## Synthesis and Antiulcer Activity of 4-Substituted 8-[(2-Benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetra-hydroquinolines and Related Compounds

Minoru Uchida,\* Masatoshi Chihiro, Seiji Morita, Hiroshi Yamashita, Katsuya Yamasaki, Toshimi Kanbe, Youichi Yabuuchi and Kazuyuki Nakagawa

Tokushima Research Institute, Otsuka Pharmaceutical Co., Ltd., Kagasuno 463-10, Kawauchi-cho, Tokushima 771-01, Japan. Received October 23, 1989

A series of 4-substituted 8-[(2-benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinolines was synthesized and examined for their  $(H^+ + K^+)$ adenosine triphosphatase (ATPase)-inhibitory and antisecretory activities against histamine-induced gastric acid secretion in rats. Many compounds tested were potent inhibitors of  $(H^+ + K^+)$ ATPase. Most compounds showed antisecretory activity. The antiulcer activity against water-immersion stress-induced gastric ulcer, aspirin-induced gastric ulcer and gastric necrosis induced by hydrochloric acid also were tested in the rat. Some of these compounds, in particular, 4-(N-allyl-N-methylamino)-1-ethyl-8-[(5-fluoro-6-methoxy-2-benzimidazolyl) sulfinylmethyl]-1-ethyl-1,2,3,4-tetrahydroquinoline (XVIIx) were found to have potent activity. The structure-activity relationships are discussed.

**Keywords** proton pump inhibitor;  $(H^+ + K^+)$  ATPase-inhibitory activity; 4-substituted 8-[(2-benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinoline; antisecretory activity; antiulcer activity; cytoprotective activity; structure-activity relationship

In two earlier papers, <sup>1,2)</sup> we reported that proton pump inhibitors, 8-[(5-fluoro-2-benzimidazolyl)sulfinyl]-3-methyl-5,6,7,8-tetrahydroquinoline (OPC-22321) and 8-[(2-benzimidazolyl)sulfinylmethyl]-1-ethyl-1,2,3,4-tetrahydroquinoline (OPC-22381) showed potent antisecretory and antiulcer activities (Chart 1). We also found that the methyl substitution at the 4-position of the 1,2,3,4-tetrahydroquinoline ring maintained potent antisecretory activity. As a continuation of our search for much more active compounds, we were interested in the synthesizing of 8-[(2-benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinoline derivatives having various substituents at the 4-po-

sition of 1,2,3,4-tetrahydroquinoline ring. We describe here the synthesis, biological activity and structure-activity relationships of 4-substituted 8-[(2-benzimidazolyl) sulfinylmethyl]-1,2,3,4-tetrahydroquinolines.

**Synthesis** Pyrido[3,2,1-*ij*]benzoxazine (V), key intermediate in the synthesis of 1,4-disubstituted 8-hydroxymethyl-1,2,3,4-tetrahydroquinolines (IXa—q), was synthesized by the pathway shown in Chart 2. Treatment of oaminobenzyl alcohol (I) with trichloromethyl chloroformate gave the benzoxazine (II), which was converted to the ester (III) by Michael addition with methyl acrylate in the presence of potassium carbonate, followed by hydrolysis with sodium carbonate to give the carboxylic acid (IV). Cyclization of IV was achieved using polyphosphoric acid (PPA) at 120 °C to give the key intermediate pyrido[3,2,1-*ij*]benzoxazine (V) in 60% yield (Chart 2).

The structure-activity relationships in the preceding paper<sup>2)</sup> indicated that ethyl and allyl groups at the 1-position of tetrahydroquinoline were most promising, and hence our synthetic work on various derivatives of tetrahydroquinoline was concentrated on 1-ethyl and 1-allyl compounds.

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TABLE I. 1H,3H,5H,6H,7H-3-Oxopyrido[3,2,1-i][3,1]bezoxazine Derivatives

Compd. No.	R¹	R <sup>2</sup>	Yield (%)	$^{1}\text{H-NMR} (CDCl_{3}, J = \text{Hz})$
VI	ОН	Н	79	2.00—2.20 (2H, m), 2.70—2.80 (1H, br s), 3.70—3.90 (1H, m), 3.90—4.10 (1H, m), 4.70—4.90 (1H, m), 5.17 (2H, s), 7.00—7.10 (2H, m), 7.30—7.40 (1H, m)
VIIa	OCH <sub>3</sub>	Н	94	1.87—2.08 (1H, m), 2.15—2.31 (1H, m), 3.42 (3H, s), 3.70—3.87 (1H, m), 4.02—4.18 (1H, m), 4.29 (1H, t, 4), 5.20 (2H, s), 6.96—7.12 (2H, m), 7.22—7.32 (1H, m)
VIIb	$OCH_2CH = CH_2$	Н	64	1.90—2.10 (1H, m), 2.10—2.30 (1H, m), 3.70—3.90 (1H, m), 4.00—4.20 (3H, m), 4.40—4.50 (1H, m), 5.19 (2H, s), 5.10—5.40 (2H, m), 5.80–6.10 (1H, m), 6.90—7.10 (2H, m), 7.20—7.30 (1H, m)
X	$\begin{bmatrix} 0 \\ 0 \end{bmatrix}$		72	2.05—2.15 (2H, m), 4.00—4.08 (2H, m), 4.09—4.26 (4H, m), 5.20 (2H, s), 7.04—7.10 (2H, m), 7.40—7.45 (1H, m)
XII	$NHCH_2CH = CH_2$	H	80	1.90—2.15 (2H, m), 3.25—3.50 (2H, m), 3.70—3.95 (2H, m), 3.95—4.10 (1H, m), 5.10—5.30 (2H, m), 5.19 (2H, s), 5.85—6.10 (1H, m), 6.95—7.10 (2H, m), 7.25—7.40 (1H, m)
XIII	$N \subset CH_3$ $CH_2CH = CH_2$	Н	95	1.80—2.00 (1H, m), 2.00—2.20 (1H, m), 2.24 (3H, s), 3.00 (1H, dd, 11, 5.5), 3.18 (1H, dd, 11, 4.5), 3.50—3.65 (1H, m), 3.84 (1H, dd, 8, 3.5), 4.25—4.35 (1H, m), 5.10—5.30 (2H, m), 5.19 (2H, s), 5.75—5.95 (1H, m), 6.90—7.10 (2H, m), 7.56 (1H, d, 5.5)

Table II. 4-Substituted 8-Hydroxymethyl-1,2,3,4-tetrahydroquinolines

Compd. No.	R <sup>1</sup>	R <sup>2</sup>	Yield (%)	$^{1}\text{H-NMR} (\text{CDCl}_{3}, J = \text{Hz})$				
VIIa <sup>a)</sup>	OCH <sub>3</sub>	Н	64	1.65—1.88 (1H, m), 2.01—2.18 (1H, m), 2.92 (1H, brs), 3.16—3.50 (2H, m), 3.33 (3H, s), 4.20 (1H, t, 3), 4.40 (2H, s), 4.95 (1H, brs), 6.55 (1H, t, 7.5), 6.91 (1H, dd, 1.5, 7.5), 7.06 (1H, dd, 1.5, 7.5)				
VIII <sup>a)</sup>	$OCH_2CH = CH_2$	H	97	1.27 (3H, t, 7), 1.98 (2H, q, 4.5), 3.10—3.35 (2H, m), 3.95—4.25 (2H, m), 4.43 (1H, t, 4.5), 4.30—4.80 (1H, m), 4.60—4.90 (2H, m), 5.10—5.40 (2H, m), 5.70—6.10 (1H, m), 6.90—7.30 (3H, m)				
XI	$\begin{bmatrix} 0 \\ 0 \end{bmatrix}$		61	1.60—1.90 (1H, m), 1.90—2.10 (2H, m), 3.40—3.50 (2H, m), 4.00—4.30 (4H, m), 4.56 (2H, s), 5.00 (1H, br s), 6.62 (1H, t, 7.5), 6.98 (1H, dd, 1.5, 7.5), 7.34 (1H, dd, 1.5, 7.5)				
XIV <sup>a)</sup>	$N = CH_3$ $CH_2CH = CH_2$	Н	91	1.70 (1H, br s), 1.93 (2H, q, 6.5), 2.24 (3H, s), 3.01 (1H, dd, 7, 14), 3.20 (1H, dd, 7, 14), 3.30—3.55 (2H, m), 3.89 (1H, t, 7), 4.55 (1H, d, 12), 4.63 (1H, d, 12), 4.80 (1H, br s), 5.05—5.30 (2H, m), 5.75—6.00 (1H, m), 6.60 (1H, t, 7.5), 6.92 (1H, dd, 1.5, 7.5), 7.42 (1H, t, 7.5)				

a) Oily compounds were purified by column chromatography.

