SAUSSUREAMINES A, B, C, D, AND E, NEW ANTI-ULCER PRINCIPLES FROM CHINESE SAUSSUREAE RADIX

Masayuki YOSHIKAWA,* Shoko HATAKEYAMA, Yasuhiro INOUE, and Johji YAMAHARA Kyoto Pharmaceutical University, 5 Nakauchi-cho, Misasagi, Yamashina-ku, Kyoto 607, Japan

Five new amino acid-sesquiterpene adducts, saussureamines A, B, C, D, and E, were isolated from Chinese Saussureae Radix, the dried root of *Saussurea lappa* Clarke, together with a new lignan glycoside, (-)-massoniresinol 4"-O- β -D-glucopyranoside. Their structures were determined on the basis of chemical and physicochemical evidence. Among of these compounds, saussureamines A, B, C showed anti-ulcer effect on HCl / ethanol-induced lesions in rats, and saussureamine A also exhibited a inhibitory activity on stress-induced ulcer formation in mice. During the course of these studies, facile conversion from sesquiterpene having an α -methylene γ -lactone function to amino acid-sesquiterpene adduct has been accomplished by means of Michael type addition reaction.

KEYWORDS Saussureae Radix; Saussurea lappa; amino acid-sesquiterpene adduct; saussureamine; anti-ulcer activity; Michael type addition

Saussureae Radix (Mokko in Japanese), the dried root of *Saussurea lappa* CLARKE (Compositae), is known as an aromatic stomachic and frequently prescribed in Chinese and Japanese traditional medicine. It has also been used as an important fragrance. Chemical studies on Saussureae Radix originating in India have been carried out by many investigators, and the presence of various sesquiterpenes such as costunolide (1) and dehydrocostus lactone (2) from the less polar fraction of the Radix has so far been reported.¹⁾ However, no work has been reported on the chemical constituents of Chinese Saussureae Radix, which is now in common use in Japan. As a part of our characterization studies on bioactive constituents of naturally occurring crude drug,²⁾ we have reported the inhibitory effect of 1 on stress-induced ulcer formation and the gastrointestinal motility-enhancing effect of 2. ³⁾ In a continuing study, we have isolated five new amino acid-sesquiterpene adducts named saussureamines A (3), B (4), C (5), D (6), and E (7) from Chinese Saussureae Radix. This paper deals with the structure elucidation of these amino acid-sesquiterpene adducts and their effects against gastric ulcerations. In addition, a new lignan glycoside, (-)-massoniresinol 4"-O-β-D-glucopyranoside (10), was chemically elucidated.

The MeOH extract of Chinese Saussureae Radix was partitioned into AcOEt and water to furnish the AcOEt soluble portion⁴⁾ and the water phase. The water phase was further partitioned with 1-BuOH, and the 1-BuOH soluble portion was subjected to XAD-2, ordinary and reversed phase silica gel, and Sephadex LH-20 column chromatography to afford 3 (0.0011% from the crude drug), 4 (0.0021%), 5 (0.0006%), 6 (0.0001%), 7 (0.0002%) and 10 (0.0009%) together with picriside B (0.0059%), 5) syringin (0.0070%), 5) and (-)-olivil 4"-O- β -D-glucopyranoside (0.0004%).

Saussureamine A (3), colorless prisms, mp 115~117°C, $[\alpha]_D$ +36.7° (MeOH), $C_{20}H_{29}O_4N$, positive FAB-MS (m/z): 348 (M+H)⁺, was positive for Dragendorff reagent. The IR spectrum of 3 showed the presence of γ -lactone (1780 cm⁻¹), and carboxyl (3400, 1630 cm⁻¹) functions. The $^{1}H_{-}^{-6}$ and ^{13}C -NMR (Table I) spectra of 3 showed signals ascribable to

