

New Withanolides, Daturataturins A and B from *Datura tatura* L.¹⁾

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Two new C₂₈ steroidal glycosides called daturataturin A (**1a**) and daturataturin B (**2a**) were isolated from the methanolic extract of the fresh aerial parts of *Datura tatura* L. (Solanaceae), and their chemical structures were characterized as (22*R*)-7 α ,27-dihydroxy-1-oxowitha-2,5,24-trienolide 27-*O*- β -D-glucopyranoside and (22*R*)-1 α ,3 β ,7 α ,27-tetrahydroxywitha-5,24-dienolide 3-*O*- β -D-glucopyranoside by spectral analysis.

Keywords *Datura tatura*; Solanaceae; withanolide; daturataturin A; daturataturin B

As a part of our studies on the constituents of solanaceous plants, we previously reported the chemical structures of C₂₈ steroidal lactones called daturametelins A, B, C, D, E, F and G-Ac from the methanolic extract of the fresh aerial parts of *Datura metel* L.²⁾

In continuing studies on the constituents of the same genus, we report here the isolation and structure elucidation of two new withanolide glucosides, daturataturins A (**1a**) and B (**2a**), from the fresh aerial parts of *D. tatura* L.

Daturataturin A (**1a**) C₃₄H₄₈O₁₀, an amorphous powder, showed a [M+Na]⁺ peak at *m/z* 639, a [M-H₂O+H]⁺ peak at *m/z* 599 and a [M-H₂O-glc]⁺ peak at *m/z* 437 in the positive fast atom bombardment mass spectrum (FAB-MS). The infrared (IR) spectrum of **1a** indicated the presence of an α,β -unsaturated δ -lactone (1698 cm⁻¹) and an α,β -unsaturated ketone (1663 cm⁻¹). Acetylation of **1a** with acetic anhydride and pyridine gave a pentaacetate (**1b**), which showed a [M-AcOH+H]⁺ peak at *m/z* 767 in the positive FAB-MS. By comparison of the proton nuclear magnetic resonance (¹H-NMR) spectrum (Table I) of **1b** with that of daturametelin A tetraacetate (**3b**), the following signals in **1b** were assigned: four methyl groups [δ 0.73 (s, H₃-18), 1.04 (d, *J*=6.6 Hz, H₃-21), 1.25 (s, H₃-19) and 2.06 (s, H₃-28)], five acetyl groups [δ 2.00, 2.01, 2.02, 2.03 and 2.09 (all s)], two methylene groups [δ 2.90 (dd, *J*=20.9 and 4.8 Hz, H-4), 3.35 (d, *J*=20.9 Hz, H'-4), 4.47 (d, *J*=11.0 Hz, H-27) and 4.59 (d, *J*=11.0 Hz, H'-27)], three olefinic protons [δ 5.92 (dd, *J*=9.9 and 2.2 Hz, H-2), 6.81 (ddd, *J*=9.9, 5.0 and 2.6 Hz, H-3) and 5.83 (dd, *J*=5.9 and 1.8 Hz, H-6)], and a characteristic H-22 proton [δ 4.41 (dt, *J*=13.2 and 3.3 Hz)]. Furthermore, one methine proton signal (δ 4.90, m) was found to be attributable to H-7 by its correlation with the olefinic proton at H-6 in the ¹H-¹H correlated spectroscopy (COSY) spectrum. The chemical shift and the coupling pattern due to the H-22 signal indicated the presence of the typical C-17 side chain without a hydroxy group at C-20.³⁾ As regards the configuration at C-22, the circular dichroism (CD) spectrum of **1a** showed a positive Cotton effect at 255 nm, suggesting *R* configuration. Moreover, the ¹H-NMR spectrum of **1b** revealed the existence of a 2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl moiety [δ 4.65 (d, *J*=8.1 Hz, H-1'), 4.94 (dd, *J*=9.6, 8.1 Hz, H-2'), 5.21 (t, *J*=9.6 Hz, H-3'), 5.08 (t, *J*=9.6 Hz, H-4'), 3.69 (ddd, *J*=9.6, 4.4, 2.6 Hz, H-5'), 4.17 (dd, *J*=12.3, 2.6 Hz, H-6') and 4.23 (dd, *J*=12.3, 4.4 Hz, H'-6')]. From the evidence of the chemical shifts and coupling patterns of H₃-28 and H₂-27, the location of the glycosidic linkage was determined to be the 27-hydroxy group such as **3b**. The carbon-13 nuclear magnetic resonance

