

Physapruins A and B, Two New Withanolides from *Physalis pruinosa* BAILEY¹⁾

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Two new withanolides, physapruins A and B, were isolated from the methanolic extract of the fresh aerial parts of *Physalis pruinosa* (Solanaceae), and their structures have been determined to be 4 β ,14 α ,17 β ,20 β -tetrahydroxy-1-oxo-20*S*,22*R*-witha-2,5,24-trienolide (1) and 6 β -acetoxy-5 α -butoxy-14 α -17 β -20 β -trihydroxy-1-oxo-20*S*,22*R*-witha-2,24-dienolide (2) on the basis of spectral evidence.

Keywords *Physalis pruinosa*; Solanaceae; withanolide; physapruin A; physapruin B

In our previous papers,²⁾ we reported the structure elucidation of seven withanolides, physagulins A, B, C, D, E, F and G, isolated from *physalis angulata* L. As an extended search for withanolides from the same genus, we have now isolated two new withanolides from the fresh aerial parts of *Physalis pruinosa* and determined their structures.

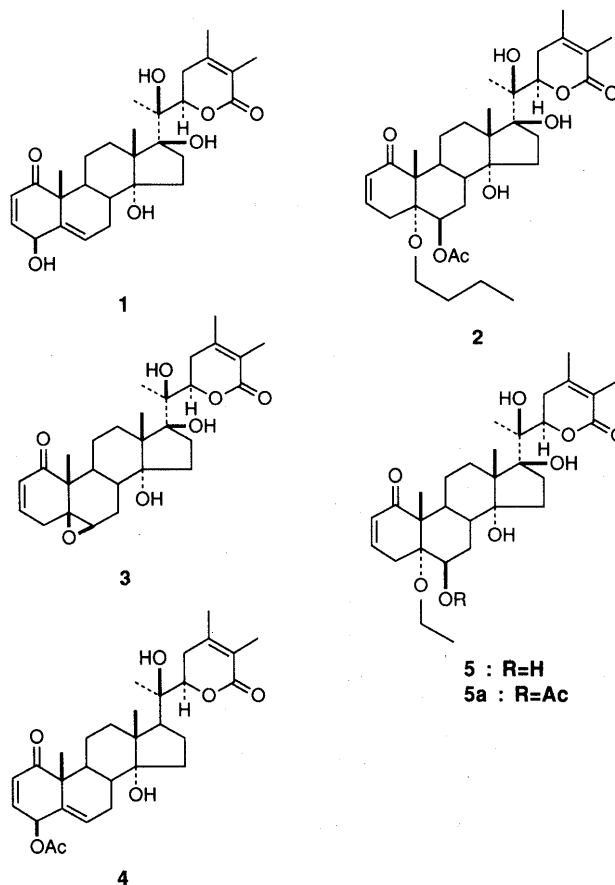
Physapruin A (1), obtained as colorless needles, with a mp of 207—209 °C and $[\alpha]_D^{25} +159.1^\circ$ (MeOH), exhibited the absorption bands at 3408 cm⁻¹ (hydroxyl), 1696 cm⁻¹ (α,β -unsaturated δ -lactone) and 1664 cm⁻¹ (α,β -unsaturated ketone) in the IR spectrum (KBr). The positive fast atom bombardment mass spectrum (FAB-MS) displayed peaks due to $[M+Na]^+$ at m/z 509 and the stepwise elimination of four hydroxy groups at m/z 469 $[M-H_2O+H]^+$, 451 $[M-2H_2O+H]^+$, 433 $[M-3H_2O+H]^+$ and 415 $[M-4H_2O+H]^+$. The mass fragments at m/z 125 and 165 were characteristic of side chain cleavage between C-20 and C-22 and between C-17 and C-20.³⁾ These peaks suggested that 1 was a 17,20-dihydroxywithanolide.

The ¹H-NMR spectrum, as listed in Table I, showed features typical of a withanolide. Five singlet signals at δ 1.13, 1.42, 1.45, 1.88 and 1.94 could be assigned to five methyl groups, and the signals of two olefinic methyl groups at δ 1.88 and 1.94, as well as the characteristic H-22 signal (dd, $J=10.8, 5.7$ Hz) at δ 4.92 suggested the presence of a typical α,β -unsaturated δ -lactone side chain. The coupling pattern and constants of the H-22 signal and the chemical shifts of two singlet methyl signals, H₃-21 and H₃-18 at δ 1.45 and 1.13, were indicative of a 14 α ,17 β ,20 β -trihydroxyl substitution by comparison with the analogous substituted withanolides, withanolide E (3) and related compounds.^{3,4)} The presence of these hydroxy groups at C-17 and 20 was supported by cleavage between C-17 and C-20, and between C-20 and C-22 in the FAB-MS.

