Novel Benzamides as Selective and Potent Gastrokinetic Agents. III. Synthesis and Structure-Activity Relationships of 4-Amino-5-chloro-2-methoxy- and 2-Ethoxy-N-[(4-substituted 2-morpholinyl)methyl]benzamides

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A series of 4-amino-5-chloro-2-methoxy- and 2-ethoxy-N-[(4-substituted 2-morpholinyl)methyl]benzamides (11-64) were prepared and evaluated for gastrokinetic activity by determining their effects on the gastric emptying of phenol red semisolid meal in rats. The N-4 substituent includes alkyl, phenoxyalkyl, (4-fluorobenzoyl)alkyl, and heteroarylmethyl groups. The benzamide derivatives, having an isopropyl, isoamyl, neopentyl, 3-(4-chlorophenoxy)propyl, or pyridylmethyl group at N-4, showed potent in vivo gastric emptying activity. In particular, 4-amino-5chloro-2-ethoxy-N-[[4-(3-pyridylmethyl)-2-morpholinyl]methyl]benzamide (57b) was equipotent to the 4-fluorobenzyl analogue 1b (AS-4370 as its citrate) in the gastrokinetic activity on phenol red semisolid meal in rats and mice, and on resin pellet solid meal in rats. Moreover, compound 57b was free from dopamine D2 receptor antagonistic activity in both in vitro ([3H]spiperone binding) and in vivo (apomorphine-induced emesis in dogs) tests. Structure-activity relationships of compounds with various substituents at N-4 are also discussed.

Keywords 2-morpholinyl benzamides; gastrokinetic agent; gastric emptying; dopamine D₂ antagonism; [3H]spiperone binding; apomorphine-induced emesis; structure-activity relationship

Our previous papers^{1,2)} reported that several benzamide derivatives represented generically by 1, which appended a new amine moiety, 2-(aminomethyl)-4-benzylmorpholine, showed potent gastrokinetic activity without dopamine D₂ receptor antagonistic activity; this morpholinyl moiety had been newly designed after consideration of the sidechain structure of cisapride which is used clinically as a gastrokinetic agent. This finding had led us to modify the benzoyl moiety and to introduce various substituents into the N-4 benzyl group, resulting in the discovery

2a:R = CH3

 $2b : R = C_2 H_5$

As an extention of that work, we have directed our effort to a search of N-4 substituents (R₁) that might cause a greater enhancement in gastrokinetic activity than the 4-fluorobenzyl group of 1b. The present paper deals with a synthesis and structure-activity relationship (SAR) of a new series of morpholinyl benzamides appending alkyl, phenoxyalkyl, (4-fluorobenzoyl)alkyl, and heteroarylmethyl groups at N-4 of the morpholine ring as shown in Table I.

Chemistry 4-Amino-5-chloro-2methoxy- and 2-ethoxy-N-[(4-substituted 2-morpholinyl)methyl]benzamides (12a— 64a), except the previously reported derivative 38a¹⁾ and the aminophenoxy derivative 52a, were synthesized by the reaction of 4-amino-5-chloro-2-methoxy- and 2-ethoxybenzoic acids (2a and 2b)1) with an appropriate 4substituted 2-(aminomethyl)morpholine (6 or 100, p) and 3-(aminomethyl)octahydropyrido [2,1-c][1,4] oxazine (10q) in the presence of 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride³⁾ as a coupling reagent (Chart 2). The aminophenoxy derivative 52a was derived by

H₂NCH₂
$$\xrightarrow{\text{N}}$$
 $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{EDC}^{a)}}$ $\xrightarrow{\text{CH}_2\text{Cl}_2}$ $\xrightarrow{\text{CI}}$ $\xrightarrow{\text{CONHCH}_2}$ $\xrightarrow{\text{N}}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{C}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{R}_1}$ $\xrightarrow{\text{R}_2}$ $\xrightarrow{\text{$

Chart 2

R₂ = H

10q: R1, R2

TABLE I. Physical Data for 2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides (12a-37b and 39a-64a)