215 January 1993

E (3-	/)	(/SMHz, pyridine-d5, oc)					
	3	4	5	6	7		
C-1	126.7	48.3	44.6	74.4	77.0		
C-2	28.5	34.2	37.4	33.7	36.4		
C-3	39.5	31.6	32.2	122.4	33.8		
C-4	140.0	154.2	150.0	133.7	144.6		
C-5	128.5	53.8	51.6	50.1	48.3		
C-6	81.5	87.0	85.3	81.5	79.2		
C-7	48.7	48.7	46.7	50.9	52.7		
C-8	26.3	30.9	29.6	23.3	23.2		
C-9	41.1	39.7	38.3	35.5	31.9		
C-10	137.5	152.3	152.1	41.2	43.1		
C-11	47.7	45.6	46.4	45.7	46.0		
C-12	178.1	179.3	177.3	178.3	178.2		
C-13	52.0	53.6	46.3	52.5	52.0		
C-14	17.1	110.0	108.1	11.4	11.7		
C-15	16.0	112.6	110.9	23.7	109.1		
C-a	67.7	68.9	59.5	67.5	67.0		
C-β	29.5	34.1	32.0	29.6	29.3		
C-γ	24.2	25.6	173.5	24.1	23.8		
C-δ	54.5	55.7	-	54.1	53.8		
C=O	176.6	177.9	175.1	176.7	176.2		

Table I. 13C NMR Data for Saussureamines A- Table II. Effect of Saussureamines A-E (3-7) and Cetraxate on HCl / Ethanol Induced Gastric Ulcer in Rats

Treatment	Dose (mg / kg)	N	Total length (mm) (Mean ± S.E.)	Inhibition (%)
Control	-	8	72.4±9.0	•
Saussureamine A (3)	50	5	42.3±8.3*	41.6
	100	6	29.1±16.6*	59.8
Saussureamine B (4)	50	6	9.3±8.6**	87.2
	100	4	9.4±3.9**	87.1
Saussureamine C (5)	50	6	57.3±16.3	20.9
, ,	100	6	23.1±5.5**	68.1
Saussureamine D (6)	100	5	63.8±13.9	11.9
Saussureamine E (7)	100	5	63.7±16.9	12.0
Cetraxate	300	6	9.3±8.6**	87.2

*p<0.05, **p<0.01.

costunolide (1) and L-proline, but they lacked the exomethylene signals in 1, indicative of the structure 3 linked with L-proline at 13-position.

Acid treatment of 3 with 1% aq. HCl at room temperature (25°C) liberated 1 and L-proline. Furthermore, the NOE correlations were observed between the 11 β -H [δ 2.35 (m)] and $\delta\beta$ -H [δ 4.75 (dd, J=9, 9Hz)] in the different NOE experiment of 3; consequently, the

structure of saussureamine A was shown to be 3. Finally, treatment of 1 with L-proline in EtOH in the presence of Et3N (90°C, 1h) furnished 3 in 76% yield. Thus, it was inferred that the Michael type addition reaction for the α -methylene γ lactone part in 1 with L-proline proceeded stereoselectively to provide thermodynamically favored addition product 3 having the α-configurated L-proline side chain.⁷⁾

Saussureamine B (4), white powder, [α]_D -25.9° (MeOH), C₂₀H₂₇O₄N, IR (KBr, cm⁻¹): 3400, 1770, 1640, positive FAB-MS (m/z): 346 $(M+H)^+$, was positive for Dragendorff reagent and liberated dehydrocostus lactone (2) and L-proline by acid treatment. The Michael type addition reaction of 2 with L-proline under the same conditions as for 1 furnished 4 (70%). Based on ¹H-⁸) and ¹³C-NMR (Table I) examinations for 4 and observation of the NOE correlation between the 11β-H and 6β-H, the structure of saussureamine B (4) was determined. 9)

Saussureamine C (5), white powder, [α]D -17.2 ° (MeOH), C₁₉H₂₆O₅N₂, IR (KBr, cm⁻¹): 3200, 1760, 1675, 1640, positive FAB-MS (m/z): 363 $(M+H)^+$ was positive for Dragendorff reagent and in Ninhidrin test. Treatment of 2 with Lasparagine in 70% aq. EtOH in the presence of Et₃N (90°C, 2h) provided 5 (48%). Comparisons in detail of ¹H-¹⁰ and ¹³C-NMR (Table I) data for 5 with those for 2 and L-asparagine and NOE observation between the 11β-H and 6β-H in 5 led us to confirm the structure of saussureamine C (5) as shown.

