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The Constituents of *Eucommia ulmoides* OLIV. III. Isolation and Structure of a New Lignan Glycoside

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A new lignan glycoside was isolated from the bark of *Eucommia ulmoides* OLIV. (Eucommiaceae) and its structure was established as (+)-1-hydroxypinoresinol 4''-O- β -D-glucopyranoside (1) on the basis of chemical evidence and spectroscopic studies. An isomer of 1, (+)-1-hydroxypinoresinol 4'-O- β -D-glucopyranoside (2), was also isolated, together with (\pm) -erythro-, and (\pm) -threo-guaiacylglycerol (3 and 4), (+)-cyclo-olivil (5) and (-)-olivil (6).

Keywords—*Eucommia ulmoides*; lignan; (+)-1-hydroxypinoresinol 4''-*O*- β -D-glucopyranoside; (+)-1-hydroxypinoresinol 4'-*O*- β -D-glucopyranoside; (±)-guaiacylglycerol; (+)-cycloolivil; (-)-olivil; ¹³C-NMR

In a previous paper,¹⁾ we reported the isolation of (-)-olivil 4',4''-di-O- β -D-gluco-pyranoside, (+)-1-hydroxypinoresinol 4',4''-di-O- β -D-glucopyranoside (7), eucommin A [(+)-medioresinol 4'-O- β -D-glucopyranoside (8)] and (+)-syringaresinol O- β -D-glucopyranoside (9) from the air-dried bark of *Eucommia ulmoides* OLIV. (Eucommiaceae), which is one of the longest-known tonic herbs in China. As a continuation of our investigation on this crude drug, this paper describes the isolation of a new lignan glycoside, (+)-1-hydroxypinoresinol 4''-O- β -D-glucopyranoside (1), together with a known lignan glycoside, (+)-1-hydroxypinoresinol 4'-O- β -D-glucopyranoside (2) and four other known compounds, (\pm)-erythro-guaiacylglycerol (3), (\pm)-threo-guaiacylglycerol (4), (+)-cyclo-olivil (5) and (-)-olivil (6), and the elucidation of their structures on the basis of chemical evidence and spectroscopic analysis.

The extraction and separation were carried out as described in the experimental section. Fraction A was obtained as an amorphous, which showed a spot at Rf 0.22 on thin layer chromatography (TLC) with CHCl₃-MeOH-water (80:20:3) as a developer. The field desorption mass spectrum (FD-MS) of A gave peaks at m/z 537 (M⁺+1) and m/z 375 (M⁺-C₆H₁₀O₅+1). Hydrolysis of A with β -glucosidase gave the aglycone (10) as an amorphous powder, $[\alpha]_D^{19}$ +21° (MeOH). Compound 10 was identified as (+)-1-hydroxypinoresinol by comparison of the ultraviolet (UV), infrared (IR), proton nuclear magnetic resonance (¹H-NMR), carbon-13 nuclear magnetic resonance (¹³C-NMR) and $[\alpha]_D$ data with those of an authentic sample.^{1,2)} The presence of glucose in the hydrolysate was detected by gas chromatography (GC). Thus, fraction A was considered to be (+)-1-hydroxypinoresinol monoglucoside. As shown in Fig. 1, fraction A was separated into two peaks by high-performance liquid chromatography (HPLC) with a reversed-phase column. Peaks 1 and 2 were each isolated and purified by preparative HPLC, yielding 1 and 2.

A new lignan glycoside (1) was isolated as an amorphous, $[\alpha]_D^{18} - 28.8^{\circ}$ (MeOH). The IR spectrum of 1 showed the presence of hydroxyl groups (3420 cm⁻¹) and aromatic rings

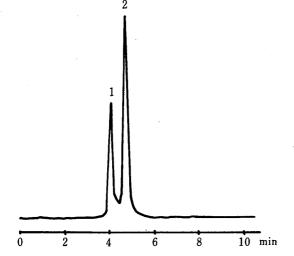


Fig. 1. High-Performance Liquid Chromatogram of Fraction A

Column, Hitachi #3056 ($4 \times 150 \text{ mm}$); eluent, CH₃CN-water (15:85, v/v); flow rate, 1.0 ml/min; detection, UV at 270 nm.

