsilane (TMS) as an internal standard. The 13 C-NMR signal assignments were carried out by means of known chemical shift rules, off-resonance decoupling studies and also by comparison with the reported data for known compounds. $^{1-11}$ The chemical shifts are given in δ values. Thin layer chromatography (TLC) was performed on Kieselgel 60 F₂₅₄ (Merck) and RP-18 F₂₅₄ (Merck). For column chromatography, Kieselgel 60 (Merck) and LiChroprep RP-18 (40—60 μ m) (Merck) were used. Saikosaponins a, c and d used in these experiments were supplied by Takeda Chemical Industries Co., Ltd.

Acidic Treatment of Saikosaponins a (1), c (4) and d (7)—Compound 1 (200 mg) was dissolved in dioxane (20 ml) and 1 N sulfuric acid (20 ml) was added. The mixture was incubated for 1 h at 60 °C with stirring. After cooling, the solution was neutralized with 10% sodium hydroxide and extracted with *n*-butanol. The butanol layer was washed with water and evaporated to dryness. The residue (170 mg) was separated into saikosaponin b₁ (2) (122 mg) and saikosaponin g (3) (40 mg) on a LiChroprep RP-18 column. The solvent system employed was methanol—water (6:1). Compound 3 was obtained as a white powder, mp 239—245 °C. [α]_D²⁵ +92.3° (c=0.2 in methanol). UV λ _{max} nm (ϵ): 281 (6600). *Anal*. Calcd for C₄₂H₆₈O₁₃·H₂O: C, 63.12; H, 8.84. Found: C, 62.80; H, 9.03.

Compound 4 (200 mg) was treated as mentioned above and the reaction product (182 mg) was separated into saikosaponin h (5) (135 mg) and saikosaponin i (6) (42 mg) on a LiChroprep RP-18 column. The solvent system employed was methanol-water (4:1). Compound 5 was obtained as a white powder, mp 204—208 °C. $[\alpha]_D^{25}$ -63.3° (c=0.5 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 242 (18600), 250 (20600), 259 (12900). Anal. Calcd for $C_{48}H_{78}O_{17} \cdot 2H_2O$: C, 59.84; H, 8.59. Found: C, 59.82; H, 8.50. Compound 6 was obtained as a white powder, mp 201—204 °C. $[\alpha]_D^{25}$ +52.6° (c=0.5 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 280 (6480). Anal. Calcd for $C_{48}H_{78}O_{17} \cdot 3H_2O$: C, 58.74; H, 8.33. Found: C, 59.22; H, 8.46.

Compound 7 (50 mg) was dissolved in dioxane (5 ml), and 1 N sulfuric acid (5 ml) was added. After incubation for 1 h at 30 °C, the solution was neutralized with 10% sodium hydroxide and extracted with *n*-butanol. The extract was washed with water and evaporated to dryness. The residue (48 mg) was purified by silica gel column chromatography to afford saikosaponin b_2 (8) (45 mg). The solvent system was the lower layer of chloroform-methanol-water (65:35:10).

Partial Hydrolysis of 1, 7, 2, 3 and 8 by Snail Enzyme—Compounds, 1, 7, 2, 3 and 8 (each 50 mg) were each dissolved in 10% ethanol solution (300 ml) and the snail enzyme (100 mg) was added. After incubation for 12 h at 37 °C, each solution was extracted with ether. The ether layer was washed with water, dried with sodium sulfate and evaporated to dryness. Each residue was chromatographed on silica gel [solvent system: the lower layer of chloroform—methanol—water (65:23:10)] to afford prosapogenins F (9) (30 mg), G (12) (16 mg), A (10) (28 mg), H (11) (23 mg) and D (13) (30 mg), respectively. Compound 10 was obtained as a white powder, mp 209—212 °C [α]_D²⁵ -63.3° (c=0.2 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ε): 242 (20620), 254 (23140), 259 (14810). Anal. Calcd for $C_{36}H_{58}O_8 \cdot 2H_2O$: C, 67.88; H, 9.50. Found: C, 67.36; H, 9.26. Compound 11 was obtained as a white powder, mp 216—221 °C. [α]_D²⁵ $+129.2^{\circ}$ (c=0.2 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ε): 282 (7880). Anal. Calcd for $C_{36}H_{58}O_8 \cdot 1/2 H_2O$: C, 68.85; H, 9.48. Found: C, 68.64; H, 8.99. Compound 13 was obtained as a white powder, mp 202—205 °C. [α]_D²⁵ -37.0° (c=0.2 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ε): 242 (20020), 254 (22250), 260 (14580). Anal. Calcd for $C_{36}H_{58}O_8 \cdot H_2O$: C, 67.88; H, 9.50. Found: C, 67.10; H, 8.81.