Saussureamine D (6), colorless needles, mp 235~237°C (MeOH-H₂O), [α]D +13.3 ° (MeOH), C₂₀H₂₉O₅N, IR (KBr, cm⁻ 1): 3200, 1765, 1634, positive FAB-MS (m/z): 370 $(M+Li)^+$, negative FAB-MS (m/z): 362 $(M-H)^-$, positive for Dragendorff reagent, had a tertiary methyl, a vinyl methyl, a secondary hydroxyl and a trisubstituted olefin group together with y-lactone and L-proline moiety as indicated by its ¹H-¹¹) and ¹³C-NMR (Table I) data. Ordinary acetylation of 6 furnished the monoacetate (6a).¹²) Detailed comparisons of ¹H- and ¹³C-NMR data for 6 with those for 3, 4 and known eudesmanolides suggested the structure 6 linked with L-proline at 13-position in santamarin (8) which was isolated from Chrysantemum parthenium. 13) Furthermore, NOE correlations were observed between the proton pairs of 6 [1α-H / 5α-H; 14β-H₃ / 6β-H; 6β-H / 11β-H]. Finally, treatment of 8 with L-proline in EtOH in the presence of Et₃N provided 6 in 91% yield. Thus, the structure of saussureamne D (6) was substantiated.

The ¹H-¹⁴) and ¹³C-NMR (Table I) spectra of saussureamine E (7), colorless needles, mp 150~153°C (MeOH-H₂O), [α]D $+33.7^{\circ}$ (MeOH), C₂₀H₂₉O₅N, IR (KBr, cm⁻¹): 3200, 1755, 1630, positive FAB-MS (m/z): 364 (M+H)⁺, positive for Dragendorff reagent, were closely similar to those of 6 except for some signals due to the olefin moiety. Treatment of reynosin (9), which was isolated from Ambrosia confertiflora¹⁵) with L-proline under the same conditions as for 8, furnished 7 (82%). Based on this evidence and NOE observation between the 6B-H and 11B-H, the structure of saussureamine E (7) was clarified.

(-)-Massoniresinol 4"-O-β-D-glucopyranoside (10), white powder, [α]_D - 67.0° (MeOH), C₂₆H₃₄O₁₃, UV [MeOH, nm (ε)] : 228 (8500), 279 (4030), IR (KBr, cm⁻¹): 3200, 1600, 1510, positive FAB-MS (m/z): 577 (M+Na)⁺, furnished the hexaacetate (10a), white powder, [α]D - 27.5° (MeOH), C38H46O19, by ordinary acetylation. Methanolysis of 10 with 9%

216 Vol. 41, No. 1

HCl-MeOH liberated (-)-massoniresinol (11) 16) and methyl D-glucopyranoside. The 1 H- and 13 C-NMR signals 17) of 10 could be analyzed completely by use of 1 H- 1 H COSY, 1 H- 13 C COSY and COLOC experiments. The NOEs were observed between the following pairs of protons in 10: anomeric-H / 5"-H; 6"-H / 4a-H₂; 2-H / 6'-H. Based on this evidence and

Table III. Effect of Saussureamines A-E (3-7), Costunolide (1), and Dehydrocostus lactone (2) and Cimetidine on Stress-induced Gastric Ulcer in Mice

Treatment	Dose (mg / 1	N kg)	Total length (mm) (Mean ± S.E.)	Inhibition (%)
Control		7	3.00±0.00	- (70)
Saussureamine A (3)	100	7	1.71±0.18**	43.0
	200	7	1.29±0.42**	57.0
Saussureamine B (4)	200	9	2.33±0.29	22.3
Saussureamine C (5)	200	9	2.78±0.15	7.3
Saussureamine D (6)	200	10	2.30±0.30	23.3
Saussureamine E (7)	200	10	2.60±0.16	13.3
Costunolide (1)	200	9	2.56±0.24	14.7
Dehydrocostus lactone (2)	200	9	2.33±0.24	22.3
Cimetidine	150	10	0.90±0.31**	70.0

**p<0.01

comparison of ¹³C-NMR data for 10 with those for 11, the structure 10 was determined as shown.