(1608 and 1518 cm⁻¹). The ¹H-NMR spectrum of 1 showed signals at δ 3.75 and 3.77 (each s) due to two aromatic methoxyl groups, and at δ 6.65—7.20 (m) due to six aromatic protons. Acetylation of 1 with acetic anhydride-pyridine gave a hexaacetate (1a) and a pentaacetate (1b). Compound 1a was isolated as an amorphous, the ¹H-NMR spectrum of which showed signals at δ 1.61 (s) due to a tertiary alcoholic acetyl group, at δ 1.96 (3H, s) and 2.00 (9H, s) due to four alcoholic acetyl groups and at $\delta 2.24$ (s) due to a phenolic acetyl group. Compound 1b was isolated as an amorphous. The IR spectrum of 1b suggested the presence of a hydroxyl group (3480 cm⁻¹). The ¹H-NMR spectrum of **1b** showed signals at δ 1.96 (3H. s) and 2.00 (9H, s) due to four alcoholic acetyl groups, and at 2.24 (s) due to a phenolic acetyl group. The above results suggested that 1 is (+)-1-hydroxypinoresinol monoglucoside and that a glucose moiety is attached to a phenolic group of the aglycone (10) at the C-4' or C-4" position. The position of the glucose linkage in 1 was investigated as follows. As shown in Table I, the ¹³C-NMR signals of the carbon atoms of the 4'-free guaiacyl group in 1 were identical with those of the corresponding atoms in 10^{1-3} On the other hand, the signals of the aromatic carbon atoms of the 4''-O- β -D-glucopyranosylguaiacyl group in 1 were identical with those of the corresponding atoms in 7.1) The shifts of corresponding carbons in going to 1 from 10 were +2.9 (C-1'') and +1.4 ppm (C-3''), which indicated that the glucosyl group in 1 was linked to the C-4" position. 1,4) The molecular optical rotation values (M_D) of 1 and 10 are -156° and $+81^{\circ}$, respectively. The difference, $\Delta M_{\rm D}$ (1 $M_{\rm D}-10~M_{\rm D}$), is -237° , which shows that the D-glucopyranosyl moiety is linked in β -form.⁵⁾ Furthermore, the ¹³C-NMR signals of the glucose moiety in 1 were consistent with β -form.⁶⁾ From the above results, 1 was established to be (+)-1-hydroxypinoresinol 4''-O- β -D-glucopyranoside. This is the first reported isolation of 1.

Glycoside 2 was isolated as an amorphous, $[\alpha]_D^{18} - 25.2^{\circ}$ (MeOH). Acetylation of 2 with acetic anhydride-pyridine gave a hexaacetate (2a) and a pentaacetate (2b). Compound 2a was obtained as a white powder (EtOH), mp 103.2 °C, and 2b was obtained as an amorphous. The IR and ¹H-NMR spectra of 2a and 2b were very similar to those of 1a and 1b. Compounds 2 and 2a were identified as (+)-1-hydroxypinoresinol 4'-O- β -D-glucopyranoside and (+)-1-hydroxypinoresinol 4'-O- β -D-glucopyranoside hexaacetate, respectively, by comparison with authentic samples.³⁾

Fraction B was obtained as a syrup, $[\alpha]_D^{28}$ 0° (MeOH). Acetylation of B with acetic anhydride–pyridine gave an amorphous powder, which showed two spots at Rf 0.31 (3a) and 0.27 (4a) on TLC with n-hexane–ether (1:1) as a developer. Compounds 3a and 4a were identified as (\pm) -erythro-guaiacylglycerol tetraacetate and (\pm) -threo-guaiacylglycerol tetraacetate, respectively, by comparison (IR and 1 H-NMR) with authentic samples. $^{7,8)}$

TABLE I. 13 C-NMR Chemical Shifts (in DMSO- d_6)

