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Solubilization of Lipid-Soluble Vitamins by Complexation with Glucosyl-β-cyclodextrin¹⁾

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Inclusion complex formation of eight kinds of lipid-soluble vitamins with 6-O-α-D-glucopyranosyl-β-cyclodextrin $(G-\beta-CD)$ in aqueous solution and in solid phase were assessed by the solubility method and thermal analysis, All lipid-soluble vitamins were highly solubilized in water by complexation with G-\beta-CD. From analysis of the phase solubility diagrams, the stoichiometric ratio of the main complex in water was estimated to be 1:2 for vitamin (V) A alcohol/G- β -CD, 1:2 for V D₂/G- β -CD, 1:1 for V D₃/G- β -CD, 1:3 for V E/G- β -CD, 1:4 for V E nicotinate/G- β -CD, 1:3 for V K₁/G-β-CD, 1:3 for V K₂/G-β-CD, and 1:1 for V K₃/G-β-CD. The stabilities of lipid-soluble vitamins in water containing G-β-CD were examined. A V E nicotinate –G-β-CD complex solution was stable even under irradiation with light.

Keywords glucosyl- β -cyclodextrin; lipid-soluble vitamin; inclusion complex; solubility method; solubilization; differential scanning calorimetry; stability

Branched cyclodextrins (CDs) which have one or more branches of an α -D-glucopyranosyl unit or a $(1\rightarrow 4)$ - α -Dglucan at carbon 6 of glucose residues in CD have many advantages over their parent CDs: high solubility both in water and in organic solvents and decreasing hemolytic activities with the elongation of the side chain. $^{2-5}$ The complexation abilities of branched and parent CDs appear to be almost the same, however, the enhancement of solubility of water-insoluble compounds by complexation with branched CDs was much more remarkable than that with parent CDs, particularly in the β -CD series.²⁻⁵⁾

This paper deals with the solubilization of lipid-soluble vitamins in water by complexation with 6-O-α-D-glucopyranosyl- β -CD (G- β -CD), and also with stabilities of these complexes in water.

Experimental

Materials G-β-CD was isolated and purified by high-performance liquid chromatography (HPLC) according to the reported method. 5) β -CD was used after recrystallization from water. All lipid-soluble vitamins used were obtained from commercial sources. Deionized and double distilled water was used throughout this experiment. Reagent-grade organic solvents used for HPLC were freshly distilled and filtered through a 0.45-µm membrane filter

General Method HPLC analyses of lipid-soluble vitamins were performed using a Familic-300S HPLC pump and a model VL-614 injector (all from JASCO, Tokyo, Japan). The columns used were a Finepak SIL C_{18} (250 × 4.6 mm i.d.) (JASCO) and a YMC-Pack A-802 C_4 (150 × 4.6 mm i.d.) (Yamamura Chemical, Kyoto, Japan).

Solubility Studies Estimation of complex-forming ability of G-β-CD by the solubility method⁶⁾ was conducted according to the procedure described previously.2)

Preparation of Solid Complexes In vitamin (V) D₂ and V D₃-G-β-CD systems which showed B_s type phase solubility diagrams, 6) the solid complexes were prepared by mixing appropriate amounts of G-β-CD and V D_2 or V D_3 in water. For example, $80\,\text{mg}$ (0.2 mmol) of V D_2 and 260 mg (0.2 mmol) of G- β -CD were added to 1 ml of water, and the mixture was shaken at 30 °C for 24 h in the dark. The complexes, precipitated as microcrystalline powders, were separated from the solution by filtration and dried in vacuo at 25 °C for 24 h. The solid complexes were dissolved in 65% ethanol and the contents of guest and host compounds were determined by HPLC. Both V D₂/G- β -CD and V D₃/G- β -CD molar ratios in the solid complexes obtained were 1/2.

Thermal Analysis The analysis was done using a Thermo Flex DSC-8230B (Rigaku, Tokyo, Japan) at a scanning speed of 5 °C/min and scanning temperature range of 30-200 °C.

