Chem. Pharm. Bull. 34(3)1128—1147(1986)

Synthesis and Angiotensin Converting Enzyme Inhibitory Activity of 1,5-Benzothiazepine and 1,5-Benzoxazepine Derivatives. I

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(Received August 23, 1985)

The design and synthesis of new structural types of angiotensin converting enzyme (ACE) inhibitors, (R)-3-amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic acids (7, 26, 33 and 37) and (S)-3-amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetic acids (8 and 27), are described. A number of compounds in these series showed potent ACE inhibitory activity in vitro and in vivo. The structure-activity relationship is also discussed.

Keywords—angiotensin converting enzyme inhibitor; ACE inhibitor; 3-amino-4-oxo-1,5-benzothiazepine-5-acetic acid derivative; 3-amino-4-oxo-1,5-benzoxazepine-5-acetic acid derivative; structure-activity relationship; conformationally restricted analog; 1,5-benzothiazepine; 1,5-benzoxazepine

Captopril (1)^{1a)} and enalapril (2a)^{1b)} are potent and orally active angiotensin converting enzyme (ACE) inhibitors. They have been confirmed to be effective for the treatment of hypertension in man.²⁾ Since the discovery of these drugs, many analogues have been prepared and the structure–activity relationships of these inhibitors have also been studied.^{2,3)} Ondetti and Petrillo proposed the active site model of ACE interaction with inhibitors.^{2b)} However, the spatial orientation of binding sites of the enzyme and the conformation of inhibitors required for the binding have not been clarified. One way to approach the problem is to restrict the possible conformations of ACE inhibitors by synthesizing rigid molecules. A few examples have been reported based on such an approach,⁴⁾ and the conformationally restricted lactams 3^{3a,5a,c)} and 4^{5b)} have been prepared.

Recently we reported two new ACE inhibitors, CV-3317 (5a)⁶⁾ and the dicarboxylic acid derivative CV-3317-COOH (5b), which is equipotent with MK-422 (2b) in vitro. We then tried to evaluate the preferred spatial arrangement⁴⁾ of the functional groups of 5 for interaction with the enzyme. We hoped that such an approach might allow us to develop more potent inhibitors.

On conformational examination by using a Dreiding model, $5c^{7}$ in Chart 2 seemed to be stable because of low levels of nonbonded interactions. Although the tricyclic fused compound (6) bridged with one atom X between the methyl group of the L-alanine moiety and the methylene group of the indan portion would clearly correspond to 5c, the structure was rather complicated to synthesize. Since the hydrophobic character of indan was shown to be important⁶ for potent *in vivo* inhibitory activity, we replaced this with benzene and designed the new benzofused 7-membered heterocyclic inhibitors,⁸ (R)-3-[(S)-1-carboxy-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic acid (7a) and (S)-3-[(S)-1-carboxy-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetic acid (8a). This paper describes the synthesis as well as ACE inhibitory activity of 7a, 8a and related compounds (26, 27, 33 and 37; Table I).

Chemistry

Our synthetic strategy involved the construction of optically active 3-amino derivative intermediates (15 and 22) starting from natural amino acids, followed by reductive alkylation with ethyl 2-oxo-4-phenylbutyrate (23a), separation of diastereomers, and removal of the protective groups on the carboxyl moieties at the final step. The key intermediates in the synthesis, the (R)-3-amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic acid (15) and (S)-3-amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetic acid (22) derivatives, were prepared as shown in Charts 3 and 4.

Chart 2

(R)-3-(2-Nitrophenyl)thio-2-aminopropionic acid (10a), prepared from 2-nitroaniline (9a) and L-cysteine, was led to the N-protected amino acid derivative 12a (R = H) by reaction with N-ethoxycarbonylphthalimide, followed by catalytic reduction over palladium carbon (Pd-C). Intramolecular condensation of 12a was achieved with the use of diethyl phosphorocyanidate (DEPC) to afford (R)-3-phthalimido-2,3,-dihydro-1,5(5H)-benzothiazepin-4-one (13a), which was allowed to react with tert-butyl chloroacetate followed by treatment with hydrazine hydrate to give the desired intermediate 15a (R = H).

We attempted to prepare the (S)-3-(2-nitrophenoxy)-2-aminopropionic acid derivative required for the synthesis of 22 from 2-halonitrobenzenes and L-serine derivatives under a variety of conditions. The preparation of the material was accomplished by the use of 2-fluoronitrobenzene (16) and N-tert-butoxycarbonyl (Boc)-L-serine in the presence of 2-fold molar excess of sodium hydride in N,N-dimethylformamide (DMF) to yield (S)-3-(2-nitrophenoxy)-2-tert-butoxycarbonylaminopropionic acid (17). Catalytic reduction of the nitro group of 17 and subsequent cyclization of the resulting 2-aminophenoxy compound (18) by treatment with DEPC gave (S)-3-tert-butoxycarbonylamino-2,3-dihydro-1,5(5H)-benzoxazepin-4-one (19). Then, (19) was allowed to react with benzyl chloroacetate followed by removal of the Boc group with hydrogen chloride (HCl) to give the benzyl ester (12)-(12)-(13)

TABLE I. 1,5-Benzothiazepine (7, 26, 33 and 37) and 1,5-Benzoxazepine (8, 27) Derivatives

• S	No. ×	~	R_2	R3	Configu- ration	Yield	Formula	An	Analysis (%) Calcd (Found)	(pun	[\alpha]D (deg.)	(c) Temp.	-	ACE inhibition in vitro (%)	E vitro (% %
					C*	(0/)		C	Н	Z		(°C)	<u> </u>	(M)	. 01) 01
26a	S	Н	$Ph(CH_2)_2$	C_2H_5	S	92	$C_{23}H_{26}N_2O_5S\cdot HCl$	57.68	5.68	5.85	-117	(0.7)	21	53	98	97
26b	S	Н	$Ph(CH_2)_2$ -	C ₂ H ₅	R	87	C ₂₃ H ₂₆ N ₂ O ₅ S·HCl	(57.48 57.68	5.86	5.76)	-173	(1.0)	(a)	1	40	84
26c		S 7-CH,		C,H,	S	93	C,H,s,N,O,S·HCl·	(57.53 57.42	5.76 6.02	5.70)	-107	(0.6)	1	46	82	95
)		n 1			$1/2 H_2 O$	(57.67	6.16	5.36)		29				
26d		7-CH ₃	S 7-CH ₃ Ph(CH ₂) ₂ -	C_2H_5	R	86	C24H28N2O5S·HCI	57.42	6.02	5.58	-161	(0.6)			13	99
		,					$1/2 H_2 O$	(57.57	6.14	5.36)		29				
7 06		7-0CH	S 7-OCH ₃ Ph(CH ₂) ₂ –	$\mathrm{C_2H_5}$	S	96	$C_{24}H_{28}N_2O_6S\cdot HCI$	56.63	5.74	5.50	- 94	(0.5)	1	18	61	87
								(56.02)	5.86	5.36)		53				
26f	S	7-0CH	S 7-OCH ₃ Ph(CH ₂) ₂ -	$\mathrm{C_2H_5}$	R	68	$C_{24}H_{28}N_2O_6S\cdot HCI$	56.63	5.74	5.50	-138	(0.0)	1		33	75
								(56.06)	5.89	5.38)		56				
26g	S	7-CI	$Ph(CH_2)_2$	$\mathrm{C_2H_5}$	S	79	$C_{23}H_{25}CIN_2O_5S$	53.80	5.10	5.46	66 –	(0.4)		41	81	96
							HCI	(53.65	5.23	5.71)		24				
26h	S	7-CI	$Ph(CH_2)_2$	$\mathrm{C_2H_5}$	R	73	$C_{23}H_{25}CIN_2O_5S$	53.80	5.10	5.46	-145	(0.4)		10	31	77
							HCI	(53.73	5.34	5.21)		24				
7 6i	S		$Ph(CH_2)_2$	$\mathrm{C_2H_5}$	$RS^{b)}$	84^{c}	$C_{26}H_{30}N_2O_5S\cdot HCI$	60.16	6.02	5.40				58	85	96
		$(CH_2)_3$	13					(59.97)	6.31	5.11)						
26j	S		$Ph(CH_2)_2$	$\mathrm{C_2H_5}$	RS	79	$C_{24}H_{25}F_3N_2O_5S$	51.02	5.00	4.96					24	<i>L</i> 9
							HCI	(51.24)	5.01	5.55)						