$$\begin{array}{c} OH \\ OH \\ OH \\ TsOH \end{array}$$

$$\begin{array}{c} OH \\ TsOH \end{array}$$

$$\begin{array}{c} OH \\ TsOH \end{array}$$

$$\begin{array}{c} NAOH \\ TsOH \end{array}$$

$$\begin{array}{c} I) \ NAOH \\ HOCH_2 \ H \end{array}$$

$$\begin{array}{c} I) \ NAOH \\ HOCH_2 \ H \end{array}$$

$$\begin{array}{c} I) \ NAOH \\ HOCH_2 \ H \end{array}$$

$$\begin{array}{c} I) \ NAOH \\ II) \ NAOH \\ III) \ NAOH \ NAOH$$

Chart 4

Table III. 4-Substituted 8-Hydroxymethyl-1,2,3,4-tetrahydroquinolines

Compd. No.	R¹	R <sup>2</sup>	R³	Yield (%)	$^{1}$ H-NMR (CDCl <sub>3</sub> , $J$ =Hz)
IXa	C <sub>2</sub> H <sub>5</sub>	OCH <sub>3</sub>	Н	50	1.25 (3H, t, 7), 1.95—2.10 (2H, m), 2.96 (2H, q, 7), 3.15—3.25 (2H, m), 3.41 (3H, s), 4.28 (1H, t, 4.5), 4.46 (1H, br s), 4.70 (1H, d, 13.5), 4.80 (1H, d, 13.5), 6.98 (1H, t, 7.5), 7.10—7.35 (2H, m)
IXb	$C_2H_5$	$OCH_2CH = CH_2$	Н	95	1.35 (3H, t, 7), 1.90—2.05 (2H, m), 2.85—3.05 (2H, m), 3.10—3.30 (2H, m), 4.00—4.20 (2H, m), 4.43 (1H, t, 4.5), 4.60—4.85 (2H, m), 5.10—5.40 (2H, m), 5.80—6.10 (1H, m), 6.90—7.10 (1H, m), 7.10—7.30 (2H, m)
IXc	$CH_2CH = CH_2$	OCH <sub>3</sub>	Н	28	1.90—2.10 (2H, m), 3.00—3.20 (2H, m), 3.42 (3H, s), 3.53 (2H, d, 6), 3.90 (1H, s), 4.27 (1H, t, 4.5), 4.64 (1H, d, 13), 4.74 (1H, d, 13), 5.20—5.40 (2H, m), 5.80—6.10 (1H, m), 6.97 (1H, t, 7.5), 7.21 (2H, d, 7)
IXd	C <sub>2</sub> H <sub>5</sub>	$\begin{pmatrix} 0 \\ 0 \end{pmatrix}$		40	1.19 (3H, t, 7), 1.90—2.00 (2H, m), 2.90 (2H, q, 7), 3.20—3.30 (2H, m), 3.90—4.20 (4H, m), 4.40 (1H, brs), 4.61 (2H, s), 6.95 (1H, t, 7.5), 7.22 (1H, d, 7.5), 7.32 (1H, dd, 1.5, 7.5)
IXe	$CH_2CH = CH_2$	$\begin{bmatrix} 0 \\ 0 \end{bmatrix}$		42	1.90—2.10 (2H, m), 3.20—3.30 (2H, m), 3.59 (2H, d, 6), 3.70 (1H, br s), 4.00—4.30 (4H, m), 4.72 (2H, s), 5.23 (1H, dd, 1.5, 10), 5.32 (1H, dd, 1.5, 17), 5.80—6.10 (1H, m), 7.03 (1H, t, 7.5), 7.26 (1H, dd, 1.5, 7.5), 7.39 (1H, dd, 1.5, 7.5)
IXf	C <sub>2</sub> H <sub>5</sub>	=O		98	1.32 (3H, t, 7), 2.73 (2H, t, 6.5), 3.16 (2H, q, 7), 3.57 (2H, t, 6.5), 3.75 (1H, br s), 4.76 (2H, s), 7.10 (1H, t, 7.5), 7.50 (1H, dd, 1.5, 7.5), 7.92 (1H, dd, 1.5, 7.5)
IXg	$CH_2CH = CH_2$	=O		95	2.60—2.70 (2H, m), 3.40—3.60 (2H, m), 3.60 (1H, br s), 3.74 (2H, d, 5.5), 4.80 (2H, br s), 5.20—5.50 (2H, m), 5.80—6.10 (1H, m), 7.05 (1H, t, 7.5), 7.55 (1H, dd, 1.5, 7.5), 7.89 (1H, dd, 1.5, 7.5)
IXh	C <sub>2</sub> H <sub>5</sub>	NH⊲	Н	81	0.30—0.60 (3H, m), 1.27 (3H, t, 7), 1.80—2.15 (2H, m), 2.15—2.30 (1H, m), 2.93 (2H, q, 7), 3.00—3.30 (2H, m), 3.85 (1H, t, 5), 4.68 (1H, d, 13), 4.73 (1H, d, 13), 6.95 (1H, t, 7.5), 7.00—7.35 (2H, m)
IXi	C <sub>2</sub> H <sub>5</sub>	NH	Н	98	1.00—1.50 (8H, m), 1.50—2.20 (7H, m), 2.55—2.75 (1H, m), 2.80—3.05 (3H, m), 3.05—3.35 (2H, m), 3.86 (1H, t, 4.5), 4.60—4.95 (3H, m), 6.90—7.40 (3H, m)
IXj	C <sub>2</sub> H <sub>5</sub>	$NHCH_2CH = CH_2$	Н	98	1.25 (3H, t, 7), 1.70—2.20 (3H, m), 2.91 (2H, q, 7), 3.00—3.50 (5H, m), 3.75 (1H, t, 5), 4.50—4.90 (3H, m), 5.00—5.30 (2H, m), 5.80—6.05 (1H, m), 6.90—7.30 (3H, m)
IXk	$CH_2CH = CH_2$	NHCH <sub>3</sub>	Н	89	1.70—2.00 (2H, m), 2.46 (3H, s), 3.00—3.20 (2H, m), 3.50 (2H, d, 6), 3.62 (1H, t, 4.5), 4.61 (1H, d, 13), 4.73 (1H, d, 13), 5.10—5.40 (2H, m), 5.80—6.10 (1H, m), 6.95 (1H, t, 7.5), 7.17 (2H, d, 8)
IXI	C <sub>2</sub> H <sub>5</sub>	N CH <sub>3</sub>	Н	80	0.35—0.60 (4H, m), 1.26 (3H, t, 7), 1.65—1.90 (1H, m), 2.00—2.30 (2H, m), 2.16 (3H, s), 2.70—3.40 (4H, m), 4.02 (1H, t, 8), 4.61 (1H, d, 13), 4.92 (1H, d, 13), 5.31 (1H, br s), 6.90—7.10 (2H, m), 7.37 (1H, dd, 1.5, 7)
IXm	C <sub>2</sub> H <sub>5</sub>	N <ch<sub>3</ch<sub>	Н	43	1.25 (3H, t, 7), 1.20—2.30 (12H, m), 2.13 (3H, s), 2.40—2.60 (1H, m), 2.80—3.30 (4H, m), 4.04 (1H, t, 8), 4.59 (1H, d, 13), 4.94 (1H, d, 13), 5.30 (1H, br s), 6.90—7.10 (2H, m), 7.55 (1H, dd, 2.5, 7)
IXn	C <sub>2</sub> H <sub>5</sub>	$N CH_2CH = CH_2$ $C_2H_5$	Н	68	1.04 (3H, t, 7), 1.25 (3H, t, 7), 1.60—1.80 (1H, m), 1.80—2.10 (1H, m), 2.30—2.70 (2H, m), 2.75—3.15 (4H, m), 3.15—3.35 (2H, m), 3.90—4.10 (1H, m), 4.55 (1H, d, 13), 4.95 (1H, d, 13), 5.00—5.35 (2H, m), 5.25 (1H, brs), 5.70—6.00 (1H, m), 6.90—7.10 (2H, m), 7.66 (1H, dd, 2.5, 6.5)
IXo	$CH_2CH = CH_2$	NCH <sub>3</sub>	Н	51	1.59—1.78 (1H, m), 1.92—2.18 (1H, m), 2.28 (6H, s), 3.00—3.28 (2H, m), 3.48—3.60 (2H, m), 3.80 (1H, t, 7.5), 4.62 (1H, d, 13), 4.90 (1H, d, 13), 5.17—5.42 (2H, m), 5.85—6.11 (1H, m), 7.00 (1H, t, 7.5), 7.10 (1H, dd, 1.5, 7.5), 7.51 (1H, dd, 1.5, 7.5)
IXp	C <sub>2</sub> H <sub>5</sub>	$N \subset CH_2CH = CH_2$ $CH_3$	Н	59	1.25 (3H, t, 7), 1.55—1.75 (1H, m), 1.85—2.15 (1H, m), 2.20 (3H, s), 2.75—3.40 (5H, m), 3.93 (1H, t, 7), 4.59 (1H, d, 13), 4.93 (1H, d, 13), 5.00—5.35 (2H, m), 5.70—6.00 (1H, m), 6.90—7.10 (2H, m), 7.55—7.65 (1H, m)
IXq	$CH_2CH = CH_2$	$N \subset CH_2CH = CH_2$ $CH_3$	Н	72	1.60—1.80 (1H, m), 1.90—2.20 (1H, m), 2.21 (3H, s), 2.90—3.30 (4H, m), 3.40—3.60 (2H, m), 3.93 (1H, t, 8), 4.60 (1H, d, 13), 4.92 (1H, d, 13), 5.00—5.40 (4H, m), 5.70—6.10 (2H, m), 6.90—7.15 (2H, m), 7.61 (1H, d, 6.5)

1,4-Disubstituted 8-hydroxymethyl-1,2,3,4-tetrahydroquinolines (IXa—q) were prepared from V by four different methods.

First, reduction of V with sodium borohydride easily gave the hydroxy derivative (VI), which was alkylated with alkyl halides in the presence of sodium hydride to the alkoxy derivatives (VIIa, b), followed by hydrolysis with sodium hydroxide to give the tetrahydroquinolines (VIIIa, b). Treatment of VIIIa, b with sodium hydride in tetrahydrofuran (THF) afforded the corresponding sodium salts, which were alkylated with alkyl halides in the presence of *n*-butyllithium to give 1-substituted 4-alkoxy-tetrahydro-

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quinolines (IXa—c) (Chart 3, Tables I and II).

Secondly, pyrido[3,2,1-ij]benzoxazine (V) was protected as the acetal (X) in the usual manner and X was hydrolyzed with sodium hydroxide to give tetrahydroquinoline (XI); these were alkylated in the same manner as described for the synthesis of IXa to give the 1-substituted tetrahydroquinolines (IXd, e), followed by removal of the acetal group with hydrochloric acid to give the 4-oxo-1,2,3,4-tetrahydroquinolines (IXf, g) (Chart 4).

Thirdly, 4-(N-alkyl-N-methylamino)-1,2,3,4-tetrahydroquinolines (IXI—o) were synthesized through reduction of the imide group with sodium borohydride. Treatment of the 4-oxo derivatives (IXf, g) with alkylamine in the presence of p-toluenesulfonic acid (TsOH) gave the imide compounds, followed by reduction with sodium borohydride to give the 4-alkylamino derivatives (IXh—k). Formylation of IXh—k with formic acid—acetic anhydride gave the formyl products, followed by reduction with lithium aluminum hydride (LAH) to give the N-alkyl-N-methylamino derivatives (IXl—o) (Chart 4). N-Allyl-N-methylamino derivatives (IXp, q) were also synthesized by this method, but did not give a satisfactory yield.

Finally, 4-(N-allyl-N-methylamino)-1,2,3,4-tetrahydro-

quinolines (IXp, q) were synthesized from V according to the procedure shown in Chart 5. Treatment of V with allylamine in the presence of TsOH gave the imide products, followed by reduction with sodium borohydride to give 7-(N-allylamino)-3-oxopyrido[3,2,1-ij]benzoxazine (XII). Methylation of XII with formic acid-formaldehyde gave the N-allyl-N-methylamino compound (XIII). Hydrolysis of XIII with sodium hydroxide gave the tetrahydroquinoline (XIV), which was alkylated in the same manner as described for the synthesis of IXa to give the N-allyl-N-methylamino derivatives (IXp, q) (Chart 5 and Table III).