Photodegradation Study A sample solution in water was prepared by mixing equimolecular amounts of G- β -CD and lipid-soluble vitamin in water. The mixture was shaken at 30 °C for 24 h in the dark, and filtered through 0.2-µm membrane filter to remove excess lipid-soluble vitamin. The concentrations of lipid-soluble vitamins in 200 mm G-β-CD solution were 14.5 mm for V A alcohol, 2.1 mm for V E, 0.5 mm for V E nicotinate, 0.4 mm for V K₁, and 1.3 mm for V K₂. V D₂ and V D₃ concentrations in 75 mm G-β-CD solution were 5.4 and 16.2 mm. In these systems, the solution was hard to make in a higher concentration of G-β-CD, since the solid complexes tended to precipitate. Then, the sample solution was irradiated with a white lamp (Tokyo Shibaura Denki Co., 20 W, 1000 lux) at a distance of 15 cm. At regular time intervals, intact lipid-soluble vitamin in G- β -CD solution was determined by HPLC.

Results and Discussion

Inclusion Complexes of Lipid-Soluble Vitamins with **G-β-CD** Figure 1 shows phase solubility diagrams which were obtained for lipid-soluble vitamins and G-β-CD systems in water at 30 °C. The water-solubilities of all lipid-soluble vitamins increased in the presence of G- β -CD. Parent β -CD was found to have little solubilizing effect on lipid-soluble vitamins except V K₃, because it formed insoluble complexes at lower β -CD concentrations. The V K_3 -G- β -CD system indicated a typical A_L type solubility curve with linearly increasing solubility, meaning that a 1:1 complex may exist in the solution. The apparent stability constants of both V K₃-G-β-CD and V K₃-β-CD systems calculated from the initial rising portion of the solubility curves according to the method of Higuchi and Connors⁶⁾ were 190 m⁻¹. On the other hand, solubility curves of the V A alcohol-, V E-, V E nicotinate-, V K₁-, and V K_2 -G- β -CD systems were A_p type, suggesting a high order complexation. Namely, complexes were not formed at lower G- β -CD concentrations, but great solubilization enhancement due to higher-order complex formations at higher G- β -CD concentrations was observed. The stability constants of higher-order complexes of these lipid-soluble vitamins in water were estimated using the nonlinear optimization least squares technique⁷⁾ modified by Uekama et al.8) (Table I). It is thought that these differences in molar composition sensitively reflect difference of chemical structures and physical properties of guest compounds. Both the V D_2 - and V D_3 -G- β -CD systems showed B_S type solubility curves in the very high concentration range of G- β -CD, a solid complex precipitating. The ratios of guest and host compounds in these microcrystalline complexes were found to be 1:2 for both V D₂/G-β-CD and V $D_3/G-\beta-CD$.

Figure 2 shows the differential scanning calorimetry thermogram of the solid complex in comparison with those

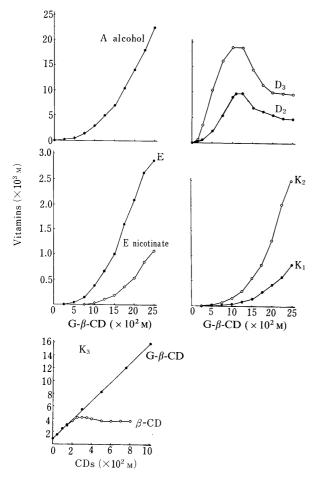


Fig. 1. Phase Solubility Diagrams of Lipid-Soluble Vitamin–G- β -CD Systems in Water at 30 °C

In cases other than that of V K_3 , solubilities of lipid-soluble vitamins in water containing β -CD did not rise as vitamin- β -CD complexes precipitated.

Table I. Stability Constants of Lipid-Soluble Vitamin–G- β -CD Complexes in Water at 30 °C

Vitamin	$K_{1:1} \choose M^{-1}$	$K_{1:2} (M^{-1})$	$K_{1:3} (M^{-1})$	$K_{1:4} \choose {M^{-1}}$	$K_{1:5} (M^{-1})$
A alcohol	20	8700	0		
D_2	1100	6500	0		
D_3	11400	2200	0		
E	0	40	6300	0	
E nicotinate	0	0	0	2800	0
\mathbf{K}_{1}	10	50	2200	0	
\mathbf{K}_{2}	20	20	5000	0	

of the physical mixture and V D_2 . V D_2 alone and the physical mixture both showed an endothermic peak at around 115 °C owing to melting; however, this peak disappeared with formation of the complex. These results imply that V D_2 interacts with G- β -CD in the solid state to form an inclusion complex.

Figure 3 shows the stabilities of lipid-soluble vitamins in G- β -CD solutions. Vitamins, D_2 , D_3 , E nicotinate, K_1 , and K_2 in aqueous solution containing G- β -CD were very stable in the dark. A V E nicotinate-G- β -CD complex solution was stable even under irradiation with light.

