

6-GINGESULFONIC ACID, A NEW ANTI-ULCER PRINCIPLE, AND GINGERGLYCOLIPIDS A, B AND C, THREE NEW MONOACYLDIGALACTOSYLGlycerOLS, FROM ZINGIBERIS RHIZOMA ORIGINATING IN TAIWAN

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By monitoring the effects on HCl / ethanol-induced gastric lesions in rats, a new anti-ulcer principle named 6-gingesulfonic acid was isolated from *Zingiberis Rhizoma*, the dried rhizome of *Zingiber officinale* ROSCOE (cultivated and processed in Taiwan) together with three new monoacyldigalactosylglycerols named gingerglycolipids A, B and C. Their chemical structures were elucidated on the basis of chemical and physicochemical evidence. 6-Gingesulfonic acid showed more potent anti-ulcer activity than 6-gingerol and 6-shogaol.

KEYWORDS *Zingiberis Rhizoma*; *Zingiber officinale*; ginger; 6-gingesulfonic acid; gingerglycolipid; anti-ulcer effect; Zingiberaceae

Zingiberis Rhizoma (Ginger; Shoukyo in Japanese), the dried-rhizome of *Zingiber officinale* ROSCOE (Zingiberaceae), is one of the best known Chinese crude drugs and it has been investigated extensively in search of its bioactive principles.¹⁾ In particular, the pungent constituents, which are the principal ingredients of *Zingiberis Rhizoma*, have been the subjects of many investigations, and various pungent constituents have been characterized.^{1,2)} During the course of our screening to find biologically active constituents contained in *Zingiberis Rhizoma*, we have found several bioactive compounds showing anticathartic, antiserotonergic, and gastrointestinal motility enhancing effect.³⁾ Furthermore, by monitoring with anti-ulcer effects in rats, we have isolated four sesquiterpenes (β -sesquiphellandrene, β -bisabolene, *ar*-curcumene and α -zingiberene), 6-gingerol and 6-shogaol as active compounds in stomachic medication from the lipophilic portion of *Zingiberis Rhizoma* originating in Taiwan.⁴⁾

In continuing studies, we have examined the water-soluble portion of the *Zingiberis Rhizoma* and have isolated a new anti-ulcer principle named 6-gingesulfonic acid (1). This paper communicates the evidence which is consistent with the structure of 1. In addition, three new monoacyldigalactosylglycerols, gingerglycolipids A (5), B (6) and C (7), were chemically elucidated.

The MeOH extract of *Zingiberis Rhizoma* was partitioned into an AcOEt-Water mixture. Repeated separation of the water-soluble portion by ordinary-phase and reversed-phase column chromatography furnished 6-gingesulfonic acid (1, 0.0013% from the crude drug) as anti-ulcer principle of the water-soluble portion together with (+)-angelicoidenol-2-*O*- β -D-glucopyranoside⁵⁾ (4, 0.0014%), gingerglycolipids A (5, 0.0013%), B (6, 0.0014%) and C (7, 0.0014%).

6-Gingesulfonic acid (1), white powder, $[\alpha]_D - 1.0^\circ$ (MeOH), $C_{17}H_{26}O_6S$,⁶⁾ UV(MeOH): 221.5 (1110), 281.5 (1000), was shown by its IR spectrum to have hydroxyl (3590 cm^{-1}), carbonyl (1710 cm^{-1}) and sulfonic acid ($1209, 1171, 1046\text{ cm}^{-1}$) functions. The negative FAB-MS of 1 showed the quasimolecular ion peak at m/z 357 (M-H)⁻, while it showed the quasimolecular ion peaks at m/z 403 (M-H+Na+Na)⁺ (glycerol matrix+NaCl) and m/z 371 (M-H+Li+Li)⁺ (glycerol matrix+LiCl) in its positive FAB-MS. The ¹H-NMR spectrum⁷⁾ of 1 showed a sulfonic acid-bearing methine proton at δ 3.32 (dddd, $J=2,6,6,7\text{ Hz}$, 5-H) together with many other signals closely resembling those of 6-gingerol (2). Acetylation of 1 with Ac₂O-pyridine yielded the monoacetate (1a), $[\alpha]_D - 0.5^\circ$ (MeOH), $C_{19}H_{28}O_7S$. Comparison of the ¹³C-NMR spectra for 1 and 1a with those of 2 let us to presume the structure of 1. (Table I) Finally, the structure of 6-gingesulfonic acid (1) was determined by chemical correlation with 6-shogaol (3). Thus, treatment of 3 with NaHSO₃ in the presence of *t*-butyl perbenzoate in refluxing methanol furnished 1, in 90% yield.