Condensation of 2-mercaptobenzimidazoles<sup>3)</sup> with 8-chloromethyl-1,2,3,4-tetrahydroquinolines, which were synthesized by treatment of 8-hydroxymethyl derivatives (IXa—q) with thionyl chloride, in the presence of potassium carbonate afforded the corresponding sulfides (XVIa—i, k, l, o—e1, i1) in good yields (Chart 6). The following sulfides (XVIf1, g1, i1) were synthesized as shown in Charts 7 and 8. N-Methylamino derivative (XVIe1) was acylated with cyclopropanecarboxylic anhydride to give the cyclopropylcarbonyl derivative, followed by reduction with LAH to give the N-cyclopropylmethyl-N-methylamino derivative (XVIf1). N-(2-Hydroxyethyl)-N-methylamino derivative

Chart 6

Chart 6

Chart 6

Chart 6

Chart 6

Chart 6

NHCH<sub>3</sub>

NHCH<sub>3</sub>

$$Et_3N$$
 $ii)$  ( $CO)_2O$ 
 $Et_3N$ 
 $ii)$  LAH

 $ii)$  CHO

 $CHO$ 
 $CH_2$ 
 $CH_3$ 
 $CH_3$ 

Chart 7

Table IV-1. 4-Substituted 8-[(2-Benzimidazolyl)thiomethyl]-1,2,3,4-tetrahydroquinolines

Compd. No.	R <sup>1</sup>	R <sup>2</sup> R <sup>3</sup>	R <sup>4</sup>	Yield (%)	$^{1}$ H-NMR (CDCl <sub>3</sub> , $J=$ Hz)
XVIa	C <sub>2</sub> H <sub>5</sub>	CH <sub>3</sub> O H	Н	43	1.25 (3H, t, 7), 1.80—2.10 (2H, m), 2.80—3.30 (4H, m), 3.35 (3H, s), 3.74 (3H, s), 4.10—4.30 (1H, m), 4.45 (2H, s), 6.70—7.40 (6H, m), 12.00—12.60 (1H, br s)
XVIb	$C_2H_5$	CH₃O H	5-CH <sub>3</sub>	58	1.29 (3H, t, 7), 1.90—2.20 (2H, m), 2.43 (3H, s), 2.90—3.20 (2H, m), 3.25 (2H, br s), 3.41 (3H, s), 4.24 (1H, br s), 4.47 (2H, s), 6.90—7.60 (6H, m)
XVIc	$C_2H_5$	CH₃O H	5-CH <sub>3</sub> O	63	1.25 (3H, t, 7), 1.95 (2H, br s), 2.80—3.20 (2H, m), 3.16 (2H, br s), 3.36 (3H, d, 6), 3.75 (3H, d, 5.5), 4.17 (1H, br s), 4.45 (2H, s), 6.70—7.50 (6H,
XVId	$C_2H_5$	CH₃O H	5-F 6-CH <sub>3</sub> O	61	m), 12.23 (1H, brs) 1.41 (3H, t, 7), 2.05—2.20 (2H, m), 2.98—3.27 (2H, m), 3.33—3.48 (2H m), 3.44(3H, s), 3.89 (3H, brs), 4.23—4.34 (3H, m), 6.79—7.33 (4H, m),
XVIe	$C_2H_5$	CH <sub>3</sub> O H	5-CH <sub>3</sub> CO	36	7.42 (1H, dd, 2, 8) 1.33 (3H, t, 7), 2.00—2.10 (2H, m), 2.60 (3H, s), 2.90—3.20 (2H, m), 3.30—3.40 (2H, m), 3.39 (3H, s), 4.23 (1H, t, 4), 4.39 (2H, d, 5.5), 6.90— 7.70 (6H, m)
XVIf	$C_2H_5$	$CH_2 = CHCH_2O$ H	Н	95	1.38 (3H, t, 7.5), 2.04—2.12 (2H, m), 3.04 (2H, q, 7.5), 3.40—3.45 (2H, m), 4.00—4.20 (2H, m), 4.29 (2H, s), 4.43 (1H, t, 0.5), 5.15—5.35 (2H,
XVIg	C <sub>2</sub> H <sub>5</sub>	CH <sub>2</sub> =CHCH <sub>2</sub> O H	5-CH <sub>3</sub> CO	96	m), 5.85—6.00 (1H, m), 7.00—7.60 (7H, m), 12.54 (1H, brs) 1.45 (3H, t, 7.5), 2.05—2.20 (2H, m), 2.65 (3H, s), 3.13 (2H, q, 7.5), 3.49 (2H, brs), 4.00—4.30 (2H, m), 4.30 (2H, s), 4.46 (1H, t, 0.5), 5.10—5.35 (2H, m), 5.85—6.05 (1H, m), 7.13 (1H, t, 8), 7.35 (2H, dd, 0.3, 8), 7.50—8.20
XVIh	C <sub>2</sub> H <sub>5</sub>	$\begin{bmatrix} 0 \end{bmatrix}$	Н	58	(3H, m), 13.00 (1H, br s) 1.22 (3H, t, 7), 1.90—2.05 (2H, m), 3.04 (2H, q, 7), 3.16—3.29 (2H, m), 3.98—4.23 (4H, m), 4.57 (2H, s), 6.94 (1H, t, 7.5), 7.10—7.22 (2H, m), 7.27—7.56 (4H, m)
XVIi	$C_2H_5$	=O	Н	57	1.40 (3H, t, 7), 2.70—2.90 (2H, m), 3.23 (2H, q, 7), 3.60—3.80 (2H, m), 4.44 (2H, s), 7.00—7.30 (4H, m), 7.30—7.60 (2H, m), 7.66 (1H, dd, 1.5, 8), 7.90 (1H, dd, 1.5, 8)
XVIj	$C_2H_5$	OH H	Н	88	1.40 (3H, t, 7), 1.90—2.30 (2H, m), 2.90—3.20 (2H, m), 3.30—3.50 (2H m), 4.12 (1H, d, 15), 4.33 (1H, d, 15), 7.00—7.50 (7H, m), 12.10—12.80 (1H, br s)
XVIk	$CH_2CH = CH_2$	CH₃O H	5-CH <sub>3</sub> CO	61	1.90—2.10 (2H, m), 2.59 (3H, s), 3.20—3.30 (2H, m), 3.40 (3H, s), 3.59 (2H, d, 5.5), 4.23 (1H, t, 4.5), 4.40 (1H, d, 14), 4.49 (1H, d, 14), 5.22 (1H, d, 10.5), 5.36 (1H, d, 17), 5.60—6.10 (1H, m), 6.98 (1H, t, 7.5), 7.21 (1H, d, 7),
XVII	$CH_2CH = CH_2$	CH₃O H	5-F 6-CH <sub>3</sub> O	64	7.36 (1H, d, 7), 7.30—7.50 (1H, m), 7.81 (1H, d, 8.5), 8.00—8.20 (1H, m) 1.90—2.10 (2H, m), 3.10—3.30 (2H, m), 3.40 (3H, s), 3.50—3.60 (2H, m), 3.82 (3H, s), 4.23 (1H, t, 4.5), 4.30—4.50 (2H, m), 5.10—5.50 (2H, m),
XVIm	$C_2^{\circ}H_5$	= N-OH	Н	61	5.80—6.00 (1H, m), 6.70—7.50 (5H, m) 1.44 (3H, t, 7), 2.93—3.19 (4H, m), 3.49 (2H, t, 6.5), 4.32 (2H, s), 7.05—7.96 (8H, m), 12.00 (1H, br s)
XVIn	$C_2H_5$	NHCH <sub>3</sub> H	Н	92	1.43 (3H, t, 7), 1.88—2.17 (2H, m), 2.48 (3H, s), 2.95—3.25 (2H, m), 3.30—3.58 (2H, m), 3.67 (1H, t, 5), 4.21 (1H, d, 15), 4.32 (1H, d, 15),
XVIo	$C_2H_5$	NHCH <sub>2</sub> CH=CH <sub>2</sub> H	5-CH <sub>3</sub>	85	7.00—7.45 (6H, m), 7.56 (1H, br s) 1.33 (3H, t, 7), 1.75—2.10 (2H, m), 2.41 (3H, s), 2.90—3.20 (2H, m), 3.20—3.50 (4H, m), 3.76 (1H, t, 5), 4.23 (1H, d, 14.5), 4.31 (1H, d, 14.5),
XVIp	C <sub>2</sub> H <sub>5</sub>	NH H	5-CH <sub>3</sub>	81	5.00—5.30 (2H, m), 5.80—6.10 (1H, m), 6.90—7.50 (6H, m), 12.67 (1H, br s 0.95—2.20 (10H, m), 1.28 (3H, t, 7), 2.39 (3H, s), 2.55—2.80 (1H, m), 2.80—3.20 (2H, m), 3.20—3.50 (2H, m), 3.82 (1H, t, 4.5), 4.20 (1H, d,
XVIq	$C_2H_5$	N(CH <sub>3</sub> ) <sub>2</sub> H	Н	31	14.5), 4.31 (1H, d, 14.5), 6.80—7.50 (6H, m) 1.42 (3H, t, 7), 1.70—1.90 (1H, m), 2.09—2.36 (1H, m), 2.26 (6H, s), 2.90—3.42 (3H, m), 3.42—3.65 (1H, m), 4.81 (1H, t, 7.5), 4.09 (1H, d, 15),
XVIr	C <sub>2</sub> H <sub>5</sub>	N(CH <sub>3</sub> ) <sub>2</sub> H	5-F 6-CH <sub>3</sub> O	40	4.42 (1H, d, 15), 7.00—7.70 (7H, m) 1.41 (3H, t, 7), 1.65—1.93 (1H, m), 2.06—2.38 (1H, m), 2.26 (6H, s), 2.88—3.12 (1H, m), 3.12—3.37 (2H, m), 3.40—3.61 (1H, m), 3.80 (1H, t, 7), 3.89 (3H, br s), 4.09 (1H, d, 15), 4.39 (1H, d, 15), 6.80—7.23 (3H, m),
XVIs	$CH_2CH = CH_2$	N(CH <sub>3</sub> ) <sub>2</sub> H	Н	95	7.36 (1H, d, 7), 7.52 (1H, d, 8) 1.70—1.90 (1H, m), 2.10—2.30 (1H, m), 2.27 (6H, s), 3.20—3.40 (1H, m), 3.40—3.90 (4H, m), 4.14 (1H, d, 15), 4.48 (1H, d, 15), 5.38 (1H, d, 11),
XVIt	CH <sub>2</sub> CH=CH <sub>2</sub>	N(CH <sub>3</sub> ) <sub>2</sub> H	5-F 6-CH <sub>3</sub> O	92	5.44 (1H, d, 18), 6.00—6.20 (1H, m), 7.00—7.70 (7H, m), 12.30 (1H, br s) 1.55—1.80 (1H, m), 1.90—2.10 (1H, m), 2.18 (6H, s), 2.95—3.15 (1H, m), 3.20—3.35 (1H, m), 3.50—3.60 (2H, m), 3.65—3.85 (1H, m), 3.78 (3H, s), 3.88 (1H, br s), 4.19 (1H, d, 14), 4.39 (1H, d, 14), 5.10—5.40 (2H, m), 5.80—6.10 (1H, m), 6.75—7.30 (4H, m), 7.39 (1H, d, 7.5)

TABLE IV-1. (continued)