66 56	73 93	88 97	64 89	84 96	30 69	84 28		70 91	_ 25	83 95	. 37	66 06	— 66	50 86		
83	20	57	23	48	16	53		1				59	91	.	96	93
39	1		1								1	4	46		77	50
(0.7)	(0.9)			(0.5)	24.5 (0.5)	24.5 (1.0)	25	(0.6)	(0.6)	(0.6)	(0.6)	(0.5)	24 (0.4)	25	(1.0)	25 (0.6) 23
-125	-161	1	1	-150	-175	- 145		-119	-151	-108	-144	-82	-1119	ļ	-137	— — 140
5.78 5.50)	5.78	5.75	5.57) 5.05	4.55)	6.04)	5.99)	5.39)	5.65	5.75	7.57	7.51	5.18	4.96) 6.76	6.79)	5.91)	6.42) 6.49 6.43)
6.86 6.86	6.86	7.24	7.31 5.63	5.75	6.79	6.81	7.23	6.50	6.42	6.21	6.21	5.40	5.44	5.36	5.29 6.89	6.91 6.30 5.91
56.95 (56.52	56.95	56.72	(56.18 62.75	(62.28 53.99	(53.98	(53.86 57.76	(57.28	53.26 (53.50	54.26 (54.09	52.53 (52.20	52.53	62.16	(61.77 60.86	(60.53 50.10	57.52	55.67 (55.52
$C_{23}H_{32}N_2O_5S\cdot HCl$	$C_{23}H_{32}N_2O_5S\cdot HCI$	$C_{23}H_{34}N_2O_5S\cdot HCI$	$C_{29}H_{30}N_2O_5S\cdot HCI$	C,0H,8N,O,S·HCl	$C_{20}H_{28}N_2O_5S\cdot HC1$	C ₂₄ H ₃₄ N ₂ O ₅ S·HCl		$C_{22}H_{30}N_2O_6S \cdot HCI \cdot 1/2H_2O$	$C_{22}H_{30}N_2O_6S\cdot HCI$	$C_{22}H_{30}N_2O_5S_2\cdot HCI$	$C_{22}H_{30}N_2O_5S_2\cdot HCl$	$C_{28}H_{28}N_2O_5S\cdot HCI$	$C_{21}H_{22}N_2O_5S$	$C_{21}H_{20}N_2Na_2O_5S\cdot$	$5/2 H_2 O C_{21} H_{28} N_2 O_5 S \cdot H_2 O$	$C_{20}H_{26}N_2O_6S$. $1/2H_2O$
92	67	99	98	84	52	79		68	50	85	74	74	92	48	77	86
S	X	RS	RS	S	R	RS		S	R	S	R	S	S	R	S	S
C_2H_5	C_2H_5	C_2H_5	C_2H_5	СН,	c_2	C_2H_5	•	$\mathrm{C_2H_5}$	C_2H_5	$\mathrm{C_2H_5}$	$\mathrm{C_2H_5}$	PhCH ₂ -	Н	Н	н	Н
$\left\langle \bigcap_{j=1}^{n} -(\operatorname{CH}_2)_2 - \bigcap_{j=1}^{n} \right\rangle$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle$ $\left\langle \left(\operatorname{CH}_{2} \right)_{2} \right\rangle$	$\mathrm{CH_3}(\mathrm{CH_2})_{7^-}$	(Ph) ₂ CHCH ₂ -	$(CH_3)_2CH(CH_3)_2$	$(CH_3)_2CH(CH_2)_2$	$\left\langle \bigcap \left\langle ^{(\mathrm{CH}_{2})_{2}-}\right.$) (H)	$0 \longrightarrow (CH_2)_{2^-}$	$\bigvee_{\bigcap} (\operatorname{CH}_2)_{2^-}$	$S \longrightarrow (CH_2)_{2^-}$	$S \longrightarrow (CH_2)_2$	$Ph(CH_2)_2-$	$Ph(CH_2)_2$	$Ph(CH_2)_2-$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle$ $\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle$	$0 \longrightarrow -(CH_2)_2 -$
S H	S H	H S	H S	S H	S H	S H		S H	S H	S H	S H	S H	H S	H S	H S	H S
26k	261	26m	26n	260	26p	26q		26r	26s	26t	26u	26v	7a	7b	Jc	7d

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ACE inhibition in vitro (%)	(M)	47 90	į	— 12 5 4 90		47 88 98	53	. 57 93	25	12 34 80 —	_ 16	34 84	_ 36		— 09 — —
(c) Temp. $_1$		$(0.2)^{d}$	23	(0.6) 24	(0.6)	24 (0.6) 25	(0.5)	(0.6)	(0.6)	(0.5)	(0.5) 24	(0.5)	(0.6)	(0.5)	(0.5)
$[\alpha]_{D}$ (deg.)	ın MeOH	-106	í	- 70	66-	-166	-134	-101	-125	-104	-123	-134	-105	-121	- 59
(%) s (%)	z	8 6.14		8 5.94 8 5.71)		7 5.74) 6 6.48 4 6.45)	9 5.97 3 5.82)	8 5.69 1 5.58)		5 6.01 8 6.00				2 5.84 8 5.38)	50 5.65
Analysis (%) Calcd (Found)	С Н	52.61 6.18		58.54 5.98 (58.45 6.08		(57.39 5.97 63.87 7.46 (64.07 7.64	58.91 7.09 (58.89 7.23	58.59 7.38 (58.29 7.41		56.70 7.35				55.06 6.72 (55.11 6.78	53.27 6.50
Formula		C ₂₀ H ₂₆ N ₂ O ₅ S ₂ ·H ₂ O	_	$C_{23}H_{26}N_2O_6 \cdot HCI \cdot 1/2 H_2O$	O ₆ ·HCl·	H ₂ O C ₂₃ H ₃₂ N ₂ O ₆	$C_{23}H_{32}N_2O_6\cdot HCl$	C ₂₄ H ₃₄ N ₂ O ₆ ·HCl· 1/2 H ₂ O	C ₂₄ H ₃₄ N ₂ O ₆ ·HCl·	$1/2 H_2^{2} O$ $C_{22} H_{32} N_2 O_6 \cdot HCl \cdot$	1/2 H ₂ O C ₂₂ H ₃₂ N ₂ O ₆ · HCl · 1/2 H O	$C_{23} + C_{34} + C_{23} + C_{34} + C$	$C_{22}H_{30}N_2O_7 \cdot HCI \cdot 1/2H_2O$	$C_{22}H_{30}N_2O_7 \cdot HCI \cdot 1/2H_2O$	C ₂₂ H ₃₀ N ₂ O ₆ S·HCl·
Yield	\Im	73		82	87	66	46	95	91	87	82	91	06	72	72
Configu- ration	*	S		S	R	S	R	S	×	S	R	RS	S	A.	S
2	₹ .	Ш		C_2H_5	C_2H_5	C_2H_5	C_2H_5	$\mathrm{C_2H_5}$	C_2H_5	$_2$ – C_2H_5	$_2$ – C_2H_5	C_2H_5	C_2H_5	C_2H_5	C ₂ H ₅
ద	7.7	CH.),-		$Ph(CH_2)_2-$	$Ph(CH_2)_2-$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle \!$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle - (\mathrm{CH_2})_2 -$	$\bigcirc \bigcirc \bigcirc (CH_2)_2 -$	$\left(\begin{array}{c} \\ \\ \end{array} \right) - \left(\operatorname{CH}_2 \right)_2 -$	$(C_2H_5)_2CH(CH_2)_2 - C_2H_5$	$(C_2H_5)_2CH(CH_2)_2-$	$\mathrm{CH_3}(\mathrm{CH_2})_{7^-}$	$0 \longrightarrow (CH_2)_2 -$	$\bigcirc \bigcirc \bigcirc \bigcirc (\mathrm{CH_2})_{2^-}$	$S \longrightarrow (CH_2)_2 -$
×		H		н о	Н О	Н О	н о	Н О	Н О	Н 0	Н 0	H 0	Н О	Н О	Н 0
Ž	.02	4	2	27a	27b	27c	27d	27e	<i>27</i> f	27g	27h	27i	27j	27k	172

	1	1	l	1	I	ſ			1	64	1	1	ı
88	92	45	66	- 9/	100	i		92	38	21	· 	88	
51	51	1	16	1	95	93	93	62			26	29	87
I	1		31	١		42	52	1			62		53
(0.5)	(0.4)	(0.6)	(0.4)	(0.3)	(0.4)	(0.4)	(0.5)						
-115	-106	-114	-87	-112	-131	-128	89 –	-	1				
5.28 - 5.03)	5.64 - 5.49)	·	6.73 6.68)	,	6.63 - 6.68)	6.74 6.36)	6.36	5.14 5.61)	5.51	5.46			
6.64	7.50				7.16	6.55	6.41 6.32	7.30		5.70			
63.33 (63.02	60.41 (60.33	56.02 (56.02	60.57 (60.44	59.29	59.70 (59.80	57.82 (57.41	54.53 (54.12	57.81 (57.76	56.74	53.85	,		
$C_{28}H_{34}N_2O_6\cdot HC1$	C ₂₅ H ₃₆ N ₂ O ₆ ·HCl	$C_{25}H_{34}N_2O_8 \cdot HCI \cdot 1/2 H_2O$	0 ₆ H ₂ 0	$C_{21}H_{22}N_2O_6 \cdot 3/2H_2O$	$C_{21}H_{28}N_2O_6\cdot H_2O$	$C_{20}H_{26}N_2O_7 \cdot 1/2 H_2O$	$C_{20}H_{26}N_{2}O_{6}S\cdot H_{2}O$	$C_{24}H_{34}N_2O_5S\cdot HCI$	$C_{24}H_{34}N_2O_5S \cdot HCI \cdot 1/2 H,O$				
71	98	94	72	23	76	93	85	29	1	99			
S	S	S	S	R	S	S	N	S	×	S			
PhCH ₂ -	$CH_3(CH_2)_{3}$	H ₅ C ₂ OCOCH ₂ -	Н	Н	Н	H	2- H	C ₂ H ₅ :00H	C_2H_5 OOH	xide C ₂ H ₅			
$\left\langle \bigcap_{j \in \mathcal{C}} H_{2} \right\rangle_{2^{-}}$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle$	$\left\langle \right\rangle$ \rightarrow $(CH_2)_2$	$Ph(CH_2)_2$	$Ph(CH_2)_2$	$\left\langle \begin{array}{c} \\ \\ \end{array} \right\rangle \!$	$ \begin{pmatrix} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $	$\left\{ \begin{array}{c} \\ \\ \end{array} \right\} - \left(\operatorname{CH}_2 \right)_2 -$	$\left\langle \begin{array}{c} \\ \\ \\ \\ \end{array} \right\rangle = C(CH_2)_2 - \\ \\ \begin{array}{c} \\ \\ \end{array} \right\rangle = C(CH_3)COOH$	CH ₂) ₂ - S-CH(CH ₃)COOH	Ph(CH ₂) ₂ -1-oxide		() ()	СООН)
Н 0	О Н	0 Н	н о	Н О	О Н	О Н	Н О	S H	S	S	2b (MK-422)	5a (CV-3317)	5b (CV-3317-COOH)
27n	270	27p	8	%	ဆိ	P8	æ	37a	37b	33	2b (N	کا (ع ا	29 (C

a) Not determined. b) Mixture of diastereomers. c) Based on 15e. d) In MeOH-1 N HCl.

$$\begin{array}{c} S \\ NOH \\ ^t BuOOC \\ \hline 15 \\ \end{array} \qquad \begin{array}{c} O \\ -NPht = -N \\ O \\ \end{array} \qquad ^t Bu = tert - Bu \\ \end{array}$$

Chart 3

Chart 4

Furthermore the *tert*-butyl ester **22b** was prepared on account of the synthetic utility of the *tert*-butoxy moiety, which is stable under alkaline and catalytic hydrogenation conditions. Thus, the Boc group of **19** was removed with HCl and the benzyloxycarbonyl (**Z**) group was introduced with **Z-Cl**. The resulting (S)-3-benzyloxycarbonylamino derivative **21** was allowed to react with *tert*-butyl chloroacetate followed by catalytic hydrogenolysis to afford **22b**.