Compd. R		$R_1^{a)}$	R_2	Yield ^{b)} (%)	mp (°C) (Recryst.	Formula			Analysis (alcd (Fo		
				Method ^{c)}	solvent ^{d)})		С	Н	N	Cl	F
12a	CH ₃	CH ₃	Н	10 ^{e)}	237—239	C ₁₄ H ₂₀ ClN ₃ O ₃	47.40	6.11	11.85	19.99	
12b	C ₂ H ₅	CH ₃	Н	C	(E)	·HCl·1/4H ₂ O	(47.33	6.07	11.73	20.11)	
120	C ₂ H ₅	Cn ₃	п	16 C	183—185 (E)	$C_{15}H_{22}CIN_3O_3$	54.96 (54.90	6.76 6.85	12.82 12.69	10.82	
13a	CH ₃	CH ₃ CH ₂	Н	23	235—237	C ₁₅ H ₂₂ ClN ₃ O ₃ ·HCl	49.46	6.36	11.54	10.77) 19.47	
				Α	(M)		(49.32	6.61	11.49	19.74)	
13b	C_2H_5	CH₃CH₂	Н	25	150—151	$C_{16}H_{24}CIN_3O_3$	56.22	7.08	12.29	10.37	
14a	CH ₃	CH ₃ (CH ₂) ₂	Н	A 48	(I) 224—228	C ₁₆ H ₂₄ ClN ₃ O ₃ ·HCl	50.80	7.11	12.24	10.24)	
	0113	0113(0112)2	•••	A .	(E)	C ₁₆ 11 ₂₄ CHV ₃ O ₃ TICI	(50.70	6.66 6.76	11.11 10.82	18.74 18.44)	
14b	C_2H_5	$CH_3(CH_2)_2$	Н	60	141—142	$C_{17}H_{26}CIN_3O_3$	57.38	7.36	11.81	9.96	
151	C 11	(011.)		A	(I)		(57.37	7.51	11.84	9.83)	
15b	C_2H_5	$(CH_3)_2CH$	Н	41 A	134—139	$C_{17}H_{26}CIN_3O_3$	47.16	7.16	9.71	18.44	
16a	CH ₃	CH ₃ (CH ₂) ₃	Н	68	(E) 231—235	·5/4HCl·7/4H ₂ O C ₁₇ H ₂₆ ClN ₃ O ₃ ·HCl	(46.91 52.05	7.53 6.94	9.61 10.71	18.79) 18.07	
	3	2,0		A	(E)	C ₁₇ 11 ₂₆ Ch(3O ₃ HC)	(52.00	7.07	10.71	17.79)	
16b	C_2H_5	$CH_3(CH_2)_3$	H	59	209—210	$C_{18}H_{28}CIN_3O_3$	52.05	7.28	10.12	17.07	
150	0.11	CII (CII)		A	(AC-W)	·HCl·1/2H ₂ O	(51.93	7.09	9.92	17.01)	
17b	C_2H_5	$CH_3(CH_2)_4$	Н	82 A	205—207	C ₁₉ H ₃₀ ClN ₃ O ₃	51.01	7.66	9.39	15.85	
18b	C_2H_5	(CH ₃) ₂ CH(CH ₂) ₂	Н	10	(M) 212—216	·HCl·3/2H ₂ O C ₁₉ H ₃₀ ClN ₃ O ₃	(51.19 48.10	7.83 7.86	9.41 8.86	15.68) 14.94	
		(3)2(2)2		A	(E)	·HCl·3H ₂ O	(48.10	7.86	8.86	15.18)	
19b	C_2H_5	(CH3)3CCH2	Н	2	210—214	$C_{19}H_{30}CIN_3O_3$	51.51	7.45	9.49	18.01	
201				A	(E)	· 5/4HCl · 3/4H ₂ O	(51.84	7.75	9.11	17.92)	
20b	C_2H_5	$CH_3(CH_2)_5$	Н	71	190—195	$C_{20}H_{32}CIN_3O_3$	49.18	8.05	8.60	14.52	
21b	C_2H_5	CH ₃ (CH ₂) ₆	Н	A 61	(I) 207—210	·HCl·3H ₂ O	(49.34	8.09	8.70	14.80)	
210	C2115	C113(C112)6	11	A	(E)	$C_{21}H_{34}ClN_3O_3 \cdot 3/2HCl \cdot 1/2C_2H_5OH^{f)}$	53.96 (54.08	7.92 7.83	8.58 8.87	18.10 17.92)	
22b	C_2H_5	CH ₃ (CH ₂) ₇	Н	74	203—206	$C_{22}H_{36}CIN_3O_3$	54.98	8.02	8.74	16.60	
				Α	(E)	5/4HCl·1/2H ₂ O	(55.30	8.03	8.74	16.70)	
23b	C_2H_5	$-(CH_2)_4-$		27 ^{g)}	192205	$C_{18}H_{26}CIN_3O_3$	58.77	7.12	11.42	9.64	
24a	CH ₃		Н	C	(M)	C II CN O	(58.70	6.94	11.25	10.00)	
27a	CII3		п	11 A	189—190 (I)	$C_{19}H_{28}CIN_3O_3$	59.76 (59.53	7.39 7.49	11.00	9.28	
25a	CH ₃	CH₂	Н	37	105—108	C ₁₇ H ₂₄ ClN ₃ O ₃ ·1/4H ₂ O		6.89	10.91 11.73	9.18) 9.89	
				A	(I)	01/11/24011/30/3 1/111/20	(56.69	6.98	11.65	9.86)	
26a	CH_3	⟨	Н	84	159—163	$C_{20}H_{28}CIN_3O_3$	56.03	6.37	8.17	6.89	
27-	CH		••	В	(E)	$C_4H_4O_4^{h} \cdot 1/4H_2O$	(55.83	6.34	8.04	6.75)	
27a	CH ₃	$CH_2 = CHCH_2$	Н	44 B	122—124	$C_{16}H_{22}CIN_3O_3$	56.55	6.53	12.37	10.43	
28a	CH ₃	$C_6H_5CH = CHCH_2$	н	58	(I–DE) 124—127	$C_{22}H_{26}CIN_3O_3$	(56.25 55.72	6.59 5.59	12.13 6.96	10.71)	
	3	0,11,011 0110112	••	B	(E-DE)	$\cdot 3/2C_4H_4O_4^{h}\cdot 3/4H_2O$	(55.69	5.82	6.83	5.87 5.82)	
29b	C_2H_5	$CH \equiv CCH_2$	Н	23	148151	$C_{17}H_{22}CIN_3O_3 \cdot 1/4H_2O$		6.36	11.94	9.95	
201	C 11	Q II (QQ) \$1(5)		A	(I)		(57.64	6.35	11.79	10.20)	
30b	C_2H_5	$C_6H_4(CO)_2N(CH_2)_4$	H	78 P	139—141	$C_{26}H_{31}CIN_4O_5$	60.64	6.07	10.88	6.88	
31a	CH ₃	C ₆ H ₅ NH(CH ₂) ₂	н	В 13	(E) 117—125	$C_{21}H_{27}CIN_4O_3$	(60.66	6.12	10.60	6.71)	
	3	-03- 111(-112/2	11	A	(E)	$C_{21}H_{27}CIN_4U_3$ $C_2H_2O_4^{i)} \cdot 5/4H_2O$	51.94 (51.98	5.99 5.97	10.54 10.15	6.67 6.90)	
32b	C_2H_5	(CH3)2N(CH2)3	Н	11	223—225	$C_{19}H_{31}CIN_4O_3$	45.08	6.03	8.37	5.69	
				Α	(M)	$\cdot 5/2C_2H_2O_4^{(i)}\cdot H_2O$	(44.90	5.97	8.73	5.52)	
33b	C ₂ H ₅	CH ₃ OCO(CH ₂) ₂	H	7	125—127	$C_{18}H_{26}ClN_3O_5 \cdot 1/4H_2O$		6.61	10.39	8.77	
34b	C ₂ H ₅	HO(CH ₂) ₃	н	A 43	(I) 162—164	C ₁₇ H ₂₆ ClN ₃ O ₄	(53.59	6.45	10.45	9.04)	
J	€2115	110(0112)3	11	43 A	(E)	$C_{17}H_{26}CIN_3U_4$ $\cdot 3/2C_4H_4O_4^{h}$	50.60 (50.46	5.91 5.89	7.70 7.52	6.49 6.38)	
35b	C_2H_5	NC(CH ₂) ₂	Н	14	182—183	$C_{17}H_{23}CIN_4O_3$	55.66	6.32	15.27	9.66	
200				В	(I)	· · · · · ·	(55.36	6.27	14.97	9.69)	
36b	C_2H_5	$(C_2H_5O)_2CHCH_2$	H	<i>77</i>	168—170	$C_{20}H_{32}CIN_3O_5$	51.70	6.34	6.96	5.87	
37ь	C ₂ H ₅	C ₆ H ₅	н	B 24 ^{j)}	(E)	·3/2C ₄ H ₄ O ₄ ^{h)}	(51.63	6.21	6.94	5.85)	
J/6	~2115	~6**5	11	24" C	163—165 (I)	$C_{20}H_{24}CIN_3O_3 \cdot 1/4H_2O$	60.91	6.26 6.11	10.56 10.57	8.99	
39a	CH ₃	C ₆ H ₅ O(CH ₂) ₃	Н	37	133—135	$C_{22}H_{28}CIN_3O_4$	60.89	6.50	9.68	9.13) 8.17	
	•		-	В	(E)	44203-4	(60.72	6.45	9.61	8.18)	

TABLE I. (continued)