(¹³C-NMR) data for **1a** (Table II) were consistent with the assumed structure. Therefore, the structure of **1a** was concluded to be (22*R*)-7 α ,27-dihydroxy-1-oxowitha-2,5,24-trienolide 27-*O*- β -D-glucopyranoside. The aglycone moiety of **1a** corresponds to compound IX, which was isolated from *Withania somnifera* DUN. by Kirson *et al.*⁴⁾

Daturataturin B (**2a**) C₃₄H₅₂O₁₁, an amorphous powder, showed a strong absorption band at 3412 cm⁻¹ (hydroxyl) and a characteristic band at 1694 cm⁻¹ (α,β -unsaturated

TABLE I. ¹H-NMR Data (CDCl₃) for Daturataturin A Pentaacetate (**1b**) and Daturataturin B Heptaacetate (**2b**)

Proton	1b	2b
1	—	5.07 (brs)
2	5.92 (dd) <i>J</i> =9.9, 2.2 Hz	
3	6.81 (ddd) <i>J</i> =9.9, 5.0, 2.6 Hz	3.88 (m)
4	2.90 (dd) <i>J</i> =20.9, 4.8 Hz	
6	3.35 (d) <i>J</i> =20.9 Hz	
7	5.83 (dd) <i>J</i> =5.9, 1.8 Hz	5.77 (d)
18	4.90 (m) ^{a)}	<i>J</i> =4.0 Hz
19	0.73 (s)	4.92 (m) ^{b)}
21	1.25 (s)	0.70 (s)
27	1.04 (d) <i>J</i> =6.6 Hz	1.26 (s)
22	4.41 (dt) <i>J</i> =13.2, 3.3 Hz	1.03 (d)
27	4.47 (d) <i>J</i> =11.0 Hz	<i>J</i> =6.6 Hz
28	4.59 (d) <i>J</i> =11.0 Hz	4.41 (br d)
Glc-1'	<i>J</i> =11.0 Hz	<i>J</i> =9.9 Hz
Glc-2'	2.06 (s)	4.91 (d)
Glc-3'	4.65 (d) <i>J</i> =8.1 Hz	4.91 (d)
Glc-4'	4.94 (dd) ^{a)} <i>J</i> =9.6, 8.1 Hz	<i>J</i> =11.5 Hz
Glc-5'	5.21 (t) <i>J</i> =9.6 Hz	4.86 (d)
Glc-6'	5.08 (t) <i>J</i> =9.6 Hz	<i>J</i> =11.5 Hz
-OAc	3.69 (ddd) <i>J</i> =9.6, 4.4, 2.6 Hz	2.05 (s) ^{c)}
	4.17 (dd) <i>J</i> =12.3, 2.6 Hz	4.58 (d)
	4.24 (dd) <i>J</i> =12.3, 4.4 Hz	<i>J</i> =8.0 Hz
	2.00, 2.01, 2.02, 2.03, 2.09	4.94 (dd) ^{b)}
		<i>J</i> =9.6, 8.0 Hz
		5.18 (t)
		<i>J</i> =9.6 Hz
		5.05 (t) ^{b)}
		<i>J</i> =9.6 Hz
		3.68 (ddd)
		<i>J</i> =9.6, 5.0, 2.5 Hz
		4.10 (dd)
		<i>J</i> =12.3, 2.5 Hz
		4.23 (dd)
		<i>J</i> =12.3, 5.0 Hz
		2.00, ^{c)} 2.02, 2.04, 2.05, 2.08