The synthetic route to 7a and 8a from 15a and 22a is illustrated in Chart 5. Reductive alkylation of the benzothiazepine derivative 15a with 23a in the presence of sodium cyanoborohydride (NaBH₃CN) afforded a mixture of two diastereomers 24a, b (R = H, $R_1 = tert$ -butyl, $R_2 =$ phenethyl), which were separated by silica gel column chromatography to give 24a with lower Rf and 24b with higher Rf. The tert-butyl ester of each isomer 24a and 24b was deprotected smoothly with HCl at room temperature to yield the monoacids 26a, b (R = H, $R_2 =$ phenethyl) respectively.

Chart 5

Fig. 1. Stereoscopic Drawing of 27a

In a manner similar to that used for the preparation of 24a, **b**, the oxazepine derivatives 25a (lower Rf) and 25b (higher Rf) were prepared by reductive alkylation of 22a with 23a followed by chromatographic separation of the resulting diastereomers. In this case, deprotection of the benzyl esters (25a, b) was carried out by catalytic hydrogenolysis using Pd-C to give 27a, **b** (R = H, $R_2 =$ phenethyl), respectively.

The monoacids (26a, b and 27a, b) obtained above were tested for *in vitro* ACE inhibitory activity. The results are shown in Table I. Compounds 26a and 27a proved to be about ten times more active than the corresponding isomers 26b and 27b. It was previously observed in studies of N-carboxymethyldipeptide inhibitors such as 2^{1b} and 5^{6} that the isomer with (S)-configuration at the chiral center in the 1-ethoxycarbonyl-3-phenylpropyl moiety is more active than the corresponding (R)-diastereomer. Therefore, the newly formed asymmetric center of 26a and 27a should have (S)-configuration. The stereochemical assignment was

confirmed by X-ray analysis of (RS)-3-[(RS)-1-ethoxycarbonyl-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic acid (racemic **27a**), ¹⁰⁾ shown in Fig. 1.

Our target compounds 7a and 8a, which were generated from the corresponding monoacids (26a and 27a) by saponification, exhibited very potent ACE inhibitory activity in vitro, as shown in Table I.

This result led us to investigate further modification of the substituents¹¹⁾ of the fused benzene and 3-amino moieties in order to find even more potent inhibitors.

First, benzothiazepine derivatives (26c—i, Table I) with a variety of substituents on the benzene ring were prepared starting from substituted 2-nitroanilines 9b (R = 4- CH_3), 9c (R = 4- CH_3), 9d (R = 4-CI) and 9e (R = 4,5-(CH_2)₃-) as illustrated in Charts 3 and 5.¹²⁾ The 7-trifluoromethyl derivative (26j) was prepared *via* another route (shown in Chart 6) starting from 9f (R = 4- CF_3).

$$F_{3}C$$

$$NO_{2}$$

$$10f$$

Thus, the amino acid 10f was protected with the Z group followed by reduction of the nitro group with zinc to give 28, which was cyclized with DEPC to give the benzothiazepin-4-one derivative (29). After the introduction of the *tert*-butoxycarbonylmethyl group at the 5-position, deprotection¹³⁾ was carried out with hydrogen bromide (HBr) to yield 30, reductive alkylation of which in the presence of NaBH₃CN gave 26j.

Secondly, modification of the substituents of the 3-amino moiety was carried out. The required α-oxoesters 23b—j (Table VI) were prepared from the corresponding esters (38b—j) by condensation reaction with diethyl oxalate followed by decarboxylation. These oxoesters (23b—j) were allowed to react with 15a or 22a under reductive conditions, and the resulting diesters 24 and 25 were led to the monoacids (26k—u and 27c—m, Table I). Some of the monoacids (26k, r, t and 27c, j, l) were converted to diacids (7c—e and 8c—e, Table I) by hydrolysis. Next, modification of the ethyl ester moiety was carried out as shown in Chart 7 in order to examine the change of *in vivo* character after oral administration. Thus, the *tert*-butyl ester derivative of thiazepine (24a) was treated with alkali to give the monoacid derivative (31a), which was alkylated with benzyl bromide to give the diester (32a) treatment of which with HCl gave 26v.

In the case of oxazepine, the (S),(S)-isomer of the diester intermediate (25q with lower Rf) was prepared from 22b by reductive alkylation with 23b (R_2 =cyclohexylethyl) using NaBH₃CN, followed by separation by silica gel column chromatography. The monoacid 31b obtained from 25q by alkaline hydrolysis was alkylated with benzyl bromide, butyl iodide and ethyl bromoacetate to give 32b, c, d, deprotection of which gave 27n, o, p, respectively.

Finally, synthesis of the S-oxide and 5-propionic acid derivatives of thiazepine was undertaken (Chart 8). Oxidation of the sulfur atom of 26a proceeded smoothly with the use of m-chloroperbenzoic acid to give 33.¹⁴⁾

No. 3

Thorsett *et al.* reported^{5c)} that the introduction of a methyl group at the α -carbon of the acetic acid moiety in the series of monolactam derivatives (3) enhanced the *in vitro* ACE inhibitory activity. Therefore we prepared the α -methylacetic acid derivative (37a)¹⁴⁾ by a method similar to those illustrated in Charts 3 and 5 by way of the intermediates 13a, 34, 35 and 36a.

Chart 8

Biological Results and Discussion

The compounds (7, 8, 26, 27, 33 and 37) described above were evaluated for *in vitro* inhibition of rabbit lung ACE using the method reported by Cushman and Cheung¹⁵⁾ with a slight modification. The results are shown in Table I. The initially designed benzofused heterocyclic lactams 7a and 8a were highly active inhibitors and their *in vitro* potency was comparable to that of CV-3317-COOH (5b) or MK-422 (2b). In the case of substituted N-carboxymethyldipeptide ACE inhibitors such as 2 and 5, it is known^{1-3,5,6)} that the monoacid is an orally well absorbed pro-drug for the corresponding diacids, and therefore the former is less active *in vitro* than the latter. In this series of benzothiazepine and benzoxazepine derivatives, the same feature was observed in comparing the *in vitro* activities of monoacids (26a and 27a) and diacids (7a and 8a). The modification of substituents on the fused benzene

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No.	Inhibi	tion of angio	•	ssor respons	e in rats 10 m	ng/kg p.o.
	1/3	1	2	3	4	5 (h)
26a	85	68	51	54	33	30
26k	92	72	68	60	45	28
26r	88	78	41	30	a)	_
26v	89	79	76	57	36	38
27a	95	83	46	27	23	11
27c	92	87	81	72	65	55
27e	96	76	60	37	34	19
27g	93	73	63	61	38	35
27n	87	78	82	84	84	80
27o	98	96	86	88	80	71
37a	81	55	13	22	24	26
CV-3317 (5a)	93	87	77	64	60	53

TABLE II. ACE Inhibitory Activity in Vivo

of benzothiazepines proved to have little effect on the potency (26c—j). Oxidation of the sulfur atom led to reduced potency (compare 33 with 26a). As regards replacement of the phenethyl group in the side chain at the 3-position with other lipophilic substituents, slightly enhanced potency was observed in the cyclohexylethyl derivatives (26k and 7c).

Selected monoacid derivatives (26a, k, r, v, 27a, c, e, g, n, o and 37a) were tested for *in vivo* ACE inhibitory activity. The activity was assessed in terms of the inhibition (percentage) by the compounds, after oral administration, of the vasopressor response induced by intravenous administration of angiotensin I in conscious normotensive rats (Table II). Most of the compounds showed potent *in vivo* inhibitory activity. A duration of activity nearly equal to that of CV-3317 (5a) was found in the cyclohexylethyl derivative 27c. The modification of the ethyl ester moiety of 27c was proved to result in an increase of duration (27n and 27o). The effect of methylation of the α -carbon in the 5-acetic acid moiety was found to be insignificant *in vivo* (compare 37a with 26k).

For effective interaction with ACE, the inhibitor should have the functional groups at the optimum positions in space. The high ACE inhibitory activity of the series of benzothiazepines and benzoxazepines shown in Tables I and II suggests that these conformationally restricted drivatives exist in a conformation suitable for binding to ACE. The result of X-ray analysis of racemic 27a (Fig. 1) indicates that the arrangement of functional groups such as carboxyl, amino, amido and hydrophobic groups corresponds well to that of the initially assumed stable conformer 5c of CV-3317. Benzothiazepine derivatives may take a conformation⁷⁾ similar to that of benzoxazepines, since the two series show little difference in in vitro and in vivo activities.

This work has provided useful information with regard to the spatial requirements of ACE inhibitors. Further work is in progress.