Gingerglycolipid A (5), white powder, $[\alpha]_D +37.7^\circ$ (MeOH), $C_{33}H_{56}O_{14}$, has hydroxyl (3569 cm^{-1}) and ester carbonyl

(1734 cm^{-1}) groups, as shown by its IR spectrum. The positive FAB-MS spectrum of **5** showed the quasimolecular ion peaks at m/z 699 ($M+\text{Na}$)⁺ and m/z 683 ($M+\text{Li}$)⁺. Treatment of **5** with 2% NaOMe in MeOH liberated a glyceryldigalactoside and methyl linolenate.

Table I. ^{13}C -NMR Data of **1**, **1a** and **2**

	1	1a	2
C- 1	30.4	30.5	33.7
C- 2	44.7	44.6	47.2
C- 3	210.6	210.2	212.8
C- 4	45.9	45.6	52.1
C- 5	57.0	57.2	69.7
C- 6	32.9	33.0	39.1
C- 7	31.9	32.0	31.0
C- 8	27.8	28.0	27.1
C- 9	23.5	23.6	24.4
C-10	14.4	14.4	15.2
C- 1'	133.9	139.4	134.8
C- 2'	113.0	113.9	113.9
C- 3'	148.8	152.4	149.6
C- 4'	145.6	141.9	146.5
C- 5'	116.0	123.5	116.9
C- 6'	121.6	121.4	122.5
OMe	56.3	56.4	57.1

(δ_{C} at 67.5 MHz, in CD_3OD).

Table II.

Effect of 6-Gingesulfonic Acid (**1**), 6-Gingerol (**2**), 6-Shogaol (**3**) and Cetraxate⁴) on HCl / ethanol Induced Gastric Ulcer in Rats

Treatment	Dose (mg / kg)	N	Total length (mm) (Mean \pm S.E)	Inhibition (%)
Control		5	101.5 \pm 19.5	-
6-Gingerol (2)	150	5	43.1 \pm 9.2*	57.5
6-Shogaol (3)	150	5	30.2 \pm 5.5*	70.2
6-Gingesulfonic acid (1)	150	5	7.4 \pm 1.6**	92.7
6-Gingesulfonic acid (1)	300	6	0.5 \pm 0.3**	99.6
Cetraxate	300	5	1.5 \pm 1.1**	98.5

* $P < 0.05$, ** $P < 0.01$.