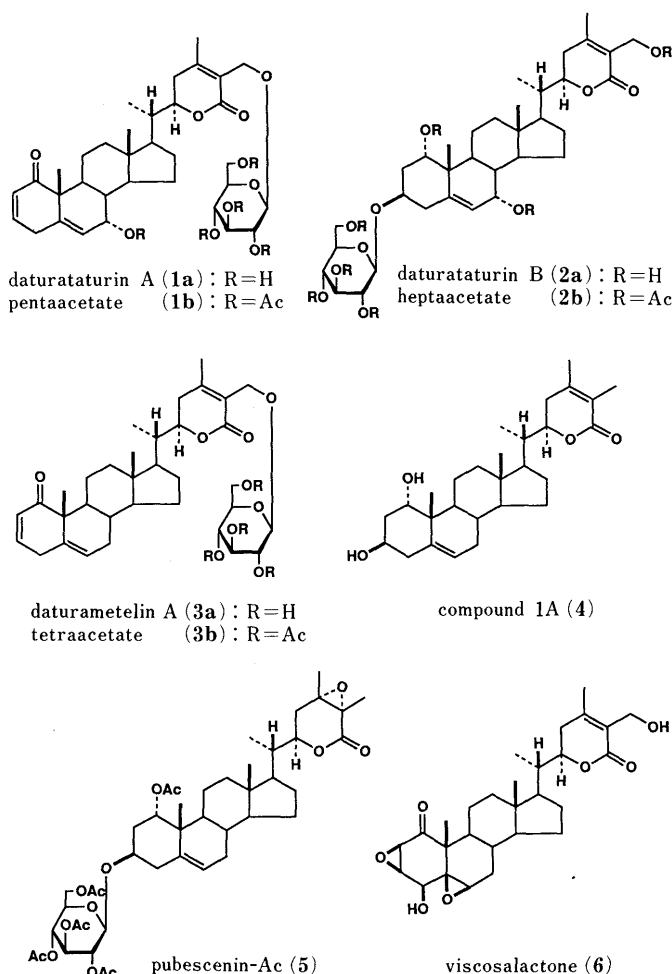
a—c) Signals overlap each other in each vertical column.

TABLE II. ^{13}C -NMR Data for Daturataturin A (**1a**),^{a)} Daturataturin B (**2a**),^{a)} Daturametelin A (**3a**),^{a)} Compound 1A (**4**),^{b)} Pubescenin Ac (**5**)^{b)} and Viscosalactone (**6**)^{b)}

	1a	2a	3a	4	5	6
C-1	203.3	72.3	203.9	72.9	74.9	206.9
C-2	127.9	37.6	127.7	38.3	33.7	54.9
C-3	145.6	73.8	145.8	66.3	74.9	55.8
C-4	33.6	38.2	33.4	41.4	38.1	74.6
C-5	139.4	142.7	136.2	137.6	136.7	64.0
C-6	128.1	127.8	124.6	125.2	124.9	59.6
C-7	63.7	64.7	30.8	31.7	28.8	29.6
C-8	35.8	34.0	33.1	31.3	31.7	31.0
C-9	39.1	39.2	43.3	41.5	42.3	42.7
C-10	50.2	42.9	50.6	41.7	40.5	48.6
C-11	23.9	20.4	23.8	20.2	20.7	20.0
C-12	39.9	39.4	39.8	39.8	39.6	27.3
C-13	42.5	42.7	42.5	43.0	42.8	42.7
C-14	52.2	52.2	56.2	56.8	56.5	56.1
C-15	24.3	24.6	24.2	23.9	24.3	24.3
C-16	27.2	27.4	26.9	22.0	27.3	40.7
C-17	51.3	49.7	51.9	54.7	52.1	51.9
C-18	11.9	11.7	11.7	13.6	11.7	11.5
C-19	18.2	18.3	18.8	19.4	19.4	14.5
C-20	38.7	38.9	38.9	75.2	38.6	38.9
C-21	13.4	13.5	13.3	20.8	13.7	13.3
C-22	78.3	78.2	78.2	81.0	76.3	78.7
C-23	29.8	29.8	29.9	31.5	31.7	29.8
C-24	157.1	153.9	157.0	149.1	59.3	153.8
C-25	In solvent	127.3	122.8	122.0	62.6	125.8
C-26	166.1	166.4	166.0	166.2	170.6	167.4
C-27	63.4	56.2	63.2	12.5	13.0	57.2
C-28	20.6	20.1	20.4	20.5	18.0	20.1
Glc-1'	104.8	102.9	104.6		99.6	
Glc-2'	75.2	75.2	75.0		71.4	
Glc-3'	78.5	78.5	78.3		72.9	
Glc-4'	71.6	71.5	71.5		68.5	
Glc-5'	78.4	78.4	78.3		71.8	
Glc-6'	62.8	62.7	62.6		62.1	
-OAc					170.3	
					170.0	
					169.4 (× 2)	
					20.7 (× 4)	
					21.1	