Compd.	R	$R_1^{a)}$	R_2	Yield ^{b)} (%)	mp (°C) (Recryst.	Formula	Analysis (%) Calcd (Found)				
Compu.	K			Method ^{c)}	solvent ^{d)})	<u>-</u>	C	Н	N	Cl	F
40a	CH ₃	C ₆ H ₅ OCH(CH ₃)CH ₂	Н	41 B	113—115 (E)	$C_{22}H_{28}CIN_3O_4$ $C_2H_2O_4^{i)}$	55.33 (55.02	5.85 5.77	8.14 8.02	6.79 6.77)	
41a	CH ₃	$4-FC_6H_4O(CH_2)_2$	Н	31 B	148—150 (I)	C ₂₁ H ₂₅ ClFN ₃ O ₄	57.60 (57.64	5.75 6.12	9.60 9.45	8.10 8.03	4.34 4.38)
41b	C_2H_5	$4-FC_6H_4O(CH_2)_2$	Н	44	182—184	C ₂₂ H ₂₇ CIFN ₃ O ₄	52.67	5.16	6.14	5.18	2.78
42a	CH ₃	4-FC ₆ H ₄ O(CH ₂) ₃	Н	B 14	(E) 127—129	·2C ₄ H ₄ O ₄ ^{h)} C ₂₂ H ₂₇ ClFN ₃ O ₄	(52.66 58.47	5.14 6.02	6.07 9.30	5.31 7.84	2.78 4.20
42b	C ₂ H ₅	4-FC ₆ H ₄ O(CH ₂) ₃	Н	B 20	(T–H) 125—126	C ₂₃ H ₂₉ ClFN ₃ O ₄	(58.63 59.29	5.96 6.27	9.21 9.02	7.80 7.61	4.14 4.08
43a	CH ₃	2-FC ₆ H ₄ O(CH ₂) ₃	Н	В 25	(I) 69—72	C ₂₂ H ₂₇ ClFN ₃ O ₄	(58.93 58.47	6.48 6.02	9.06 9.30	7.84 7.84	3.81 4.20
44a	CH ₃	3-FC ₆ H ₄ O(CH ₂) ₃	Н	B 33	(E) 78—81	C ₂₂ H ₂₇ ClFN ₃ O ₄	(58.08 58.47	6.02 6.02	9.10 9.30	7.88 7.84	4.09
	_			B 64	(E) 158—161	$C_{23}H_{29}CIFN_3O_4$	(58.11 54.42	5.97 5.51	9.12 6.57	7.92 5.54	4.09 2.97
45a	CH ₃	4-FC ₆ H ₄ O(CH ₂) ₄	H 	В	(E-DE)	$\cdot 3/2C_4H_4O_4^{h}$	(54.31	5.79	6.40	5.25	2.92
46a	CH ₃	4-FC ₆ H ₄ O(CH ₂) ₅	Н	48 B	164—166 (E)	$C_{24}H_{31}ClFN_3O_4$ $\cdot C_2H_2O_4^{i)} \cdot 1/4H_2O$	54.26 (54.11	6.04 5.84	7.30 7.29	6.16 5.98	3.30 3.41
47a	CH ₃	$4-FC_6H_4O(CH_2)_6$	Н	39 B	120—122 (E-DE)	$C_{25}H_{33}CIFN_3O_4$ $\cdot 3/2C_4H_4O_4^{h}$	55.73 (55.49	5.88 5.91	6.29 6.27	5.31 5.16	2.84 2.85
48a	CH ₃	$4-ClC_6H_4O(CH_2)_3$	Н	51 B	123—126	$C_{22}H_{27}Cl_2N_3O_4$	56.42 (56.27	5.81 5.73	8.97 8.98	15.14 15.33)	
48b	C_2H_5	4-ClC ₆ H ₄ O(CH ₂) ₃	Н	59	(I) 149—151	$C_{23}H_{29}Cl_2N_3O_4$	57.27	6.06	8.71	14.70	
49a	CH ₃	4-ClC ₆ H ₄ O(CH ₂) ₂	Н	B 28	(E) 158—162	$C_{21}H_{25}Cl_2N_3O_4$	(57.01 50.01	6.13 4.74	8.42 5.64	14.56) 9.52	
49b	C_2H_5	4-ClC ₆ H ₄ O(CH ₂) ₂	н	В 31	(E-DE) 190—192	$\cdot 5/2C_4H_4O_4^{h}$ $C_{22}H_{27}Cl_2N_3O_4$	(50.05 51.62	4.84 5.23	5.48 7.53	9.17) 12.70	
50a	CH ₃	4-CNC ₆ H ₄ O(CH ₂) ₃	Н	B 46	(E) 170—172	$\cdot \tilde{C}_{2}H_{2}O_{4}^{i)}$ $C_{23}H_{27}CIN_{4}O_{4}\cdot 1/4H_{2}O$	(51.76 59.61	5.33 5.98	7.46 12.09	12.46) 7.65	
				В	(I)		(59.49 60.95	5.90 6.18	11.88 11.85	7.76) 7.50	
50b	C_2H_5	4-CNC ₆ H ₄ O(CH ₂) ₃	Н	50 B	157—158 (E)	C ₂₄ H ₂₉ ClN ₄ O ₄	(60.89	6.22	11.84	7.47)	
51a	CH ₃	$4-NO_2C_6H_4O(CH_2)_3$	Н	68 B	149—153 (E)	$C_{22}H_{27}CIN_4O_6\cdot 1/5H_2O$	54.76 (54.98	5.72 5.64	11.61 11.28	7.35 7.52)	
51b	C_2H_5	$4-NO_2C_6H_4O(CH_2)_3$	Н	65 B	137—139 (E)	$C_{23}H_{29}ClN_4O_6\cdot 1/4H_2O$	55.53 (55.77	5.98 6.13	11.26 10.97	7.13 6.98)	
52a	CH ₃	4-NH ₂ C ₆ H ₄ O(CH ₂) ₃	Н	51 K	212—216 (I)	C ₂₂ H ₂₉ ClN ₄ O ₄ ·3/2C ₂ H ₂ O ₄ ⁱ⁾ ·4/5(CH ₃) ₂ CHOH ^{f)}	52.07 (52.28	6.12 6.30	8.86 8.73	5.61 5.69)	
53a	CH_3	$4-FC_6H_4S(CH_2)_3$	Н	60 B	127—130 (I)	$C_{22}H_{27}ClFN_3O_3S^{1}$	56.46 (56.45	5.82 5.71	8.98 8.90	7.58 7.63	4.0 4.0
53b	C_2H_5	$4-FC_6H_4S(CH_2)_3$	Н	66	158—160	C ₂₃ H ₂₉ ClFN ₃ O ₃ S	54.22	5.56	7.03	5.93	3.1
54a	CH ₃	4-FC ₆ H ₄ CO(CH ₂) ₃	Н	В 15	(E) 148—152	$C_4H_4O_4^{m,h}$ $C_{23}H_{27}CIFN_3O_4$	(54.48 55.48	5.53 5.43	7.18 7.19	6.10 6.07	3.3
54b	C ₂ H ₅	4-FC ₆ H ₄ CO(CH ₂) ₃	Н	A 13	(E) 135—138	$\cdot C_4 H_4 O_4^{h_1} \cdot 1/4 H_2 O$ $C_{24} H_{29} ClF N_3 O_4$	(55.20 52.67	5.62 5.98	7.11 7.23	6.26 6.10	3.1 3.2
55a	CH ₃	4-FC ₆ H ₄ COCH ₂	Н	A 70	(M) 180—183	$\cdot \frac{3}{4} C_{2} H_{2} O_{4}^{ij} \cdot 2 H_{2} O$ $C_{21} H_{23} CIFN_{3} O_{4}$	(52.39 57.87	5.69 5.32	7.12 9.64	6.46 8.13	3.0 4.3
	_	• • •	Н	A 77	(E) 187—189	C ₂₂ H ₂₅ ClFN ₃ O ₄	(57.78 55.00	5.31 5.35	9.60 7.70	8.21 6.49	4.2 3.4
55b	C ₂ H ₅	4-FC ₆ H ₄ COCH ₂		Α	(E)	$\cdot 3/4C_4H_4O_4^{h}\cdot 1/2H_2O$	(54.69	5.47	7.48	6.77	3.2
56a	CH ₃	2-Pyridylmethyl	Н	64 A	88—91 (E-DE)	$C_{19}H_{23}CIN_4O_3$ $\cdot 3/2C_4H_4O_4^{h_1}\cdot H_2O$	51.51 (51.23	5.36 5.28	9.61 9.35	6.08 6.12)	
56b	C_2H_5	2-Pyridylmethyl	Н	51 A	182—185 (I)	$C_{20}H_{25}ClN_4O_3 \cdot C_4H_4O_4^{h)}$	55.33 (55.11	5.61 5.90	10.75 10.45	6.81 6.70)	
57a	CH ₃	3-Pyridylmethyl	Н	26 A	126—128 (E-W)	$C_{19}H_{23}CIN_4O_3$ $\cdot C_2H_2O_4^{(1)}$ $\cdot 2/5C_2H_5OH^{(f)}$	52.44 (52.27	5.53 5.65	11.22 10.95	7.10 7.20)	
57b	C_2H_5	3-Pyridylmethyl	Н	34 A	150—152 (I)	C ₂₀ H ₂₅ ClN ₄ O ₃ ·C ₄ H ₄ O ₄ ^{h)} ·1/4(CH ₃) ₂ CHOH ^f)	55.46 (55.33	5.83 5.90	10.45 10.15	6.61 6.69)	
58a	CH ₃	4-Pyridylmethyl	Н	50	167—170	$C_{19}H_{23}CIN_4O_3$	58.38 (58.12	5.93 5.93	14.33 14.19	9.07 9.35)	
58b	C_2H_5	4-Pyridylmethyl	Н	A 51	(I) 175—176	$C_{20}H_{25}ClN_4O_3$	59.33	6.22	13.84	8.76	
59a	CH ₃	2-Furylmethyl	Н	A 60	(I) 131—134	$C_{18}H_{22}ClN_3O_4$	(59.13 56.92	6.45 5.84 5.82	13.58 11.06	8.70) 9.33	

TABLE I. (continued)

