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9-ANTHRYLDIAZOMETHANE-HPLC METHOD FOR DETECTION OF PROSTAGLANDINS
AND THROMBOXANE: AN APPLICATION TO THE MEASUREMENT OF
THE PRODUCTS OF STIMULATED RABBIT PLATELETS

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The cyclooxygenase products of rabbit platelets stimulated with arachidonic acid were measured by HPLC. Prostaglandins and thromboxane were extracted, derivatized with 9-anthryldiazomethane, and then loaded on an ODS-column. The fluorescence of the eluate was monitored. Thromboxane B₂ was a major product, which returned to the control level in the presence of indomethacin or the thromboxane synthetase inhibitor OKY-046. Pretreatment with OKY-046 shifted the cyclooxygenase products to increased levels of PGE₂, PGD₂ and PGF_{2α}.

KEYWORDS—TXB₂; PGE₂; PGD₂; 9-anthryldiazomethane; OKY-046; platelet aggregation; HPLC

Prostaglandins (PGs) and thromboxane (TX) in biological fluids have been assayed by radioimmunoassay with good sensitivity and selectivity. They also have been separated by high-performance liquid chromatography (HPLC), but the sensitivity of UV-monitoring is not sufficient to detect a trace amount of the metabolites in biological samples.¹⁾ The fluorescent labeling of prostaglandins with 9-anthryldiazomethane (ADAM) was reported by Hatsumi et al,²⁾ but frequent noise levels interfered with the procedure. We previously reported an improved ADAM-method with a clean-up procedure and applied it to the products of isolated rat pleural cells stimulated with phorbol myristate acetate.³⁾ With this clean-up procedure, better resolution of the chromatogram was obtained. This method can be applied to various other biological samples. Here we report the application of this method to the measurement of the products of rabbit platelet-rich plasma (PRP) stimulated with arachidonic acid (AA).

Arachidonic acid (>99%, Nu Chek Prep, Inc., Elysian, Minn.) and indomethacin (Sigma) were purchased. OKY-046 ((E)-3-[4-(1-imidazolylmethyl)phenyl]-2-propenoic acid hydrochloride) was a gift from Kissei Pharmaceutical Co. PGE₂, 6-keto-PGF_{1α}, PGD₂, PGF_{2α}, TXB₂ and anti-TXB₂ antiserum were gifts from Ono Pharm. Co. Osaka. ADAM was prepared by Funakoshi Pharm Co. Tokyo. [5,6,8,11,12,14,15(n)-³H]-PGE₂ (160 Ci/mmol, Amersham Int. Plc.) and [5,6,8,9,11,12,14,15-³H(N)]-TXB₂ (100 Ci/mmol, New England Nuc., Boston), and anti-PGE₂ antiserum (Institut Pasteur

Production, Paris) were purchased. Blood was collected from the carotid artery of male albino rabbits (Japanese white, 3-5 kg, Doken, Ibaragi), under light ether anesthesia, into a polypropylene tube containing 1/10 volume of 3.8% trisodium citrate solution. PRP was obtained by centrifugation at 280 x g for 20 min. The supernatant plasma was further centrifuged at 2,200 x g to yield platelet-poor plasma (PPP). The platelets were counted with a Sysmex platelet counter (PL-110, Toa Electric Co.). The number of PRP platelets was finally adjusted with PPP to 5×10^5 cells/ μ l.

Generation of cyclooxygenase metabolites: One hundred μ l of Hanks' balanced salt solution containing 0.1% bovine serum albumin and 800 μ l of PRP were taken into polypropylene tube (Falcon #2306), and preincubated at 37°C for 2 min with gentle shaking in the presence or absence of 1 mM OKY-046 or indomethacin. AA was then added to the platelet suspensions to a final concentration of 0.3 mM, and further incubated for 10 min. The reaction was terminated by the addition of a chilled solution of indomethacin (0.1 mM) and the sample was processed for extraction.