a) In pyridine- d_5 , b) in CDCl_3 .

δ -lactone) in its IR spectrum. The positive FAB-MS of **2a** showed a $[\text{M} + \text{Na}]^+$ peak at m/z 659. Next, the ^1H - and ^{13}C -NMR spectral data (Tables I and II, respectively) of **2a** and its heptaacetate (**2b**) were compared with those of compound 1A (**4**)⁵⁾ and pubescenin-Ac (**5**)⁶⁾ (for the A, B-ring moiety), and viscosalactone (**6**)⁷⁾ (for the side chain). The ^1H -NMR spectrum (Table I) of **2b** disclosed the occurrence of four methyl groups [δ 0.70 (H_3 -18), 1.03 (H_3 -21), 1.26 (H_3 -19) and 2.05 (H_3 -28)], one oxymethyl group [δ 4.91 and 4.86 (H_2 -27)] and two oxygenated methine protons [δ 3.88 (H -3) and 4.41 (H -22)] together with the presence of the β -D-glucopyranosyl moiety. The chemical shifts of this oxymethyl signal in **2b** indicated that the glucosyl moiety did not link to the C-27 hydroxymethyl group, in contrast to **1b**. Furthermore, in the ^1H -NMR spectrum of **2b**, the olefinic protons at C-2 and C-3 in ring A of **1b** disappeared, and instead, two new oxygenated methine proton signals occurred at δ 3.88 (1H, m) and 5.07 (1H, brs). Thus, the steroidal 2-ene-1-one system in **1a** no longer existed in **2a**. These new methine protons were assigned to the 5-ene-1 α ,3 β -diol system, which is known as



the precursor of the 2-ene-1-one system in the biogenesis of the withanolides.³⁾ The ^{13}C -NMR data of the side chain moiety (Table II) of the side chain part for **2a** were in good accordance with those of **6** except for C-25. Further, by comparison of the ^{13}C -NMR data of **2a** with those of **4** and **5**, the presence of a 5-ene-1 α ,3 β -diol structure (*vide supra*) and the β -D-glucopyranosyl group linked to the 3 β -hydroxy group in **2a** (and also **2b**) were reasonably clarified. Based on these spectral data and from the positive Cotton effect at 252 nm, the structure of **2a** having the (22*R*)-configuration was shown in the formula.

Experimental

Optical rotations were measured on a JASCO DIP-360 automatic digital polarimeter and CD spectra on a JASCO J-50A spectropolarimeter. The IR spectra were recorded with a Hitachi IR spectrometer, model 270-30. The ^1H - and ^{13}C -NMR spectra were measured with a JEOL JNM-GX 400 NMR spectrometer and chemical shifts are given on a δ (ppm) scale with tetramethylsilane as an internal standard. The FAB-MS were taken in a glycerol matrix containing NaI. Thin layer chromatography was performed on precoated Kieselgel 60 F₂₅₄ (Merck) and detection was achieved by spraying 10% H_2SO_4 followed by heating. Column chromatography was carried out on Kieselgel (70–230 mesh and 230–400 mesh, Merck) and Sephadex LH-20 (Pharmacia Co.).