Compd.	R	$R_1^{a)}$	R ₂	Yield ^{b)} mp (°C) (%) (Recryst. Method ^{c)} solvent ^{d)})	- ' '	Formula	Analysis (%) Calcd (Found)				
						С	Н	N	Cl	F	
60a	CH ₃	3-Furylmethyl	Н	50 B	166—168 (E-DE)	C ₁₈ H ₂₂ ClN ₃ O ₄ ·1/2C ₄ H ₄ O ₄ ^{h)} ·1/4H ₂ O	54.30 (54.35	5.58 5.58	9.50 9.33	8.01 7.88)	
61a	CH ₃	2-Thienylmethyl	Н	76 B	158—160 (E-DE)	C ₁₈ H ₂₂ ClN ₃ O ₃ S ·1/2C ₄ H ₄ O ₄ ^h) ·3/10H ₂ O ⁿ)	52.30 (52.58	5.40 5.26	9.33 9.15 9.04	7.72 7.40)	
62a	CH ₃	3-Thienylmethyl	Н	38 B	146—147 (I–DI)	C ₁₈ H ₂₂ ClN ₃ O ₃ S ·1/5H ₂ O°)	54.12 (54.23	5.65 5.40	10.52 10.50	8.87 9.07)	
63a	CH ₃	3-Benzisoxazolylmethyl	Н	86 A	128—129 (E)	$C_{21}H_{23}CIN_4O_4$ ·1/2C ₄ H ₄ O ₄ ^{h)} ·1/2H ₂ O	55.48 (55.62	5.26 5.14	11.25 10.95	7.12 6.97)	
64a	CH ₃	2-Naphthylmethyl	Н	62 B	155—158 (E-DE)	$C_{24}H_{4}O_{4}^{h_{1}}O_{3}$ $\cdot C_{4}H_{4}O_{4}^{h_{1}}\cdot 3/5H_{2}O$	59.33 (59.62	5.55 5.83	7.41 7.15	6.25 5.95)	

a) Alkylating agents R₁X are obtained from commercial suppliers or synthesized according to the literatures. b) Total yields of the free bases are based on 2-[(acetylamino)methyl]-4-benzylmorpholine (methods A and B) or 2-substituted aminoethanol (C). c) Capital letters refer to the procedures described in the experimental section. d) Abbreviations for the solvents used are as follows: E=ethanol, M=methanol, I=isopropanol, AC=acetone, W=water, DE=diethyl ether, T=toluene, H=n-hexane, DI=diisopropyl ether. e) 2-(Methylamino)ethanol is used as the starting material. f) The presence of crystallization solvent is shown by ¹H-NMR. g) 2-Piperidinemethanol is used as the starting material. h) Fumaric acid. i) Oxalic acid. j) 2-Anilinoethanol is used as the starting material. k) See the experimental section. l) Calcd for S: 6.85, Found: 7.12. m) Calcd for S: 5.36, Found: 5.63. n) Calcd for S: 6.98, Found: 6.67. o) Calcd for S: 8.03, Found: 7.76.

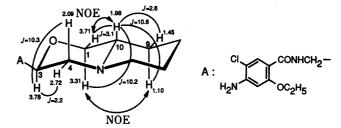
Chart 3

method C

Chart 4

hydrogenation of the corresponding nitro compound 51a in the presence of Raney nickel.

Most of the morpholine derivatives were derived as shown in Chart 3. Thus, hydrogenolysis of 2-[(acetylamino)methyl]-4-benzylmorpholine (3) over palladium on carbon gave 2-[(acetylamino)methyl]morpholine (4), which was subsequently treated with appropriate alkyl and aralkyl halides to give 4-substituted 2-[(acetylamino)methyl]morpholines (5). Acid or base catalized hydrolysis of the acetylamino group of 5 produced the desired 4-substituted 2-(aminomethyl)morpholines (6). 2-(Aminomethyl)-4-methylmorpholine (100), 2-(aminomethyl)-4phenylmorpholine (10p), and 3-(aminomethyl)octahydropyrido[2,1-c][1,4] oxazine (10q) were prepared according to the method described in the previous paper²⁾ (Chart 4). The reactions of 2-(methylamino)ethanol (70), 2-anilinoethanol (7p), and 2-(hydroxymethyl)piperidine (7q) with N-(2,3-epoxypropyl)phthalimide (8) at 80 °C, followed by the treatment of the corresponding intermediate diols 90q with concentrated sulfuric acid, gave (aminomethyl)-



 $R_1, R_2 = -(CH_2)_4$

Fig. 1. ¹H-NMR Chemical Shifts (ppm), Coupling Constants (Hz), and NOE Correlation of the Octahydropyrido[2,1-c][1,4]oxazine Ring Protons of 23b (¹H-NMR in CDCl₃)

morpholines 100—q, respectively, accompanied by the formation of phthalic anhydride. Acid treatment of the diols caused not only the cyclization to the morpholine ring but the concurrent hydrolysis of the phthalimide moiety to an amino group, thereby conveniently yielding the desired (aminomethyl)morpholines. The structures of all compounds thus prepared were confirmed by their proton nuclear magnetic resonance (1H-NMR) spectra

Table II. Effect of 2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides (11a—64a) on Gastric Emptying of Phenol Red Semisolid Meal in Rats

Gastric emptying rate Acute toxicity d) 2.0 mg/kg, p.o. Control Compd. $(Mean \pm S.E.M.)$ $(Mean \pm S.E.M.)$ % change (1.0g/kg, p.o.) $(N^{a)}$ ND 119 e 55.6 ± 5.2 (5) 71.6 ± 5.4 (4) ND 16 52.7 ± 2.1 (5) 61.1 ± 4.6 (4) 12a 30^{b)} $55.1 \pm 4.6 (4)$ 71.7 ± 3.7 (3) ND 12b 26b) ND 70.5 ± 4.6 (4) 13a $55.8 \pm 3.4 (5)$ 39c) 76.6 ± 7.3 (3) ND 13b 55.1 ± 4.6 (4) 35c) ND 75.2 ± 2.0 (4) 14a $55.8 \pm 3.4 (5)$ 47^{c)} 81.2 ± 3.8 (3) ND 55.1 ± 4.6 (4) 14b 78.3 ± 1.9 (4) 54°) 7/10 15b 50.8 ± 3.1 (5) 73.0 ± 3.2 (4) 286 ND $57.2 \pm 4.2 (5)$ 16a 75.2 ± 3.8 (4) 48c) 10/10 16b 50.8 ± 3.1 (5) 49^{c)} 17b 50.8 ± 3.1 (5) 75.9 ± 4.6 (4) 10/10 58c) 79.5 ± 2.5 (4) 10/10 18b $50.2 \pm 3.4 (5)$ 82.7 ± 7.0 (3) 50° ND 19b 55.1 ± 4.6 (4) 47°) 73.3 ± 3.4 (4) 8/10 20b 50.0 ± 1.6 (5) ND 67.5 ± 3.6 (3) 25 21b 54.2 ± 2.6 (4) 54.2 ± 2.6 (4) 58.7 ± 7.9 (3) 8 ND 22b ND $51.3 \pm 4.9 (5)$ 54.4 ± 4.9 (4) 6 23b 52.8 ± 3.4 (5) 63.2 ± 4.9 (4) 20 ND 249 ND 24 258 52.8 ± 3.4 (5) 65.6 ± 4.5 (4) ND $54.5 \pm 3.9 (5)$ 61.6 ± 3.4 (4) 13 26a ND 53.2 ± 1.9 (5) 60.3 ± 5.1 (4) 13 27a 19^{b)} ND 56.6 ± 2.1 (5) 67.2 ± 1.8 (4) 28a 32c) ND 29b 51.2 ± 1.7 (5) 67.6 ± 1.8 (4) 35c) ND 30h 45.0 ± 5.2 (6) 60.8 ± 5.6 (4) 57.2 ± 3.0 (4) 8 ND 53.1 ± 3.1 (5) 31a 28c) ND 52.4 ± 1.8 (5) 67.1 ± 2.9 (4) 32h $28^{b)}$ ND 70.6 ± 4.3 (3) 33b $55.1 \pm 4.6 (4)$ 67.8 ± 4.2 (3) 23 ND 34b 55.1 ± 4.6 (4) 67.6 ± 3.4 (3) 23 ND 35h $55.1 \pm 4.6 (4)$ 29^{b)} ND 70.1 ± 5.5 (3) 54.2 ± 1.6 (4) 36b 21 ND 66.0 + 4.9 (4)37b 54.5 ± 1.0 (5) 67.1 ± 4.4 (4) 19 ND 38ae) 56.6 ± 2.1 (5) 24b) ND 39a 55.1 ± 4.6 (4) 68.3 ± 1.6 (3) ND 19 40a $52.6 \pm 3.9 (5)$ 62.8 ± 2.6 (4) $31^{b)}$ ND 72.6 ± 1.2 (4) 55.6 ± 5.2 (5) 41a 37^{c)} ND 55.1 ± 4.6 (4) 75.5 ± 5.4 (3) 41b $55.4 \pm 1.4 (15)$ 23°) ND 68.1 ± 2.0 (4) 429 37c) ND 55.1 ± 4.6 (4) 75.5 ± 5.1 (3) 42h $23^{b)}$ ND 66.7 ± 3.9 (3) 54.2 ± 2.6 (4) 43a 19 ND 54.2 ± 2.6 (4) 64.6 ± 3.6 (3) 44a $20^{b)}$ 45a 56.6 ± 2.1 (5) 67.9 ± 3.4 (4) ND ND 50.7 ± 2.3 (5) 57.5 ± 3.0 (4) 13 46a 9 ND 55.6 ± 5.2 (5) 60.6 ± 4.2 (4) 479 53°) 0/10 48a 55.8 ± 3.4 (5) 85.1 ± 1.2 (4) $34^{b)}$ ND 51.2 ± 1.7 (5) 68.5 ± 4.9 (4) 48h 32b) ND 49a 54.2 ± 2.6 (4) 71.5 + 1.1(3)35c) ND 51.8 + 2.1 (5) 70.0 ± 1.6 (4) 49h 29c) ND 50a 52.6 ± 1.7 (5) 67.9 ± 2.8 (4) 48°) ND 50b 54.2 ± 2.6 (4) 80.3 ± 4.2 (3) 28c) ND 63.8 ± 3.0 (4) 51a 50.0 ± 1.8 (5) 27^{b)} 51b 54.2 ± 2.6 (4) 69.0 ± 6.7 (3) ND ND 58.0 ± 2.8 (4) 12 51.7 ± 3.4 (5) 52a 36c) ND 50.0 ± 1.8 (5) 68.1 ± 1.9 (4) 53a 40c) ND 55.1 ± 4.6 (4) 77.1 ± 0.7 (3) 53b 24c) ND 56.6 ± 2.1 (5) 70.0 ± 1.9 (4) 548 37c) ND 51.1 ± 1.2 (5) 69.9 ± 3.5 (4) 54h 33b) ND 55a 55.8 ± 3.4 (5) 74.3 ± 4.1 (4) 48c) 54.2 ± 2.6 (4) 80.0 ± 2.3 (3) ND 55b 30b) ND 52.8 ± 3.4 (5) 56a 68.4 ± 2.6 (4) 51°) ND 56b 54.1 ± 1.5 (5) 81.7 ± 1.2 (4) 35°) ND 71.7 ± 2.5 (4) 57a $53.1 \pm 3.1 (5)$ 66c) 4/10 57b 50.8 ± 2.9 (5) 84.1 ± 3.7 (4) 70.0 ± 6.4 (4) 326) ND 58a 53.1 ± 3.1 (5)