Experimental

All melting points were determined on a Yanagimoto micro melting point apparatus (a hot stage type) and are uncorrected. The infrared (IR) spectra were recorded with a Hitachi 260-10 spectrophotometer. The proton nuclear magnetic resonance (1 H-NMR) spectra were recorded in the indicated solvents on Varian EM-360, EM-390 and XL-100A instruments. Chemical shifts are reported as δ -values relative to tetramethylsilane (TMS) as an internal standard. Mass spectra (MS) were obtained on a JEOL JMS-01SC mass spectrometer. [α]_D values were determined in

a) Not determined.

TABLE III. (R)-2-Amino-3-(2-nitrophenyl)thiopropionic Acid Derivatives (10 and 11)

No.	R	R_1	mp (dec.) °C	Yield (%)	Formula		nalysis (cd (Fou	., .,	$[\alpha]_{\mathrm{D}}$ $(\mathrm{deg})^{a)}$	(c) Temp.
				(/0)		С	Н	N	(ucg)	(°C)
10a	Н	NH_2	167 169	60	$C_9H_{10}N_2O_4S$	44.63	4.16	11.57	+68	(0.6)
						(44.51	4.43	11.43)		24
10b	$4-CH_3$	NH_2	156—158	27	$C_{10}H_{12}N_2O_4S \cdot 1/2H_2O$	45.29	4.92	10.56	+44	(0.2)
						(44.90	4.60	10.86)		26.5
10c	4 -OCH $_3$	NH_2	166168	14	$C_{10}H_{12}N_2O_5S$	44.11	4.44	10.29	+24	(0.2)
						(43.67	4.52	9.94)		26
10d	4-Cl	NH_2	169—171	47	$C_9H_9ClN_2O_4S \cdot 1/2H_2O$	37.84	3.53	9.81	+46	(0.4)
						(37.58	3.26	9.65)		29
10e	$4,5-(CH_2)_3$	NH_2	157—158	6.1	$C_{12}H_{14}N_2O_4S \cdot 1/2H_2O$	49.47	5.19	9.62	+33	(0.2)
						(49.13	5.00	9.58)		27
10f	4-CF ₃	NH_2	181—183	41	$C_{10}H_9F_3N_2O_4S$	38.71	2.92	9.03	+53	(0.1)
						(38.43	2.86	9.02)		24
11a	Н	NPht ^{c)}	220—222	81	$C_{17}H_{12}N_2O_6S$	54.84	3.25	7.53	-79	(0.9)
						(54.46	3.26	7.46)		24
11b	$4-CH_3$	NPht	b)	81	$C_{18}H_{14}N_2O_6S$				_	
11c	4 -OCH $_3$	NPht	157—159	89	$C_{18}H_{14}N_2O_7S$	53.73	3.51	6.96	-120	(0.7)
						(53.82	3.50	6.65)		26
11d	4-C1	NPht	183—185	37	$C_{17}H_{11}CIN_2O_6S$	50.19	2.73	6.89	-116	(0.6)
						(50.18	2.74	6.80)		28
11e	$4,5-(CH_2)_3$	NPht	219—222	58	$C_{20}H_{16}N_2O_6S$	58.25	3.91	6.79	-149	(0.1)
						(58.25	3.96	6.73)		26.5

a) In 1 N HCl. b) Used for the next step without purification. c) Phthalimido group.

the indicated solvents on a JASCO DIP-181 4-4822.

Reactions were run at room temperature unless otherwise noted, and followed by thin-layer chromatography (TLC) on Merck F-254 silica gel plates. Standard work-up procedures were as follows. The reaction mixture was partitioned between the indicated solvent and water. The organic extract was washed successively with the following aqueous solutions: water, NaOH solution (NaOH) and hydrochloric acid (aq. HCl). The extract was dried over MgSO₄, filtered and evaporated *in vacuo*. Chromatographic separation of the residue was done on Merck Silica gel 60 with the indicated eluents.

(R)-2-Amino-3-(2-nitrophenyl)thiopropionic Acids (10, Table III)—(R)-2-Amino-3-(2-nitrophenyl)thiopropionic acid (10a) was prepared according to the reported procedure. Substituted derivatives (10b—f) were obtained from substituted 2-nitroaniline (9b—f) and cysteine by a procedure similar to that used for 10a.

(R)-3-(2-Nitrophenyl)thio-2-phthalimidopropionic Acids (11, Table III)—N-Ethoxycarbonylphthalimide (3.5 g) and 10a (2.9 g) were added to an aqueous solution (200 ml) of Na₂CO₃ (1.4 g). After being stirred for 5 h, the mixture was filtered and filtrate was made acidic with conc. HCl. The deposited crystals were collected by filtration and recrystallized from EtOH to give 11a (3.6 g) as pale yellow needles. Compounds 10b—e were allowed to react with N-ethoxycarbonylphthalimide similarly to yield 11b—e.

(R)-3-Phthalimido-2,3-dihydro-1,5-(5H)-benzothiazepin-4-ones (13, Table IV)—A mixture of 11a (10 g) and MeOH (300 ml) was hydrogenated over 5% Pd-C (3.5 g) at atmospheric pressure. After absorption of the calculated amount of hydrogen, the catalyst was removed by filtration and the filtrate was concentrated *in vacuo*. The residue was crystallized from Et₂O-petroleum ether to give 12a (8.4 g, 91%) as pale yellow crystals. DEPC (5.5 g) was added dropwise to a stirred solution of 12a (8.4 g) in DMF (50 ml) at ice bath temperature. The mixture was stirred for 5 min, then Et₃N (2.28 g) was added dropwise at ice bath temperature. The stirring was continued for 30 min at ice bath temperature and for another 1 h at room temperature, then the mixture was diluted with water (200 ml) and allowed to stand overnight. The deposited solid was collected by filtration and purified by silica gel column chromatography (CH₂Cl₂: AcOEt = 2:1) to give 13a (5.4 g) as colorless prisms. MS m/z: 324 (M⁺). IR v_{max}^{Nujol} cm⁻¹:

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TABLE IV. (R)-3-Phthalimido-2,3-dihydro-1,5(5H)-benzothiazepin-4-one Derivatives (13)

13	R	mp (°C)	Yield (%)	Formula		alysis (, 0,	$[\alpha]_{D}$ (deg)	(c) Temp.
		(C)	(/₀)		C	Н	N	(deg)	(°C)
a	Н	202—205	68	$C_{17}H_{12}N_2O_3S$	62.95	3.73	8.64	$-164^{b)}$	(0.9)
					(63.15	4.02	8.49)		21
b	7-CH ₃	222—225	$39^{a)}$	$C_{18}H_{14}N_2O_3S$	63.89	4.17	8.28	-180^{b}	(0.5)
					(63.96	4.27	8.20)		27
c	7-OCH ₃	255-258	43	$C_{18}H_{14}N_2O_4S$	61.01	3.98	7.90	$-34^{c)}$	(0.5)
	Ü			10 11 2 1	(60.96	3.92	7.64)		26
d	7-Cl	256—258	22	$C_{17}H_{11}CIN_2O_3S$	56.91	3.09	7.81	$-169^{b)}$	(0.1)
				2 3	(57.15	3.22	7.80)		25
e	$7.8-(CH_2)_3$	240-243	39	$C_{20}H_{16}N_2O_3S$	65.92	4.43	7.69	$-136^{b)}$	(0.1)
	2.3			20 10 M	(65.97	4.53	7.56)		27

a) Based on 10b. b) In MeOH. c) In CHCl₃.

Table V. tert-Butyl (R)-3-Amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetate Derivatives (14 and 15)

No.	R	R ₁	mp	Yield	Formula		alysis (cd (Fou		$[\alpha]_{D}^{c)}$	(c) Temp.
		1	(°C)	(%)		С	Н	N	(deg)	(°C)
14a	Н	NPht	181—184	74 ^{a)} 71 ^{b)}	$C_{23}H_{22}N_2O_5S$	63.01	5.06	6.39	-164	(0.4)
						(62.95	5.10	6.34)		24.5
14b	7-CH ₃	NPht	140—143	$75^{a)}$	$C_{24}H_{24}N_2O_5S$	63.70	5.35	6.19	-151	(0.6)
	J					(63.49	5.43	6.13)		27
14c	7-OCH ₃	NPht	155—157	$66^{a)}$	$C_{24}H_{24}N_2O_6S$	61.53	5.16	5.98	-139	(0.8)
	J					(61.57	5.20	6.00)		26
14d	7 - Cl	NPht	182—184	$85^{a)}$	$C_{23}H_{21}CIN_2O_5S$	58.41	4.48	5.92	-148	(0.4)
						(58.36	4.67	5.83)		24
14e	$7.8-(CH_2)_3$	NPht	195—198	$69^{a)}$	$C_{26}H_{26}N_2O_5S$	65.26	5.48	5.85	-114	(0.6)
						(65.48	5.61	5.82)		27
15a	Н	NH_2	8689	71	$C_{15}H_{20}N_2O_3S$	58.42	6.54	9.08	-238	(1.0)
						(58.73	6.48	9.13)		20
15b	7-CH ₃	NH_2	159—160 ^{d)}	96	$C_{16}H_{22}N_2O_3S$	50.22	6.09	6.51	-146	(0.5)
					$C_2H_2O_4 \cdot H_2O$	(49.84	5.66	6.14)		27
15c	7-OCH ₃	NH_2	175—178	92	$C_{16}H_{22}N_2O_4S\cdot HCl\cdot$	50.07	6.30	7.30	– 147	(0.5)
					$1/2\mathrm{H_2O}$	(49.88	6.21	7.26)		26.5
15d	7-C1	NH_2	$158 - 160^{d}$	77	$C_{15}H_{19}CIN_2O_3S$	45.29	5.14	6.21	-102	(0.4)
					$C_2H_2O_4 \cdot H_2O$	(45.15	4.75	6.33)		24
15e	$7.8-(CH_2)_3$	NH ₂	Oil	69 ^{e)}	$C_{18}H_{24}N_2O_3S$					

a) Method A. b) Method B. c) In MeOH. d) Oxalic acid salt. e) Used for the next step without purification.