Procedure for ADAM-HPLC method: PGs and TX were extracted by the previously reported³⁾ modified method of Powell.⁴⁾ ADAM-derivatization and the clean-up method were also reported previously.³⁾ Briefly, samples were extracted through Sep-pak C₁₈ (Waters), evaporated, then dissolved in methanol. The solution was mixed with 0.2% ADAM solution in ethylacetate (1:1), and left overnight at room temperature. The reaction mixture was evaporated and cleaned up through Sep-pak silica cartridge (Waters). Finally ADAM-PGs were eluted with acetonitrile/methanol (4:1). The recoveries of TXB₂, PGF_{2 α} and PGE₂ were 84.7 \pm 2.0% (n=4), 89.5 \pm 3.8% (n=4) and 92.7 \pm 1.3% (n=4) respectively. The HPLC instruments used were a TriRoter V (Jasco, Tokyo); Uvidec-V (Jasco) and RF-530 (Shimadzu) detectors, Zorbax-ODS (250x4.6 mm, Dupont), ERC-ODS (30x6 mm, Erma) and Develosil-C₁₈ (Nomura Chem., Seto) columns with particle size of 5 μ m. The solvent, acetonitrile/water/phosphoric acid (60:40:0.1), flowed at 0.7 ml/min at 30°C. The fluorescence of the ADAM-derivatives were monitored at 412 nm with excitation at 365 nm. Radioimmunoassay (RIA) was performed as previously described.¹⁾

An HPLC chromatogram HPLC of ADAM derivatives of 5 ng of authentic 6-keto-PGF_{1 α} , PGF_{2 α} , PGE₂, PGD₂ and TXB₂ is shown in Fig. 1. TXB₂ and PGF_{2 α} migrated at a similar retention time when ODS (Zorbax) was eluted with acetonitrile/phosphoric acid system. TXB₂ formed a rather broad peak, but other PGs were clearly separated. Figure 3 shows typical chromatograms of ADAM derivatives of the AA metabolites of rabbit PRP stimulated with 0.3 mM AA for 10 min. The major peaks

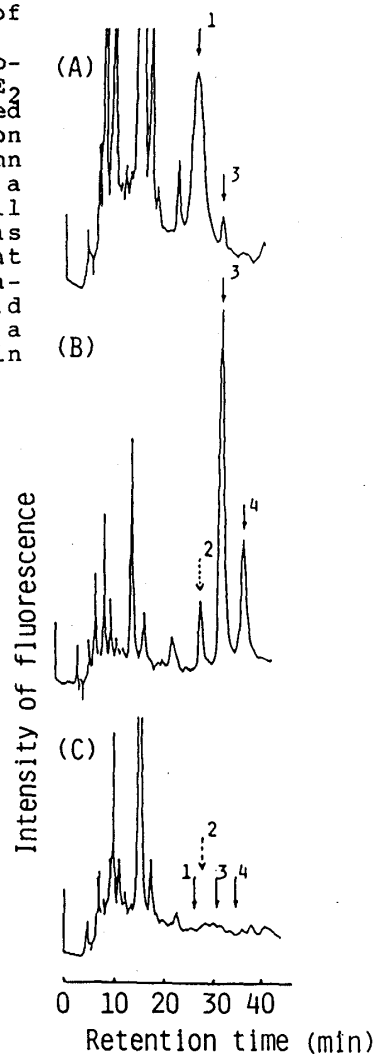
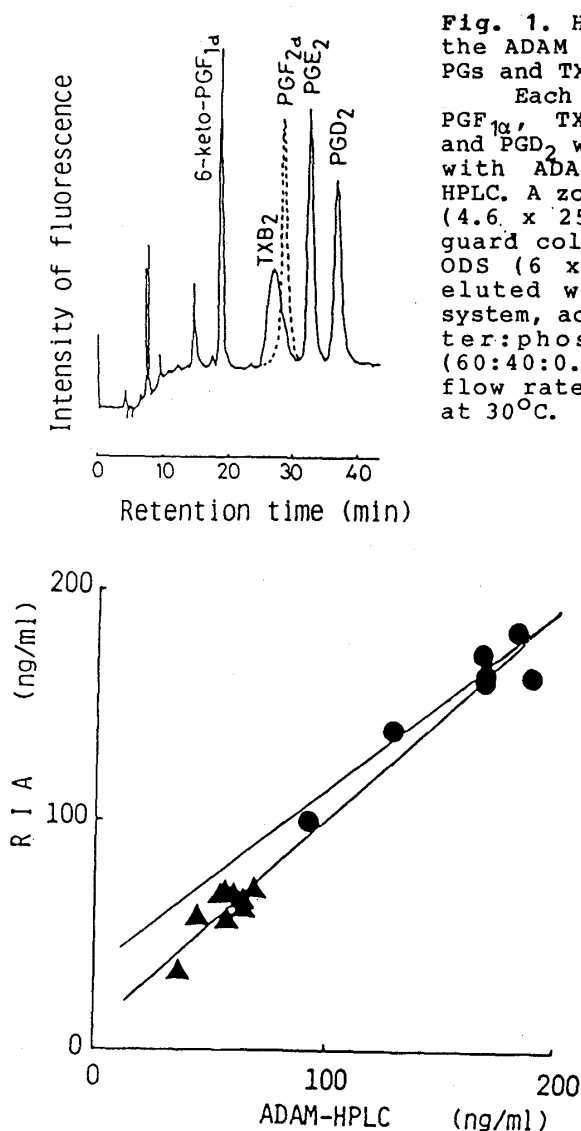


