## Communications to the Editor

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ABSOLUTE STEREOSTRUCTURES OF REHMAGLUTIN C AND GLUTINOSIDE

A NEW IRIDOID LACTONE AND A NEW CHLORINATED IRIDOID GLUCOSIDE

FROM CHINESE REHMANNIAE RADIX

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A new iridoid lactone, rehmaglutin C (4), and a new chlorinated iridoid glucoside, glutinoside (5), were isolated from Chinese Rehmanniae Radix, the dried root of Rehmannia glutinosa Libosch. [Kan-jiō (in Japanese) from China]. Their absolute stereostructures were determined on the basis of chemical and physicochemical evidence and by use of the exciton chirality method for allylic benzoyl derivatives.

KEYWORDS — Rehmannia glutinosa; Scrophulariaceae; iridoid lactone; iridoid glucoside chlorinated; rehmaglutin C; glutinoside; exciton chirality method

In the preceding paper, 1) we reported the isolation of four iridoids named rehmaglutins A, B, C, and D from Chinese Rehmanniae Radix, the dried root of Rehmannia glutinosa Libosch. (Scrophulariaceae), which is now in common use in Chinese medicinal preparations in Japan, and described the absolute stereostructures of rehmaglutins A (1), B (2), and D (3). In a continuing study, we isolated a new chlorinated iridoid glucoside named glutinoside from the water-soluble portion of the same Rehmanniae Radix. This paper deals with the absolute stereostructure elucidation of rehmaglutin C (4) and glutinoside (5).

The 50% aq. acetone extract of the radix was partitioned with ethyl acetate to furnish the organic phase soluble portion and the aqueous phase soluble portion as described previously. Repeated chromatography of the organic phase afforded acteoside, cerebroside, rehmaglutins A (1), B (2), C (4), and D (3). The aqueous phase, after repeated chromatographic purification with active charcoal, reversed-phase silica gel and ordinary silica gel, furnished catalpol (6)  $^{3}$ ) (0.004% from the radix), leonuride  $^{4,5}$ ) (0.003%), monomelittoside  $^{6}$ ) (0.002%), melittoside  $^{4}$ ) (0.019%), rehmannioside D $^{4}$ ) (0.031%), and glutinoside (5, 0.05%) together with new glucosides named rehmaionosides A, B, and C and rehmapicroside.  $^{7}$ )

Rehmaglutin C (4), colorless oil,  $C_9H_{12}O_5$ ,  $^{8}$  [ $\alpha$ ]  $^{24}_D$  -51.4° (MeOH), CI-MS m/z (%): 201 [(M+H)<sup>+</sup>, 12], 183 [(M+H-H<sub>2</sub>O)<sup>+</sup>, 100], had one trisubstituted olefin moiety, one secondary hydroxyl group, two primary hydroxyl groups, and one  $\gamma$ -lactone moiety as shown by the IR  $\nu$  (film) cm<sup>-1</sup>: 3340, 1758,  $^1$ H NMR and  $^{13}$ C NMR spectra (Table I). Ordinary acetylation (Ac<sub>2</sub>O-pyridine, r.t., 12 h) of rehmaglutin C (4) afforded the triacetate (4a), colorless oil,  $C_{15}H_{18}O_8$ , [ $\alpha$ ]  $^{20}_D$  -42.6° (MeOH), CI-MS m/z (%): 327 [(M+H)<sup>+</sup>, 47], 267 [(M+H-AcOH)<sup>+</sup>, 100], IR  $\nu$  (CHCl<sub>3</sub>) cm<sup>-1</sup>:

1775, 1739, 1235.