1770, 1710, 1670 (C=O). 1 H-NMR (CDCl₃) δ : 3.55 (1H, dd J=6, 12 Hz, C₂-H), 4.55 (1H, t, J=12 Hz, C₃-H), 5.05 (1H, dd J=6, 12 Hz, C₂-H). Substituted benzothiazepin-4-one derivatives (13b—e) were prepared similarly from 11b—e.

tert-Butyl (R)-4-Oxo-3-phthalimido-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetate (14, Table V)——Method A) Compound 13a (4g) was added to a cooled mixture of NaH (60% in oil, 0.5g) and DMF (50 ml) in an ice bath. The mixture was stirred for 5 min, ClCH₂COO'Bu (2g) was added, and stirring was continued for 15 min. The mixture was diluted with ice-water (200 ml) and deposited crystals were collected by filtration, dried and purified by silica gel column chromatography (hexane: AcOEt = 3:1) to give 14a (4g) as colorless crystals. Recrystallization from ethyl ether gave colorless prisms.

Method B) *tert*-Butyl chloroacetate (39 g), K_2CO_3 (36 g) and KI (2 g) were added to a solution of **13a** (70 g) in DMF (300 ml). The mixture was stirred overnight and worked up (AcOEt; 0.1 N aq. HCl, water). The residue was crystallized from EtOH to give **14a** (67.4 g) as colorless crystals. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1780, 1740, 1730, 1690 (C=O). ¹H-NMR (CDCl₃) δ: 1.40 (9H, s, 'Bu), 3.50 (1H, dd, J=6, 12 Hz, C_2 -H), 4.10 (1H, d, J=17 Hz, N_5 -CH), 4.65 (1H, t, J=12 Hz), 4.60 (1H, d, J=17 Hz, N_5 -CH), 5.08 (1H, dd, J=6, 12 Hz, C_2 -H), 7.20—7.55, 7.65—7.95 (8H, m, phenyl protons).

Substituted benzothiazepine-5-acetate derivatives (14b—e) were prepared from 13b—e according to method A. *tert*-Butyl (*R*)-3-Amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetates (15, Table V)—A mixture of 14a (4g), $N_2H_4 \cdot H_2O$ (1.4g) and EtOH (100 ml) was heated under reflux for 1 h, concentrated *in vacuo* and worked up (AcOEt; NaOH, water). The oily residue was crystallized from Et₂O-petroleum ether to give 15a (2g) as colorless prisms. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3380, 3320 (NH), 1740, 1670 (C=O). ¹H-NMR (CDCl₃) δ : 1.50 (9H, s, ¹Bu), 1.75 (2H, s, NH₂), 2.80 (1H, t, J=12 Hz, C₃-H), 3.45—3.75 (2H, m, 2 × C₂-H), 3.95 (1H, d, J=17 Hz, N₅-CH), 4.85 (1H, d, J=17 Hz, N₅-CH), 6.1—7.75 (4H, m, phenyl protons). Compounds 14b—e were treated with N₂H₄ similarly to yield 15b—e.

(S)-2-tert-Butoxycarbonylamino-3-(2-nitrophenoxy)propionic Acid (17)—A solution of Boc-L-serine (25 g) in DMF (10 ml) was added dropwise to a stirred mixture of NaH (60% in oil, 10.1 g) and DMF (200 ml) in a stream of N₂ at 0 °C. Stirring was continued until the evolution of hydrogen stopped, then 16 (19 g) was added dropwise to the mixture. After being stirred for 4 h, the mixture was poured into ice-aq. HCl and worked up (AcOEt; water). The residue was purified by silica gel column chromatography (hexane: AcOEt = 1:1) to give 17 (30 g, 75%) as a colorless liquid.

(S)-3-(2-Aminophenoxy)-2-tert-butoxycarbonylaminopropionic Acid (18)—A mixture of 17 (30 g) and MeOH (500 ml) was hydrogenated over 10% Pd–C (50% wet, 1 g) under ordinary pressure. The catalyst was filtered off, and the filtrate was concentrated in vacuo. The residue was crystallized from AcOEt to give 18 (23 g, 84%) as colorless crystals, mp 90—91 °C. Anal. Calcd for $C_{14}H_{20}N_2O_5$: C, 56.75; H, 6.80; N, 9.45. Found: C, 56.48; H, 6.82; N, 9.43.

(S)-3-tert-Butoxycarbonylamino-2,3-dihydro-1,5(5H)-benzoxazepin-4-one (19)—A solution of 18 (21.4 g) in DMF (120 ml) was treated with DEPC (14 g) and Et₃N (7 g) as described for the preparation of 13 to give 19, which was recrystallized from AcOEt-hexane to yield colorless plates (12.3 g, 61%), mp 202—203 °C. [α]_D^{24.5} – 19.5 ° (c = 0.9, MeOH). Anal. Calcd for C₁₄H₁₈N₂O₄: C, 60.42; H, 6.52; N, 10.07. Found: C, 60.69; H, 6.71; N, 9.99. IR ν (CBr max cm⁻¹: 1720, 1680 (C=O). ¹H-NMR (CDCl₃) δ : 1.4 (9H, s, 'Bu), 4.2 (1H, t, J = 12 Hz, CH), 4.6 (2H, q, J = 6 Hz, OCH₂), 5.55 (1H, m, NHBoc), 6.9—7.3 (4H, m, phenyl protons), 8.4 (1H, br s, N₅-H).

Benzyl (S)-3-tert-Butoxycarbonylamino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetate (20) — The reaction of 19 (12.3 g) with ClCH₂COOCH₂Ph (8.7 g) in DMF (150 ml) in the presence of K₂CO₃ (8.7 g) and KI (1 g) was carried out according to method B. The product was recrystallized from AcOEt–hexane to give 20 (11.7 g, 62%) as colorless prisms, mp 122—124 °C. [α]_D²⁴ – 180 ° (c = 1, MeOH). Anal. Calcd for C₂₃H₂₆N₂O₆: C, 64.78; H, 6.14; N, 6.57. Found: C, 64.65; H, 6.21; N, 6.69. IR $v_{\rm max}^{\rm KBF}$ cm ⁻¹: 1730, 1710, 1690, 1680 (C = O). ¹H-NMR (CDCl₃) δ: 1.4 (9H, s, ¹Bu), 4.3 (1H, d, J = 17 Hz, N₅-CH), 4.75 (1H, d, J = 17 Hz, N₅-CH), 4.1—4.8 (3H, m), 5.2 (2H, s, CH₂Ph), 5.45 (1H, m, NHBoc), 7.15 (4H, m, phenyl protons).

Benzyl (S)-3-Amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetate (22a) — A mixture of 20 (7.6 g) in 5 N HCl–AcOEt (30 ml) was allowed to stand for 3 h and then concentrated *in vacuo*. The residue was crystallized from AcOEt–Et₂O to give 22a·HCl (6.2 g, 96%) as a colorless crystalline powder, mp 169—172 °C. *Anal.* Calcd for C₁₈H₁₈N₂O₄·HCl: C, 59.59; H, 5.28; N, 7.72. Found: C, 59.09; N, 5.12; N, 7.55. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1730, 1680 (C=O). MS m/z: 326 (M⁺). [α]_D²⁴ – 202 ° (c=0.6, MeOH). ¹H-NMR (DMSO- d_6 +D₂O) δ: 4.3—5.0 (5H, m), 5.33 (2H, s, CH₂Ph), 7.4—7.7 (9H, m, phenyl protons).

tert-Butyl (S)-3-Amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetate (22b) —A mixture of 19 (5 g) and 5 N HCl-AcOEt (30 ml) was allowed to stand for 3 h. The deposited crystals were collected by filtration to give (S)-3-amino-2,3-dihydro-1,5(5H)-benzoxazepin-4-one · HCl (3.8 g, 98%) as colorless needles, mp 230—240 °C (dec.). Anal. Calcd for $C_9H_{10}N_2O_2$ · HCl: C, 50.36; H, 5.17; N, 13.05. Found: C, 50.30; H, 5.18; N, 13.02. $[\alpha]_D^{24}$ —227 ° (c=0.4, MeOH). Z-Cl (1.5 ml) was added to a stirred mixture of the above 3-amino derivative (1.5 g), AcOEt (100 ml), K_2CO_3 (excess) and water (50 ml) at ice bath temperature. After being stirred for 1 h, the mixture was worked up (AcOEt; water). The reasidue was crystallized from Et₂O to give 21 (2 g, 92%), which was recrystallized from AcOEt-Et₂O to yield colorless needles, mp 157—159 °C. Anal. Calcd for $C_{17}H_{16}N_2O_4$: C, 65.38; H, 5.16; N, 8.97. Found: C, 65.54; H,

23	R_2	Method	Yield (%)	bp (°C) (mmHg)
a	Ph(CH ₂) ₂ -	С	76	123—141 (3)
b	(CH ₂) ₂ -	D	52	105110 (1.5)
c	$CH_3(CH_2)_7-$	D	63 ^{a)}	_
d	(Ph) ₂ CHCH ₂ -	D	20	165—175 (1)
e	$(CH_3)_2CH(CH_2)_2-$	D	52	100—110 (26)
f	CH_3	D	$40^{a)}$	-
g	(CH ₂) ₂ -	E	70	115—125 (2)
h	S—(CH ₂) ₂ -	E	72	148—151 (3)
i	(CH ₂) ₂ -	D	50	118—123 (2)
j	$(C_2H_5)_2CH(CH_2)_2-$	D	65	102 (27)

Table VI. α-Oxoesters (23a—j)
R₂COCOOCH₂CH₃

5.19; N, 8.95. $[\alpha]_D^{23} - 175^{\circ}$ (c = 0.7, MeOH). This compound **21** (1.9 g) was allowed to react with ClCH₂COO'Bu (1.1 g) as described for the preparation of **14** to give *tert*-butyl (S)-3-benzyloxycarbonylamino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetate (2.5 g, 96%) as a pale yellow liquid. IR v_{max}^{neat} cm⁻¹: 3350, (NH); 1730, 1680 (C=O). MS m/z: 426 (M⁺). Catalytic hydrogenolysis of the above 5-acetate (2.5 g) was carried out in amanner similar to that described in the preparation of **12** to give **22b** (1.2 g, 67%) as a colorless liquid. *Anal*. Calcd for $C_{15}H_{20}N_2O_4$: C, 61.63; H, 6.90; N, 9.58. Found: C, 61.75; H, 6.91; N, 9.37. MS m/z: 292 (M⁺). $[\alpha]_D^{22} - 253^{\circ}$ (c = 0.9, MeOH). IR v_{max}^{neat} cm⁻¹: 3380, 3310 (NH); 1730, 1670 (C=O). ¹H-NMR (CDCl₃) δ : 1.45 (9H, s, ¹Bu), 1.7 (2H, s, NH₂), 3.6—4.8 (5H, m), 7.1 (4H, s, phenyl protons).