On the other hand, in the low field, *cis* olefinic proton signals at δ 6.78 (1H, dd, $J=10.2, 4.5$ Hz) and 5.93 (1H, d, $J=10.2$ Hz), an olefinic proton signal at δ 5.92 (1H, d, $J=5.9$ Hz) and a hydroxymethine proton signal at δ 4.62 (d, $J=4.5$ Hz) could be assigned to H-3, H-2, H-6 and H_x-4, respectively. These signals suggested a 2,5-diene-1-one-4 β -hydroxyl system for rings A and B.

In the ¹³C-NMR spectrum of 1, as listed in Table II, signals for rings C and D and the side chain moiety were similar to those of 3. Moreover, the remaining signals due to rings A and B could be reasonably assigned, taking into

consideration the effect of an acetyl group, by comparison with those of withanolide U monoacetate (4).^{4,5)} Based on these spectral data and the positive Cotton effect curve at 256 nm⁶⁾ in the CD spectrum, the structure of 1 having a 22*R*-configuration could be expressed as shown in the formula.



Physapruin B (2), was isolated as an acetyl derivative, C₃₄H₅₀O₉, an amorphous powder, $[\alpha]_D^{25} +47.0^\circ$ (MeOH), in order to achieve easy separation. In the IR spectrum of 2, absorption bands appeared at 3452 cm⁻¹ (hydroxy), 1740 cm⁻¹ (acetyl carbonyl), 1714 cm⁻¹ (α,β -unsaturated δ -lactone) and 1690 cm⁻¹ (α,β -unsaturated ketone). The positive FAB-MS of 2 gave $[M+Na]^+$, $[M+H]^+$ and $[M-H_2O+H]^+$ ion peaks at m/z 625, 603 and 585, respectively, and peaks at m/z 165 and 125 indicated the

TABLE I. ¹H-NMR Data (δ /ppm, in CDCl₃) for Physapruin A (1), B (2), Withanolide E (3), and 5 α -Ethoxy-6 β ,14 α ,17 β ,20-tetrahydroxy-1-oxo-20S,22R-witha-2,24-dienolide Monoacetate (5a)

H	1	3	2	5a
2	5.93 d (10.2)	6.03 dq (10.0, 2.5, 1.0)	5.82 dd (10.0, 2.7)	5.82 dd (10, 3)
3	6.78 dd (10.2, 4.5)	6.87 dq (10.0, 5.5, 2.5)	6.43 ddd (10.0, 5.3, 2.0)	6.44 dq (10, 3)
4	4.62 d (4.5)		2.33 dd (19.7, 5.7) 2.55 d (19.7)	2.34 dd (22, 6) 2.55 br d (22, 4)
6	5.92 d (5.9)	3.20 m	5.15 d (2.7)	5.15 br s (W _{1/2} 8.2)
18	1.13 s	1.10 s	1.14 s	1.15 s
19	1.42 s	1.25 s	1.26 s	1.27 s
21	1.45 s	1.42 s	1.41 s	1.41 s
22	4.92 dd (10.8, 5.7)	4.88 m	4.93 m	4.94 br t (W _{1/2} 20)
27	1.88 s	1.88 s	1.88 s	1.88 s
28	1.94 s	1.93 s	1.94 s	1.94 s
-OAc			2.11 s	2.11 s
5 α -Butoxyl, ethoxyl			3.05 dd (14.2, 6.6) 3.28 dd (14.2, 6.1) 0.81 t (7.3)	3.14 dq (8, 7) 3.34 dq (8, 7) 1.05 t (7)

Chemical shifts are in δ /ppm; coupling constants in Hz are in parentheses; abbreviations: s=singlet; d=doublet; t=triplet; q=quartet; m=multiplet; br=broad.

TABLE II. ¹³C-NMR Data (δ /ppm, in CDCl₃) for Physapruin A (1), B (2), Withanolide E (3), Withanolide U Monoacetate (4) and 5 α -Ethoxy-6 β ,14 α ,17 β ,20-tetrahydroxy-1-oxo-20S,22R-witha-2,24-dienolide (5)

