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Angiotensin Converting Enzyme-Inhibitory Triterpenes from *Ganoderma lucidum*

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The 70% MeOH extract of *Ganoderma lucidum* had an inhibitory effect on angiotensin converting enzyme activity, and from this extract, five new triterpenes, named ganoderol A, ganoderols A and B, and ganoderic acids K and S, were isolated. Their structures were determined on the basis of spectral evidence.

Keywords—*Ganoderma lucidum*; Polyporaceae; lanostane; triterpene; ganoderol A; ganoderol B; ganoderol C; ganoderic acid K; ganoderic acid S; angiotensin converting enzyme inhibitor

The fungus *Ganoderma lucidum* (Polyporaceae) has many pharmaceutical effects and has long been used as a home remedy. A number of triterpene constituents have been isolated and characterized from this fungus in relation to the biological activities.¹⁾ In the course of our research on anti-hypertensive substances we found that the 70% MeOH extract of *G. lucidum* exhibited an inhibitory activity on angiotensin converting enzyme (ACE). From this extract we have now isolated and characterized five new triterpenes,²⁾ named ganoderol A (1), ganoderols A (2) and B (3),³⁾ and ganoderic acids K (8) and S (4), along with five known triterpenes. In this paper we describe the structure determination of the new compounds and the ACE-inhibitory activity of those triterpenes.

The dried fruiting body of *G. lucidum* KARST. produced in Nagano was extracted with 70% MeOH. The extract was adjusted to pH 2–3 with 1 N HCl and partitioned with ethyl acetate. The residue obtained by evaporation of the organic layer was separated on a column of silica gel into 14 fractions (Fr). Bioassays showed that Fr I, II, IV, V, VII, and XIII were ACE-inhibitory. Each bioactive fraction was repeatedly purified by silica gel column and/or preparative layer chromatography. From Fr I and II, compounds 1, 2, 3, 4 and 5 were isolated. From Fr IV and V, compound 6, from Fr VII, compounds 7, 8 and 9, and from Fr XIII, compound 10 were obtained. Compounds 5, 6, 7, 9 and 10 were characterized as ganoderic acids Y,⁴⁾ F,⁵⁾ H,⁶⁾ B,⁷⁾ and D (C)^{5,8)} (in that order) based on the published physical and spectral data. The structures of the new triterpenes were determined as follows.

Compound 2, named ganoderol A, showed characteristic peaks (236, 243 and 252 nm) of a 7,9(11)-diene tetracyclic triterpene in the ultraviolet (UV) spectrum, and gave the molecular (M⁺) ion peak at *m/z* 438 (C₃₀H₄₆O₂) together with fragment peaks at 423 (M–CH₃), 337(c–2⁹), 309(b–2), 269(a–1). Further, the M⁺ ion peak at *m/z* 510 of the trimethylsilyl derivative indicated the presence of a hydroxyl group. The proton nuclear magnetic resonance (¹H-NMR) spectrum (Table I) showed signals attributable to one *sec*- and six *tert*-methyl groups, three olefinic protons and an allylic hydroxymethylene group. From the ¹H-NMR data the partial structural features of 3-oxo-7,9(11)-diene nucleus and 24-en-26-ol side chain

were easily assigned. The chemical shifts of 24-H (δ 5.40), 27-H₃ (δ 1.67) and 26-H₂ (δ 4.01) were in good agreement with the data published for a (24*E*)-isomer.⁴⁾ In view of these data, ganoderol A was determined to be (24*E*)-3-oxo-5 α -lanosta-7,9(11), 24-trien-26-ol.

Compound 3, named ganoderol B, showed the same UV spectrum as ganoderol A and gave the M⁺ ion peak at m/z 440 (C₃₀H₄₈O₂), along with a characteristic fragment peak at 311 ($b-2$) in the mass spectrum (MS). The ¹H-NMR spectrum was closely similar to that of ganoderol A except for the presence of a new signal (δ 3.24) assignable to the 3 α -proton of a 3 β -alcohol derivative and the slight upfield shift of the 30- and 31-methyl signals. In view of these data, ganoderol B was established to be (24*E*)-5 α -lanosta-7,9(11), 24-triene-3,26-diol.