Extraction and Separation Fresh aerial parts (1.78 kg) of *Datura tatur* L. (Solanaceae) harvested at the botanical garden in Kumamoto University in July 1988 were extracted with MeOH and the extract (40.3 g) was partitioned between *n*-BuOH and H_2O . The *n*-BuOH layer (16.3 g) was subjected to column chromatography repeatedly over silica gel using CHCl_3 -MeOH- H_2O = 1:0:1 → 8:2:0.1 → 0:1:0 and over Sephadex LH-20 using MeOH to give daturataturins A (**1a**, 1.70 g) and B (**2a**, 20.7 mg).

Daturataturin A (1a) An amorphous powder, $[\alpha]_D^{21} -38.1^\circ$ ($c=0.72$, pyridine). IR ν_{\max}^{KBr} cm^{-1} : 3442, 1698, 1663, 1464, 1404, 1344, 1323. Positive FAB-MS m/z : 639 $[\text{M}+\text{Na}]^+$, 599 $[\text{M}-\text{H}_2\text{O}+\text{H}]^+$, 437 $[\text{M}-\text{glc}-\text{H}_2\text{O}+\text{H}]^+$, 419 $[\text{M}-\text{glc}-2\text{H}_2\text{O}+\text{H}]^+$, 401 $[\text{M}-\text{glc}-3\text{H}_2\text{O}+\text{H}]^+$, 265, 171. CD ($c=0.94$, MeOH) $[\theta]$ (nm): -25000 (325) (negative max.), $+47000$ (255) (positive max.).

Acetylation of Daturataturin A (1a) A solution of **1a** (109 mg) in pyridine-acetic anhydride (1:1, v/v, 4 ml) was kept at room temperature overnight and the product was chromatographed on silica gel (*n*-hexane-EtOAc=1:1, v/v) to afford a pentaacetate (**1b**, 53.4 mg). An amorphous powder, $[\alpha]_D^{33} -42.0^\circ$ ($c=0.59$, CHCl_3). Positive FAB-MS m/z : 767 $[\text{M}-\text{AcOH}+\text{H}]^+$, 331.

Daturataturin B (2a) An amorphous powder, $[\alpha]_D^{15} -17.7^\circ$ ($c=0.52$, pyridine). IR ν_{\max}^{KBr} cm^{-1} : 3412, 1694, 1466, 1400, 1386, 1320, 1306. Positive FAB-MS m/z : 659 $[\text{M}+\text{Na}]^+$, 619 $[\text{M}-\text{H}_2\text{O}+\text{H}]^+$, 439 $[\text{M}-\text{glc}-2\text{H}_2\text{O}+\text{H}]^+$, 289, 232, 176. CD ($c=0.60$, MeOH) $[\theta]$ (nm): -4770 (315) (negative max.), $+48700$ (252) (positive max.).

Acetylation of Daturataturin B (2a) Acetylation was performed with pyridine and acetic anhydride (2:1, v/v, 3 ml) overnight at room temperature. The product was purified on silica gel column chromatography (benzene-acetone=20:1, v/v), and provided a heptaacetate (**2b**, 11.1 mg). An amorphous powder, $[\alpha]_D^{25} -31.9^\circ$ ($c=0.53$, CHCl_3). Positive FAB-MS m/z : 953 $[\text{M}+\text{Na}]^+$, 871 $[\text{M}-\text{AcOH}+\text{H}]^+$, 811 $[\text{M}-2\text{AcOH}+\text{H}]^+$, 601 $[\text{M}-\text{glc}(\text{Ac})+\text{H}]^+$, 550, 522, 331, 289, 246.

References and Notes

- 1) This work is part XX in the series of studies on the constituents of solanaceous plants. Part XIX: K. Shingu, Y. Furusawa, N. Marubayashi, I. Ueda, S. Yahara and T. Nohara, *Chem. Pharm. Bull.*, **38**, 2866 (1990).
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