	1	2	10	7	Δδ (1—10)	Δδ (2—10)	1a	1b	2a	2b
C-1	91.0	91.1	91.1	91.1			96.9	91.2	96.8	91.1
C-5	60.8	60.8	60.9	60.8			58.1	60.9	58.2	60.9
C-4	70.2	70.3	70.2	70.3			71.7	70.4	71.8	70.4
C-8	74.7	74.7	74.9	75.0			73.7	74.7	74.0	74.7
C-2	87.1	86.9	87.1	86.8			85.9	86.5	85.7	86.5
C-6	85.1	85.4	85.5	85.2			83.8	84.8	84.0	84.8
C-1'	127.9	131.1	128.1	131.2		+3.0	135.5	136.2	132.5	133.2
C-1''	135.3	132.3	132.4	135.5	+2.9		136.3	137.5	138.6	138.5
C-2'	112.3	112.5	112.5	112.9		0	113.0	112.1	113.6	112.7
C-2''	110.9	110.7	110.9	111.2	0		111.1	111.0	110.6	110.5
C-3′	146.8	148.3	146.9	148.5		+1.4	150.1	150.0	149.1	149.1
C-3''	148.9	147.5	147.5	149.0	+1.4		149.7	149.8	150.7	150.7
C-4'	145.0	146.0	146.0	146.0		0	138.9	138.5	145.4	145.1
C-4''	145.9	146.0	146.0	146.0	-0.1		145.2	145.0	139.4	140.5
C-5′	114.5	114.7	114.6	115.1		+0.1	118.1	110.2	117.3	117.3
C-5''	115.3	115.1	115.2	115.7	+0.1		118.1	118.2	117.9	117.9
C-6'	120.1	119.8	120.2	119.8		-0.4	122.0	121.6	120.7	119.7
C-6''	118.3	118.8	118.9	118.4	-0.6		118.3	119.5	122.6	122.5
OCH ₃	55.6 55.7	55.6 55.7	55.7	55.9						
Glc 1	100.1	100.3		100.5	•					
Glc 2	73.2	73.2		73.3		OCOCH ₃	{ 20.2	20.2	20.2	20.2
Glc 3	76.9	76.9		76.8		ососн ₃	20.3			
Glc 4	69.7	69.7		69.8						
Glc 5	76.9	76.9		77.0			f 168:3	168.4	168.4	168.5
Glc 6	60.8	60.8		60.8		ососн3	168.7	168.9	168.7	168.9
						OCOCII	168.9	169.1	168.9	169.2
		•				OCOCH3	169.2	169.4	169.2	169.5
							169.5	169.8	169.4	169.8
							l 169.8		169.8	

Compounds 5 and 6 were identified as (+)-cyclo-olivil and (-)-olivil, respectively, by comparison (IR, ¹H-, ¹³C-NMR) with authentic samples.^{2,9)}

Experimental

All melting points are uncorrected. The following instruments were used; melting point, Mettler FP-61; optical rotation value, JASCO DIP-4; UV spectra, Hitachi 200-20; IR spectra, Hitachi 270-30; GC, Hitachi 063 with a hydrogen flame ionization detector; HPLC, Hitachi 638 with a UV detector; EI-MS, Hitachi RMU-7L; FD-MS, JEOL 01-SG2; 1 H-NMR spectra, JEOL JNM-FX-90; 13 C-NMR spectra, JEOL JNM-FX-60, with tetramethylsilane (δ =0) as an internal reference. The abbreviations used for NMR data are as follows: s, singlet; d, doublet; dt, doublet-triplet; t, triplet; m, multiplet.

Precoated TLC plates (Silica gel 60 F_{254} , Merck) were used for TLC. The spots were detected by spraying the plates with 20% H_2SO_4 and by heating. Silica gel (Wako gel C-300, Wako Pure Chemical Co.), polyamide C-200 (Wako Pure Chemical Co.) and Diaion HP-20 (Nippon Rensui Co.) were used for column chromatography. TSK gel HW-40 (Toyo Soda Co.) was used for gel filtration.