The detailed  $^1$ H NMR decoupling experiments (500 MHz, CDCl $_3$ ) with  $^4$ a resulted in the following assignment (J in Hz):  $\delta$  4.25, 4.47 (both d, J=ll.9; 1-H $_2$ ), 2.65 (dd, J=5.2, 18.6; 4 $\alpha$ -H), 3.05 (dd, J=ll.3, 18.6; 4 $\beta$ -H), 2.86 (ddd, J=1.8, 5.2, 11.3; 5 $\beta$ -H), 5.35 (br s,  $^9$ ) 6 $\alpha$ -H), 6.07 (dd, J=1.5, 4.0; 7-H), 4.76, 4.80 (AB in ABX, J $_{AB}$ =13.4, J $_{AX}$ =J $_{BX}$ =1.5; 10-H $_2$ ). The NOE's were observed between the following pairs of protons<sup>10</sup>):  $\frac{4}{\alpha}$ -H & 4 $\beta$ -H (18%),  $\frac{4}{\alpha}$ -H & 6 $\alpha$ -H (11%),  $\frac{4}{\beta}$ -H & 5 $\beta$ -H (18%),  $\frac{5}{\beta}$ -H & 4 $\beta$ -H (11%),  $\frac{1}{\alpha}$ -H (at  $\delta$  4.25) & 5 $\beta$ -H (7%),  $\frac{1}{\alpha}$ -H (at  $\delta$  4.47) & 4 $\beta$ -H (9%), and  $\frac{1}{\alpha}$ -H (at  $\delta$  4.47) & 5 $\beta$ -H (8%). Based on these spectral data, the stereostructure  $\frac{4}{\alpha}$  was assigned to rehmaglutin C triacetate and subsequently the structure  $\frac{4}{\alpha}$  to rehmaglutin C.

The absolute configuration of rehmaglutin C (4) was determined by applying the exciton chirality method 11) to the allylic benzoyl derivative of 4. Thus, tritylation of 4 with p-anisylchlorodiphenylmethane (MMTrCl) in pyridine (r.t., 48 h) and subsequent benzoylation (benzoyl chloride-pyridine, r.t., 8 h) furnished 4b, which, by detritylating with p-toluenesulfonic acid in MeOH-tetrahydrofuran (r.t., 1 h), was converted to 6-0-benzoylrehmaglutin C (4c), colorless oil,  $C_{16}^{H}_{16}^{O}_{6}^{O}$ , [ $\alpha$ ]  $C_{16}^{H}_{16}^{O}_{16}^{O}$ , [ $\alpha$ ]  $C_{16}^{H}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{O}_{16}^{$ 

Glutinoside (5), a hygroscopic white powder,  $[\alpha]_D^{20}$  -79.2° (MeOH), IR  $\nu$  (KBr) cm<sup>-1</sup>: 3388, 2922, 1047, was positive in the Beilstein test. The secondary ion mass spectrum (SIMS Xe<sup>+</sup>, glycerol matrix, m/z) of 5 had isotope ion peaks due to a chlorine atom: (M+H)<sup>+</sup> at 399 and 401, (M+Na)<sup>+</sup> at 421 and 423, and (M+H+glycerol)<sup>+</sup> at 491 and 493. The <sup>1</sup>H NMR and <sup>13</sup>C NMR (Table I) data for 5 indicated the presence of two acetal moieties, one each of secondary and tertiary hydroxyl groups in the iridoid glucoside skeleton.

