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VOLTAMMETRIC METHOD AS A TOOL IN DETECTING SURFACTANT-INDUCED DISRUPTION OF A LANGMUIR-BLODGETT FILM

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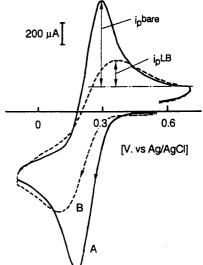
Tetsuo KAWAGUCHI, Yuji YAMAUCHI, Hatsuo MAEDA and Hidenobu OHMORI* Faculty of Pharmaceutical Sciences, Osaka University, 1-6 Yamadaoka Suita, Osaka 565, Japan

The interaction of surfactants with a stearic acid LB membrane on a glassy-carbon electrode is shown to be conveniently evaluated by cyclic voltammetry. The membrane disrupting effect of surfactants depended on their structure: the length of hydrophobic carbon chain and the charge of hydrophilic group.

KEYWORDS voltammetry; LB membrane; stearic acid; membrane disruption; surfactant

The interaction of surfactants with lipid membranes has received considerable attention from the standpoint of enhancement in percutaneous drug absorption¹⁾ and solubilization of membrane components.²⁾ In order to investigate such interaction, it will be worthwhile to develop a simple procedure for evaluating the membrane-disrupting ability of surfactants,3) where artificial lipid membranes, often employed as models for biomembranes, can be used conveniently. In this paper, we approach the subject by an electrochemical method based on the concept similar to that reported for ionchannel sensors.⁴⁾ Stearic acid Langmuir-Blodgett(LB) membrane transferred onto a glassy-carbon (GC) plate electrode as Y-type trilayer⁵⁾ was used as the model membrane, and the voltammetric behavior of Fe(CN)₆³- at the electrode was examined.

Figure 1 shows the cyclic voltammograms of Fe(CN)₆³- at the GC electrode with and without the stearic acid LB membrane. The distorted voltammetric wave at the coated electrode indicates that the LB membrane inhibits the access of Fe(CN)63- to the electrode surface. As reported for the voltammetry of Fe(CN)₆³- at membrane-coated electrodes,⁶) the ratio $R(=i_p^{LB}/i_p^{bare})$ of anodic currents (see Fig.1) for the coated electrode will represent the degree of this inhibition. If the structure of the lipid membrane is disrupted by a surfactant, the flux of $Fe(CN)_6^{3-}$ to the electrode surface will be enhanced and the shape of the Fig. 1. Cyclic Voltammograms of 1mM Fe(CN)6³voltammogram will be restored to that observed at the Coated Electrode(B) in 10mM Acetate Buffer (pH 6) bare electrode, i.e., the R value will approach 1.0. Thus, the membrane-coated electrode was immersed in



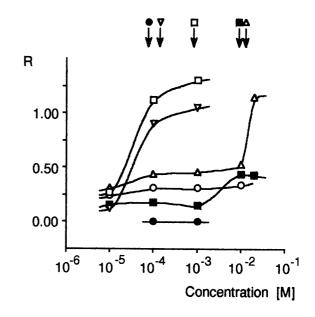
at the Bare GC Electrode(A) and the Membrane-Containing 50mM K2SO4 at 15 °C (Voltage scan rate, $0.5Vs^{-1}$)

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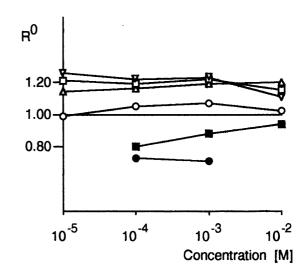
a solution containing one of the following surfactants for 5 min and then rinsed with distilled water for 10 s : $CH_3(CH_2)_7N(CH_3)_3Br$ (1a), $CH_3(CH_2)_{11}N(CH_3)_3Br$ (1b), $CH_3(CH_2)_{15}N(CH_3)_3Br$ (1c), CH₃(CH₂)₁₇N(CH₃)₃Cl (1d), CH₃(CH₂)₁₁SO₄Na (2), CH₃(CH₂)₁₁(OCH₂CH₂)₂₃OH (3). All procedures were performed at 15 °C.

The R values obtained by the coated electrode after the treatment with surfactants(1~3) are shown in Fig. 2. Among the surfactants with hydrophobic C12-alkyl chain in common [a cationic(1b), an anionic(2), and a nonionic(3) surfactant], the cationic one (1b) induced a significant increase in the R value, indicating that disruption of the stearic acid LB membrane is brought about by the surfactant(1b) at the concentration near 10⁻²M. However, greatly pronounced disrupting effects were exhibited by cationic surfactants(1c and 1d) with hydrophobic alkyl chaines (C16 and C18) similar to that of stearic acid (C17). It can be seen that the disrupting effects are not directly correlated to the critical micelle concentration⁷⁾ of the surfactants (vertical arrows in Fig. 2). The structure dependence, observed by the present method, of the ability of surfactants to disrupt a lipid membrane is comparable to that of the effectiveness of saturated straight fatty acids⁸⁾ and alkane sulfonates⁹⁾ to enhance percutaneous drug absorption.

The R values for the cationic surfactants with C12-, C16-, and C18-alkyl chain(1b~d) at their highest concentration employed were unexpectedly beyond unity. To identify the origin of this phenomenon, the bare GC electrode was treated with surfactants by the same method as described for the membrane coated electrode, and the voltammograms of Fe(CN)63- were obtained by the treated electrode. As seen in Fig. 3, the Fig. 3. surfactants(1a~d), where $i_p{}^s$ represents the (o)(1a), (Δ)(1b), (\square)(1c), (∇) (1d), (\square)(2), (Φ)(3). corresponding anodic current of Fe(CN)₆³-



Effects of Surfactants on the Cyclic Voltammograms of Fe(CN)₆³⁻ at the GC Electrode Coated with Stearic Acid LB Membrane [in 10mM Acetate Buffer (pH 6) Containing 50mM K2SO4] Vertical arrows represent the c.m.c. values of the corresponding surfactants. The c.m.c. value of 1a is not shown. (a)(1a), (b), $(\Box)(1c), (\nabla)(1d), (\Box)(2), (\bullet)(3).$



Effects of Surfactants on the Cyclic $R^0(=i_p^s/i_p^{bare})$ values for the cationic Voltammograms of Fe(CN)6³⁻ at the Bare Electrode [in 10] mM Acetate Buffer (pH 6) Containing 50mM K2SO4]

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(cf. Fig. 1), were all larger than unity and essentially independent of the surfactant concentration. In this case, the shape of the voltammogram became sharper than that at the untreated bare electrode: the cathodic and the anodic peaks were shifted to positive and negative directions, respectively. On the other hand, the R⁰ values were less than unity for the anionic and nonionic surfactants. These results suggest that the surfactants are adsorbed on the electrode surface, and that those with cationic hydrophilic group enhance the access of Fe(CN)6³⁻ to the electrode by electrostatic attraction while those with nonionic or anionic group inhibit the access by steric and/or electrostatic repulsion. When the LB membrane on the coated electrode is disrupted by a cationic surfactant, similar adsorption of the surfactant can be expected and the R value will then exceed unity.

Although the observed effects of surfactants on the stearic acid LB membrane (Fig. 2) should be explained by considering the adsorption of the surfactants (Fig. 3), we believe that the present electrochemical method will serve as a useful tool to investigate the interaction between surfactants and lipid membranes. Application of the present electrochemical method to other lipid membranes and markers other than $Fe(CN)_6^{3-}$ is in progress.

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