Compd. No.	R <sup>1</sup>	$R^2$ $R^3$	R <sup>4</sup>	Yield (%)	$^{1}\text{H-NMR} (CDCl_{3}, J=\text{Hz})$
XVIu	$CH_2CH = CH_2$	N(CH <sub>3</sub> ) <sub>2</sub> H	5-CH <sub>3</sub> CO	22	1.72—1.93 (1H, m), 2.09—2.40 (1H, m), 2.28 (6H, s), 2.64 (3H, s), 3.20—3.41 (1H, m), 3.44—3.77 (3H, m), 3.82 (1H, t, 8), 4.15 and 4.19 (1H, d, 15), 4.50 (1H, d, 15), 5.30—5.55 (2H, m), 5.98—6.24 (1H, m), 7.11 (1H, t, 8), 7.29 (0.5H, d, 3.5), 7.38 (1H, d, 7.5), 7.50—7.66 (1.5H, m), 7.73—7.90 (1H, m), 7.94 and 8.21 (1H, br s)
XVIv	C <sub>2</sub> H <sub>5</sub>	NCH <sub>2</sub> CH=CH <sub>2</sub> CH <sub>3</sub>	5-CH <sub>3</sub>	73	1.41 (3H, t, 7), 1.70—1.90 (1H, m), 2.15—2.30 (1H, m), 2.19 (3H, s), 2.43 (3H, s), 2.60—3.30 (6H, m), 3.45—3.60 (1H, m), 3.95 (1H, t, 8), 4.06 (1H, d, 15), 4.41 (1H, d, 15), 5.05—5.25 (2H, m), 5.70—6.00 (1H, m), 6.90—7.6 (6H, m), 12.48 (1H, br s)
XVIw	$C_2H_5$	$N \subset CH_2CH = CH_2$ $CH_3$ $H$	5-CH <sub>3</sub> CO	60	1.40 (3H, t, 7), 1.70—1.95 (1H, m), 2.05—2.30 (1H, m), 2.20 (3H, s), 2.67 (3H, s), 2.80—3.35 (5H, m), 3.45—3.70 (1H, m), 3.97 (1H, t, 8.5), 4.2′ (1H, d, 14.5), 4.50 (1H, d, 14.5), 5.05—5.35 (2H, m), 5.70—6.00 (1H, m), 7.00—8.30 (6H, m)
XVIx	C <sub>2</sub> H <sub>5</sub>	NCH <sub>2</sub> CH=CH <sub>2</sub> CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	66	1.40 (3H, t, 7), 1.70—1.95 (1H, m), 2.05—2.30 (1H, m), 2.20 (3H, s), 2.67 (3H, s), 2.80—3.35 (5H, m), 3.45—3.70 (1H, m), 3.97 (1H, t, 8.5), 4.27 (1H, d, 14.5), 4.50 (1H, d, 14.5), 5.05—5.35 (2H, m), 5.70—6.00 (1H, m), 7.00—8.30 (6H, m)
XVIy	C <sub>2</sub> H <sub>5</sub>	N <sup>C</sup> CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	75	0.30 - 0.60 (4H, m), 1.38 (3H, t, 7), 1.80 - 2.00 (1H, m), 2.00 - 2.40 (2l m), 2.14 (3H, s), 2.80 - 3.15 (1H, m), 3.15 - 3.35 (2H, m), 3.35 - 3.60 (1H, m), 3.87 (3H, s), 3.97 (1H, t, 6.5), 4.15 (1H, d, 14.5), 4.40 (1H, d, 14.5), 6.80 - 7.60 (5H, m), 12.90 (1H, br s)
XVIz	C <sub>2</sub> H <sub>5</sub>	NCH <sub>3</sub>	5-CH <sub>3</sub> CO	56	0.30—0.65 (4H, m), 1.44 (3H, t, 7), 1.80—2.45 (4H, m), 2.15 (3H, s), 2.65 (3H, s), 2.90—3.10 (1H, m), 3.10—3.40 (2H, m), 3.50—3.75 (1H, m), 4.02 (1H, t, 8), 4.13 (1H, d, 15), 4.43 (1H, d, 15), 7.06 (1H, t, 7.5), 7.25—8.30 (5H, m), 13.33 (1H, br s)
XVIa1	C <sub>2</sub> H <sub>5</sub>	N CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	89	1.00—1.50 (5H, m), 1.36 (3H, t, 7), 1.50—2.00 (6H, m), 2.11 (3H, s), 2.10—2.35 (1H, m), 2.40—2.60 (1H, m), 2.85—3.10 (1H, m), 3.10—3. (2H, m), 3.35—3.60 (1H, m), 3.88 (3H, s), 4.05 (1H, t, 7), 4.19 (1H, d), 4.45 (1H, d, 14.5), 6.90—7.60 (5H, m)
XVIb1	$C_2H_5$	N CH2CH = CH2 $ C2H5 $ H	5-F 6-CH <sub>3</sub> O	83	1.03 (3H, t, 7), 1.41 (3H, t, 7), 1.60—1.95 (1H, m), 2.00—2.20 (1H, m), 2.30—2.70 (2H, m), 2.80—3.05 (2H, m), 3.05—3.35 (3H, m), 3.45—3. (1H, m), 3.88 (3H, s), 3.95—4.20 (2H, m), 4.40 (1H, d, 15), 5.00—5.30 (2H, m), 5.70—5.90 (1H, m), 6.80—7.40 (4H, m), 7.69 (1H, d, 7.5), 12.70 (1H, br s)
XVIc1	$CH_2CH = CH_2$	NCH <sub>2</sub> CH=CH <sub>2</sub> CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	93	1.60—1.90 (1H, m), 1.90—2.30 (1H, m), 2.20 (3H, s), 2.90—4.00 (7H, m), 3.88 (3H, s), 4.00—4.20 (1H, m), 4.45 (1H, d, 15), 5.00—5.50 (4H, m) 5.70—6.20 (2H, m), 6.80—7.40 (4H, m), 7.62 (1H, d, 7.5), 12.00—12.30 (1H, br s)
XVId1	$C_2H_5$	=O	5-F 6-CH₃O	95	1.42 (3H, t, 7), 2.80 (2H, t, 6), 3.25 (2H, q, 7), 3.68 (2H, t, 6), 3.90 (3) s), 4.41 (2H, s), 6.87—7.47 (2H, br s), 7.12 (1H, t, 7.5), 7.65 (1H, d, 7.7.00 (1H, d, 7.5)
XVIe1	$C_2H_5$	NHCH <sub>3</sub> H	5-F 6-CH₃O	87	1.42 (3H, t, 7.5), 1.98 (2H, br s), 2.49 (3H, s), 2.95—3.26 (2H, m), 3.30—3. (2H, m), 3.88 (3H, br s), 4.10—4.40 (2H, m), 6.75—7.45 (5H, m)
XVIf1	C <sub>2</sub> H <sub>5</sub>	N <sup>CH</sup> <sub>2</sub> CH <sub>3</sub> H	5-F 6-CH <sub>3</sub> O	48	0—0.20 (2H, m), 0.35—0.62 (2H, m), 0.76—0.99 (1H, m), 1.41 (3H, t 7), 1.66—1.90 (1H, m), 2.02—2.47 (3H, m), 2.29 (3H, s), 2.83—3.09 (1H, m), 3.09—3.34 (2H, m), 3.34—3.62 (1H, m), 3.88 (3H, s), 4.02 (1H, t, 6.5), 4.08 (1H, d, 15), 4.38 (1H, d, 15), 6.78—7.22 (3H, m), 7.34 (1H, 7.5), 7.64 (1H, d, 8), 12.55 and 12.83 (1H, br s)
XVIg1	$C_2H_5$	$N \subset (CH_2)_2OH$ $CH_3$	5-F 6-CH <sub>3</sub> O	45	1.42 (3H, t, 7), 1.51—1.98 (2H, m), 2.24 (3H, s), 2.58—2.74 (2H, m), 2.87—4.10 (7H, m), 3.88 (3H, s), 4.10 (1H, d, 15), 4.39 (1H, d, 15), 6.74—7.61 (5H, m), 12.20—12.70 (1H, br s)
XVIh1	$CH_2CH = CH_2$	=0	5-F 6-CH <sub>3</sub> O	66	2.78 (2H, t, 6.5), 3.63 (2H, t, 6.5), 3.83 (2H, d, 6), 3.90 (3H, s), 4.45 (2H, s), 5.30—5.51 (2H, m), 6.00—6.20 (1H, m), 6.80—8.00 (5H, m), 10.28 and 10.68 (1H, br s)
XVIi1	$CH_2CH = CH_2$	NH <sub>2</sub> H	5-F 6-CH <sub>3</sub> O	38	1.70—1.90 (1H, m), 2.09—2.32 (1H, m), 3.23—3.51 (2H, m), 3.66 (2H, d, 6), 3.89 (3H, s), 4.05 (1H, t, 5.5), 4.26 (1H, d, 14.5), 4.36 (1H, d, 14.5), 5.30—5.52 (2H, m), 5.97—6.23 (1H, m), 6.77—7.43 (5H, m)

Chart 8

TABLE IV-2. 4-Substituted 8-[(2-Benzimidazolyl)thiomethyl]-1,2,3,4-tetrahydroquinolines

						Analy	/sis (%)		
Compd. No.	Appearance (Recrystn. solv.)	mp (°C)	Formula		Calcd	•	. ,	Found	
NO.	(Recrystii. solv.)	( C)		С	Н	N	C	Н	N
XVIa	Pale yellow powder(AcOEt-hexane)	136—137	C <sub>20</sub> H <sub>23</sub> N <sub>3</sub> OS	67.96	6.59	11.89	67.83	6.42	11.80
XVIh	White powder (MeOH)	193—194	$C_{22}H_{21}N_3O_2S$	67.50	5.41	10.73	67.55	5.35	10.80
XVIi	Yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	165—167	$C_{19}H_{19}N_3OS$	67.63	5.68	12.45	67.69	5.71	12.08
XVII	White powder (AcOEt-hexane)	154—155	$C_{22}H_{24}FN_3O_2S$	63.90	5.85	10.16	63.72	5.89	10.04
XVIm	Colorless prisms (MeOH)	201202	$C_{19}H_{20}N_4OS$	64.75	5.72	15.90	64.34	5.69	15.67
XVIn	Yellow powder (AcOEt)	144145	$C_{20}H_{24}N_4S \cdot 1/4H_2O$	67.29	6.92	15.69	67.51	6.80	15.37
XVIq	Pale yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	150151.5	$C_{21}H_{26}N_4S$	68.82	7.15	15.29	68.63	6.95	15.04
XVIs	Yellow powder (Et <sub>2</sub> O-hexane)	145—146	$C_{22}H_{26}N_4S \cdot 1/4H_2O$	68.98	6.97	14.63	69.12	6.85	14.60
XVIt	Pale yellow powder (Et <sub>2</sub> O-hexane)	131—132	$C_{23}H_{27}FN_4OS$	64.76	6.38	13.13	64.58	6.28	13.15
XVIu	Yellow powder (Et <sub>2</sub> O-hexane)	143—145	$C_{24}H_{28}N_4OS \cdot 1/4H_2O$	67.81	6.76	13.18	67.79	6.51	13.31
XVIv	White powder (Et <sub>2</sub> O)	132—133	$C_{24}H_{30}N_4S \cdot 1/4H_2O$	70.12	7.48	13.63	70.08	7.35	13.80
XVIz	Yellow powder (AcOEt-hexane)	167.5—168.5	$C_{25}H_{30}N_4OS \cdot 1/4H_2O$	68.38	7.00	12.76	68.75	6.69	12.28
XVIb1	Colorless needles (AcOEt-hexane)	146—147	$C_{25}H_{31}FN_4OS \cdot 1/4H_2O$	65.40	6.92	12.20	65.57	6.90	12.04

(XVIg1) was synthesized by reduction of the formyl derivative which was prepared from XVIe1 by reaction with glyoxal. Treatment of 4-oxo derivative (XVIh1) with ammonia gave the imide product, followed by reduction with sodium borohydride to give 4-amino derivative (XVIi1) (Tables IV-1 and IV-2).