α-Oxoesters (23, Table VI)—Preparation of Materials (38): Ethyl nonanate (38c) was purchased from Wako Pure Chemical Ind. Ethyl 3-phenylpropionate (38a, bp 145—148 °C, 25 mmHg), ethyl 3,3-diphenylpropionate (38d) and ethyl 4-methylpentanoate (38e) were prepared from the corresponding acids purchased from Wako or Aldrich Chemical Co. by usual esterification using EtOH and H₂SO₄. Ethyl 3-cyclohexylpropionate (38b) was prepared as follows: a mixture of cyclohexanecarbaldehyde (Aldrich, 35 g), Ph₃PCHCOOEt (119 g) and benzene (500 ml) was refluxed for 1 h and then concentrated *in vacuo*. The residue was triturated with petroleum ether (1 l) and the insoluble solid was removed by filtration. The filtrate was concentrated and distilled *in vacuo* to give ethyl 3-cyclohexylacrylate (51 g, 90%) as a colorless liquid, bp 86—91 °C (3 mmHg). A solution of this ester (51 g) in EtOH (300 ml) was hydrogenated under atmospheric pressure using 10% Pd–C (50% wet, 5 g) as a catalyst. After absorption of hydrogen had stopped the catalyst was removed by filtration and the filtrate was concentrated *in vacuo*. The residue was purified by vacuum distillation to yield 38b (27 g, 91%; bp 65—70 °C, 3 mmHg) as a colorless liquid.

Ethyl 3-(1-methylcyclohexyl)propionate (**38f**), ethyl 3-(3,4,5,6-tetrahydro-2*H*-pyran-4-yl)propionate (**38g**, bp 121—123 °C, 16 mmHg), ethyl 3-(4-thianyl)propionate (**38h**, bp 155—157 °C, 15 mmHg), ethyl 3-cycloheptyl-propionate (**38i**, bp 142—145 °C, 27 mmHg) and ethyl 4-ethylhexanoate (**38j**, bp 102 °C, 27 mmHg) were prepared from the corresponding aldehydes, 1-methylcyclohex-3-enecarbaldehyde, 3,4,5,6-tetrahydro-2*H*-pyran-4-carbaldehyde, 1-10 dehyde, 1-10 dehyde,

Preparation of 23: Method C) A mixture of 38a (14.3 g), (COOEt)₂ (23.4 g) and 28% NaOMe–MeOH (15.4 ml) was evaporated under reduced pressure at 60—70 °C for 1.5 h. After cooling, 15% H₂SO₄ (130 ml) was added. The mixture was stirred for 15 h under reflux and worked up (AcOEt; water) to give 2-oxo-4-phenylbutyric acid (14.4 g), which was dissolved in a mixture of H₂SO₄ (1.3 ml) and EtOH (65 ml). The resulting solution was heated under reflux for 5 h, concentrated *in vacuo* to a half the initial volume and worked up (AcOEt; water). The residue was purified by vacuum distillation to yield 23a (11.4 g) as a pale yellow liquid.

a) Used for the next step without purification.

TABLE VII. Diesters of Benzothiazepine and Benzoxazepine Derivatives (24 and 25)

No.	X	R	R_i	R_2	Configu- ration	Yield (%)	IR NH	$v_{\text{max}}^{\text{neat}} \text{ cm}^{-1}$ $C = O$	MS (<i>m</i> / <i>z</i>) M +
					C*				
24a	S	Н	¹Bu	$Ph(CH_2)_2-$	S	31	3320	1740, 1670	498
24b	S	Н	^t Bu	$Ph(CH_2)_2-$	R	23	3320	1730, 1670	498
24c	S	7-CH ₃	t Bu	$Ph(CH_2)_2$	S	43	3330	1740, 1675	512
24d	S	$7-CH_3$	^t Bu	$Ph(CH_2)_2$	R	22	3330	1740, 1675	512
24e	S	$7-OCH_3$	^t Bu	$Ph(CH_2)_2^2$	s	30	3320	1730, 1660	528
24f	S	7-OCH ₃	^t Bu	$Ph(CH_2)_2$	R	17	3320	1720, 1660	528
24g	Š	7-Cl	'Bu	$Ph(CH_2)_2$	S	36	3330	1740, 1680	532
24h	Š	7-Cl	^t Bu	$Ph(CH_2)_2$	R	26	3330	1740, 1680	532
24i	S	$7.8-(CH_2)_3$	'Bu	$Ph(CH_2)_2$ $Ph(CH_2)_2$	RS	a)	3330	1740, 1000	538
241	3	7,0-(C11 ₂) ₃	Du	$\frac{1}{1}$	ΛS				330
24k	S	Н	^t Bu	(CH ₂) ₂ -	S	18	3320	1730, 1670	504
241	S	Н	$^{t}\mathrm{Bu}$	(CH ₂) ₂ -	R	12	3320	1730, 1670	504
24m	S	Н	t D	CH (CH)	r.~				
24m 24n			^t Bu	$CH_3(CH_2)_7$	RS	7.7	3325	1730, 1690	506
	S	Н	'Bu	(Ph) ₂ CHCH ₂ -	RS	21	3325	1740, 1680	574
24o	S	H	^t Bu	$(CH_3)_2CH(CH_2)_2-$	\boldsymbol{S}	22	3330	1730, 1670	464
24p	S	Н	'Bu	$(CH_3)_2CH(CH_2)_2-$	R	13	3330	1740, 1680	464
24q	S	Н	^t Bu	$\left(\begin{array}{c} \left(\text{CH}_{2}\right)_{2}^{-} \\ \text{CH}_{3} \end{array}\right)$	RS	28	3335	1740, 1680	518
24r	S	Н	^t Bu	(CH ₂) ₂ -	S	35	3330	1745, 1680	506
24s	S	Н	^t Bu	O(CH ₂) ₂	R	35	3320	1740, 1680	506
24t	S	Н	'Bu	(CH ₂) ₂ -	S	36	3320	1740, 1675	522
24u	S	Н	^t Bu	$S \longrightarrow (CH_2)_2 -$	R	23	3320	1740, 1680	522
25a	О	Н	PhCH ₂ -	$Ph(CH_2)_2$	\boldsymbol{S}	40			516
25b	Ö	H	PhCH ₂ -	$Ph(CH_2)_2$ -	R	30			
230	O	11	1 110112	$1 \text{ II}(C11_2)_2^{-}$	Λ	30			516
25c	O	Н	PhCH ₂ -	$\left\langle \right\rangle$ $\left(\text{CH}_2 \right)_2$ $\left(\text{CH}_2 \right)_2$	S	19	3330	1740, 1680	522
25d	О	Н	PhCH ₂ -	(CH ₂) ₂ -	R	19	3330	1740, 1680	522
25e	О	Н	PhCH ₂ -	(CH ₂) ₂ -	S	14	3330	1740, 1670	536
25f	О	Н	PhCH ₂ -	(CH ₂) ₂ -	R	9	3330	1730, 1680	536
25g	O	Н	PhCH ₂ -	$(C_2H_5)_2CH(CH_2)_2-$	S	14	3330	1740, 1670	510
25h	Ö	H	PhCH ₂ -	$(C_2H_5)_2CH(CH_2)_2-$	R	9.5	3330	1730, 1680	510
25i	ŏ	Н	PhCH ₂ -	$CH_3(CH_2)_7$	RS	11	3330	1740, 1680	524
	~		2	-113(-112)7	110		2230	1770, 1000	524

					TABLE VII. (continu	ued)				
N	lo.	X	R	R_1	R_2	Configuration	Yield (%)	IR NH	$v_{\text{max}}^{\text{neat}} \text{ cm}^{-1}$ $C = O$	MS (<i>m</i> / <i>z</i>) M ⁺
25	5j	0	Н	PhCH ₂ -	O(CH ₂) ₂ -	S	34	3330	1740, 1680	524
25	5k	О	Н	PhCH ₂ -	O	R	26	3330	1740, 1680	524
25	51	O	Н	PhCH ₂ -	S	S	35	3330	1740, 1680	540
25	5m	O	Н	PhCH ₂ -	S—(CH ₂) ₂ -	R	19	3330	1740, 1680	540
25	5q	O	Н	^t Bu	(CH ₂) ₂ -	S	20	3330	1740, 1680	488
25	5r	O	Н	^t Bu	(CH ₂) ₂ -	R	15	3325	1740, 1680	488

a) Used for the next step without purification.