Compound 4, named ganoderic acid S, showed a peak at 226 nm, indicating a conjugated carboxylic group, in addition to the peaks due to 7,9(11)-diene in the UV spectrum and the M⁺ ion peak at m/z 452 (C₃₀H₄₄O₃), together with fragment ion peaks at m/z 437 (M - CH₃), 419 (M - CH₃ - H₂O), 353 (d), 309 ($b-2$), 269 ($a-1$) in the MS. Comparison of the ¹H-NMR spectrum with that of ganoderol A indicated the presence of a 24-en-26-oic acid unit instead of the 24-en-26-ol of ganoderol A. The stereochemistry of the 24-ene was deduced to be *E*, since the chemical shifts of 24-H (δ 6.91) and 27-H₃ (δ 1.84) of 4 resembled those of a (24*E*)-isomer such as ganoderic acid Y (3-oxo derivative of compound 4) rather than a (24*Z*)-isomer such as tyromisic acid.^{4,10)} Therefore, the structure was determined to be (24*E*)-3-oxo-5 α -lanosta-7,9(11), 24-trien-26-oic acid.

Compound 1, named ganoderol A, showed the M⁺ ion peak at m/z 436 (C₃₀H₄₄O₂), together with fragment peaks at m/z 337 ($c-2$), 309 ($b-2$), 281 and 269 ($a-1$). The ¹H-NMR spectrum was similar to that of ganoderol A, except for the presence of the aldehyde proton (δ 9.40) and a downfield shift of the 24-proton (δ 6.50) and the 27-methyl (δ 1.76) signals, indicating the presence of an unsaturated aldehyde in the side chain. The (24*E*)-stereochemistry was deduced from the published ¹H-NMR data for a triterpene having a (24*E*)-26-al moiety.⁴⁾ In view of these data, ganoderol A was deduced to be (24*E*)-3-oxo-5 α -lanosta-7,9(11),24-trien-26-al.

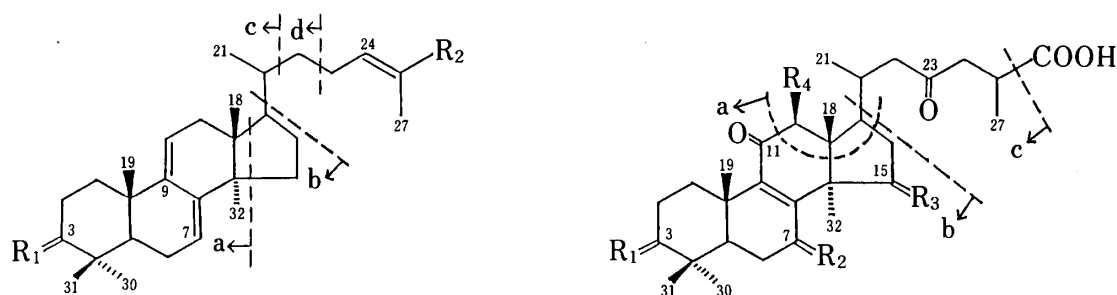
Compound 8, named ganoderic acid K, showed a peak (252 nm) indicative of a

TABLE I. ¹H-NMR (200 MHz) Spectral Data for Ganoderol A (1), Ganoderols A (2) and B (3), and Ganoderic Acids S (4) and K (8) (in CDCl₃)

Assignment/ multiplicity (<i>J</i> in Hz)	Compounds				
	1	2	3	4	8
18-H ₃ s	0.60	0.59	0.57	0.59	0.96
19-H ₃ s	1.13	1.13	0.88	1.13	1.26
21-H ₃ d (6)	0.96	0.93	0.92	0.94	0.96
27-H ₃ s	1.76	1.67	1.67	1.84	1.22 ^{a)}
30-H ₃ s	1.09	1.09	1.01	1.09	1.03
31-H ₃ s	1.21	1.20	0.98	1.20	0.85
32-H ₃ s	0.89	0.88	0.88	0.88	1.49
OAc s	—	—	—	—	2.26
2 β -H dt (14.5, 6)	2.79	2.78	—	2.78	—
3 α -H dd (10.5, 5)	—	—	3.24	—	3.20
7-H d (6)	5.52	5.51	5.47	5.51	4.80 ^{b)}
11-H d (6)	5.40	5.38	5.32	5.39	—
12 α -H s	—	—	—	—	5.62
24-H t (7)	6.50	5.40	5.40	6.91	—
CHO s	9.40	—	—	—	—
CH ₂ OH s	—	4.01	3.99	—	—

a) (d, $J=7$ Hz). b) (t, $J=9$ Hz).