Various sulfinyl compounds (XVIIa—f1) were prepared from XVIa—c1, f1—h1 by oxidation with *m*-chloroperbenzoic acid (*m*-CPBA). These compounds (XVIIa—h, j—l, n—f1) are the mixture of the diastereomer; however, we were not able to separate these isomers (Chart 9 and Table V).

**Biological Activity and Discussion** The *in vitro*  $(H^+ + K^+)$  adenosine triphosphatase (ATPase) assay and *in vivo* rat gastric acid secretory study described in Table V suggested the following structure–activity relationships. Most of the compounds inhibited both  $(H^+ + K^+)$ ATPase activity and gastric acid secretion stimulated by histamine.

As regards the substituents in the benzimidazole ring, non-substituted compounds (XVIIq, s) inhibited (H<sup>+</sup>+K<sup>+</sup>)ATPase in gastric microsomes more potently than substituted compounds (XVIIr, u), while these substituted compounds still showed a significant inhibition

of gastric acid secretion.

The inhibitory effect of a number of substituted tetrahydroquinolines including different functional groups at the 4-position on  $(H^+ + K^+)ATP$ ase in gastric microsomes and on gastric acid secretion stimulated by histamine were investigated. It was found that  $(H^+ + K^+)ATP$ ase was inhibited potently by methoxy and hydroxy derivatives (XVIIa, j) and the order of potency was  $CH_3O(XVIIa) \ge OH(XVIIj) > = N-OH(XVIIm) > NHCH_3$   $(XVIIn) \ge OOM(XVIIh) \ge OOM(XVIIh) \ge OOM(XVIIh) \ge OOM(XVIIh) \ge OOM(XVIIh)$ . These substituted

derivatives also possessed inhibiting effect on histamineinduced gastric acid secretion except for the compound containing oxime group.

The primary (XVIIf1) and secondary (XVIIn) amine substituted compounds were more potent than tertiary amine substituted compounds (XVIIr, x) in inhibiting the  $(H^+ + K^+)$ ATPase. Since the secondary and tertiary amine compounds, however, showed potent antisecretory activity, the inhibitory effects of N-substitutents were further investigated. The potency of antisecretory activities of N-substituents was in the following order.

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TABLE V. 4-Substituted 8-[(2-Benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinolines

$$R^{2} \xrightarrow{\frac{5}{6}} \begin{pmatrix} \frac{4}{3} & \frac{3}{1} & \frac{1}{1} \\ \frac{7}{1} & \frac{1}{1} & \frac{1}{1} & \frac{1}{1} \\ \frac{7}{1} & \frac{1}{1} & \frac{1}{1} & \frac{1}{1} \end{pmatrix}$$

Compd.	$\mathbb{R}^1$	$\mathbb{R}^2$	H <sup>+</sup> /K <sup>+</sup> ATPase	Histamine- stimulated rat %	Yield		mp	Formula		ılysis ( d (Fo	. ,
No.			IC <sub>50</sub> (M) <sup>a)</sup>	inhibn. (at i.v. dose, mg/kg) <sup>b)</sup>	(%)	(Recrystn. solv.)	(°C)		С	Н	N
XVIIa	1-C <sub>2</sub> H <sub>5</sub> 4-CH <sub>3</sub> O	Н	$1.1 \times 10^{-6}$	88.9 (10)	28	Yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	118—118.5 (dec.)	$C_{20}H_{23}N_3O_2S$ $\cdot 1/4H_2O$	64.23 (64.27		11.24 11.24)
XVIIb	1-C <sub>2</sub> H <sub>5</sub> 4-CH <sub>3</sub> O	5-CH <sub>3</sub>	$1.3 \times 10^{-6}$	35.7 (3) 87.7 (10)	17	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	148.5—149.5	$C_{21}H_{25}O_2N_3S$	65.77	6.57	,
XVIIc	1-C <sub>2</sub> H <sub>5</sub> 4-CH <sub>3</sub> O	5-CH <sub>3</sub> O	$3.1 \times 10^{-6}$		21		149—150	$C_{21}H_{25}O_3N_3S$	63.14 (63.16	6.31	10.52
XVIId	1-C <sub>2</sub> H <sub>5</sub> 4-CH <sub>3</sub> O	5-F 6-CH <sub>3</sub> O	$4.5 \times 10^{-6}$	31.5 (10) 101.9 (30)	42	Yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	131—132	$C_{21}H_{24}FN_3O_3S$ 1/4 $H_2O$		5.85	9.96 9.64)
XVIIe	1-C <sub>2</sub> H <sub>5</sub> 4-CH <sub>3</sub> O	5-CH <sub>3</sub> CO			42	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	149—150.5	$C_{22}H_{25}N_3O_3S$	64.21 (63.99		
XVIIf	$1-C_2H_5$ $4-CH_2 = CHCH_2O$	Н	$2.9 \times 10^{-6}$	31.2 (3) 73.7 (10)	26	Colorless needles (AcOEt-hexane)	128.5—129	$C_{22}H_{25}N_3O_2S$	66.81 (66.52		10.62 10.49)
XVIIg	$1-C_2H_5$ $4-CH_2CH=CH_2O$	5-CH <sub>3</sub> CO	$8.2 \times 10^{-6}$	10.4 (10) 85.4 (30)	36	Colorless needles (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	131—133	$C_{24}H_{27}N_3O_3S$	65.88 (65.80		9.60 9.29)
XVIIh	$1-C_2H_5$ $4 O$	Н	$1.4 \times 10^{-5}$	83.1 (10)	67	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	141—142	$C_{21}H_{23}N_3O_3S$	63.46 (63.53		
XVIIi	$1-C_2H_5$ 4,4-=0	H	$2.2 \times 10^{-5}$	57.7 (10)	32	Yellow powder (CHCl <sub>3</sub> -Et <sub>2</sub> O)	168—169	$C_{19}H_{19}N_3O_2S$	64.57 (64.20		11.89 11.50)
XVIIj	1-C <sub>2</sub> H <sub>5</sub> 4-OH	Н	$1.7 \times 10^{-6}$	13.6 (3) 81.8 (10)	39	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	117—118	$C_{19}H_{21}N_3O_2S$ $1/2H_2O$	62.61 (62.52	6.08	11.53
XVIIk	1-CH2CH = CH2 $4-CH3O$	5-CH <sub>3</sub> CO	$1.1 \times 10^{-5}$	37.8 (10) 84.4 (30)	36	White needles (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	129—130	$C_{23}H_{25}N_3O_3S$	65.23 (64.87		9.92 9.68)
XVIII	1-CH2CH = CH2 $4-CH3O$	5-F 6-CH <sub>3</sub> O	$8.7 \times 10^{-6}$	22.2 (10) 100.0 (30)	30	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	143—144	$C_{22}H_{24}FN_3O_3S$	61.52 (61.50		9.78 9.73)
XVIIm	$1-C_2H_5$ 4,4-=NOH	Н	$4.7 \times 10^{-6}$	38.6 (30)	77	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	168—169	$C_{19}H_{20}N_4O_2S$ $\cdot H_2O$			14.50 14.49)
XVIIn	1-C <sub>2</sub> H <sub>5</sub> 4-NHCH <sub>3</sub>	Н	$1.2 \times 10^{-5}$	86.8 (10)	30	White powder (pet. ether)	118—120	$C_{20}H_{24}N_4OS$			15.20 15.18)
XVIIo	$1-C_2H_5$ $4-NHCH_2CH=CH_2$	5-CH <sub>3</sub>	$3.3 \times 10^{-6}$	108.3 (10)	30	White powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	162—164	$C_{23}H_{28}N_4OS$			13.71 13.48)
XVIIp	$1-C_2H_5$ 4-NH-	5-CH <sub>3</sub>		46.9 (30°)	53	White powder (pet. ether)	135—138	$C_{26}H_{34}N_4OS$ · $1/4H_2O$	68.61 (68.80		12.31 11.95)
XVIIq	1-C <sub>2</sub> H <sub>5</sub> 4-N(CH <sub>3</sub> ) <sub>2</sub>	Н	$2.6 \times 10^{-5}$		31	Yellow needles (CH <sub>2</sub> Cl <sub>2</sub> -pet. ether)	141—142	$C_{21}H_{26}N_4OS$	(65.45	6.86	14.65 14.33)
XVIIr	1-C <sub>2</sub> H <sub>5</sub> 4-N(CH <sub>3</sub> ) <sub>2</sub>	5-F 6-CH <sub>3</sub> O	$7.0 \times 10^{-5}$		47	$(CH_2Cl_2-Et_2O)$	162—163	$C_{22}H_{27}FN_4O_2S$	61.37 (61.11		
XVIIs	1-CH2CH = CH2 $4-N(CH3)2$	Н	$5.5 \times 10^{-6}$	70.8 (30)	52	Pale yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -pet. ether)	)	$C_{22}H_{26}N_4OS$	(66.58	6.52	14.20 13.82)
XVIIt	1-CH2CH = CH2 $4-N(CH3)2$	.5-F 6-CH <sub>3</sub> O	$5.2 \times 10^{-6}$		47	$(CH_2Cl_2-Et_2O)$		$C_{23}H_{27}FN_4O_2S$	(62.47	6.18	12.54)
	1-CH2CH = CH2 $4-N(CH3)2$	5-CH <sub>3</sub> CO	>10 <sup>-4</sup>	38.1 (10) 95.2 (30)		White powder (Et <sub>2</sub> O-hexane)	114—116	$C_{24}H_{28}N_4O_2S$	(65.70	6.44	12.83 12.79)
XVIIv	$ \begin{array}{c} 1-C_2H_5\\ 4-N < CH_2CH = CH_2\\ CH_3 \end{array} $	5-CH <sub>3</sub>	$1.3 \times 10^{-5}$	NE	21	White powder (pet. ether)	109111	$C_{24}H_{30}N_4OS$			13.26 13.14)
XVIIw	$ \begin{array}{c} 1-C_2H_5\\ 4-N \end{array} CH_2CH = CH_2 $	5-CH <sub>3</sub> CO	$6.1 \times 10^{-5}$	49.0 (10)	28	Pale yellow powder (pet. ether)	115—118	$C_{24}H_{30}N_4O_2S$			12.43 12.27)
XVIIx	$CH_3$ $1-C_2H_5$ $CH_2CH=CH_2$ $4-N_2CH=CH_2$	5-F 6-CH <sub>3</sub> O	$3.8 \times 10^{-5}$	64.9 (3)	54	White powder (pet. ether)	111—115	C <sub>24</sub> H <sub>29</sub> FN <sub>4</sub> O <sub>2</sub> S			12.27 11.96)
XVIIy	$CH_3$ $1-C_2H_5$ $4-N \stackrel{\frown}{\smile} CH_3$	5-F 6-CH <sub>3</sub> O	$4.0 \times 10^{-6}$	28.8 (3) 88.8 (10)	41	White powder (Et <sub>2</sub> O-hexane)	141—143	C <sub>24</sub> H <sub>29</sub> FN <sub>4</sub> O <sub>2</sub> S			12.27 11.72)

TABLE V. (continued)