Isolation—The air-dried bark of Eucommia ulmoides OLIV. (10 kg, commercial crude drug produced in China) was chopped and extracted with hot water (20 1×3). The extract was filtered and the filtrate was evaporated under reduced pressure to a small volume, which was suspended in water. This suspension was extracted with EtOAc (3 1×2) and then with n-BuOH (3 1×2), successively. The n-BuOH extract was evaporated under reduced pressure, then the residue was taken up in water (2 l) and filtered. The filtrate was subjected to chromatography (Diaion HP-20), eluting with water (2 l), 10%, 50% MeOH (each 3 l) and then 100% MeOH (4 l), successively. The 10% MeOH eluate was concentrated and subjected to silica gel chromatography, eluting with CHCl3-MeOH-water (100:10:1, 90:10:1, 80:20:3, 70:30:5). The fractions were monitored by TLC with CHCl₃-MeOH-water (80:20:3) as a developer. The fractions showing a TLC spot at Rf 0.29 were collected and purified by silica gel chromatography and gel filtration on TSK gel HW-40 with MeOH, yielding fraction B (0.2 g). The 50% MeOH eluate was concentrated and subjected to silica gel chromatography, eluting with CHCl₃-MeOH-water (100:10:1, 80:20:3). The fractions were monitored in the same way as described for B. The fractions showing a TLC spot at Rf 0.22 were collected and purified by repeated silica gel chromatography and gel filtration on TSK gel HW-40 with MeOH-water (1:1), yielding fraction A (0.3 g), which was separated into two peaks (1 and 2) by HPLC. Conditions: column, Hitachi gel #3056, 4 mm × 15 cm; eluent, CH₃CN-water (1:9); flow rate, 1 ml/min; detector, UV detector (270 nm). Compounds 1 and 2 were each collected by repeated HPLC under the above conditions, yielding 1 (30 mg) and 2 (60 mg). The 100% MeOH eluate was concentrated. The concentrate showed spots at Rf 0.22 (fraction A), 0.36 [(+)-pinoresinol $O-\beta$ -D-glucopyranoside], 10) 0.42 (8), 0.43 (5), 0.44 (9) and 0.52 (6) on TLC with CHCl₃-MeOH-water (80:20:3) as a developer, and was subjected to silica gel chromatography with CHCl₃-MeOH (10:1). The fractions showing TLC spots at Rf 0.43 (5) and 0.52 (6) were each collected and purified by silica gel chromatography and on TSK gel HW-40, yielding 5 (0.1 g) and 6 (0.2 g).

Fraction A (1 and 2)—Amorphous, FD-MS m/z: 537 (M⁺+1), 375 (M⁺-C₆H₁₀O₅+1). IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3396 (OH), 1604, 1514 (aromatic ring).

Enzymatic Hydrolysis of Fraction A —Fraction A (100 mg) was hydrolyzed with β-glucosidase (50 mg, Sigma) in acetate buffer (0.1 n HOAc–0.1 m NaOAc=1:1, pH=5.0) for 1 d at 37 °C. The reaction mixture was extracted with Et₂O (50 ml × 2) and the residue obtained from the Et₂O layer was chromatographed on silica gel. Elution with CHCl₃–MeOH (20:1) gave a pure aglycone (7, 30 mg). The aqueous layer was evaporated under reduced pressure to give a residue. This residue was trimethylsilylated with TMS-PZ (Tokyo Kasei Co.) and left for 10 min, then the reaction mixture was extracted with CHCl₃. The CHCl₃ layer was washed with water and concentrated. The presence of trimethylsilylated α-glucose [t_R (min) 4.8] and β-glucose [t_R (min) 6.9] in this residue was detected by GC. Conditions: column, 1.5% OV-17, 3 mm × 1.5 m; column temperature, 200 °C; carrier gas, N₂, 30 ml/min; injection temperature, 220 °C.