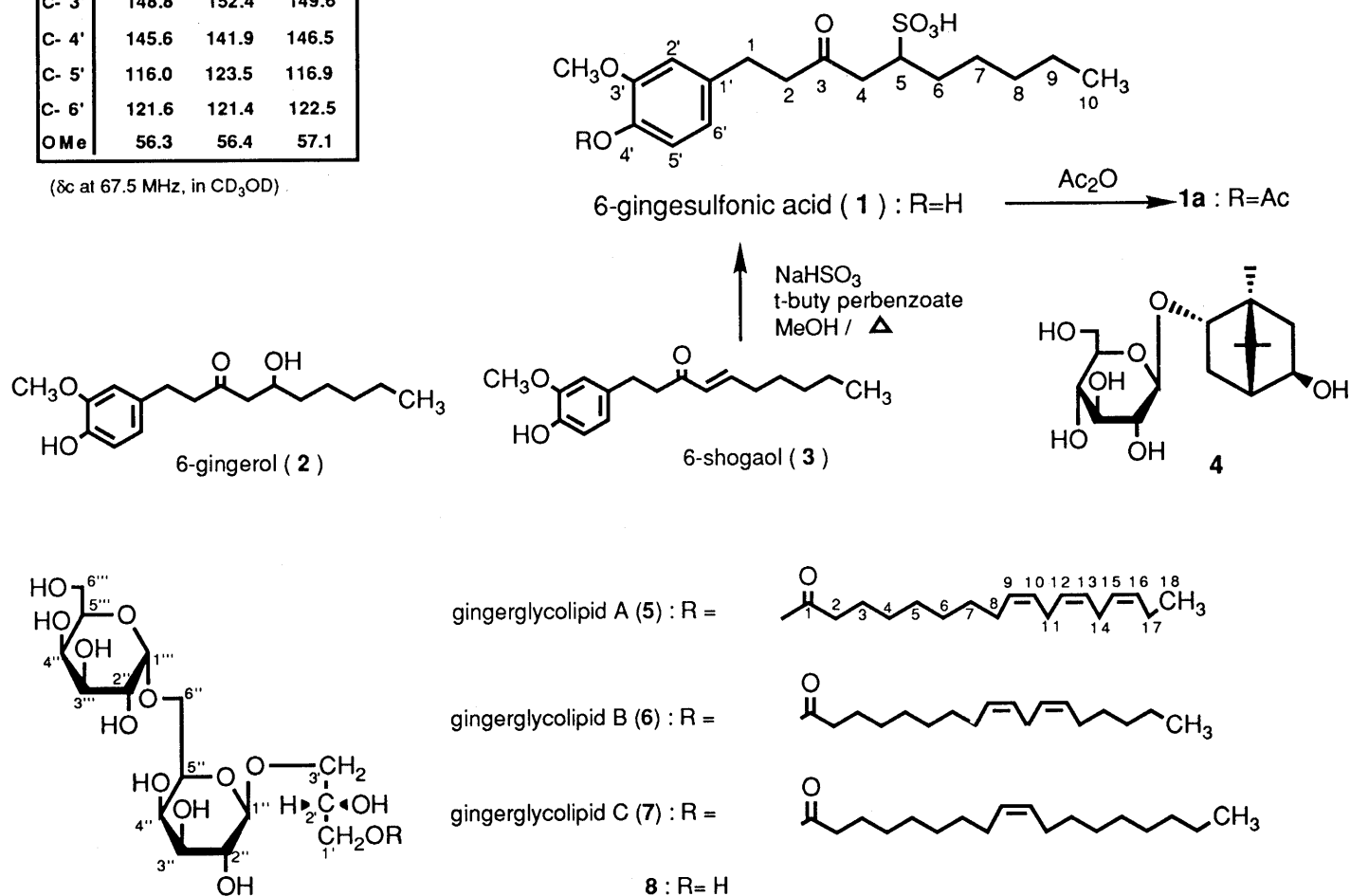


Chart 1

The glyceryldigalactoside, $[\alpha]_D + 80.0^\circ$ (H₂O), was shown to be identical in all respects with 3-*O*-(α -D-galactopyranosyl-(1-6)-*O*- β -D-galactopyranosyl-*sn*- glycerol (8)⁸). Comparisons in detail of ¹H-NMR and ¹³C-NMR data⁹) for **5** with those for **8** have shown that the fatty acid residue in **5** attach to C-1' in the glycerol moiety. Consequently, the chemical structure of gingerlycolipid A (**5**) was determined as shown.

The structures of the other two gingerlycolipids have been elucidated in the same way. Gingerlycolipid B (**6**), white powder, $[\alpha]_D + 50.9^\circ$ (MeOH), C₃₃H₅₈O₁₄, IR (KBr): 3590, 3569, 1719 cm⁻¹, showed quasimolecular ion peaks at *m/z* 701 (M+Na)⁺ and *m/z* 685 (M+Li)⁺ in its positive FAB-MS. The ¹H-NMR and ¹³C-NMR spectra¹⁰) of **6** closely resembled those spectra of gingerlycolipid A (**5**) expected for signals due to the fatty acid moiety. Treatment of **6** with NaOMe as carried out for **5** furnished the same glyceryldigalactoside (**8**) and methyl linoleate. Finally, the ¹H-NMR and ¹³C-NMR analysis of **6** in comparison with **5** have led to the formulation of gingerlycolipid B (**6**) as shown.

Alkaline treatment of gingerlycolipid C (**7**), white powder, $[\alpha]_D + 26.9^\circ$ (MeOH), C₃₃H₆₀O₁₄, IR (KBr): 3571, 1736 cm⁻¹. FAB-MS: *m/z* 703 (M+Na)⁺ and *m/z* 687 (M+Li)⁺, liberated **8** and methyl oleate. These findings together with ¹H-NMR and ¹³C-NMR data¹¹) for **7** led us to formulate gingerlycolipid C as **7**.

