

Communications to the Editor

[Chem. Pharm. Bull.]
34(11)4887—4888(1986)

FLUOROMETRIC DETERMINATION OF UNSATURATED DISACCHARIDES PRODUCED
ENZYMATICALLY FROM GLYCOSAMINOGLYCANS AND SEPARATED BY HIGH
PERFORMANCE LIQUID CHROMATOGRAPHY

Kazufusa Shinomiya, Ichiro Koshiishi, Toshio Imanari,*,^a
Motoko Takeda, Masako Maeda and Akio Tsuji^b

Faculty of Pharmaceutical Sciences, Chiba University,^a 1-33, Yayoi-
cho, Chiba-shi, Chiba 260, Japan and School of Pharmaceutical Sciences,
Showa University,^b 1-5-8, Hatanodai, Shinagawa-ku, Tokyo 142, Japan

Unsaturated disaccharides produced enzymatically from glycosamino-
glycans were converted to fluorescent derivatives by treatment with
dansylhydrazine. They were separated by high performance liquid
chromatography with a TSK-gel NH₂-60 column (Toyo Soda Co. Ltd.) using
acetonitrile-0.1 M acetate buffer (pH 5.6) (90/20: v/v) as the mobile
phase. The detection limit was a few pmol.

KEYWORDS ——— unsaturated disaccharide; glycosaminoglycan;
high performance liquid chromatography; dansylhydrazine

Although paper chromatography has been used for many years to analyze unsatu-
rated disaccharides produced from glycosaminoglycans (GAGs) by digestion with
enzymes such as chondroitinase AC and ABC, high performance liquid chromatography
(HPLC) has been shown to be a faster and more sensitive method.¹⁾ Recently,
Kodama et al.²⁾ developed a sensitive determination method using HPLC with a pre-
column derivatization process. In their method, unsaturated disaccharides were
converted to the corresponding pyridylamino derivatives and submitted to HPLC
equipped with a fluorometric detector. However, this method was very complicated
and the derivatizing was time-consuming.

In this communication, we describe the chromatographic separation of unsatu-
rated disaccharides using dansylhydrazine (N-dimethylaminonaphthalene-5-sulphonic
acid hydrazide) as a prelabeling reagent. This process was applied to the deter-
mination of reducing sugars^{3,4,5)} and uronic acids.⁶⁾

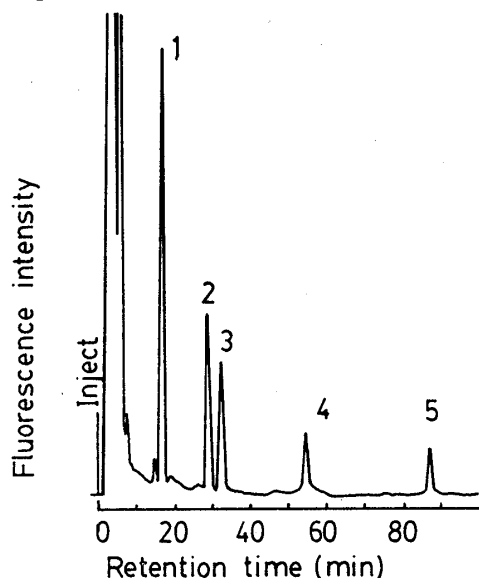
The derivatization procedure was as follows: To 10 μ l of the aqueous 90%
methanol sample solution containing 8 pmol to 40 nmol of unsaturated disaccharides,
20 μ l each of 0.75% (w/v) trichloroacetic acid-ethanol solution and 1.0% (w/v)
dansylhydrazine ethanol solution were added. The mixture was incubated at 40°C
for 45 min and then cooled to room temperature. An aliquot of the resulting
solution was submitted to HPLC. A typical chromatogram of unsaturated disaccha-
rides and the conditions of HPLC are shown in Fig.1.

The calibration curve for 4Di-OS was linear in the range of 0.8 μ M to 4 mM

with a sample size of 10 μ l. The curves for Δ Di-6S and Δ Di-4S were 2.8 μ M to 4 mM. The detection limits (S/N = 2) were 2-7 pmol.

The retention times of the dansylhydrazine derivatives of unsaturated disaccharides are summarized in Table I.

The proposed method may be useful for the separation and quantification of nmol-pmol levels of the unsaturated disaccharides produced enzymatically from GAGs.



1, Δ Di-0S; 2, Δ Di-6S; 3, Δ Di-4S;
4, Δ Di-diS_D; 5, Δ Di-diS_E.

Conditions

Column: TSK-gel NH₂-60 (4.6 mm i.d. x 250 mm).

Mobile phase: acetonitrile-0.1 M acetate
buffer (pH 5.6) (90/20: v/v).

Flow rate: 1.0 ml/min.

Detector: Shimadzu FLD-1 fluorometric
detector (Cut filter EM-5).

Sample size: 10 μ l.

Sample: 0.2 nmol each of unsaturated
disaccharides.

Fig.1 Chromatogram of Dansylhydrazine Derivatives of Unsaturated Disaccharides

Table I. Retention Times of Dansylhydrazine Derivatives of Unsaturated Disaccharides

| Compound | Retention time (min) | Compound | Retention time (min) |
|----------------|----------------------|------------------------------|----------------------|
| Δ Di-HA | 14.0 | Δ Di-UA2S | 25.6 |
| Δ Di-0S | 17.0 | Δ Di-diS _B | 79.6 |
| Δ Di-4S | 32.0 | Δ Di-diS _D | 54.0 |
| Δ Di-6S | 28.4 | Δ Di-diS _E | 87.0 |

Abbreviations used: Δ Di-HA, 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-D-glucose; Δ Di-0S, 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-D-galactose; Δ Di-4S, 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-4-O-sulfo-D-galactose; Δ Di-6S, 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-6-O-sulfo-D-galactose; Δ Di-UA2S, 2-acetamido-2-deoxy-3-O-(2-O-sulfo- β -D-glucopyranosyluronic acid)-D-galactose; Δ Di-diS_B, 2-acetamido-2-deoxy-3-O-(2-O-sulfo- β -D-glucopyranosyluronic acid)-4-O-sulfo-D-galactose; Δ Di-diS_D, 2-acetamido-2-deoxy-3-O-(2-O-sulfo- β -D-glucopyranosyluronic acid)-6-O-sulfo-D-galactose; Δ Di-diS_E, 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-4,6-di-O-sulfo-D-galactose.

ACKNOWLEDGEMENT The authors gratefully acknowledge the supply of the unsaturated disaccharides from Dr. K. Sakurai and Dr. K. Yoshida, Seikagaku Kogyo Co.

REFERENCES

- 1) K. Murata and Y. Yokoyama, *Anal. Biochem.*, **146**, 327 (1985).
- 2) C. Kodama, N. Ototani, M. Isemura and Z. Yosizawa, *J. Biochem.*, **96**, 1283 (1984).
- 3) G. Avigad, *J. Chromatogr.*, **139**, 343 (1977).
- 4) W. F. Alpenfels, *Anal. Biochem.*, **114**, 153 (1981).
- 5) M. Takeda, M. Maeda and A. Tsuji, *J. Chromatogr.*, **244**, 347 (1982).
- 6) M. Takeda, M. Maeda and A. Tsuji, *Bunseki Kagaku*, **33**, 681 (1984).

(Received September 11, 1986)