- (+)-1-Hydroxypinoresinol (10): Amorphous powder, $[\alpha]_D^{19} + 21^\circ$ (c = 1.0, CHCl₃). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 232, 281. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3416 (OH), 1606, 1520 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 2.80—3.04 (1H, m, C₅-H), 3.46—4.08 (3H, m, C_{4a}-H, C₈-H), 3.66 (6H, s, 2 × OCH₃), 4.20—4.48 (1H, m, C_{4e}-H), 4.52 (1H, s, C₂-H), 4.76 (2H, d, J = 5 Hz, C₆-H), 6.60—7.04 (6H, m, arom. H). ¹³C-NMR: Table I.
- (+)-1-Hydroxypinoresinol 4"-*O*-β-D-Glucopyranoside (1)—Amorphous, $[\alpha]_D^{18}$ -28.8° (c=1.0, MeOH). UV $\lambda_{\max}^{\text{MeOH}}$ nm: 228.5, 278. IR ν_{\max}^{KBr} cm⁻¹: 3420 (OH), 1608, 1518 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 3.75, 3.77 (6H, each s, 2 × OCH₃), 6.65—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.
- (+)-1-Hydroxypinoresinol 4'-*O*-β-D-Glucopyranoside (2)—Amorphous, $[\alpha]_D^{18}$ -25.2° (c=1.0, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 228.5, 279. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3416 (OH), 1606, 1518 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 3.76 (6H, s, 2 × OCH₃), 6.70—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.

Acetylation of 1—1 (20 mg) was acetylated with acetic anhydride-pyridine. The crude acetate was subjected to silica gel chromatography, with benzene-EtOAc (2:1) as a developer. The fractions were monitored by TLC with benzene-EtOAc (1:1) as a developer. The fractions showing TLC spots at Rf 0.25 (1a) and Rf 0.13 (1b) were each collected, yielding 1a (3.6 mg) and 1b (5.6 mg).

(+)-1-Hydroxypinoresinol 4''-O-β-D-Glucopyranoside Hexaacetate (1a): Amorphous powder. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1758, 1230 (OCOCH₃), 1610, 1516 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 1.61 (3H, s, tert. alcoholic OCOCH₃),

1.96, 2.00 (3H, 9H, each s, $4 \times$ alcoholic OCOCH₃), 2.24 (3H, s, phenolic OCOCH₃), 3.76 (6H, s, $2 \times$ OCH₃), 6.70—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.

(+)-1-Hydroxypinoresinol 4''-O-β-D-Glucopyranoside Pentaacetate (**1b**): Amorphous. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3480 (OH), 1758, 1228 (OCOCH₃), 1608, 1514 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 1.96, 2.00 (3H, 9H, each s, 4 × alcoholic OCOCH₃), 2.24 (3H, s, phenolic OCOCH₃), 3.76 (6H, s, 2 × OCH₃), 6.70—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.

Acetylation of 2—2 (50 mg) was acetylated and the crude acetate was treated in the same way as described for 1a and 1b, yielding 2a (Rf 0.27, 23 mg) and 2b (Rf 0.15, 21 mg).

- (+)-1-Hydroxypinoresinol 4'-*O*-β-D-Glucopyranoside Hexaacetate (**2a**): A white powder (EtOH), mp 103.2 °C. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1760, 1226 (OCOCH₃), 1610, 1516 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 1.64 (3H, tert. alcoholic OCOCH₃), 1.96, 2.00 (3H, 9H, each s, 4 × alcoholic OCOCH₃) 2.24 (3H, s, phenolic OCOCH₃), 3.73, 3.79 (6H, each s, 2 × OCH₃), 6.85—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.
- (+)-1-Hydroxypinoresinol 4'-*O*-β-D-Glucopyranoside Pentaacetate (**2b**): Amorphous. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3460 (OH), 1760, 1226 (OCOCH₃), 1608, 1514 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ: 1.97, 2.02, (3H, 9H, each s, 4 × alcoholic OCOCH₃), 2.24 (3H, s, phenolic OCOCH₃), 3.73, 3.79 (6H, each s, 2 × OCH₃), 6.85—7.20 (6H, m, arom. H). ¹³C-NMR: Table I.
- (±)-erythro- and (±)-threo-Guaiacylglycerol (Fraction B; 3 and 4)—Colorless syrup, $[\alpha]_D^{27}$ 0° (c=2.24, MeOH). IR v_{max}^{KBr} cm⁻¹: 3400 (OH), 1608, 1520 (aromatic ring). MS m/z: 214 (M⁺), 196 (M⁺ H₂O), 167, 152, 137, 125, 97, 85.