conjugated carbonyl group, presumably 8-en-11-one, in the UV spectrum and fragment ion peaks at m/z 556 ($M - H_2O$, $C_{32}H_{46}O_9$ for the molecular formula), 538 ($M - 2H_2O$), 528 ($c - 1$), 514 ($M - AcOH$), 468 ($c - 1 - AcOH$), 417 (b), 345, 306 ($a + 1$). The 1H -NMR spectrum had the combined features of the spectra of ganoderic acids B^7) and H^6) showing characteristic signals assignable to two hydroxyl groups (δ 3.20, 3α -H; 4.80, 7β -H), an acetoxy methyl (δ 2.26) and a methine (δ 5.62, 12α -H). The chemical shifts of the *tert*-methyl signals were rationalized by assuming the nucleus to contain a 15-oxo group. The 27-methyl signal at δ 1.22 indicated the 23-oxo-26-acid side chain. On the basis of these data the structure of ganoderic acid K was assigned as $3\beta,7\alpha$ -dihydroxy- 12β -acetoxy-11,15,23-trioxo- 5α -lanost-8-en-26-oic acid.



	R ₁	R ₂	IC ₅₀ (M)		R ₁	R ₂	R ₃	R ₄	IC ₅₀ (M)
1	=O	-CHO	10 ⁻⁵ <	6	=O	=O	=O	-OAc	4.7 × 10 ⁻⁶
2	=O	-CH ₂ OH	10 ⁻⁴ <	7	α -H, β -OH	=O	=O	-OAc	2.0 × 10 ⁻⁵
3	α -H, β -OH	-CH ₂ OH	2.2 × 10 ⁻⁴	8	α -H, β -OH	α -H, β -OH	=O	-OAc	2.6 × 10 ⁻⁵
4	=O	-COOH	9.5 × 10 ⁻⁵	9	α -H, β -OH	α -H, β -OH	=O	-H	2.9 × 10 ⁻⁵
5	α -H, β -OH	-COOH	4.3 × 10 ⁻⁵	10	α -H, β -OH	α -H, β -OH	α -OH, β -H	-H	3.0 × 10 ⁻⁵

Chart 1

The inhibitory effects of compounds 1–10 on ACE activity were determined by a modification of the method described by Friedland and Silverstein,¹¹ and expressed in terms of IC₅₀ (the amount of samples needed to inhibit 50% of ACE activity). Eight compounds isolated in the present study were inhibitory (data included in Chart 1). Ganoderic acid F had the highest inhibitory effect, IC₅₀ = 4.7 × 10⁻⁶ M, whereas the IC₅₀ values of the other compounds were of the order of 10⁻⁵ M.

In summary, we have isolated and characterized from *G. lucidum* ten lanostane triterpenes which have ACE-inhibitory activity. Our results are in contrast to the conclusion by Arichi *et al.* that the compounds responsible for anti-hypertensive activity have molecular weights of more than 100000 daltons, based on their *in vivo* data in SHR rats.¹²

Experimental

Apparatus and Isolation—Melting points were measured on a Yazawa hot stage microscope and are uncorrected. UV spectra were recorded on a Shimadzu UV 200 spectrometer as a EtOH solution. The 1H -NMR spectra were obtained on a JEOL FX-200 spectrometer at 200 MHz, with a tetramethylsilane as an internal standard. Mass spectra (70 eV) were measured on a Shimadzu GC-MS 9020 DF spectrometer using a direct inlet system. The *Rf* values reported were obtained on E. Merck Kiesel gel 60 F₂₅₄ pre-coated plates (0.25 mm thickness). Preparative layer chromatography was done on E. Merck Kiesel gel 60 F₂₅₄ pre-coated plates (0.5 mm thickness). Solvent systems for the silica gel (E. Merck Silica gel 60, 70–230 mesh) column chromatography of the ethyl acetate extract were CHCl₃ (Fr I and II), CHCl₃–methanol, 50:1 (Fr IV, V, and VII) and CHCl₃–methanol, 9:1 (XIII). Approximate yields (starting from 10 g of 70% methanol extract) of the compounds were 1 (7 mg), 2 (19 mg), 3 (12 mg), 4 (7 mg), 5 (34 mg), 6 (24 mg), 7 (20 mg), 8 (43 mg), 9 (18 mg), 10 (17 mg).