The inhibitory activities of saussureamines A (3), B (4), C (5), D (6), and E (7) on HCl / ethanol-induced gastric lesions in rats and on the formation of gastric ulcer in restrained and water-immersed mice are summarized in Table II and III, respectively. Among compounds tested for their effect on HCl / ethanol-induced gastric lesions, 4 showed a more potent anti-ulcer effect than cetraxate, and the inhibitory effect of 3 and 5 was comparable to cetraxate. As given in Table III, 3 showed potent inhibitory effect on stress-induced gastric ulcer formation, while 4 and 6 were found to exhibit a weak inhibitory activity.

REFERENCES AND NOTES

1) R. S. Dhillon, P. S. Kalsi, W. P. Singh, V. K. Gautam, and B. R. Chhabra, Phytochemistry, 26, 1209 (1987), and the literatures sited therein. 2) M. Yoshikawa, S. Hatakeyama, K. Taniguchi, H. Matsuda, and J. Yamahara, Chem. Pharm. Bull., 40, 2239 (1992). 3)a) J. Yamahara, M. Kobayashi, K. Miki, M. Kozuka, T. Sawada, and H. Fujimura, Chem. Pharm. Bull., 33, 1285 (1985); b) J. Yamahara, T. Chisaka, Q. Huang, K. Kishi, H. Kobayashi, and Y. Kawahara, Phytothr. Res., 4, 160 (1990). 4) Costunolide (1, 1.070%), dehydrocostus lactone (2, 1.430%), α and β-costol mixture (0.005%), and (-)elema-1,3,11(13)-trien-12-ol (0.002%) were isolated from the AcOEt soluble portion. 5)a) The chemical and physicochemical properties of these compounds were identical with those of picriside B 5b) syringin, 5c) and (-)-olivil 4"-O-β-D-glucopyranoside, ^{5d)}: b) K. Nishimura, T. Miyase, A. Ueno, T. Noro, M. Kuroyanagi, and S. Fukushima, Chem. Pharm. Bull., 34, 2518 (1986); c) C. P. Falshaw, W. D. Ollis, and K. L. Ormand, Phytochemistry, 8, 913 (1969); d) T. Deyama, T. Ikawa, S. Kitagawa, and S. Nishibe, Chem. Pharm. Bull., 34, 4933 (1986). 6) The ${}^{1}H$ -NMR (pyridine-d₅) of 3: δ 1.34 (s, 14-H₃), $1.60 \text{ (s, } 15\text{-H}_3), 2.35 \text{ (m, } 11\text{-H)}, [3.22 \text{ (dd, } J=3,13\text{Hz}), 3.41 \text{ (dd, } J=6,13\text{Hz}), 13\text{-H}_2], 4.70 \text{ (m, } 1\text{-H)}, 4.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H)}, 1.60 \text{ (s, } 15\text{-H}_3), 1.60 \text{ (m, } 1\text{-H)}, 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H)}, 1.60 \text{ (m, } 1\text{-H)}, 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H)}, 1.60 \text{ (m, } 1\text{-H)}, 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H)}, 1.60 \text{ (m, } 1\text{-H)}, 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H)}, 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-H}), 1.75 \text{ (dd, } J=9,9\text{Hz, } 6\text{-Hz, } 6\text{-H$ 4.82 (m, 5-H). 