TABLE II. (continued)

	Gast	ric emptying rate		- Acute
Compd.	Control (Mean ± S.E.M.) (Na)	% change	toxicity ^{d)}	
58b	54.1 ± 1.5 (5)	82.7 ± 3.0 (4)	53°)	0/10
59a	$52.6 \pm 1.7 (5)$	59.0 ± 3.1 (4)	12	ND
60a	50.5 ± 1.3 (5)	$57.1 \pm 5.8 (4)$	13	ND
61a	51.1 ± 2.7 (5)	63.2 ± 4.8 (4)	$24^{b)}$	ND
62a	51.1 ± 2.7 (5)	62.5 ± 3.6 (4)	$22^{b)}$	ND
63a	$50.5 \pm 1.3 (5)$	62.6 ± 2.6 (4)	24 ^{c)}	ND
64a	52.6 ± 1.7 (5)	69.3 ± 1.8 (4)	32^{c}	ND
Cisapride	$55.2 \pm 1.9 (5)$	$81.3 \pm 1.2 (5)$	47 ^{c)}	0/10
Metoclopr- amide	$58.9 \pm 2.1 (5)$	$71.3 \pm 3.8 $ (5)	21 ^{b)}	10/10
1a	$54.5 \pm 3.8 (5)$	75.9 ± 3.6 (4)	39c)	6/10
1b	$51.8 \pm 2.1 (5)$	83.6 ± 2.4 (4)	61°)	4/10

a) Number of rat used. A statistically significant difference from the control group; b) p < 0.05; c) p < 0.01 (Duncan's multiple range test). d) Number of dead mouse/number of mouse used. e) See ref. 1. ND, not done.

and elemental analyses.

The relative stereochemistry of the bicyclic octahydropyrido[2,1-c][1,4]oxazine ring of 23b was confirmed on the basis of coupling constants and the nuclear Overhauser effect (NOE) in their ¹H-NMR experiments. The chemical shifts, coupling constants, and NOE correlation were shown in Fig. 1. The signal for the H-10 appeared as two pairs of a double doublet centered at δ 1.98 with coupling constants of 10.2/3.1 and 10.6/2.6 Hz owing to its spinspin couplings to axial/equatorial protons at C-1 and C-9, respectively. NOEs were observed between the axial H-9 $(\delta 1.10)$ and the axial H-1 $(\delta 3.31)$, and the H-10 $(\delta 1.98)$ and the equatorial H-1 (δ 3.71) on irradiation at each former proton. The NOE experiment between the H-10 and the axial H-4 (δ 2.09), however, provided no information because their signals appeared quite closely. In infrared (IR) spectrum of 23b, Bohlmann bands⁴⁾ (2700— 2800 cm⁻¹) were observed. The signals for the axial and equatorial H-4's appeared as a double doublet centered at δ 2.09 and 2.72 with coupling constants of 10.3 and 2.2 Hz $(J_{aem H-4} = 11.4 \text{ Hz})$, respectively, owing to H-3; the configuration of H-3 therefore is axial. These data collectively indicate that the octahydropyrido [2,1-c][1,4] oxazine ring is of the trans configuration with the axial H-10.

Biological Results and Discussion

Compounds 12a—64a were evaluated for gastrokinetic activity by determining their effects on the gastric emptying rates of phenol red semisolid meal through the stomach. The activity data at an oral dose of 2.0 mg/kg in rats are given in Table II, which includes, for comparison, data for 4-amino-5-chloro-2-methoxy-N-[(2-morpholinyl)methyl]-benzamide¹⁾ (11a), cisapride, metoclopramide, 4-amino-N-[(4-benzyl-2-morpholinyl)methyl]-5-chloro-2-methoxy-benzamide²⁾ (1a) and 4-amino-5-chloro-2-ethoxy-N-[[4-(4-fluorobenzyl)-2-morpholinyl]methyl]benzamide¹⁾ (1b, AS-4370 as its citrate). The series of benzamide derivatives, which have commonly a 2-alkoxy-4-amino-5-chlorobenzoyl group, are classified into the following two series for convenience: series a, 2-methoxybenzamides (analogues of 1b).

The SARs associated with modification of the N-4 substituent (R₁) is first discussed. In series a, either removal of the benzyl group from 1a (giving 11a1) or replacement with an alkyl group at N-4 (giving 12a-14a and 16a) resulted in reducing activity; thus the decreasing order is $n-C_3H_7$ (14a)>H (11a)= $n-C_4H_9$ (16a) $\geq C_2H_5$ (13a)>metoclopramide>CH₃ (12a). In series b, replacement of the 4-fluorobenzyl group of 1b with an alkyl group (yielding 12b-22b) caused a decrease in activity. However, compounds 14b-20b bearing an alkyl group with three to six carbon atoms are practically comparable or somewhat superior to cisapride, and, in particular, the isoamyl substitution (18b) conferred the highest activity. Compounds bearing an n-alkyl group exhibit activity in the decreasing order pentyl (17b)≥butyl (16b)≥hexyl (20b) = propyl (14b) > ethyl (13b) > methyl (12b) > heptyl(21b) > metoclopramide » octyl (22b). Overall, the 2-ethoxy compounds (12b-14b, 16b) are clearly more active than the 2-methoxy counterparts (12a-14a, 16a). Formation of a bicyclic octahydropyrido[2,1-c][1,4]oxazine ring as in 23b (an n-butylene bridge between N-4 and C-5 of the morpholine ring) caused a profound decrease in activity as compared to the N-4 butyl derivative 16b. Introduction of a cycloalkyl group as in 24a-26a deteriorated the activity. Introduction of a double bond such as allyl (27a) and cinnamyl (28a) groups and a triple bond such as a propargyl (29b) group into the N-4 side chain reduced the activity. The N-4 alkyl group bearing substituted amino (30b, 31a, 32b), methoxycarbonyl (33b), hydroxy (34b), cyano (35b), and acetal (36b) groups also was deleterious. Replacement of the N-4 alkyl group by phenyl (37b) and ethoxycarbonyl (38a) groups caused a decrease in activity.