Hydrolysis of 1, 4 and 7 by Mouse Intestinal Flora—Intestinal flora obtained from fifty male mice (ddY) was added to 500 ml of Davis' medium ($K_2HPO_4 \cdot 2H_2O \cdot 7g$, $KH_2PO_4 \cdot 2g$, $MgSO_4 \cdot 7H_2O \cdot 0.1g$, (NH_4)₂SO₄ 1g, $Na_3C_6H_5O_7 \cdot 2H_2O$ and glucose 2g were made up to 1000 ml and the pH was adjusted to 7.0), and the mixture was filtered through cotton. Compound 1, 4 or 7 (each 50 mg) in ethanol (10 ml) was added to the medium and incubated for 12 h at 37 °C. Each solution was extracted with ether and the ether layer was washed with water. After evaporation of the organic layer, the residue was chromatographed on silica gel [solvent system: chloroform—methanol (8:1 for 14 and 20, 10:1 for 17)] to afford saikogenin F (14) (20 mg), saikogenin E (17) (12 mg) or saikogenin G (20) (11 mg), respectively.

Acidic Hydrolysis of 1, 4 and 7—Compound 1 (100 mg) was dissolved in dioxane (20 ml), and 1 N sulfuric acid (20 ml) was added. The mixture was refluxed for 3 h at 120 °C. After cooling, the solution was extracted with chloroform. The organic layer was washed with water and evaporated to dryness. The residue was separated into saikogenin A (15) (39 mg) and saikogenin H (16) (12 mg) on a LiChroprep RP-18 column [solvent system: methanolwater (8:1)]. Compound 16 was obtained as a white powder, mp > 300 °C. [α]_D²⁵ + 209.8° (c=0.5 in methanol). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ε): 282 (9060). Anal. Calcd for $C_{30}H_{48}O_4 \cdot 1/2H_2O$: C, 74.79; H, 10.26. Found: C, 74.59; H, 10.02. The peracetate (22) of 16 was obtained by dissolving 16 (2 mg) in pyridine (1 ml) and acetic anhydride (1 ml) for 12 h at room temperature. Compound 22 was obtained as a white powder, mp 197—201 °C. [α]_D²⁵ +218° (c=0.4 in chloroform). IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1740, 1240, 1032. UV $\lambda_{\text{max}}^{\text{CHCl}_3}$ nm (ε): 282 (7870). ¹H-NMR (CDCl₃): 3.69 (1H, d, J=12 Hz), 3.98 (1H, d, J=12 Hz), 4.03 (1H, d, J=12 Hz), 4.21 (1H, d, J=12 Hz), 4.70—4.90 (1H, m), 5.42—5.64 (1H, m), 5.62 (2H, s). Anal. Calcd for $C_{38}H_{56}O_8$: C, 71.21; H, 8.81. Found: C, 70.64; H, 8.92.

Compound 4 (200 mg) was dissolved in dioxane (20 ml), and 1 N sulfuric acid (20 ml) was added. The mixture was refluxed for 6 h at 120 °C. After cooling, the solution was extracted with chloroform, washed with water and evaporated to dryness. The residue was separated into saikogenin C (18) (58 mg) and saikogenin B (19) (20 mg) on a LiChroprep RP-18 column [solvent system: methanol-water (15:1)].

Compound 7 (100 mg) was dissolved in dioxane (20 ml) and 1 N sulfuric acid (20 ml) was added. The mixture was refluxed for 2 h at 120 °C. After cooling, the solution was extracted with chloroform, washed with water and evaporated to dryness. The residue was chromatographed on silica gel to afford saikogenin D (21) (53 mg) [solvent system: chloroform—methanol (8:1)].

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References

- 1) S. Shibata, I. Kitagawa and H. Fujimoto, Tetrahedron Lett., 1965, 3783.
- 2) S. Shibata, Chem. Pharm. Bull., 14, 1022 (1966).
- 3) T. Kubota, F. Tonami and H. Hinoh, Tetrahedron, 23, 3333 (1967).
- 4) T. Kubota, F. Tonami and H. Hinoh, Tetrahedron, 24, 675 (1968).
- 5) T. Kubota, F. Tonami and H. Hinoh, Tetrahedron Lett., 1968, 303.
- 6) A. Shimaoka, S. Seo and H. Minato, J. Chem. Soc., 1975, 2043.
- 7) H. Ishii, M. Nakamura, S. Seo, K. Tori, T. Tozyu and Y. Yoshimura, Chem. Pharm. Bull., 28, 2367 (1980).
- 8) H. Kimata, C. Hiyama, S. Yahara, O. Tanaka, O. Ishikawa and M. Aiura, Chem. Pharm. Bull., 27, 1836 (1979).
- 9) T. Kubota and F. Tonami, Tetrahedron, 23, 3353 (1967).
- 10) K. Takeda, "Metabolism and Disease," Vol. 10, ed. by Nakayama-Shoten, Tokyo, 1973, p. 682.
- 11) J. G. Diaz, B. M. Fraga, A. G. Gonzalez, P. Gonzalez, M. G. Hernandez and J. M. Miranda, *Phytochemistry*, 23, 1471 (1984).