7) a) C. R. Narayanan and N. K. Venkatasubramanian, J. Org. Chem., 33, 3156 (1968); b) M. Yoshikawa, Y. Okaichi, B. C. Cha, and I. Kitagawa, Tetrahedron, 46, 7459 (1990). 8) The ¹H-NMR (pyridine-d₅) of 4: δ 2.85 (m, 11-H), $[3.14 \text{ (dd, } J=4, 13 \text{ Hz)}, 3.38 \text{ (dd, } J=5,13 \text{ Hz)}, 13-\text{H}_2], 4.00 \text{ (dd, } J=9,9\text{Hz, } 6-\text{H)}, [4.70 \text{ (br s)}, 4.80 \text{ (br s)}, 14-\text{H}_2], [5.10 \text{ (br s)}, 14-\text{Hz}_2], [5.10 \text{ (br s)}, 14$ s), 5.37 (br s), 15-H₂]. 9)a) The plane structure of saussureamine B (4) is identical with that proposed for involucratin. 9b); b) Y. Li and Z. J. Jia, Phytochemistry, 28, 3395 (1989). 10) The ¹H-NMR (pyridine-d₅) of 5; δ 2.67 (m, 11-H), [3.23 (m), $3.45 \text{ (dd, } J=6, 13\text{Hz)}, 13\text{-H}_2], 3.96 \text{ (dd, } J=9.9\text{Hz, } 6\text{-H}), [4.29 \text{ (br s)}, 4.70 \text{ (br s)}, 14\text{-H}_2], [5.06 \text{ (br s)}, 5.31 \text{ (br s)}, 15\text{-H}_2].$ 11) The ${}^{1}\text{H-NMR}$ (pyridine-d₅) of **6**: δ 1.07 (s, 14-H₃), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 1.94 (s, 15-H₃), 2.70 (m, 11-H), [3.12 (dd, J=4, 13Hz), 3.48 (dd, J=6, 13Hz), 3.48 (dd, Hz), 13-H₂], 3.64 (m, 1-H), 4.07 (dd, J=10, 10Hz, 6-H), 5.36 (br s, 3-H). 12) 6a: white powder, $[\alpha]_D + 12.9^\circ$ (MeOH), C₂₂H₃₁O₆N, IR (KBr, cm-1): 3500, 1760, 1740, 1640, ¹H-NMR: 0.94 (s, 14-H₃), 1.86 (s, 15-H₃), 3.30 (ddd, *J*=5, 5, 12Hz, 11-H), [3.44 (dd, J=5, 13Hz), 3.17 (dd, J=5, 13Hz), 13-H₂], 3.99 (dd, J=10, 10Hz, 6-H), 4.97 (dd, J=7, 10Hz, 1-H), 5.21 (br s, 3-H), positive FAB-MS (m/z): 428 (M+Na)+. 13) A. R. Vivar, and H. Jimenez, Tetrahedron, 21, 1741 (1965). 14) The ¹H-NMR (pyridine-d₅) of 7: δ 1.03 (s, 14-H₃), 2.78 (ddd, J=5, 5, 12Hz, 11-H), [3.15 (dd, J=5, 13Hz), 3.48 (dd, J=5, 13Hz, 13-H₂], 3.65 (m, 1-H), 4.22 (dd, J=10, 10Hz, 6-H), [5.03 (br s), 5.40 (br s), 15-H₂]. 15) H. Yoshioka, W. Renold, N.H. Fischer, A. Higo, and T. J. Mabry, Phytochemistry, 9, 823 (1970). 16) Z. Shen, and O. Thean-der, Phytochemistry, 24, 364 (1985). 17) 10: 1 H-NMR (CD₃OD, δ) 2.95 (ABq, J=9, 12Hz, 4a-H₂), 3.82, 3.83 (s, O-Me), 5.00 (s, 2-H), 6.75 (d, J=8Hz, 5'-H), 6.79 (dd, J=2, 8Hz, 6'-H), 6.83 (dd, J=2, 8Hz, 6"-H), 7.02 (d, J=2Hz, 2', 2"-H), 7.09 (d, J=8Hz, 5"-H), $^{13}C=^{13}$ NMR: &c 40.2 (C-4a), 56.4, 56.7 (O-Me x 2), 62.5 (glu-6), 64.6 (C-3a), 74.8 (glu-4), 74.9 (glu-2), 75.0 (C-5), 77.8 (glu-3), 78.2 (glu-5), 82.1 (C-3), 82.3 (C-4), 86.1 (C-2), 102.9 (glu-1), 112.9 (C-2'), 115.5 (C-5'), 116.3 (C-2"), 117.7 (C-5"), 121.7 (C-6'), 124.2 (C-6"), 131.1 (C-1'), 133.8 (C-1"), 146.6 (C-4"), 147.2 (C-4'), 148.6 (C-3'), 150.3 (C-3").

(Received October 23, 1992)