Ordinary acetylation (Ac<sub>2</sub>O-pyridine, r.t., 3 h) of glutinoside (5) provided the pentaacetate (5a), colorless prisms, mp 185-186°C,  $C_{25}^{H}_{33}^{O}_{15}^{C1}$ ,  $[\alpha]_{D}^{20}$  -18.5°  $(CHCl_3)$ , IR  $\vee$   $(CCl_4)$  cm<sup>-1</sup>: 3483, 2940, 1755, 1224, 1036,  $l_{H}$  NMR (500 MHz, CDCl<sub>3</sub>,  $\delta$ ) : 2.01 (3H), 2.03 (6H), 2.10, 2.11 (3H each) (all s, OCOCH<sub>3</sub> x 5), 5.55 (d, J=1.8 Hz;  $1\alpha-H$ ), 5.33 (d, J=3.4 Hz;  $3\beta-H$ ), 2.16 (dd, J=3.4, 14.7 Hz;  $4\alpha-H$ ), 12) 2.22 (ddd, J= 3.1, 9.8, 10.3 Hz;  $5\beta$ -H), 4.98 (dd, J=3.1, 7.9 Hz;  $6\alpha$ -H), 4.28 (d, J=7.9 Hz;  $7\beta$ -H), 2.66 (br d, J=ca. 9.8 Hz; 9 $\beta$ -H), 3.67 (d, J=12.2 Hz;  $10\beta$ -H), 4.07 (d, J=12.2 Hz,  $10\alpha-H$ ), and the hexaacetate (5b), colorless prisms, mp 150-153°C,  $C_{27}^{H}_{35}^{O}_{16}^{Cl}$ ,  $[\alpha]_{D}^{20}$  -34.4° (CHCl<sub>3</sub>), IR  $\nu$  (CCl<sub>4</sub>) cm<sup>-1</sup>: no OH, 2942, 1750, 1222, 1036,  $\widetilde{l}_{H}$  NMR (500 MHz, CDCl<sub>3</sub>,  $\delta$ ): 2.01 (3H), 2.03 (6H), 2.08 (3H), 2.11 (6H)(all s, OCOCH<sub>3</sub> x 6), 5.55 (d, J=2.2 Hz;  $1\alpha$ -H), 5.34 (d, J=3.1 Hz,  $3\beta$ -H), 2.17 (dd, J=3.1, 14.0 Hz;  $4\alpha$ -H), 12) 2.26 (ddd, J=2.7, 10.4, 10.4 Hz; 5 $\beta$ -H), 4.91 (dd, J=2.7, 7.9 Hz; 6 $\alpha$ -H), 5.31 (dd, J=1.5, 7.9 Hz; 7 $\beta$ -H), 3.51 (br d, J= $\alpha$ 2. 10.4 Hz; 9 $\beta$ -H), 3.87 (dd, J=1.5, 12.5 Hz; 108-H), 4.26 (d, J=12.5 Hz;  $10\alpha$ -H). Detailed comparison of the  $^{13}\text{C}$  NMR data (Table I) for 5a and 5b with those for rehmaglutins A (1), B (2), C (4), and D (3) 1) indicated the presence of 6,8-dihydroxyl groups and a 7-chloro residue in the iridoid skeleton.

Methanolysis of glutinoside (5) with 9% HCl-dry MeOH (r.t., 4 h) yielded

Chart 1

Table I.  $^{13}$ C NMR Data for Rehmaglutin C (4), the Triacetate (4a), Glutinoside (5), and the Derivatives (5a, 5b, 5c, 7, 7a, 7b)

	4 <sup>a)</sup> ∼	4a <sup>b)</sup>	7 <sup>a)</sup>	7a <sup>a)</sup>	7.b <sup>a)</sup>
C-1	59.0	59.3	104.9	104.6	104.7
3	177.0	174.3	107.8	107.5	107.5
4	35.4	34.0	32.8	32.4	32.5
5	50.5	46.6	40.1	38.6	38.4
6	81.0	82.0	75.3	79.3	80.0
7	134.7	131.8	79.9	71.6	67.0
8	147.4	143.6	86.1	85.9	91.0
9	99.6	94.9	57.8	57.7	55.8
10	65.8	65.5	73.9	73.5	71.9

a)~c) Measured at 22.5 MHz in a)  $d_6$ -acetone, b) CDCl $_3$ , or c)  $d_5$ -pyridine.

d) The assignment may be interchanged.