Fig. 2. Relationship between the Estimates Obtained by ADAM-HPLC Method and Those by RIA

Values are expressed as ng/ml incubation mixture of rabbit PRP and AA (0.3 mM) with or without OKY-046 (1 mM). (●) TXB₂ and (▲) PGE₂. Correlation coefficients are 0.94 ($p < 0.01$) and 0.80 ($p < 0.01$) for TXB₂ and PGE₂, respectively.

TABLE I. The Effect of Pretreatment with OKY-046 (1mM) on the Arachidonic Acid Metabolites of Rabbit Platelets (ng/ml)

| | TXB ₂ | PGE ₂ | PGD ₂ | PGF _{2α} |
|---------|------------------|------------------|------------------|-------------------|
| Control | 161.8 ± 9.4 | 12.3 ± 0.6 | <3 | <3 |
| OKY-046 | <5 | 192.5 ± 14.4 | 76.6 ± 5.3 | 35.8 ± 2.4 |

Values are means and standard errors of the determinations by the ADAM-HPLC method (n=5).

were TXB₂ and PGE₂. In the presence of 1 mM OKY-046, a TXA₂ synthetase inhibitor,⁵⁾ the TXB₂ peak disappeared (Fig. 3B). Instead the PGE₂ peak height increased to 192.5 ± 14.4 ng/ml (n=5) as shown in Table I. Simultaneously the PGD₂ peak (76.6 ± 5.3 ng/ml, n=5) appeared, but there was no detectable 6-keto-PGF_{1α} peak. Platelet aggregation was monitored with a light-transmission detector (Rikadenki, Tokyo) simultaneously with the experiment of Fig. 3. OKY-046 and indomethacin completely inhibited the AA-induced aggregation of rabbit PRP. The values of PGE₂ and TXB₂ in the incubation mixtures of rabbit PRP with AA (0.3 mM) were also measured by RIA and plotted against those obtained by ADAM-HPLC as shown in Fig. 2. The actual values of the samples used in Fig. 2 were: for TXB₂, 155.8±12.9 ng/ml (n=7, ADAM method) and 155.2±10.5 ng/ml (n=7, RIA), and for PGE₂, 56.2±3.0 ng/ml (n=10, ADAM method) and 61.1±3.4 ng/ml (n=10, RIA). For the PGE₂ assay the sample was incubated in the presence of OKY-046 (1 mM). By this HPLC the separation of the PGs and TX was demonstrated within 30 min. The detection limit is about 100 pg for standard PGs and TXB₂. Although the PGF_{2α} peak coeluted with the broad TXB₂ peak in this HPLC condition, fortunately significant amount of PGF_{2α} was not generated in the rabbit platelet aggregation, even when assayed with the solvent of the methanol/acetonitrile system, which was previously reported for the separation of the two metabolites.³⁾ The results coincided nicely with the reports of Smith et al.⁶⁾ and Rajtar et al.⁷⁾ The former reference describes how in rabbit platelets, OKY-1581, a selective inhibitor of TX synthetase, inhibits the generation of TX and increases PGE₂ and PGF_{2α}. The second paper reports that dazoxiben also inhibited TX synthesis and caused reorientation of cyclooxygenase products towards PGE₂, PGD₂ and PGF_{2α}. As shown in Fig. 3C, the sample incubated with indomethacin (1 mM) did not yield a detectable amount of PGs and TXB₂ even when stimulated with 0.3 mM AA, further proving the validity of ADAM-HPLC method.

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