## Communications to the Editor

Chem. Pharm. Bull. 34(2) 931—934 (1986)

SYNTHESES OF CHOLESTEROL ANALOGS WITH A CARBENE-GENERATING SUBSTITUENT ON THE SIDE CHAIN

Takeshi Terasawa, a Nobuo Ikekawa, a and Masuo Morisaki \*,b

Department of Chemistry, Tokyo Institute of Technology, Meguro-ku,
Tokyo 152, and Kyoritsu College of Pharmacy, Minato-ku, Tokyo 105, Japan

As photoaffinity-labelled analogs of cholesterol, the diazoacetate 1, the aryldiazirines 2 and 3, and the fluorodiazirine 4 were prepared.

KEYWORDS-cholesterol analog; photoaffinity label; diazoacetate; aryldiazirine; fluorodiazirine; phospholipid-cholesterol interaction

Since cholesterol is a major lipid constituent of biological membrane, knowledge of its interaction with phospholipid is essential in understanding the architectural and functional roles of cholesterol. 1) Although the molecular interaction of cholesterol with phospholipids has been studied extensively by physicochemical techniques such as differential scanning calorimetry, X-ray diffraction, NMR and ESR, there has been no definite proof of direct interaction between the two components. 2) One approach to this problem is photoaffinity labelling, and cholesteryl diazoacetate has recently been reported as a useful membrane photolabelling reagent. 3,4) However, in view of the importance of the  $3\beta$ -hydroxyl group of cholesterol for interaction with phospholipids,  $^{2)}$  cholesteryl diazoacetate does not appear to be ideal as a membrane probe. describe the syntheses of cholesterol analogs  $\frac{1}{2}$ ,  $\frac{2}{2}$ ,  $\frac{3}{2}$  and  $\frac{4}{2}$  having a carbenegenerating substituent on the side chain,  $\frac{5}{1}$  and in the accompanying paper, the photochemical behavior of these compounds is reported. 6) nature of these substituents and also our previous observations 7) that relatively large modification of the side chain structure of cholesterol causes little or no

Vol. 34 (1986)

perturbation in the properties of liposomes, suggest that compounds 1-4 would be promising alternatives as membrane probes.

932

We first attempted to prepare diazoacetate 1 from 24-hydroxy-3-tetrahydropyranyl(THP)- or 3-t-butyldimethylsilyl(TBDMS)-ether 5a<sup>8</sup> or 6a.<sup>9</sup> When 5a or 6a were treated with glyoxylic acid chloride p-toluenesulfonyl hydrazone and triethylamine according to the published method, 3,10) the sulfinate 5b and 6b, instead of the desired diazoacetate 5c and 6c, were the major products (64% and 51% yield, respectively). Substitution of triethylamine for the weaker base dimethylaniline, 11) gave 5c and 6c in 40% and 49% yield, respectively without formation of 5b and 6b. However, various attempts to deprotect the 3-THP or 3-TBDMS groups of 5c and 6c, induced a concomitant decomposition of the diazoacetyl Accordingly, we turned our attention to selective diazoacetylation of the 3,24-diol 7a. Treatment of 7a with glyoxylic acid p-toluenesulfonyl hydrazone in the presence of dicyclohexylcarbodiimide and 4-dimethylaminopyridine, 12) again afforded the sulfinate 7b as a major product accompanied with a trace of the expected diazoacetate 1. A modest success finally came when the 3,24-diol was subjected to esterification with glyoxylic acid chloride ptoluenesulfonyl hydrazone( l eq) in the presence of dimethylaniline in  $CH_2Cl_2$ -dimethylformamide (10:1) at O°C, followed by treatment with tiethylamine(5 eq). Flash chromatography  $^{13}$ ) of the crude product gave the recovered diol 7a(40%), the less polar 3-diazoacetate 7c(4%), the more polar 24-diazoacetate 1 [13%, mp 83-85°C, UV(EtOH)  $\lambda_{\text{max}} 247 \text{nm}(\xi, 1.0 \times 10^4)$ ; IR(CHCl<sub>3</sub>) 2105(diazo), 1680cm<sup>-1</sup>(carbonyl); H-NMR(CDCl<sub>3</sub>, 270 MHz): 4.13(2H, td, J=6.8 and 3Hz, 24-H<sub>2</sub>), 4.68ppm(lH, s, -CHN<sub>2</sub>)], and a mixture of 1 and 7c (25%).