Recently, Uekama et al.⁸⁾ investigated the inclusion complexes of V E esters with heptakis $(2,6-di-O-methyl)-\beta$ -

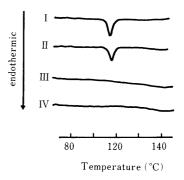


Fig. 2. DSC Thermograms of V D₂-G-β-CD Systems
I, V D₂ alone; II, physical mixture of V D₂ and G-β-CD in 1:2 molar ratio; III,

1:2 complex of V D_2 with G- β -CD; IV, G- β -CD alone.

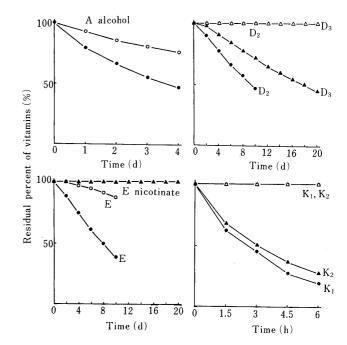


Fig. 3. Stabilities of Lipid-Soluble Vitamins in Water Containing $G-\beta$ -CD

Open symbols indicate in the dark. Closed symbols indicate under light irradiation.

CD (DM- β -CD). Horiuchi *et al.*⁹⁾ also reported on V K group–DM- β -CD complexes. Both papers described that the enhanced bioavailability by complexation seemed mainly ascribable to the increase in dissolution of V E esters and V K groups due to higher-order complex formation with DM- β -CD.

It is expected that lipid-soluble vitamin-branched CD complexes give rather better results than DM- β -CD complexes, on the basis of following facts. Although solubilizing effects on lipid-soluble vitamins in water at ordinary temperature with DM- β -CD complex formation were excellent, DM- β -CD complexes would tend to precipitate at high temperature owing to the solubility of DM- β -CD in water which becomes significantly lower with rising temperature. On the contrary, branched CDs are extremely easily dissolved in water at various temperatures, and therefore it is possible to use them as solubilizing agents over a wide range of temperature. Moreover, the hemolytic activity of branched CDs is lower than that of their parent CDs^{3,4,12} and DM- β -CD. (12)

These facts suggested that lipid-soluble vitamin-branched CD complexes may be practically applicable to injectable preparations. In addition, the lyophilized samples of inclusion complexes of lipid-soluble vitamins with G- β -CD may facilitate the development of solid dosage forms, and may resolve the undesirable problems encountered with the storage and handling of most lipid-soluble vitamins because of their viscous oily character.

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References and Notes

- A part of this study was presented at the 108th Annual Meeting of the Pharmaceutical Society of Japan, Hiroshima, April 1988.
- K. Koizumi, Y. Okada, Y. Kubota, and T. Utamura, Chem. Pharm. Bull., 35, 3413 (1987).
- 3) Y. Okada, Y. Kubota, K. Koizumi, S. Hizukuri, T. Ohfuji, and K.

- Ogata, Chem. Pharm. Bull., 36, 2176 (1988).
- M. Yamamoto, A. Yoshida, F. Hirayama, and K. Uekama, *Int. J. Pharm.*, 49, 163 (1989).
- Y. Okada, K. Koizumi, K. Ogata, and T. Ohfuji, *Chem. Pharm. Bull.*, 37, 3096 (1989).
- T. Higuchi and K. A. Connors, Adv. Anal. Chem. Instr., 4, 117 (1965).
- J. A. Nelder and R. Mead, Computer J., 7, 308 (1965); K. Yamaoka, Y. Tanigawara, T. Nakagawa, and T. Uno, J. Pharmcobio-Dyn., 4, 879 (1981).
- K. Uekama, H. Horiuchi, M. Kikuchi, and F. Hirayama, J. Inclusion Phenomena, 6, 167 (1988).
- Y. Horiuchi, M. Kikuchi, F. Hirayama, K. Uekama, M. Ueno, and T. Ijitsu, Yakugaku Zasshi, 108, 1093 (1988).
- T. Imai, T. Irie, M. Otagiri, and K. Uekama, J. Inclusion Phenomena, 2, 597 (1984).
- Y. Kubota, T. Tanimoto, S. Horiyama, and K. Koizumi, Carbohydr. Res., 192, 159 (1989).
- T. Irie, M. Otagiri, M. Sunada, K. Uekama, Y. Ohtani, Y. Yamada, and Y. Sugiyama, J. Pharmcobio-Dyn., 5, 741 (1982).