Compd. No.	$\mathbb{R}^1$	R²	H <sup>+</sup> /K <sup>+</sup> ATPase	Histamine- stimulated rat % inhibn. (at	Yield (%)	Appearance (Recrystn. solv.)	mp (°C)	Formula	Analysis (%) Calcd (Found)		
,			$IC_{50} (M)^{a}$	i.v. dose, mg/kg) <sup>b)</sup>	(70)	(Recrystin. solv.)	( )		С	Н	N
XVIIz	1-C <sub>2</sub> H <sub>5</sub> 4-N ← CH <sub>3</sub>	5-CH <sub>3</sub> CO	$5.2 \times 10^{-6}$	58.1 (10)	46	White powder (pet. ether)	125—128	$C_{25}H_{30}N_4O_2S$	66.64 (66.43		
XVIIa1	1-C <sub>2</sub> H <sub>5</sub> 4-N CH <sub>2</sub>	5-F 6-CH <sub>3</sub> O	$6.5 \times 10^{-6}$	17.4 (30°)	46	Colorless needles (AcOEt-hexane)	164—168 (dec.)	$\begin{array}{c} \mathrm{C_{27}H_{35}FN_4O_2S} \\ \cdot 1/4\mathrm{H_2O} \end{array}$	64.45 (64.41		
XVIIb1	$ \begin{array}{c} 1-C_2H_5\\ 4-N \\ C_2H_5 \end{array} $	5-F 6-CH <sub>3</sub> O	$1.0 \times 10^{-4}$	60.5 (30°)	54	Colorless prisms (Et <sub>2</sub> O-hexane)	110—114	$C_{25}H_{31}FN_4O_2S$			11.91 11.45)
XVIIc1	1-CH <sub>2</sub> CH=CH <sub>2</sub> 4-N/CH <sub>2</sub> CH=CH <sub>2</sub> CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	$4.4\times10^{-6}$	15.1 (30°)	37	Colorless needles (Et <sub>2</sub> O)	135—137.5	$C_{25}H_{31}FN_4O_2S$			11.96 11.70)
XVIId1	1-C <sub>2</sub> H <sub>5</sub> 4-N CH <sub>2</sub> CCH <sub>3</sub>	5-F 6-CH <sub>3</sub> O		43.6 (10) 101.5 (30)	22	Yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -pet. ether)	116—118	$C_{25}H_{31}FN_4O_2S$	63.81 (63.56		
XVIIel	1-C <sub>2</sub> H <sub>5</sub> 4-N (CH <sub>2</sub> ) <sub>2</sub> OH CH <sub>3</sub>	5-F 6-CH <sub>3</sub> O	$1.8 \times 10^{-5}$	26.3 (10) 90.4 (30)	72	White powder (CH <sub>2</sub> Cl <sub>2</sub> -pet. ether)	149—150	$C_{25}H_{29}FN_4O_2S$	59.98 (59.71		
XVIIf1	1-CH2CH = CH2 $4-NH2$	5-F 6-CH <sub>3</sub> O	$1.6 \times 10^{-5}$	42.2 (30)	61	Yellow powder (CH <sub>2</sub> Cl <sub>2</sub> -Et <sub>2</sub> O)	101—103	$C_{21}H_{23}FN_4O_2S$ $\cdot 1/3H_2O$	59.98 (59.75		

a) Omeprazole,  $2.0 \times 10^{-6}$  (M). b) Omeprazole, 78.6% (1 mg/kg). c) Compound administered intraperitoneally. NE = no effect.

$$R^3$$
 $R^4$ 
 $R^4$ 
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 $R^4$ 

The best compound (XVIIx) has a methyl group as  $R^2$  and an allyl group as  $R^3$ . The compounds (XVIIq, r) in which both  $R^2$  and  $R^3$  were methyl showed high potency, whereas in the compounds (XVIIa1, b1), in which both  $R^2$  and  $R^3$  were methyl and cyclohexyl, ethyl and allyl showed less antisecretory activity. The secondary amine compounds (XVIIo, n) also showed potent antisecretory activity.

On the other hand, 8-[(5-acetyl-2-benzimidazolyl)sulfinylmethyl]-1-allyl-4-(N,N-dimethylamino)-1,2,3,4-tetrahydroquinoline (XVIIu) having very weak inhibitory effect on ( $H^+ + K^+$ )ATPase decreased gastric acid secretion. Therefore, it is possible that XVIIu has a different mechanism from omeprazole and it seems worthy of further pharmacological evaluation.

Some compounds were selected for further study. The antiulcer activity against water-immersion stress-induced gastric ulcer,<sup>4)</sup> aspirin-induced gastric ulcer and gastric necrosis induced by 0.6 N hydrochloric acid was investigated in the rat (Table VI). Most compounds possessing the potent antisecretory activity inhibited the production of gastric mucosal lesions induced by aspirin. Only the 4-amino substituted analogues showed inhibitory effect on stress-induced gastric ulcer. Many compounds showed

TABLE VI. Antiulcer and Cytoprotective Activities 4-Substituted 8-[(2-Benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinolines

Compd. No.	Aspirin-induced ulcer ED <sub>50</sub> (mg/kg) p.o.	Stress ulcer ED <sub>50</sub> (mg/kg) p.o.	Cytoprotective activity ED <sub>50</sub> (mg/kg) p.o.		
XVIIa	7.7	NE	5.4		
XVIIb	16.0	NE	ND		
XVIIc	13.6	>30	8.3		
XVIIe	7.1	15.1	9.2		
XVIIf	16.8	NE	ND		
XVIIj	> 30	NE	12.0		
XVIIh	> 30	NE	16.5		
XVIIr	3.5	9.3	20.1		
XVIIo	15.5	>30	17.0		
XVIIu	11.9	17.6	14.6		
XVIIx	5.5	7.6	13.3		
XVIIy	7.2	9.3	17.5		
Omeprazole	8.3	9.8	30.2		

NE = no effect. ND = not determined.

cytoprotective activity. It appears from these results that the presence of an amino group necessary for activity and the basicity are probably important contributing factors.

Among these compounds, 4-(N-allyl-N-methylamino)-1-ethyl-8-[(5-fluoro-6-methoxy-2-benzimidazolyl)sulfinyl-methyl]-1,2,3,4-tetrahydroquinoline (XVIIx, OPC-22575) showed the most potent activity. It was confirmed that the amino substitution at the 4-position of the 1,2,3,4-tetrahydroquinoline ring leads to interesting products.

## **Experimental**

Melting points were determined with a Yamato MP-21 apparatus and

are uncorrected. Infrared (IR) spectra were recorded on a JASCO IRA-2 spectrometer. Nuclear magnetic resonance (NMR) spectra were recorded in CDCl<sub>3</sub> on a Bruker AC-200 spectrometer with tetramethylsilane as an internal standard. Mass spectra (MS) were obtained on a Varian MAT-312 instrument.

**4H-1,2-Dihydro-2-oxo-3,1-benzoxazine (II)** To a solution of 2-aminobenzyl alcohol (5.0 g, 41 mmol) and Et<sub>3</sub>N (32 ml) in THF (250 ml) was added trichloromethyl chloroformate (2.7 ml, 22 mmol) at 0—10 °C. The mixture was stirred at room temperature for 2 h. After removal of the insoluble meterial by filtration, the filtrate was concentrated *in vacuo*. The residue was recrystallized from Et<sub>2</sub>O-hexane to give II (4.5 g, 74%) as white needles, mp 120—121 °C. NMR  $\delta$ : 5.33 (2H, s), 6.86 (1H, d, J=6.5 Hz), 7.00—7.16 (2H, m), 7.20—7.30 (1H, m), 8.59 (1H, br s). IR  $\nu$  (KBr): 3160, 3104, 1716, 1606 cm<sup>-1</sup>. *Anal.* Calcd for C<sub>8</sub>H<sub>7</sub>NO<sub>2</sub>: C, 64.42; H, 4.73; N, 9.39. Found: C, 64.65; H, 4.79; N, 9.26.

Methyl 3-(4*H*-1,2-Dihydro-2-oxo-3,1-benzoxazin-1-yl)propionate (III) A mixture of II (792 g, 4.06 mol), methyl acrylate (440 ml, 4.87 mol) and  $K_2CO_3$  (840 g) in *N*,*N*-dimethylformamide (DMF) (61) was stirred at 50 °C for 5 h. The reaction mixture was poured into water and extracted with AcOEt-benzene (3:1). The extract was washed with water, dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was recrystallized from AcOEt-hexane to give III (870 g, 90%) as colorless needles, mp 66—67 °C. NMR δ: 2.81 (2H, t, J=6.2 Hz), 3.70 (3H, s), 4.20 (2H, t, J=6.2 Hz), 5.19 (2H, s), 6.99 (1H, d, J=6.5 Hz), 7.05—7.20 (2H, m), 7.30—7.40 (1H, m). IR ν (KBr): 2965, 1730, 1700, 1180 cm<sup>-1</sup>. *Anal.* Calcd for  $C_{12}H_{13}NO_4$ : C, 61.27; H, 5.57; N, 5.95. Found: C, 61.35; H, 5.63; N, 5.88.

3-(4*H*-1,2-Dihydro-2-oxo-3,1-benzoxazin-1-yl)propionic Acid (IV) A mixture of III (870 g, 37 mol) and saturated Na<sub>2</sub>CO<sub>3</sub> solution (1.51) in MeOH (5.51) was refluxed for 8 h. After removal of the solvent, the residue was dissolved in water, neutralized with citric acid and extracted with Et<sub>2</sub>O. Then, the aqueous layer was acidified with citric acid and extracted with AcOEt. The combined extracts were dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was recrystallized from AcOEt—hexane to give IV (770 g, 92%) as colorless needles, mp 147—148.5 °C. NMR  $\delta$ : 2.87 (2H, t, J=7.5 Hz), 4.22 (2H, t, J=7.5 Hz), 5.20 (2H, s), 6.90—7.50 (4H, m). IR  $\nu$  (KBr): 2970, 1720, 1660, 1185 cm<sup>-1</sup>. *Anal*. Calcd for C<sub>11</sub>H<sub>11</sub>NO<sub>4</sub>: C, 59.73; H, 5.01; N, 6.33. Found: C, 59.69; H, 5.03; N, 6.23.

1H,3H,5H,6H,7H-3,7-Dioxopyrido]3,2,1-ij||3,1|benzoxazine (V) IV (200 g, 0.97 mol) was added to PPA prepared from phosphorus pentoxide (1.5 kg) and phosphoric acid (1.5 l) and the mixture was heated at 120 °C for 1 h. The reaction mixture was poured into ice-water and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was washed with saturated Na<sub>2</sub>CO<sub>3</sub> solution, dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was recrystallized from AcOEt-hexane to give V (110 g, 60%) as colorless needles, mp 130—130.5 °C. NMR δ: 2.85 (2H, t, J=6.8 Hz), 4.30 (2H, t, J=7.8 Hz), 5.32 (2H, s), 7.16 (1H, t, J=7.5 Hz), 7.25—7.45 (1H, m), 7.92 (1H, d, J=7.8 Hz). IR  $\nu$  (KBr): 1715, 1685, 1605, 1490 cm<sup>-1</sup>. *Anal.* Calcd for C<sub>11</sub>H<sub>9</sub>NO<sub>3</sub>: C, 65.02; H, 4.46; N, 6.89. Found: C, 65.11; H, 4.29; N, 6.79.