As given in Table II, 6-gingesulfonic acid (**1**) showed more potent anti-ulcer effect than 6-gingerol (**2**) and 6-shogaol (**3**). It is interesting to note that 6-gingesulfonic acid (**1**) exhibits weak pungency and higher water-solubility compared to **2** and **3**, and these property may be important for the use as a stomachic.

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- 5) S. Inoshiri, M. Saiki, H. Kohda, H. Otsuka and K. Yamasaki, *Phytochemistry*, **27**, 2869 (1988).
- 6) The molecular composition of the compound given with the chemical formula was determined either by elemental analysis or by high resolution mass spectrometry.
- 7) ¹H-NMR (270MHz, CD₃OD, δ) of **1**: 0.78 (t, *J*=7Hz, 10-H), 0.97-1.26 (m, 7, 8, 9-H), {1.34 (m), 1.80 (td, *J*=7, 11Hz), 6-H}, 2.41, 2.94 (both dd, *J*=6, 18Hz, 4-H), 2.70 (br s, 1, 2-H), 3.32 (dddd, *J*=2, 6, 6, 7Hz, 5-H), 6.51 (dd, *J*=2, 8Hz, 6'-H), 6.59 (d, *J*=8Hz, 5'-H), 6.68 (d, *J*=2Hz, 2'-H).
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- 9) **5**: ¹H-NMR (CD₃OD, δ): 0.97 (t, *J*=7Hz, 18-H), 2.35 (t, *J*=8Hz, 2-H), 4.14 (d, *J*=5Hz, 1''-H), 4.24 (d, *J*=7Hz, 1''-H). ¹³C-NMR (CD₃OD, δ c): 15.4 (C-18), 22.3, 26.7, 27.2, 27.3, 28.9, 31.0x2, 31.1, 31.5, 35.7 (methylene carbons), 63.5 (C-6'''), 67.4 (C-6''), 68.5 (C-3'), 70.4 (C-2'''), 70.8x2 (C-2',4'''), 71.0 (C-4''), 71.8 (C-3'''), 72.2 (C-5'''), 72.9 (C-1'), 73.3 (C-2''), 75.3x2 (C-3'',5''), 101.3 (C-1'''), 106.0 (C-1''), 129.0, 129.6, 130.0x2, 131.8, 133.5, 176.2 (C-1).
- 10) **6**: ¹H-NMR (CD₃OD, δ): 0.89 (t, *J*=7Hz, 18-H), 2.45 (t, *J*=7Hz, 2-H), 2.86 (m, 11-H), 4.23 (d, *J*=5Hz, 1'''H), 4.34 (d, *J*=7Hz, 1''-H). ¹³C-NMR (CD₃OD, δ c): 15.2 (C-18), 24.4, 26.8, 27.3, 29.0, 31.0x2, 31.1, 31.3, 31.5x2, 33.5, 35.7 (methylene carbons), 63.5 (C-6'''), 67.4 (C-6''), 68.5 (C-3'), 70.4 (C-2'''), 70.8x2 (C-2',4'''), 71.0 (C-4''), 71.8 (C-3'''), 72.2 (C-5'''), 72.9(C-1'), 73.3 (C-2''), 75.3x2 (C-3'',5''), 101.3 (C-1'''), 106.0 (C-1''), 129.9x2, 131.7x2, 176.2 (C-1).
- 11) **7**: ¹H-NMR (CD₃OD, δ): 0.90 (t, *J*=7Hz, 18-H), 2.36 (t, *J*=7Hz, 2-H), 4.15 (d, *J*=5Hz, 1'''-H), 4.25 (d, *J*=7Hz, 1''-H), 5.35 (td, *J*=11, 15Hz, 9, 10-H). ¹³C-NMR (CD₃OD, δ c): 15.5 (C-18), 24.6, 27.0, 29.0x2, 30.8x2, 31.0x2, 31.1, 31.3, 31.8x2, 34.0, 36.0 (methylene carbons), 63.5 (C-6'''), 67.4 (C-6''), 68.5 (C-3'), 70.4 (C-2'''), 70.8x2 (C-2',4'''), 71.0 (C-4''), 71.8 (C-3'''), 72.2 (C-5'''), 72.9 (C-1'), 73.3 (C-2''), 75.3x2 (C-3'',5''), 101.3 (C-1'''), 106.0 (C-1''), 131.5, 131.8, 176.4 (C-1).

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