Method D) A mixture of 38b (30 g), NaOEt solution (prepared from 4.5 g of Na and 100 ml of EtOH) and (COOEt)₂ (29 g) was heated at 70 °C for 30 min, evaporated *in vacuo* at 70 °C for 30 min, and then allowed to cool. Water (500 ml), Et₂O (200 ml) and petroleum ether (100 ml) were added, and the mixture was thoroughly shaken. The aqueous layer was separated, acidified with H_2SO_4 and worked up (AcOEt; water). The residue was dissolved in a mixture of aqueous dimethyl sulfoxide (DMSO) (water: DMSO=1:9, 110 ml) and NaCl (10 g), then the whole was heated at 140 °C for 2.5 h, allowed to cool, and worked up (AcOEt; water) to yield 23b (18 g) as a pale yellow liquid.

Ethyl 2-oxooctanoate (23c), ethyl 2-oxo-4,4-diphenylbutyrate (23d), ethyl 2-oxo-5-methylhexanoate (23e), ethyl 4-(1-methylcyclohexyl)-2-oxobutyrate (23f), ethyl 4-cycloheptyl-2-oxobutyrate (23i) and ethyl 2-oxo-5-ethylheptanoate (23j) were prepared similarly from the corresponding esters, 38c—f, i and j.

Method E) Ethyl 2-oxo-4-(3,4,5,6-tetrahydro-2*H*-pyran-4-yl)butyrate (23g) and ethyl 2-oxo-4-(4-thianyl)butyrate (23h) were prepared in a manner similar to that described in method D using LiCl instead of NaCl.

Diesters of Benzothiazepines and Benzoxazepines (24 and 25, Table VII)——A mixture of 3-amino compound (15a, 1.5 g), AcOH (0.3 g), α-oxoester (23a, 4.2 g), EtOH (50 ml) and molecular sieves 4A (8 g) was stirred for 30 min. A solution of NaBH₃CN (0.6 g) in EtOH (40 ml) was added dropwise to the stirred mixture over a period of 2 h. After stirring of the mixture overnight, α-oxoester (2.1 g) was added, then a solution of NaBH₃CN (1.3 g) in EtOH (40 ml) was added dropwise over a period of 2 h. After removal of the insoluble material by filtration, the solution was concentrated *in vacuo* and worked up (AcOEt). After addition of Et₂O (50 ml) and (COOH)₂ (2 g) to the residue, the mixture was shaken thoroughly, diluted with petroleum ether (300 ml) and allowed to stand overnight. The supernatant layer was removed by decantation. Water (50 ml), AcOEt (300 ml) and NaHCO₃ (excess) were added to the precipitate. The AcOEt layer was separated, dried (MgSO₄) and concentrated *in vacuo* to give an oily residue, which was subjected to silica gel column chromatography (hexane: AcOEt = 5: 1—10: 3) to yield firstly 24b (0.55 g) as a colorless liquid. *Anal*. Calcd for C₂₇H₃₄N₂O₅S: C, 65.04; H, 6.87; N, 5.62. Found: C, 65.36; H, 6.91; N, 5.61. From the second fraction, 24a (0.75 g) was obtained as a colorless liquid. *Anal*. Calcd for C₂₇H₃₄N₂O₅S: C, 65.04; H, 6.87; N, 5.62. Found: C, 64.90; H, 6.63; N, 5.66. ¹H-NMR (CDCl₃) δ: 1.10 (3H, t, J = 7 Hz, CH₃), 1.50 (9H, s, ¹Bu), 1.7—4.2 (12H, m), 4.8 (1H, d, J = 16 Hz, N₅-CH), 6.9—7.7 (9H, m, phenyl protons).

Other diesters in Table VII were prepared similarly. In the cases of 24m, n, q and 25i, the diesters were isolated as mixtures of diastereomers.

(R)-3-(1-Ethoxycarbonyl-3-phenylpropyl)amino-4-oxo-7-trifluoromethyl-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic Acid (26j, Table I)—Z-Cl (2.7 ml) and 1 n NaOH (19 ml) were simultaneously added to a stirred mixture of 10f (5.3 g) and 2.5 n NaOH (67 ml) at ice bath temperature over a period of 30 min. The resulting mixture was stirred for a further 2.5 h and extracted with Et₂O (50 ml). The aqueous layer was acidified with 1 n aq. HCl and worked up (AcOEt) to give (R)-2-benzyloxycarbonylamino-3-(2-nitro-4-trifluoromethylphenyl)thiopropionic acid (5.5 g, 72%) as pale yellow crystals, mp 150—153 °C. Anal. Calcd for $C_{18}H_{15}F_3N_2O_6S$: C, 48.65; H, 3.40; N, 6.30. Found: C, 48.68; H, 3.41; N, 6.27. Powdered Zn (4 g) was added to a mixture of the above acid (4.3 g), AcOH (50 ml) and water (50 ml). The resulting mixture was stirred for 50 min, diluted with water (150 ml) and worked up (AcOEt; water). The residue was dissolved in Et₂O (50 ml) and treated with HCl to deposit 28 HCl (3.4 g) as a pale yellow powder. The

ring closure reaction of $28 \cdot \text{HCl}$ (3.4 g) was carried out using DEPC (1.83 g) in the presence of Et₃N (1.56 g) in a manner similar to that used in the preparation of 13a to give $29 \cdot (1.3 \text{ g}, 42\%)$ as colorless crystals, mp $120-123 \,^{\circ}\text{C}$. [α]²³ $-161 \,^{\circ}$ (c=0.4, MeOH). Anal. Calcd for $C_{18}H_{15}F_3N_2O_3S$: C, 54.54; H, 3.81; N, 7.07. Found: C, 54.79; H, 3.90; N, 7.09. Compound $29 \cdot (1.1 \text{ g})$ was allowed to react with $\text{ClCH}_2\text{COO'Bu}$ (0.46 g) according to method B for the preparation of 14 to yield tert-butyl (R)-3-benzyloxycarbonylamino-4-oxo-7-trifluoromethyl-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetate (1.4 g, 98%) as a colorless viscous liquid. A solution of HBr in AcOH (30%, 10 ml) was added to a solution of the above acetate (1.4 g) in AcOH (5 ml). The resulting solution was allowed to stand for 4h and diluted with petroleum ether (100 ml) to yield $30 \cdot \text{HBr}$ (0.75 g, 60%) as colorless crystals, mp 176—180 °C. Anal. Calcd for $C_{12}H_{11}F_3N_2O_3S \cdot \text{HBr} \cdot \text{H}_2O$: C, 34.38; H, 3.37; N, 6.68. Found: C, 34.40; H, 3.60; N, 6.66. A mixture of $30 \cdot \text{HBr}$ (0.65 g), EtOH (50 ml), NaOAc (0.2 g), AcOH (0.19 g), $23a \cdot (1.67 \text{ g})$ and molecular sieves $4A \cdot (5 \text{ g})$ was treated with a solution of NaBH₃CN (0.56 g) in EtOH (40 ml) as described for 24. After evaporation of the solvent, the residue was worked up (AcOEt) to yield $26j \cdot (0.7 \, \text{g})$, which was isolated as the hydrochloride; colorless powder. MS m/z: 510 (M⁺).

tert-Butyl (R)-3-[(S)-1-Carboxy-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetate (31a) and tert-Butyl (S)-3-[(S)-1-Carboxy-3-cyclohexylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzoxazepine-5-acetate (31b)——A mixture of 24a (0.8 g) and 1 N NaOH (3 ml) was stirred for 2 h, diluted with water (200 ml) and extracted with Et₂O (100 ml). The aqueous layer was acidified with aq. HCl to deposit 31a (0.5 g, 66%) as colorless crystals, mp 165—167 °C. [α]_D²⁴ -101 ° (c=0.5, MeOH). Anal. Calcd for C₂₅H₃₀N₂O₅S: C, 63.81; H, 6.43; N, 5.95. Found: C, 63.69; H, 6.38; N, 5.87. IR $v_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 1740, 1680 (C=O). ¹H-NMR (DMSO- d_6 + D₂O) δ: 1.6 (9H, s, ¹Bu), 1.8—2.15 (2H, m), 2.7—3.8 (6H, m), 4.35 (1H, d, J=17 Hz, N₅-CH), 4.8 (1H, d, J=17 Hz, N₅-CH), 7.2—7.9 (9H, m, phenyl protons).

Hydrolysis of **25q** (1.5 g) was carried out similarly to yield **31b** (1.2 g, 85%) as colorless needles, mp 180—183 °C. [α]_D²⁵ –122 ° (c=0.5, MeOH). MS m/z: 460 (M⁺). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1740, 1700, 1625 (C=O). Anal. Calcd for C₂₅H₃₆N₂O₆: C, 65.20; H, 7.88; N, 6.08. Found: C, 65.18; H, 7.83; N, 6.14. ¹H-NMR (CDCl₃) δ: 1.45 (9H, s, ¹Bu), 0.7—2.0 (15H, m), 3.1 (1H, t, J=5 Hz, C₃-N-CHCOO), 3.55 (1H, dd, J=7, 12 Hz, C₂-H), 4.15 (1H, d, J=17 Hz, N₅-CH), 4.1—4.3 (1H, m, C₃-H), 4.45 (1H, dd, J=7, 12 Hz, C₂-H), 4.6 (1H, d, J=17 Hz, N₅-CH), 7.0—7.3 (4H, m, phenyl protons).

tert-Butyl (R)-3-[(S)-1-Benzyloxycarbonyl-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetate (32a) and tert-Butyl (S)-3-[(S)-Alkoxycarbonyl-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzo-xazepine-5-acetate (32b—d)——A mixture of 31a (0.3 g), NaHCO₃ (0.5 g), PhCH₂Br (0.15 g) and DMF (10 ml) was stirred overnight, diluted with water (100 ml) and worked up (AcOEt; 0.1 N aq. HCl, water) to give 32a (0.35 g, 98%) as a colorless liquid. IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3330 (NH), 1740, 1680 (C=O). MS m/z: 560 (M⁺). ¹H-NMR (CDCl₃) δ : 1.45 (9H, s, 'Bu), 1.9—3.7 (10H, m), 3.95 (1H, d, J=17 Hz, N₅-CH), 4.75 (1H, d, J=17 Hz, N₅-CH), 5.20 (2H, s, CH₂-Ph), 6.9—7.6 (14H, m, phenyl protons).