1*H*,3*H*,5*H*,6*H*,7*H*-7-Hydroxy-3-oxopyrido[3,2,1-*ij*][3,1]benzoxazine (VI) To a solution of V (1.0 g, 5 mmol) in MeOH (20 ml) was added NaBH<sub>4</sub> (186 mg, 5 mmol) in small portions at room temperature and the mixture was stirred at the same temperature for 30 min. After removal of the solvent, the residue was poured into water and extracted with CHCl<sub>3</sub>. The extract was dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was recrystallized from AcOEt to give VI (0.9 g, 79%) as colorless needles, mp 133—135 °C. NMR δ: 2.00—2.20 (2H, m), 2.70—2.80 (1H, br s), 3.70—3.90 (1H, m), 3.90—4.10 (1H, m), 4.70—4.90 (1H, m), 5.17 (2H, s), 7.00—7.10 (2H, m), 7.30—7.40 (1H, m). IR ν (KBr): 3450, 1690, 1600, 1480, 1400, 1290, 1110, 1090, 810 cm<sup>-1</sup>. *Anal*. Calcd for C<sub>11</sub>H<sub>11</sub>NO<sub>3</sub>: C, 64.38; H, 5.40; N, 6.83. Found: C, 64.51; H, 5.43; N, 6.68.

1H,3H,5H,6H,7H-7-Alkoxy-3-oxopyrido[3,2,1-ij][3,1]benzoxazines (VIIa, b). A Typical Procedure To a solution of VI (4.3 g, 21 mmol) in DMF (50 ml) was added 60% NaH (0.94 g, 24 mmol) at room temperature. The mixture was stirred at the same temperature for 30 min, then CH<sub>3</sub>I (3.35 g, 24 mmol) was added dropwise with stirring at room temperature. After being stirred for 1 h, the reaction mixture was poured into water and extracted with benzene—AcOEt (1:3). The extract was washed with water, dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, CH<sub>2</sub>Cl<sub>2</sub>: MeOH = 300:1) and gave VIIa (4.3 g, 94%) as a yellow oil. IR  $\nu$  (neat): 2930, 1720, 1610, 1490, 1410, 1270, 1100, 770 cm<sup>-1</sup>. MS m/z (%): 219 (M<sup>+</sup>, 24), 144 (40), 143 (100), 142 (25). NMR data are given in Table I.

Compound VIIb was obtained by a similar procedure to that described for VIIa; the yields and NMR data are listed in Table I.

1H,3H,5H,6H,7H-7,7-Ethylenedioxy-3-oxopyrido[3,2,1-ij][3,1]benz-

**oxazine (X)** A mixture of V (2.0 g, 9.8 mmol), ethylene glycol (2.2 ml) and *p*-TsOH (0.1 g) in toluene (60 ml) was refluxed on Dean-Stark for 3 h. The reaction mixture was washed with aqueous NaHCO<sub>3</sub> solution, dried over MgSO<sub>4</sub> and concentrated *in vacuo* to give an oily X (1.9 g, 72%). IR  $\nu$  (neat): 2900, 1720, 1610, 1490, 1460, 1410, 1220, 1130, 1040, 750 cm<sup>-1</sup>. MS m/z (%): 247 (M<sup>+</sup>, 15), 203 (28), 159 (33), 131 (53), 130 (100), 77 (26), 51 (33). NMR data are given in Table I.

8-Hydroxymethyl-1,2,3,4-tetrahydroquinolines (VIIIa, b, XI, XIV). A Typical Procedure A mixture of X (2.2 g, 8.9 mmol) and NaOH (0.43 g) in MeOH (10 ml) was refluxed for 5 h. The reaction mixture was poured into water and extracted with  $\mathrm{CH_2Cl_2}$ . The extract was washed with water, dried over  $\mathrm{MgSO_4}$  and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt=4:1) to give XI (1.2 g, 61%). Recrystallization from AcOEt–hexane gave XI as pale yellow prisms, mp 125—128 °C. IR v (KBr): 3450, 3350, 1610, 1505, 1120, 1020 cm<sup>-1</sup>. Anal. Calcd for  $\mathrm{C_{12}H_{15}NO_3}$ : C, 65.14; H, 6.83; N, 6.33. Found: C, 65.08; H, 6.82; N, 6.35. NMR data are given in Table II.

Compounds VIIIa, b and XIV were obtained by a similar procedure to that described for XI; the yields and NMR data are listed in Table II.

1-Substituted 8-Hydroxymethyl-1,2,3,4-tetrahydroquinolines (IXa-e, p, q). A Typical Procedure To a stirred solution of XI (11.0 g, 50 mmol), in THF (120 ml) was added NaH (60%, 2.0 g, 50 mmol) at room temperature. The reaction mixture was stirred at  $60 \,^{\circ}$ C for 1 h and cooled at  $-60 \,^{\circ}$ C. Then, n-butyllithium (16% solution in hexane, 21.2 ml) was added dropwise with stirring. After being stirred at the same temperature for 30 min, allyl bromide (6.0 g, 50 mmol) was added dropwise and the reaction mixture was stirred at -65---40°C for 4h. After removal of the solvent, the residue was poured into water and extracted with CHCl<sub>2</sub>. The extract was dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt = 3:1) to give IXe (6.6 g, 51%). Recrystallization from AcOEt-hexane gave IXe as pale yellow prisms, mp 76—78 °C. IR ν (KBr): 3320, 1440, 1350, 1145, 1105, 1080, 1020, 950, 930, 790 cm<sup>-1</sup>. Anal. Calcd for C<sub>15</sub>H<sub>19</sub>NO<sub>3</sub>: C, 68.94; H, 7.33; N, 5.36. Found: C, 69.07; H, 7.33; N, 5.28. NMR data are given in Table III.

Compounds IXa—d, p and IXq were obtained by a similar procedure to that described for IXa, the yields and NMR data are shown in Table III.

1-Substituted 8-Hydroxymethyl-4-oxo-1,2,3,4-tetrahydroquinolines (IXf, g). A Typical Procedure A mixture of IXe (6.6 g, 25 mmol) and 10% HCl (20 ml) in THF (100 ml) was stirred at room temperature for 2 h. The reaction mixture was adjusted to pH 9 (aqueous  $\rm Na_2CO_3$  solution) and extracted with  $\rm CH_2Cl_2$ . The extract was dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt=4:1) to give IXg (6.0 g, 92%) as a yellow oil. IR v (neat): 3420, 1680, 1595, 1480, 1440, 1370, 1280, 1100 cm<sup>-1</sup>. MS m/z (%): 218 (79), 217 (M<sup>+</sup>, 100), 200 (30), 199 (50), 198 (46), 176 (35), 148 (64), 130 (69), 77 (37), 41 (70). NMR data are given in Table III.

Compound IXg was obtained by a similar procedure to that described for IXf; the yields and NMR data are listed in Table III.

*N*-Substituted 4-Amino-8-hydroxymethyl-1,2,3,4-tetrahydroquinolines (IXh—k). A Typical Procedure A mixture of IXg (34 g, 0.16 mol), methylamine (40% MeOH solution, 243 ml, 1.6 mol), molecular sieves 3A (40 g) and *p*-TsOH (0.5 g) in MeOH (100 ml) was stirred at 65 °C for 5 h. The mixture was cooled and the molecular sieves were removed by filtration. To the stirred and ice-cooled filtrate was added NaBH<sub>4</sub> (8.9 g, 0.24 mol) in small portions. The mixture was stirred at room temperature for 1 h, then water was added and the mixture was concentrated *in vacuo*. The residue was extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt = 2:1) to give IXk (32 g, 89%) as a yellow oil. IR  $\nu$  (neat): 3320, 2950, 2870, 1450, 1420, 1060, 760 cm<sup>-1</sup>. MS m/z (%): 232 (M<sup>+</sup>, 3), 201 (100), 200 (47), 172 (41), 160 (62), 158 (44), 142 (62), 130 (42), 86 (81), 84 (76), 47 (42). NMR data are given in Table III.

Compounds IXh—j were obtained by a similar procedure to that described for IXk; the yields, elemental analysis and NMR data are listed in Table III.

*N,N*-Disubstituted 4-Amino-8-hydroxymethyl-1,2,3,4-tetrahydroquinolines (IXI—o). A Typical Procedure A mixture of acetic anhydride (42 ml) and formic acid (21 ml) was stirred at 60 °C for 2 h. Then, IXk (32 g, 0.14 mol) was added dropwise with stirring at 0—10 °C and stirred at room temperature for 16 h. The reaction mixture was made alkaline with 30% NaOH and extracted with  $CH_2Cl_2$ . The extract was dried over MgSO<sub>4</sub> and concentrated *in vacuo*. This residue was added to a suspension of LiAlH<sub>4</sub> (11.3 g, 0.3 mol) in THF (800 ml) dropwise at 0—10 °C. The mixture was then refluxed for 2 h. Saturated Rochelle solution was carefully added

to destroy the excess LiAlH<sub>4</sub>. After removal of the insoluble material by filtration, the filtrate was dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt: MeOH = 2:5:0.5) to give IXo (43.6 g, 57%) as a yellow oil. IR  $\nu$  (neat): 3400, 2940, 2870, 2820, 1450, 1360, 1080, 1040, 930, 780, 760 cm<sup>-1</sup>. MS m/z (%): 246 (M<sup>+</sup>, 2.4), 201 (100), 200 (43), 160 (42), 158 (33), 142 (32), 130 (29), 86 (39), 84 (64). NMR data are given in Table III.

Compounds IXI—n were obtained by a similar procedure to that described for IXo; the yields and NMR data are listed in Table III.

**7-(N-Allylamino)-1H,3H,5H,6H,7H-3-oxopyrido[3,2,1-ij][3,1]benzoxazine (XII)** A mixture of V (6.1 g, 30 mmol), allylamine (50 ml), moleular sieves 3A (12 g) and p-TsOH (0.1 g) in MeOH (100 ml) was refluxed overnight. The mixture was cooled and the molecular sieves removed by filtration. To the stirred and ice-cooled filtrate was added NaBH<sub>4</sub> (1.7 g, 45 mmol) in small portions. The reaction mixture was stirred at room temperature for 1 h, then water (50 ml) was added, and the mixture was concentrated in vacuo. The residue was extracted with  $CH_2Cl_2$ , dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt = 3:1) to give XII (5.9 g, 80%) as a yellow oil. IR v (neat): 1720, 1610, 1490, 1460, 1410, 1300, 1290, 770 cm<sup>-1</sup>. MS m/z (%): 244 (M<sup>+</sup>, 4), 227 (5), 187 (7), 144 (37), 143 (100), 142 (22), 117 (10), 115 (12), 58 (13), 41 (18). NMR data are given in Table I.