The next discussion concerns the SAR of a series of N-4 phenoxyalkyl derivatives (39—52). The 3-phenoxypropyl derivative (39a) is more potent than the 2-phenoxypropyl analogue (40a). Introduction of a fluoro group into the phenyl group of 39a provided no favorable influence on activity, thus retaining (as in 42a, 43a) or slightly decreasing the activity (as in 44a). As for the position of the fluoro group on the phenyl ring, the decreasing order of activity is para (42a)=ortho (43a)>meta (44a) positions. There has previously been observed a similar SAR concerning the position of the fluoro group on the

phenyl ring of 1a and 1b.¹⁾ With an increase in size of the alkyl moiety of the (4-fluorophenoxy)alkyl group in series a, the activity decreases in the order $(CH_2)_2$ $(41a) > (CH_2)_3$ $(42a) > (CH_2)_4$ $(45a) > (CH_2)_5$ $(46a) > (CH_2)_6$ (47a). In series b, on the other hand, the *n*-propylene group (42b) contributes to activity as same as the ethyl group (41b). It is noteworthy that compounds 41a and 41b with the shortest methylene chain are even more potent than metoclopramide, while compound 47a with the longest methylene chain $(-(CH_2)_6-)$ is the least active and practically inactive at this screening dose.

In general, replacement of the fluoro group of the 3-(4-fluorophenoxy)propyl derivative 42a by an electronwithdrawing group such as chloro (48a), cyano (50a), and nitro (51a) groups tended to increase activity, whereas replacement by an electron-donating amino group (52a) led to a decrease in activity. Thus, variation of the para substituent causes a decrease in activity in the order chloro $(48a) \gg \text{cyano } (50a) \ge \text{nitro } (51a) > \text{hydrogen } (39a) \ge \text{fluoro}$ (42a) » amino (52a) groups. In series b, the para substitution reduced activity in the order cyano (50b)>fluoro (42b)>chloro (48b)>nitro (51b) groups. Although the 3-(4-chlorophenoxy)propyl group gave compound 48a with a greater activity compared with cisapride as well as 42a, the substitution by this group in series b (giving 48b) gave little effect on activity (cf. 48a vs. 48b). Similarly, replacement of the propylene moiety of the 3-(4-chlorophenoxy)propyl derivatives 48a and 48b by an ethylene moiety (giving 49a and 49b, respectively) provided no favorable influence on activity.

Replacement of the oxygen atom of the 3-(4-fluorophenoxy)propyl derivatives 42a and 42b by a sulfur atom (yielding 53a, 53b, respectively) resulted in an increase in activity. Replacement by a carbonyl group (yielding 54a, 54b) substantially retained the activity. Conversion of the propylene moiety of the 3-(4-fluorobenzoyl)propyl derivatives 54a and 54b into a methylene moiety gave compounds 55a and 55b, respectively, with a considerably increased activity.

In order to know the influence of a heteroaryl ring, instead of the phenyl ring of the *N*-benzyl groups of 1a and 1b, on the gastric emptying activity, compounds 56—64 were prepared. In series a, substitution by pyridyl

TABLE III. Effect of Selective 2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides on Gastric Emptying of Phenol Red Semisolid Meal in Rats

			Gastric emptying	g rate ^{a)}		
Compd.	Control (N ^{b)})	0.1	Control (N)	0/ -1	Control (N)	% change
	0.2 mg/kg, p.o. (N)	% change	0.5 mg/kg, p.o. (N)	% change	2.0 mg/kg, p.o. (N)	70 change
48a	53.2±1.9 (5)	, , , , , , , , , , , , , , , , , , , ,	53.2±1.9 (5)	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	55.8 ± 3.4 (5)	
	$72.4 \pm 2.1 (4)$	36 ^{d)}	66.3 ± 3.2 (4)	25°)	85.1 ± 1.2 (4)	53 ^{d)}
57b	$50.8 \pm 2.9 (5)$		50.8 ± 2.9 (5)		50.8 ± 2.9 (5)	
55	77.1 ± 0.3 (4)	52 ^{d)}	81.3 ± 2.1 (4)	60 ^{d)}	84.1 ± 3.7 (4)	66 ^d)
1b ^{e)}	52.5±2.6 (5)		49.3±3.6 (5)		51.8 ± 2.1 (5)	
	75.4 ± 2.1 (4)	44 ^{d)}	$72.0 \pm 3.4 (4)$	46 ^d)	83.6 ± 2.4 (4)	61 ^{d)}

a) Each value represents the mean ± S.E.M. b) Number of rat used. A statistically significant difference from the control group; c) p < 0.05; d) p < 0.01 (Duncan's multiple range test). e) The figures shown are the data for the 1b hydrochloride.

Table IV. Effect of Selective 2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides on Gastric Emptying of Phenol Red Semisolid Meal in Mice and Resin Pellet Solid Meal in Rats

	Gastric emp	tying rate of phe	nol red semisolid meal in	mice ^{a)}	Gastric emptying rate of resin pelle meal in rats ^{a)}			
Compd.	Control $(N^{b)}$)	0/ ahanaa	Control (N)	0/ 1	Control (N)			
	$0.5 \mathrm{mg/kg}, p.o. (N)$	% change	1.0 mg/kg, p.o. (N)	% change	2.0 mg/kg, p.o. (N)	% change		
48a	50.3 ± 3.0 (14)		50.3±3.0 (14)		32.6±2.6 (5)			
	$62.8 \pm 4.9 (5)$	25	$80.9 \pm 3.8 (5)$	61 ^d)	22.5 ± 3.5 (4)	31°)		
57b	52.5 ± 2.1 (7)		52.5 ± 2.1 (7)	•	32.6 ± 2.6 (5)			
	$87.8 \pm 3.1 (5)$	67 ^{d)}	94.2 ± 3.8 (5)	79 ^{d)}	13.0 ± 2.2 (4)	60 ^d)		
1b	$53.1 \pm 4.1 \ (7)^{e}$		$52.7 \pm 2.1 \ (7)^{e}$		27.0 ± 2.7 (5)			
	$76.7 \pm 4.1 (5)$	44 ^d)	72.0 ± 1.9 (5)	37 ^{d)}	$5.8 \pm 2.1 (4)$	79 ^{d)}		

a) Each value represents the mean \pm S.E.M. b) Number of rat used. A statistically significant difference from the control group; c) p < 0.05; d) p < 0.01 (Duncan's multiple range test). e) The figures shown are the data for the 1b-hydrochloride.

TABLE V. Dopamine D₂ Receptor Antagonistic Activity of Selective 2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides

Compound	[³ H]Spiperone binding (μ M)	Apomorphine-induced emesis % inhibitory activity (3.0 mg/kg, p.o.)
48a	>1	7
57b	>1	0
1b	>1	0
Metoclopramide	$0.63^{a)}$	100 ^{b)}

a) The figure shown indicates the IC $_{50}$ value, which is obtained by log-logit regression analysis. b) The ED $_{50}$ value is 0.45 mg/kg, p.o.

(56a-58a) and naphthyl (64a) rings caused a slight decrease in activity. Furyl (59a, 60a), thienyl (61a, 62a), and benzisoxazolyl (63a) rings generally are deleterious to activity, although such heteroaryl rings except the furyl ring conferred slightly more active derivatives than metoclopramide in the gastric emptying activity. Overall, the decreasing order of contribution to activity is thus phenyl > pyridyl ≥ naphthyl > benzisoxazolyl ≥ thienyl > furyl and, hence, 6-membered > bicyclic > 5-membered heteroaryl rings. In series b, on the other hand, compounds 56b-58b with a pyridyl ring are more potent than metoclopramide and even cisapride. In particular, compound 57b, appending a 3-pyridylmethyl group, has a comparable activity to that of 1b (AS-4370) which is the most potent compound in our previous study.1) In a comparison between the regioisomeric pyridyl derivatives 56—58, the 3-pyridyl analogues (57a, 57b) are very slightly more potent than the 2- and 4-pyridyl (56a, 56b, 58a, 58b) isomers.