Compound **31b** (0.25 g) was allowed to react with PhCH₂Br, CH₃(CH₂)₃I and BrCH₂COOEt as described in the case of **32a** to yield **32b** (0.25 g, 84%), **32c** (0.26 g, 91%) and **32d** (0.26 g, 88%) as colorless liquids, respectively. **32b**: 1 H-NMR (CDCl₃) δ : 1.4 (9H, s, 1 Bu), 0.5—2.7 (15H, m), 3.2 (1H, t, J=6Hz, C₃-N-CHCOO), 3.6 (1H, dd, J=9, 12 Hz, C₂-H), 4.2 (1H, d, J=18 Hz, N₅-CH), 3.9—4.2 (1H, m, C₃-H), 4.4 (1H, dd, J=9, 12 Hz, C₂-H), 4.6 (1H, d, J=18 Hz, N₅-CH), 5.65 (2H, s, CH₂Ph), 6.8—7.6 (9H, m, phenyl protons). [α]_D²⁵ - 155° (c=0.6, MeOH), MS m/z: 550 (M⁺). **32c**: 1 H-NMR (CDCl₃) δ : 0.85 (3H, t, CH₃), 1.4 (9H, s, 1 Bu), 0.7—2.3 (15H, m), 3.15 (1H, t, J=6 Hz, C₃-H), 4.6 (1H, dd, J=8, 11 Hz, C₂-H), 4.9 (1H, dd, J=8, 11 Hz, C₂-H), 7.0—7.4 (4H, m, phenyl protons). [α]_D²⁵ - 145° (c=0.7, MeOH). MS m/z: 516 (M⁺). **32d**: 1 H-NMR (CDCl₃) δ : 1.2 (3H, t, J=6 Hz, CH₃), 1.4 (9H, s, 1 Bu), 0.6—2.7 (15H, m), 3.3 (1H, t, C₃-N-CHCOO), 3.7 (1H, dd, J=7, 11 Hz, C₂-H), 4.05 (2H, q, COOCH₂), 4.2 (1H, d, J=18 Hz, N₅-CH), 3.9—4.2 (1H, m, C₃-H), 4.3 (1H, d, J=7, 11 Hz, C₂-H), 4.5 (2H, s, COOCH₂CO), 4.6 (1H, d, J=18 Hz, N₅-CH), 7.0—7.3 (4H, m, phenyl protons). [α]_D²³ - 200° (c=0.5, MeOH). MS m/z: 546 (M⁺).

General Procedures for Deprotection of Esters (7, 8, 26, 27 and 37; Table I)—Method F) A mixture of tert-butyl ester (24a—u, 32a—d and 36a, b; 0.5 g) and 5 N HCl-AcOEt (5 ml) was allowed to stand overnight. Et₂O (20 ml) and petroleum ether (100 ml) were added to the mixture to deposit a colorless powder, which was collected by filtration to give the corresponding 5-acetic acid derivative (26a—i, k—v, 27n—p and 37a, b).

Method G) A mixture of benzyl ester (25a—m, $0.5 \, \mathrm{g}$) and EtOH (50 ml) was hydrogenated over $10\% \, \mathrm{Pd-C}$ (50% wet, $0.5 \, \mathrm{g}$) under atmospheric pressure. After the absorption of hydrogen had stopped, the catalyst was removed by filtration and the filtrate was concentrated *in vacuo*. The residue was dissolved in $\mathrm{Et_2O}$ and treated with HCl to yield the hydrochloride of the corresponding 5-acetic acid derivative (27a, b, d—p) as a colorless powder. In the case of 27c, colorless crystals of free base were obtained from AcOEt–Et₂O, mp 146—148 °C.

Method H) A mixture of EtOH (1 ml), 1 N NaOH (3 ml) and a monoacid derivative (26a, b, k, r, u and 27a, b, c, j, m; 0.1 g) was allowed to stand for 30—120 min. In the case of 7b, the disodium salt crystallized from the reaction mixture. In other cases, free acids 7a, 7c and 8a—c were deposited as colorless crystals after concentration of the reaction mixture followed by neutralization with aq. HCl. In the cases of 7d, e and 8d, e, the mixture was neutralized and subjected to XAD-2 column chromatography (acetone: water = 1:1). The eluate was lyophilized to give the

corresponding diacid derivative as a colorless powder.

(R)-3-[(S)-1-Ethoxycarbonyl-3-phenylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-acetic Acid S-Oxide (33, Table I)—m-Chloroperbenzoic acid (0.5 g) was added to a stirred solution of 26a (0.5 g) in CH₂Cl₂ (50 ml) over a period of 2.5 h. The resulting mixture was stirred for an additional 1 h, diluted with water (200 ml) and worked up (CH₂Cl₂; water). The residue was dissolved in Et₂O and treated with HCl to yield 33 ·HCl (0.3 g) as a colorless powder. MS m/z: 458 (M⁺).

tert-Butyl (R)-3-[(S)-1-Ethoxycarbonyl-3-cyclohexylpropyl]amino-4-oxo-2,3,4,5-tetrahydro-1,5-benzothiazepine-5-α-methylacetate (36a, b)— The reaction of 13a (6.48 g) with tert-butyl 2-bromopropionate (6.27 g) was carried out according to method B. The product was purified by silica gel column chromatography (hexane: AcOEt = 3:1—2:1) to give 34 (7.8 g) as a colorless powder. IR $v_{\text{max}}^{\text{neat}}$ cm⁻¹: 1770, 1730, 1720, 1680 (C=O). Anal. Calcd for $C_{24}H_{24}N_2O_5S\cdot 1/2H_2O$: C, 62.46; H, 5.46; N, 6.07. Found: C, 62.62; H, 5.14; N, 6.13. H-NMR (CDCl₃) δ: 1.1—1.7 (12H, m, 'Bu and CH₃), 3.25—3.6 (1H, m), 4.0—5.6 (4H, m), 6.9—7.9 (8H, m, phenyl protons). A solution of 34 (7.6 g) in EtOH (250 ml) was treated with $N_2H_4\cdot H_2O$ (2.5 g) as described for the preparation of 15a to yield 35 (5.4 g, 99%) as a pale yellow liquid. IR $v_{\text{max}}^{\text{neat}}$ cm⁻¹: 1735, 1670 (C=O). MS m/e: 322 (M⁺). H-NMR (CDCl₃) δ: 1.15, 1.6 (3H, d, J=7.5 Hz, CH₃), 1.4—1.5 (9H, s, 'Bu), 1.8 (2H, br s, NH₂), 2.7 (1H, m), 3.2—3.7 (2H, m), 4.4—5.3 (1H, q, J=7.5 Hz), 7.0—7.7 (4H, m, phenyl protons). Reductive alkylation of 35 (2.5 g) with 23b (6.6 g) in the presence of NaBH₃CN (0.59 g) was carried out in a manner similar to that used for the preparation of 24. The product was purified by silica gel column chromatography (hexane: AcOEt=4:1) to give 36b (0.74 g) firstly as a colorless liquid. MS m/z: 518 (M⁺). Compound 36a (0.28 g, 7%) was obtained from the second fraction. IR $v_{\text{max}}^{\text{neat}}$ cm⁻¹: 1740, 1670 (C=O). [α]₂²³ -223° (c=0.4, MeOH). MS m/z: 518 (M⁺).

ACE Inhibitory Activity in Vitro—The supernatant (20000 g, 25 min) of albino rabbit lung homogenates in 4 volumes of 100 mm borate-HCl buffer containing 300 mm NaCl (pH 8.3) was prepared as a source of crude ACE according to a slight modification of the method of Wallace et al.²⁰⁾ The ACE inhibitory activity was determined in terms of percent inhibition based on the amount of hippuric acid produced from the synthetic substrate hippuryl-L-histidyl-L-leucine (HHL) by ACE. Each assay mixture contained the following components: borate-HCl buffer (100 mm), NaCl (300 mm), HHL (5 mm), test compound (0.01 to 10 μ m) and crude ACE (0.1 ml) in a volume of 0.25 ml. The reaction tubes were incubated at 37 °C for 1 h. After termination of the reaction by the addition of 1 N aq. HCl (0.15 ml), hippuric acid was extracted with AcOEt and the hippuric acid concentration was determined from the absorbance at 288 nm.

ACE Inhibitory Activity in Vivo—On the day before the experiments, 8- to 10-week-old male Sprague Dawley rats were anesthetized with sodium pentobarbital (50 mg/kg) by i.p. injection. The animals were surgically prepared with an aortic catheter inserted via the left femoral artery and a caval catheter inserted via the right femoral vein. Both catheters were passed subcutaneously to the neck and exposed there. The rats were placed in individual plastic cages after the surgery. At the first stage of the experiment, the aortic cannula was connected with a pressure transducer and the mean blood pressure was recorded on the polygraph. Next, angiotensin I (A-I, 300 ng) and angiotensin II (A-II, 100 ng) were injected into the femoral vein in order to measure their hypertensive activities. The A-I and A-II challenges were repeated 1/3, 1, 2, 3, 4 and 5 h after the oral administration of the test compound (10 mg/kg). The result of A-II challenge was used for correcting the percent inhibition value based on the change of vascular responsiveness during the course of the experiment.

Acknowledgment The authors are grateful to Dr. T. Fujino for his encouragement and support of this work. Thanks are also due to Dr. K. Kamiya and Mr. K. Wada for X-ray analysis, and Miss M. Ojima and Mr. H. Nakagawa for biological assays.

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