7-(N-Allyl-N-methylamino)-1H,3H,5H,6H,7H-3-oxopyrido[3,2,1-ij[[3,1]-benzoxazine (XIII) A solution of XII (5.9 g, 24 mmol) in formic acid (7 ml) and formalin (7 ml) was stirred at 90 °C for 2 h. After removal of the formic acid and formalin, the residue was poured into water, made alkaline with Na<sub>2</sub>CO<sub>3</sub> and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, CH<sub>2</sub>Cl<sub>2</sub>: MeOH = 100:1) to give XIII (5.9 g, 95%) as an orange oil. IR v (neat): 1720, 1610, 1480, 1460, 1400, 1300, 1290, 1090, 770 cm<sup>-1</sup>. MS m/z (%): 258 (M+, 20), 227 (6), 188 (18), 144 (82), 143 (100), 142 (26), 117 (10), 72 (36), 70 (16). NMR data are given in Table I.

4-Substituted 8-[(2-Benzimidazolyl)thiomethyl]-1,2,3,4-tetrahydroquinolines (XVIa—i, k, l, o—e1, i1). A Typical Procedure Thionyl chloride (17 ml, 0.23 mol) was added dropwise to a stirred and ice-cooled solution of IXo (19.2 g, 78 mmol) in  $CH_2Cl_2$  (460 ml). The reaction mixture was stirred at 0—10 °C for 1 h and concentrated *in vacuo*. The residue was dissolved in DMF (380 ml), then to this solution was added 5-acetyl-2-mercaptobenzimidazole (15.0 g, 78 mmol) and  $K_2CO_3$  (21.6 g, 0.16 mol). The mixture was stirred at room temperature for 24 h, poured into water and extracted with toluene—AcOEt (1:4). The extract was washed with water and dried over MgSO<sub>4</sub>. After removal of the solvent, the residue was purified by column chromatography (silica gel; eluent,  $CH_2Cl_2$ : MeOH = 100:1) to give XVIu (28.0 g, 85%). Recrystallization from Et<sub>2</sub>O—hexane gave XVIu as yellow powder, mp 143—145 °C. IR  $\nu$  (KBr): 2930, 2820, 1670, 1450, 1420, 1360, 1280 cm<sup>-1</sup>. The elemental analysis and NMR data are given in Tables IV-1 and IV-2.

Compounds XVIa—i, k, l, o—t, v—el and XVIil were obtained by a similar procedure to that described for XVIu; the yields, mp, elemental analyses and NMR data are listed in Tables IV-1 and IV-2.

**8-[(2-Benzimidazolyl)thiomethyl]-1-ethyl-4-hydroxy-1,2,3,4-tetrahydroquinoline (XVIj)** Sodium borohydride (0.1 g, 2.6 mmol) was added to a stirred and ice-cooled solution of XVIi (0.84 g, 2.5 mmol) in MeOH (40 ml). The mixture was stirred at room temperature for 1 h, then water was added, and the mixture was concentrated *in vacuo*. The residue was extracted with  $CH_2Cl_2$ , dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt=1:1) to give XVIj (0.74 g, 88%) as a white amorphous powder. IR  $\nu$  (KBr): 2970, 1450, 1410, 1350, 1270, 770, 750 cm<sup>-1</sup>. MS m/z (%): 339 (M<sup>+</sup>, 1.5), 191 (36), 190 (63), 174 (40), 173 (67), 150 (100), 146 (42), 144 (43), 130 (32). NMR data are given in Table IV-1.

8-[(2-Benzimidazolyl)thiomethyl]-1-ethyl-4-oxo-1,2,3,4-tetrahydroquinoline Oxime (XVIm) A mixture of XVIi (1.5 g, 4.4 mmol), hydroxylamine hydrochloride (0.49 g, 6.6 mmol) and sodium acetate (1.7 g) in EtOH (20 ml) and  $\rm H_2O$  (3 ml) was refluxed for 3 h. After removal of the solvent, the residue was poured into water. The precipitated crystals were collected by filtration. Recrystallization from MeOH gave XVIm (0.94 g, 61%) as colorless columns, mp 201—202 °C. IR  $\nu$  (KBr): 3050, 2970, 2820, 1450, 1410, 1270, 1000, 770, 750 cm<sup>-1</sup>. The elemental analysis and NMR data are listed in Tables IV-1 and IV-2.

8-[(2-Benzimidazolyl)thiomethyl]-1-ethyl-4-(N-methylamino)-1,2,3,4-tetrahydroquinolines (XVIn) Compound XVIn was obtained by the same procedure as described for IXk. Yield 92%, as yellow powder from AcOEt,

mp 144—145 °C. IR  $\nu$  (KBr): 2970, 2850, 1440, 1400, 1350, 1270, 750 cm<sup>-1</sup>. The elemental analysis and NMR data are given in Tables IV-1 and IV-2.

4-(N-Cyclopropylmethyl-N-methylamino)-1-ethyl-8-[(5-fluoro-6-methoxy-2-benzimidazolyl)thiomethyl]-1,2,3,4-tetrahydroquinoline (XVIf1) Cyclopropanecarbonyl chloride (0.35 g, 3.3 mmol) was added to a stirred solution of cyclopropanecarboxylic acid (0.29 g, 3.4 mmol) and Et<sub>3</sub>N (0.33 g, 3.3 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 ml) at 0—10 °C. The mixture was stirred at the same temperature for 30 min, then XVIe1 (0.7 g, 1.8 mmol) was added, and stirring continued at room temperature for 2h. The reaction mixture was washed with saturated NaHCO3 solution, dried over MgSO4 and concentrated in vacuo. To the stirred and ice-cooled solution of residue (0.9 g) in THF (20 ml) was added LiAlH<sub>4</sub> (0.2 g, 5.3 mmol) in small portions. The mixture was stirred at room temperature for 1h. Saturated Rochelle solution was carefully added to destroy the excess LiAlH<sub>4</sub>. After removal of the insoluble material by filtration, the filtrate was dried over MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified by column chromatography (silica gel; eluent, CH<sub>2</sub>Cl<sub>2</sub>:hexane=99:1) to give XVIf1 (0.6 g, 48%) as a yellow oil. IR  $\nu$  (neat): 3080, 2970, 2940, 2870,  $1490, 1470, 1450, 1430, 1350, 1200, 1160, 1130, 1030, 890, 760 \, \text{cm}^{-1}$ . MS m/z (%): 455 (M<sup>+</sup> +1, 31), 370 (17), 307 (9), 257 (52), 172 (100), 136 (71), 107 (23), 89 (22), 79 (38). NMR data are given in Table IV-1.

**1-Ethyl-8-[(5-fluoro-6-methoxy-2-benzimidazolyl)thiomethyl]-4-[N-(2-hydroxyethyl)-N-methylamino]-1,2,3,4-tetrahydroquinoline (XVIg1)** Glyoxal (5 ml) was added to a solution of XVIe1 (1.3 g, 3 mmol) in MeOH (20 ml). The mixture was stirred at room temperature for 3 h. After removal of the solvent, the residue was dissolved in MeOH (20 ml) and NaBH<sub>4</sub> (0.7 g) was added to this solution in small portions. The reaction mixture was stirred at room temperature for 1 h, then water was added and the mixture was concentrated *in vacuo*. The residue was extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over MgSO<sub>4</sub> and concentrated *in vacuo*. The residue was purified by column chromatography (silica gel; eluent, CH<sub>2</sub>Cl<sub>2</sub>: MeOH = 99: 1) to give XVIg1 (0.6 g, 45%) as a white amorphous powder. IR  $\nu$  (KBr): 3400, 3070, 2950, 1490, 1450, 1430, 1350, 1200, 1160, 1130, 890, 760 cm<sup>-1</sup>. MS m/z (%): 445 (M<sup>+</sup> + 1, 1.1), 370 (1.5), 307 (7), 289 (7), 247 (7), 172 (48), 154 (100), 136 (86), 107 (32), 89 (38), 77 (38). NMR data are given in Table IV-1.

**1-Allyl-4-amino-8-[(5-fluoro-6-methoxy-2-benzimidazolyl)thiomethyl] 1,2,3,4-tetrahydroquinoline (XVIi)** Compound XVIi was obtained by the same procedure as described for IXk. Yield 38%, a pale yellow amorphous powder. IR  $\nu$  (KBr): 3070, 2940, 1490, 1450, 1430, 1350, 1200, 1170, 1130, 890, 760 cm<sup>-1</sup>. MS m/z (%): 398 (M<sup>+</sup>, 3.4), 365 (10), 338 (23), 201 (53), 198 (58), 185 (31), 184 (74), 183 (100), 182 (67), 172 (40), 168 (30), 158 (38), 155 (33). NMR data are given in Table IV-1.

4-Substituted 8-[(2-Benzimidazolyl)sulfinylmethyl]-1,2,3,4-tetrahydroquinolines (XVIIa-f1). A Typical Procedure A solution of 80% m-CPBA (8.6 g, 40 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (60 ml) was added dropwise to a stirred and dry ice acetone-cooled solution of XVIx (17.5 g, 40 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (170 ml) and the reaction mixture was stirred at the same temperature for 1 h. The CH<sub>2</sub>Cl<sub>2</sub> solution was washed with Na<sub>2</sub>CO<sub>3</sub> aqueous solution and dried over MgSO<sub>4</sub>. After removal of the solvent, the residue was purified by column chromatography (silica gel; eluent, hexane: AcOEt= 2.5:1) and recrystallized from petroleum ether to give XVIIx (15.0 g, 83%) as a white powder, mp 111—115 °C. NMR δ: 1.10—1.40 (3H, m), 1.40—1.75 (1H, m), 1.75—2.00 (1H, m), 2.14 (3H, s), 2.50—3.20 (6H, m), 3.68 (1H, t, J=7.5 Hz), 3.91 (3H, s), 4.30—4.90 (2H, m), 5.00—5.30 (2H, m), 5.80—6.00 (1H, m), 6.75—7.70 (5H, m), 12.47 (1H, br s). IR  $\nu$  (KBr): 2945, 1450, 1200, 1045 cm<sup>-1</sup>. The elemental analysis data are given in Table V.

Compounds XVIIa—w and XVIIy—f1 were obtained by a similar procedure to that described for XVIIx; the yields, mp, and elemental analyses data are listed in Table V.

**Biological Methods** The  $(H^+ + K^+)$ ATPase-inhibitory, gastric antisecretory, antiulcer and cytoprotective activities were tested by the reported method <sup>1)</sup>

Water-Immersion Stress Ulcer<sup>4</sup>) Male Wistar rats weighing 160—200 g were fasted but allowed free access to water for 24 h before the experiment. The test compounds and the vehicle were given orally 30 min before the water immersion suspended in 0.5% carboxymethyl cellulose (CMC). Each rat was placed in a stress cage and immersed to the level of the xiphoid process for 7h in a water bath maintained at 23 °C. The animals were sacrificed and the stomachs were removed. The stomach was fixed by instilling it with 10 ml of 1% formalin solution and then immersing it in the formalin solution for 30 min, after which it was incised along the greater curvature. The length of each lesion in the glandular portion was measured under a dissecting microscope. The sum of the length (mm) of

all lesions for each rat was used as an ulcer index. Percent inhibition was calculated as follows: [(ulcer index of control—ulcer index of test compound/ulcer index of control]  $\times\,100$ . The doses inhibiting water-immersion stress ulcer by 50% (ED $_{50}$ ) were calculated by linear regression analysis.

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