Acetylation of Fraction B ——Fraction B (0.2 g) was acetylated with acetic anhydride—pyridine. The crude acetate was subjected to silica gel chromatography with *n*-hexane—Et₂O (2:1). The fractions were monitored by TLC with *n*-hexane—Et₂O (1:1) as a developer. The fractions showing spots at Rf 0.31 (3a) and at 0.27 (4a) were each collected, yielding 3a (60 mg) and 4a (150 mg).

- (±)-erythro-Guaiacylglycerol Tetraacetate (**3a**): Amorphous, $[\alpha]_D^{27} 2.5^\circ (c = 2.02, \text{MeOH})$. IR $\nu_{\text{max}}^{\text{KBr}} \text{cm}^{-1}$: 1754, 1224 (OCOCH₃), 1608, 1514 (aromatic ring). ¹H-NMR (in CDCl₃) δ: 2.02, 2.11, 2.16 (9H, each s, 3 × alcoholic OCOCH₃), 2.29 (3H, s, phenolic OCOCH₃), 3.83 (3H, s, OCH₃), 4.24 (2H, d, J = 4.83 Hz, C₉-H), 5.38 (1H, dt, J = 5.71, 4.83 Hz, C₈-H), 6.00 (1H, d, J = 5.71 Hz, C₇-H), 6.86—7.05 (3H, m, arom. H). ¹³C-NMR (in CDCl₃): 55.9 (OCH₃), 61.6 (9), 72.3, 72.7 (8, 7), 111.2 (2), 119.3 (5), 122.8 (6), 134.6 (1), 140.0 (4), 151.2 (3).
- (±)-threo-Guaiacylglycerol Tetraacetate (4a): Colorless prisms (EtOH), mp 115.6 °C, $[\alpha]_D^{27}$ -7.0 ° (c = 3.14, MeOH). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1768, 1224 (OCOCH₃), 1608, 1514 (aromatic ring). ¹H-NMR (in CDCl₃) δ: 2.04, 2.06, 2.08 (9H, each s, 3 × alcoholic OCOCH₃), 2.29 (3H, s, phenolic OCOCH₃), 3.83 (3H, s, OCH₃), 4.15—4.40 (1H, m, C₉-H_e), 5.25—5.55 (1H, m, C₈-H), 5.95 (1H, d, J = 7.48 Hz, C₇-H), 6.90—7.10 (3H, m, arom. H). ¹³C-NMR (in CDCl₃): 56.0 (OCH₃), 62.1 (9), 72.3, 73.4 (8, 7), 111.4 (2), 119.7 (5), 123.1 (6), 134.7 (1), 140.3 (4), 151.5 (3).
- (+)-Cyclo-olivil (5)—A white powder (EtOH), mp 165 °C, $[\alpha]_{0}^{25}$ + 50.0 ° (c = 1.0, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 228, 283. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3400 (OH), 1604, 1516 (aromatic ring). MS m/z: 376 (M⁺), 358 (M⁺ H₂O), 340 (M⁺ 2 × H₂O), 327, 309, 297, 265. ¹H-NMR (in MeOH- d_4) δ : 1.80—2.06 (1H, m, C₂-H), 2.58 (1H, d, J = 17 Hz, C₄-H_a), 3.20 (1H, d, J = 17 Hz, C₄-H_e), 3.73, 3.76 (6H, each s, 2 × OCH₃), 6.16—6.85 (5H, m, arom. H).
- (-)-Olivil (6)—A white powder (EtOH), mp 120 °C, $[\alpha]_D^{25}$ -25.2 ° (c = 2.5, MeOH). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 231, 282. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3368 (OH), 1606, 1516 (aromatic ring). ¹H-NMR (in DMSO- d_6) δ : 3.75 (6H, s, 2 × OCH₃), 6.52—7.08 (6H, m, arom. H).

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