Syntheses of the aryldiazirines 2 and 3 were patterned after the published preparation of the diazirinophenoxy derivatives of fatty acids. Thus, 22-iodide  $8a^{15}$ ) and sodium  $\underline{m}$ -(3H-diazirino)phenoxide were coupled in hexamethyl-phosphoramide-tetrahydrofuran (3 : 2) to give the ether 8b (40%), which on acid treatment yielded 2 [mp 133-134°C, UV(EtOH)  $\lambda_{max}$  363nm(£, 340); IR(CHCl<sub>3</sub>) 1585cm<sup>-1</sup> (diazo);  ${}^{1}H$ -NMR(CDCl<sub>3</sub>): 1.99(1H, s, CH $^{N}_{N}$ ), 3.64(1H, dd, J=9 and 7Hz, 22-H<sub>a</sub>), 3.87 ppm(1H, dd, J=9 and 3Hz, 22-H<sub>b</sub>)]. Similarly, 24-iodide 9a, which was prepared by the iodide substitution reaction on the corresponding 24-tosylate, was converted into the ether 9b (78%), and then into 3 [mp 65-67°C, UV(EtOH)  $\lambda_{max}$  363 nm(£, 340); IR(CHCl<sub>3</sub>) 1585cm<sup>-1</sup>(diazo);  ${}^{1}H$ -NMR: 1.99(1H, s, CH $^{N}_{N}$ ), 3.89ppm(2H, t, J=7Hz, 24-H<sub>2</sub>)].

For synthesis of the fluorodiazirine 4, the 22-aldehyde  $10a^{16}$  was subjected to the Horner-Emmons reaction with  $(\text{MeO})_2\text{P(O)CH(OTHP)CO}_2\text{Me/lithium diisopropyl-amide}^{17}$  in tetrahydrofuran to give the enol ether 10b (1: 1 mixture of E and Z isomers) in 85% yield, which on refluxing with zinc acetate in acetic acetic acid afforded the oxoester  $11a^{18}$  in 72% yield. Reaction of 11a with diethylaminosulfur trifluoride in  $\text{CH}_2\text{Cl}_2^{19}$  gave the difluoride  $11b^{18}$  (60%). This was converted into the aldehyde hydrate 12a in 57% overall yield by the sequences: 1) hydrolysis ( $K_2\text{CO}_3$ /methanol); 2) esterification( $\text{CH}_2\text{N}_2$ /ether); 3) THP ether formation (dihydropyran/p-toluenesulfonic acid); 4) reduction (diisobutyl aluminum hydride/ether). Transformation of the aldehyde hydrate 12a into the diazirine 12c was patterned after the method of Khorana. The compound 12a was dehydrated by azeotropic refluxing with benzene using a Soxhlet apparatus filled

with 4A molecular sieves. The resulting free aldehyde was refluxed with excess tert-butylamine to give the tert-butylimine derivative. This was unstable and so, it was directly treated with hydroxylamine O-sulfonic acid in ethanol-benzene-triethylamine (10 : 2 : 5) at O°C to yield the N-tert-butyldiazirine 12b (12% yield from 12a), together with the recovered aldehyde hydrate 12a (60%). The diazirine 12b was oxidized with tert-butyl hypochlorite in tert-butanol-ethanol-tetrahydrofuran in the presence of triethylamine at O°C to yield the diazirine 12c in 90% yield. Acid treatment (d.HCl/tetrahydrofuran-methanol) furnished the fluorodiazirine 4 [amorph. UV(EtOH)  $\lambda_{max}$  306, 314 and 319nm( $\varepsilon_{306-318}$ 160); IR (CHCl<sub>3</sub>) 1600cm<sup>-1</sup> (diazo);  $^{1}_{H-NMR}$ (CDCl<sub>3</sub>): 0.72(3H, s, 18-H<sub>3</sub>), 1.00(3H, s, 19-H<sub>3</sub>), 3.5(1H, m, 3d-H), 5.3ppm(1H, m, 6-H)].