Compound 1 (Ganoderic A)—Compound 1 was obtained as colorless leaflets, mp 127–128 °C (from MeOH). $[\alpha]_D^{25} + 27^\circ$ ($c = 0.21$, CHCl₃). UV λ_{max}^{EtOH} nm (log ϵ): 235 (4.17). *Rf* value in TLC: 0.50 (hexane–ethyl ace-

tate, 4:1)

Compound 2 (Ganoderol A)—Compound 2 was obtained as colorless leaflets, mp 99—101 °C (from MeOH). $[\alpha]_D^{25} + 33^\circ$ ($c=0.31$, CHCl_3). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 236 (4.13), 243 (4.18), 252 (4.00). *Rf* value in TLC: 0.30 (hexane-ethyl acetate, 4:1).

Compound 3 (Ganoderol B)—Compound 3 was obtained as colorless needles, mp 171—173 °C (from MeOH). $[\alpha]_D^{25} + 61^\circ$ ($c=0.18$, CHCl_3). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 237 (4.10), 244 (4.17), 252 (3.98). *Rf* value in TLC: 0.20 (hexane-ethyl acetate, 4:1).

Compound 4 (Ganoderic Acid S)—Compound 4 was obtained as colorless sticks, mp 168—169 °C (from MeOH). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 226 (4.14), 235 (4.19), 243 (4.15), 252 (3.98). *Rf* value in TLC: 0.25 (hexane-ethyl acetate, 4:1).

Compound 8 (Ganoderic Acid K)—Compound 8 was obtained as an amorphous powder. $[\alpha]_D^{25} + 48^\circ$ ($c=0.29$, CHCl_3). UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 251.5 (4.14). *Rf* value in TLC: 0.47 (CHCl_3 -MeOH, 9:1).

Bioassay—Purified ACE (9.2 units/mg protein) prepared from hog kidney was provided by Dr. T. Suzuki. Samples were dissolved either in water or in ethanol (ethanol concentration in the assay mixture was held constant at 10%). Inhibitory effects on ACE activity were determined by a modification of the published method, at least in duplicate.¹¹⁾

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References and Notes

- 1) For a recent review, S. Mizuno and S. Itamura, *Kagaku To Seibutsu*, **23**, 797 (1985).
- 2) This work was presented at the 3rd Annual Meeting of the Shoyaku Gakkai of Japan, Okayama, Oct. 1985, Abstract, p. 42.
- 3) M. Arisawa *et al.* have independently isolated ganoderols A and B from *G. lucidum*, and proposed the names ganodermenonol and ganodermediol, respectively; personal communication from Dr. M. Arisawa, Toyama Medical and Pharmaceutical University, and Abstracts of Papers, of the 65th Meeting of the Hokuriku Branch of the Pharmaceutical Society of Japan, Toyama, June 1985, p. 12.
- 4) J. O. Toth, B. Luu, J. Beck, and G. Ourisson, *J. Chem. Research (M)*, **1983**, 2722.
- 5) T. Kikuchi, S. Matsuda, S. Kadota, Y. Murai, and Z. Ogita, *Chem. Pharm. Bull.*, **33**, 2624 (1985).
- 6) T. Kikuchi, S. Matsuda, Y. Murai, and Z. Ogita, *Chem. Pharm. Bull.*, **33**, 2628 (1985).
- 7) T. Kubota, U. Asaka, I. Miura, and H. Mori, *Helv. Chim. Acta*, **65**, 611 (1982).
- 8) H. Kohda, W. Tokumoto, K. Sakamoto, M. Fujii, Y. Hirai, K. Yamasaki, Y. Komoda, H. Nakamura, S. Ishihara, and M. Uchida, *Chem. Pharm. Bull.*, **33**, 1367 (1985).
- 9) For abbreviations used in assignments of fragment ions, see the structures in Chart 1.
- 10) A. Gaudemer, J. Polonsky, and R. Gmelin, *Bull. Soc. Chim. Fr.*, **5**, 1844 (1967).
- 11) J. Friedland and E. Silverstein, *Am. J. Clin. Pathol.*, **66**, 416 (1976).
- 12) S. Arichi, T. Tani, M. Kubo, H. Matsuda, N. Yoshimura, and N. Kirigaya, *Kiso To Rinsho*, **13**, 4239 (1979).