	5 <sup>℃</sup>	5a <sup>C)</sup>	5b <sup>c)</sup>	5c <sup>c)</sup>
C-1	94.6	94.8	94.8	94.3
3	92.6	93.5	93.1	93.1
4	33.9	33.5	33.8	29.4
5	35.5	33.5	33.8	29.8
6	84.3	86.4	86.2	61.8
7	75.6	70.3	64.5	58.6
8	79.3	79.1	86.0	79.3
9	47.5	47.4	43.1	47.5
10	61.9	62.1	59.9	64.4
C-1'	98.6	96.5	96.3	98.5
2 '	74.2	71.5	71.8	74.2
3'	77.9	73.2	73.3	77.8 <sup>d)</sup>
4 '	70.9	68.9	69.0	70.9
5'	77.9	72.3	72.5	77.9 <sup>d)</sup>
6'	61.9	62.1	62.0	61.5

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methyl glucoside and the acetal  $(\frac{7}{2})$ , colorless oil,  $C_{12}H_{21}O_6Cl$ ,  $[\alpha]_D^{20}$  +31.8° (MeOH), IR  $\nu$  (CHCl<sub>3</sub>) cm<sup>-1</sup>: 3411, 2929, 1100, CI-MS m/z (%): (M+H)<sup>+</sup> at 297 (3) and 299 (1);  $(M+H-\dot{O}CH_3)^{\frac{1}{4}}$  at 265 (100) and 267 (34);  $(M+H-\dot{O}CH_3-CH_3OH)^{\frac{1}{4}}$  at 233 (98) and 235 (34), <sup>1</sup>H NMR (90 MHz,  $d_6$ -acetone, δ): 1.62-2.09 (m, 4-H<sub>2</sub>), 2.38-2.55 (m, 5β-H & 9β-H), 3.29 (6H), 3.31 (3H) (both s, OCH<sub>3</sub> x 3), 3.72 (dd, J=1.5, 9.5 Hz,  $10\beta-H$ ), 4.25 (d, J=9.5 Hz,  $10\alpha-H$ ),  $4.08 (dd, J=1.5, 9.9 Hz; <math>7\beta-H$ ), 4.48 (t, J=5.9 Hz; 3-H), 4.93 (br)s, 1-H),  $^{13}$ C NMR (Table I). Acetylation of 7 (Ac20-pyridine, r.t., 5 h) provided the monoacetate (7a) and the diacetate (7b). The comparison of the spectral data for 7, 7a, and 7b with those for rehmaglutin D  $(3)^{1}$  indicated the presence of 6,8-dihydroxyl groups and a 7-chloro residue in the structure. Finally, methanolysis of catalpol (6) under the same reaction conditions described for the methanolysis of glutinoside (5) yielded 7 in 35% yield (major) together with the several minor products. Thus the structure of the methanolysis product of glutinoside (5) was proposed as 7.

Treatment of glutinoside (5) with 10% aq. KOH under reflux for 2.5 h afforded the 6 $\beta$ ,7 $\beta$ -epoxide (5c) (in 53% yield), <sup>13</sup>) colorless prisms, mp 139-140°C,  $C_{15}^{H}_{22}^{O}_{10}$ °C,  $C_{15}^{H}_{2$ (Table I) for 5c indicated the presence of the 6-hydroxyl group and the 7-chloro residue in the structure of glutinoside (5). On the other hand, treatment of catalpol (6) with 0.6% HCl-dry MeOH at room temperature for 24 h provided glutinoside (5) in 55% yield.

Based on the above evidence, the absolute stereostructure of glutinoside (5) was determined as shown.

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8) The molecular compositon of the compound given with the chemical formula was determined by elemental analysis or by high resolution mass spectrometry.

- termined by elemental analysis or by high resolution mass spectrometry.

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- 10) The magnitude of the NOE (%) shown in the parenthesis was obtained when the underlined proton was irradiated.

underlined proton was irradiated.
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12) The signal due to 4β-H overlapped the signal of the acetoxyl methyls.
13) a) As we reported before, <sup>13b</sup> the alkaline treatment (2% KOH/MeOH-H<sub>2</sub>O, 45-50°C, 20 min) of linarioside, which had the 6β,8β-dihydroxyl groups and the 7α-chloro residue in the iridoid glucoside structure, gave the 7β,8β-epoxide derivative. However, the present alkaline treatment of 5 yielded the 6β,7β-epoxide (5c), presumably due to stereochemical reason; b) I. Kitagawa, T. Tani, K. Akita, and I. Yosioka, Chem. Pharm. Bull., 21, 1978 (1973).

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