In light of the gastric emptying activity, eight compounds (15b—18b, 20b, 48a, 57b, 58b) were selected and subjected to the acute toxicity test in mice (Table II). The N-4 alkyl (15b—18b, 20b) derivatives show a potent acute toxicity. The 3-(4-chlorophenoxy)propyl (48a), 3- and 4-pyridylmethyl (57b, 58b) derivatives, however, are weak in acute toxicity.

Compounds 48a and 57b, which showed potent gastric emptying activity as well as weak acute toxicity, were

therefore selected for further biological assays including the gastric emptying rates of the phenol red semisolid meal in mice and rats and of resin pellet solid meal in rats at different doses (Tables III and IV). Overall, compound 57b compares very favorably with 1b, whereas compound 48a is somewhat less potent than 1b.

The activity for dopamine D_2 receptor antagonism was also determined by the [3H]spiperone binding test and the suppression of apomorphine-induced emesis in dogs (Table V). All the selected compounds show no dopamine D_2 receptor antagonistic activity (the [3H]spiperone binding test at the concentration of 10^{-6} M and antagonism of the apomorphine-induced emesis at an oral dose of $3.0 \, \text{mg/kg}$). On the contrary, metoclopramide shows potent dopamine D_2 receptor antagonistic activity. As for compound 48a, unfortunately, some unfavorable actions on the central nervous system, such as catalepsy and ptosis, were observed at an oral dose of $100 \, \text{mg/kg}$ in mice $^{5)}$; a similar side effect has been observed for metoclopramide and cisapride at an oral dose of $10 - 100 \, \text{mg/kg}$.

In summary, most compounds (52 of 70 compounds examined in the present study) displayed a gastric emptying activity superior to that of metoclopramide, and eleven compounds having alkyl, 4-substituted phenoxyalkyl, or pyridylmethyl group at N-4 were more potent than cisapride. The SAR study, including acute toxicity correlation, of the N-4 substituent of 1b revealed that the 3-pyridylmethyl group was optimal. 4-Amino-5-chloro-2-ethoxy-N-[[4-(3-pyridylmethyl)-2-morpholinyl]methyl]-benzamide (57b), on the whole, was found to possess the most favorable activity profile of gastric emptying enhancement without the dopamine D₂ receptor antagonistic activity and is worthy of further biological evaluation as a potent and selective gastrokinetic agent.

Experimental

Chemistry All melting points were determined on a Yanagimoto micromelting point apparatus and are uncorrected. IR spectra were recorded on a Hitachi 260-10 spectrometer with KBr disks and electron-impact mass (EIMS) spectra were recorded on a JEOL JMS D-300. $^1\text{H-NMR}$ spectra were taken at 200 MHz with a Varian GEMINI-200 spectrometer and at 300 MHz with a Varian XL-300 spectrometer. Chemical shifts are expressed as δ (ppm) values with

tetramethylsilane as an internal standard, and coupling constants (J) are given in hertz (Hz). Organic extracts were dried over anhydrous MgSO₄. The solvent was evaporated under reduced pressure. Merck Kieselgel 60 was used for column chromatography.

The following known alkyl and aralkyl halides were prepared according to the cited literature: 3-cyclohexenylmethyl bromide, ⁷⁾ 2-(4-fluorophenoxy)ethyl chloride, ⁸⁾ 3-(4-fluorophenoxy)propyl chloride, ⁸⁾ 3-(3-fluorophenoxy)propyl chloride, ⁸⁾ 4-(4-fluorophenoxy)butyl bromide, ⁸⁾ 5-(4-fluorophenoxy)pentyl bromide, ⁸⁾ 6-(4-fluorophenoxy)hexyl bromide, ⁸⁾ 2-(4-chlorophenoxy)ethyl chloride, ⁸⁾ 3-(4-chlorophenoxy)propyl bromide, ⁸⁾ 3-(4-nitrophenoxy)propyl bromide, ⁸⁾ 3-(4-fluorophenoxy)propyl bromide, ⁸⁾ 3-chloromethylfuran, ¹⁰⁾ 2-chloromethylthiophen, ¹¹⁾ 3-bromomethylthiophen, ¹²⁾ and 3-benzisoxazolylmethyl bromide. ¹³⁾

The preparation of compounds 11a and 38a was reported in the previous paper. $^{1)}$

2-Alkoxy-4-amino-5-chloro-N-[(4-substituted 2-morpholinyl)methyl]benzamides (12—37, 39—51, and 53—65); Method A. 4-Amino-5-chloro-2-ethoxy-N-[[(4-pyridylmethyl)-2-morpholinyl]methyl]benzamide (58b) A solution of 2-[(acetylamino)methyl]-4-benzylmorpholine²⁾ (3, 10.0 g, 40 mmol) in an EtOH (70 ml)-AcOH (5 ml) mixture was hydrogenated over 10% palladium on carbon (0.5 g) at 60 °C. After the theoretical amount of hydrogen was absorbed, the catalyst was removed by filtration. The filtrate was concentrated to dryness, giving ca. 7 g of [2-(acetylamino)methyl]morpholine acetate (4).

A mixture of crude 4 (7g), 4-picolyl chloride hydrochloride (7.9g, 48 mmol), K_2CO_3 (55.6g, 397 mmol), KI (1g), and methyl ethyl ketone (100 ml) was heated to reflux for 17h and cooled to room temperature. The insoluble materials were removed by filtration, and the filtrate was concentrated to dryness. The residue was diluted with water and extracted with CHCl₃. The extract was washed with brine. The solvent was evaporated to give a solid, which was recrystallized from toluene to afford 9.4g of 2-[(acetylamino)methyl]-4-(4-pyridylmethyl)morpholine [5 (R_1 =4-pyridylmethyl)].

A solution of 5 (2.9 g, 12 mmol) in 10% HCl (60 ml) was heated to reflux for 4h and cooled to room temperature. The reaction mixture was basified with 10% NaOH and then extracted with CHCl₃. The extract was washed successively with water and brine. The solvent was evaporated to give 2.0 g of 6 ($R_1 = 4$ -pyridylmethyl) as an oil. EIMS m/z: 195 (M⁺). To a stirred solution of 6 (2.0 g, 10 mmol) in CH₂Cl₂ (50 ml) were added 4-amino-5-chloro-2-ethoxybenzoic acid1) (2b, 2.1 g, 10 mmol) and 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (1.9 g, 10 mmol). The mixture was stirred at room temperature for 4h and then washed successively with water, 10% NaOH, water, and brine. The organic phase was dried and concentrated to dryness. The residue was chromatographed on silica gel with CHCl₃-MeOH (9:1) to give a solid, which was recrystallized from iso-PrOH to afford 2.5 g (53%) of 58b. ¹H-NMR (200 MHz, CDCl₃): 1.48 (3H, t, J=6, OCH₂CH₃), 2.06 (1H, t, J=10.5, $3-H_{ax}$), 2.21 (1H, td, J=11, 3.5, $5-H_{ax}$), 2.65 (1H, dd, $J=11, 1, 5-H_{eq}$), 2.78 (1H, d, $J=11, 3-H_{eq}$), 3.38 (1H, ddd, J=14, 8, 3, CONHCH), 3.51 (2H, d, J=3, CH₂C₅H₄N), 3.6—3.8 (3H, m, CONHCH, 2-H, and 6-H_{ax}), 3.85 (1H, ddd, J=11.5, 3.5, 2, 6-H_{eq}), 4.07 (2H, q, J=6, 15, OC \underline{H}_2 CH₃), 4.39 (2H, br s, NH₂), 6.28 (1H, s, arom 3-H), 8.10 (1H, s, arom 6-H), 7.28 (2H, d, J=5, C_5H_4N), 8.55 (2H, dd, J=4, 1.5, C_5H_4N), 8.22 (1H, t, J=6, CONH). EIMS m/z: 404 (M⁺), 312 (M⁺ -92). IR ν cm⁻¹: 3355, 3300, 3190, 1620, 1510.

Method B. General Procedure In a manner similar to that described for the preparation of 2-[(acetylamino)methyl]-4-(4-pyridylmethyl)morpholine [5 (R_1 =4-pyridylmethyl)], 3 (10.0 g, 40 mmol) was converted into 4-substituted 2-[(acetylamino)methyl]morpholines 5. A solution of 5 (10 mmol) in 10% NaOH (60 ml) was heated to reflux for 20 h and cooled to room temperature. The reaction mixture was extracted with CHCl₃, and the combined organic layers were washed successively twice with water and brine. The organic phase was dried and evaporated to give 4-substituted 2-(aminomethyl)morpholines 6 as an oil. The reaction of 4-substituted 2-(aminomethyl)morpholines with 4-amino-5-chloro-2-methoxy- or 2-ethoxybenzoic acid (2a or 2b) was performed according to method A.