ACKNOWLEDGEMENT This research was supported by grant(No. 58570867) from the Ministry of Education, Science and Culture.

## REFERENCES

- 1) G. F. Gibbons, K. A. Mitropoulos, and N. B. Myrant, "Biochemistry of Cholesterol," Chapter 9, Elsevier, Amsterdam, 1982.
- 2) R. A. Demel and B. deKruyff, Biochem. Biophys. Acta,  $\underline{457}$ , 109(1976).
- 3) S. A. Kailbaugh and E. R. Thornton, J. Am. Chem. Soc.,  $\underline{105}$ , 3283(1983).
- 4) S. A. Kailbaugh and E. R. Thornton, Biochemistry, 22, 5063(1983).
- 5) Recently, synthesis of cholesterol analog possessing a flourescent side chain

- for use as a membrane probe has been reported: J. Drew, G. Gowda, P. Morand, P. Proulx, A. G. Szabo, and D. Williamson, J. Chem. Soc., Chem. Commun., 1985, 901.
- 6) T. Terasawa, N. Ikekawa and M. Morisaki, Chem. Pharm. Bull., in press.
- 7) H. Hagiwara, T. Nagasaki, Y. Inada, Y. Saito, T. Yasuda, H. Kojima, M. Morisaki and N. Ikekawa, Biochem. Internat., 5, 329(1982); R. Goto, M. Morisaki and N. Ikekawa, Chem. Pharm. Bull., 31, 3528(1983).
- 8) M. Morisaki, M. Shibata, C. Duque, N. Imamura and N. Ikekawa, Chem. Pharm. Bull., 28, 606(1980).
- 9) J. H. Dygos and B. N. Desai, J. Org. Chem., 44, 1590(1979).
- 10) J. A. Katzenellenbogen, H. N. Myers and H. J. Johnston Jr., J. Org. Chem., 38, 3525(1973).
- 11) E. J. Corey and A. G. Myers, Tetrahedron Lett., 25, 3559(1984).
- 12) R. Sen, J. D. Carriker, V. Balogh-Nair and K. Nakanishi, J. Am. Chem. Soc., 104, 3214(1982).
- 13) W. C. Still, M. Kahn and A. Mitra, J. Org. Chem., 43, 2923(1978).
- 14) R. Radhakrishnan, R. J. Robson, Y. Takagaki and H. G. Khorana, Methods in Enzymology, 72D, 408(1981).
- 15) S. Sato, A. Akaiwa, Y. Fujimoto, M. Ishiguro and N. Ikekawa, Chem. Pharm. Bull., 29, 406(1981).
- 16) G. D. Anderson, T. J. Powers, C. Djerassi, J. Fayos and J. Clardy, J. Am. Chem. Soc., <u>97</u>, 388(1975).
- 17) E. Nakamura, Tetrahedron Lett., 22, 663(1981).
- 18) T. Taguchi, S. Mitsuhashi, A. Yamanouchi, Y. Kobayashi, H. Sai and N. Ikekawa, Tetrahedron Lett., <u>25</u>, 4933(1984).
- 19) W. J. Middleton, J. Org. Chem., 40, 574(1975).
- 20) B. Erni and H. G. Khorana, J. Am. Chem. Soc., 102, 3888(1980).

(Received November 28, 1985)