Method C. 4-Amino-5-chloro-2-methoxy-N-[(4-methyl-2-morpholinyl)-methyl]benzamide (12a) A mixture of 2-(methylamino)ethanol (7o, R_1 =CH₃, R_2 =H, 5.0 g, 67 mmol), N-(2,3-epoxypropyl)phthalimide (8, 16.2 g, 80 mmol) was stirred at 80 °C for 1h. To the reaction mixture consisting of the resultant diol 9o (R_1 =CH₃, R_2 =H) was gradually added concentrated H₂SO₄ (39.2 g, 40 mmol), and the mixture was

rapidly heated to 150 °C and kept at the same temperature for 2h. The resulting brown solution was cooled, poured into ice-water, and washed with CHCl₃. The aqueous layer was basified with 25% NaOH, and then extracted with CHCl₃. The extract was washed successively with water and brine. The solvent was evaporated to give 1.3 g of 10o as an oil, which was used in the next step without further purification. The reaction of 10o with 4-amino-5-chloro-2-methoxybenzoic acid (2a) was performed according to method A to give 12a. ¹H-NMR (200 MHz, dimethylsulfoxide (DMSO)- d_6): 2.77 (3H, d, J=4, NCH₃), 2.9—3.2 (1H, m), 3.2—3.6 (4H, m), 3.7—4.1 (4H, m), 3.85 (3H, s, OCH₃), 6.00 (2H, br s, NH₂), 6.49 (1H, s, arom 3-H), 7.70 (1H, s, arom 6-H), 8.09 (1H, t, J=6 CONH). EIMS m/z: 313 (M⁺). IR ν cm⁻¹: 3340, 3290, 3190, 1620, 1530.

4-Amino-5-chloro-2-ethoxy-N-[3-(octahydropyrido[2,1-c][1,4]oxazinyl)methyl]benzamide (23b) 3-(Aminomethyl)octahydropyrido[2,1-c][1,4]oxazine 10q was prepared from 2-(hydroxymethyl)piperidine (7q, R_1 , R_2 =-(CH₂)₄-) and N-(2,3-epoxypropyl)phthalimide 8 according to the same method. ¹H-NMR (300 MHz, CDCl₃): 1.10 (1H, dddd, J=12.8, 12.4, 10.6, 3.8, 9-H_{ax.}), 1.30 (1H, m, 8-H_{ax.}), 1.45 (1H, dd, J=12.8, 2.6, 9-H_{eq.}), 1.50 (3H, t, J=6.9, OCH₂CH₃), 1.53—1.68 (2H, m, 7-H), 1.79 (1H, m, 8-H_{eq.}), 1.98 (1H, dddd, J=10.6, 10.2, 3.1, 2.6, 10-H), 2.04 (1H, dd, J=11.4, 11.2, 6-H_{ax.}), 2.09 (1H, dd, J=11.4, 10.3, 4-H_{ax.}), 2.72 (1H, dd, J=11.4, 2.2, 4-H_{eq.}), 2.79 (1H, dddd, J=11.4, 3.8, 3.6, 1.4, 6-H_{eq.}), 3.31 (1H, dd, J=11.2, 10.2, 1-H_{ax.}), 3.34 (2H, s, NH₂), 3.36 (1H, ddd, J=13.8, 6.1, 3.6, CH₂), 3.69 (1H, ddd, J=13.8, 7.3, 4.3, CH₂), 3.71 (1H, dd, J=11.2, 3.1, 1-H_{eq.}), 3.78 (1H, m, 3-H), 4.08 (2H, q, J=6.9, OCH₂CH₃), 6.27 (1H, s, arom 3-H), 8.11 (1H, s, arom 6-H), 8.23 (1H, dd, J=6.1, 4.3, CONH). EIMS m/z: 367 (M⁺). IR v cm⁻¹: 3450, 3360, 3280, 3150, 1620, 1525.

4-Amino-N-[[4-[3-(4-aminophenoxy)propyl]-2-morpholinyl]methyl]-5-chloro-2-methoxybenzamide (52a) A solution of compound 51a (1.7 g, 3.5 mmol) in MeOH (50 ml) was hydrogenated over 5% palladium on carbon (0.3 g) at room temperature. The catalyst was removed by filtration and the filtrate was evaporated to give 0.8 g (51%) of 52a as an oil. This oil was converted to the oxalate in the usual manner. 1 H-NMR (200 MHz, DMSO- d_6): 3.83 (3H, s, OCH₃), 5.99 (2H, br s, 4-NH₂), 6.49 (1H, s, arom 3-H), 7.70 (1H, s, arom 6-H), 6.54 and 6.73 (each 2H, d, J=9, OC₆H₄NH₂), 8.05 (1H, t, J=6, CONH). EIMS m/z: 448 (M⁺). IR ν cm⁻¹: 3380, 3310, 1750, 1605, 1515.

Pharmacology Male mice of Std-ddY strain (Japan SLC Inc.) weighing 30—40 g and male rats of Wister strain (Japan SLC Inc.) weighing 130—150 g were used. The mice and rats were fasted for 18 h before the experiments.

Gastric Emptying of Semisolid Meal A test meal (0.05% phenol red (PR) in 1.5% aqueous methylcellulose solution) of 0.2 ml per mouse and 1.5 ml per rat was given with a gastric tube. Fifteen minutes later, the animals were sacrificed. The stomachs were removed, and the amount of PR remaining in them was measured according to the method of Scarpignato et al.¹⁴; the isolated stomachs were homogenated by 0.1 N NaOH solution (100 ml) and centrifuged. The resultant supernatant (5 ml) was deproteinized by 20% trichloroacetic acid solution (0.5 ml) and centrifuged. Thereafter, the supernatant (3 ml) obtained was added to 0.5 N NaOH solution (4 ml). The PR contents were determined by measuring the absorbance at 560 nm wave length. The gastric emptying rate was calculated according to the following equation. The test compounds, suspended in a 0.5% tragacanth solution, were orally administered 60 min before administration of the test meal.

the rate (%)=

$$\left(1 - \frac{\text{stomach PR contents 15 min after PR solution}}{\text{stomach PR contents immediately after PR solution}}\right) \times 100$$

Gastric Emptying of Solid Meal Gastric emptying of solid meal (resin pellets) was measured according to the method of Jacoby. ¹⁵⁾ Small resin pellets (Amberlite IRA-93, 1-mm diameter, 40 pellets per rat) were administered through a polyethylene tube (PE-200) into the stomach. One hour later, the animals were sacrificed and the number of pellets remaining in the stomach was counted. The test compounds were orally administered 30 min before administration of the resin pellets.

Inhibition on Apomorphine-Induced Emesis in Dogs The antiapomorphine test in dogs was carried out according to the method of Janssen¹⁶) with modification. Male beagle dogs, weighing 10—16 kg, were used. Groups of three to six dogs received a subcutaneous injection of apomorphine hydrochloride (0.3 mg/kg) 2 h after pretreatment with test compounds. The frequency of emesis was then counted for 1 h.

Acute Toxicity Male ddY mice, weighing 18—25 g, were used in groups of 10 animals each. The test compounds, dissolved or suspended in a 0.5% tragacanth solution, were administered at an oral dose of 1.0 g/kg to the animals. The mortality was observed for 7 d after the administration.

Inhibition of [3 H]Spiperone Binding The test compounds at concentrations of $0.01-1000\,\mu\text{M}$ were tested in binding assays using rat brain synaptic membranes for competition with [3 H]spiperone in the striatum. 17 The assay was started by the addition of tissue preparations (10 mg wet tissue) and terminated by rapid filtration through Whatman GF/B glass-fiber filters under reduced pressure. The filters were washed two or three times with 5 ml of ice-cold buffer and transferred to scintillation vials that contained 1 ml of Soluene-350. After 1 h of incubation at 25 °C, the solubilized filters were shaken vigorously with 10 ml of toluene scintillator, and the radioactivity in the filters was counted with a Packard Tris-carb scintillation counter (B-2450).

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