Carbon	1	3	4	2	5
C- 1	203.4	203.2	202.9	203.1	204.9
C- 2	129.2	129.8	134.2	129.6	129.4
C- 3	143.2	143.9	140.0	138.0	139.6
C- 4	69.3	32.9	70.0	32.1	31.0
C- 5	138.2	62.2	133.9	77.0	81.5
C- 6	131.5	64.2	130.1	71.6	69.9
C- 7	25.9	26.2	25.6	26.5	28.5
C- 8	35.8	34.1	34.4	33.4	33.9
C- 9	36.9	36.9	36.4	34.8	34.1
C-10	49.5	48.9	49.4	52.8	53.2
C-11	22.5	22.9	21.8	22.6	22.9
C-12	34.2	34.3	32.4	37.7	37.7
C-13	54.5	54.5	47.5	54.9	55.1
C-14	81.9	82.3	84.9	82.3	83.0
C-15	30.2	30.1	32.0	30.8	30.1
C-16	37.7	37.7	20.6	34.2	34.5
C-17	88.0	87.8	49.4	87.9	88.2
C-18	20.5	20.6	17.4	21.0	21.2
C-19	22.6	14.6	21.8	13.8	15.6
C-20	78.9	80.0	75.3	79.0	79.2
C-21	19.7	19.5	21.2	20.1	20.1
C-22	80.1	80.3	81.3	79.9	80.5
C-23	32.5	32.5	31.8	32.7	32.8
C-24	150.7	151.1	149.1	150.5	151.3
C-25	121.4	121.4	122.0	121.5	121.5
C-26	166.2	166.6	166.2	165.8	166.6
C-27	12.3	12.3	12.5	12.4	12.5
C-28	20.6	20.6	20.6	20.6	20.9
5 α -Butoxy, ethoxy				61.7 28.0 19.3 15.0	57.5 16.0
-OAc			170.3 21.4	170.2 21.4	

presence of the hydroxy groups at C-17 and 20.

A comparative study of the ¹H-NMR spectrum of 2 (Table I) with that of 1 suggested that 2 has the same C and D rings and side chain moiety as 1. Signals at δ 2.33 (1H, dd, $J=19.7, 5.7$ Hz), δ 2.55 (1H, d, $J=19.7$ Hz) and δ 5.15 (1H, d, $J=2.7$ Hz) were similar to those of the acetate (5a) of an analog withanolide, 5 α -ethoxy-6 β ,14 α ,17 β ,20-tetrahydroxy-1-oxo-20S,22R-witha-2,24-dienolide (5)⁷ obtained from *Withania somnifera*, and were attributable to methylene protons at C-4 and an acetoxy-methine proton at C-6. Moreover, a deshield signal due to H₃-19 at δ 1.26 suggested the presence of a hydroxy group at C-5. Furthermore, the existence of a 1-butoxy group was suggested by the ¹H-¹H correlated spectroscopy (COSY) of 2. It was determined that the 1-butoxy group was connected to C-5 by comparing the ¹³C-NMR spectrum with that of 5.

In comparing the ¹³C-NMR spectrum of 2 with that of 5, the chemical shifts, except for those at C-3—7, 9, 10 and 19 on the A, B rings and the 1-butoxy group were in good correlation. These chemical shifts of C-3—7, 9, 10 and 19 were presumed to occur because of the introduction of an acetoxy group at C-6. The positive Cotton effect at 255 nm in the CD spectrum of 2 indicated a 22R configuration, and the negative Cotton effect at 334 nm showed a *trans* fusion of rings A and B.⁶ Thus, the structure of 2 was concluded to be as shown in the formula. The 1-butoxy group at C-5 was suspected to be artificially introduced into the 5 β ,6 β -epoxy ring during separation.

Experimental

The melting point was measured using a Yanagimoto micromelting point apparatus and was uncorrected. Optical rotations were measured on a JASCO DIP-360 automatic digital polarimeter and CD spectrum on a JASCO J-50A spectropolarimeter. The IR spectra were recorded with a Hitachi IR spectrometer, model 270-30. The ¹H- and ¹³C-NMR spectra were measured with a JEOL JNM-GX 400 NMR spectrometer and chemical shifts are given on a δ (ppm) scale with tetramethylsilane (TMS) as an internal standard. The FAB-MS were measured with a JEOL DX-303 HF spectrometer and taken in a glycerol matrix containing NaI. Thin layer chromatography was performed on pre-coated Kieselgel 60 F₂₅₄ (Merck) and detection was achieved by spraying it with 10% H₂SO₄, followed by heating. Column chromatography was carried out on Kieselgel (270—300 mesh and 230—400 mesh, Merck) and Sephadex LH-20 (Pharmacia Fine Chemicals).

Extraction and Separation The fresh aerial parts (2.2 kg) of *Physalis pruinosa* BAILEY (Solanaceae), harvested at the botanical garden of Kyushu University in December 1988, were extracted with MeOH, and the extract was partitioned between 1-BuOH and H₂O, then the 1-BuOH layer was partitioned between 1-BuOH-EtOAc and H₂O. The organic layer (30 g) was subjected repeatedly to column chromatography over silica gel using CHCl₃-MeOH-H₂O=1:0:0→8:3:0.5 and Sephadex LH-20 using MeOH to give physapruins A (1, 34.6 mg), B (2, 13.7 mg), withanolide E (3, 113.0 mg), 4 β -hydroxywithanolide E⁸) (76.9 mg) and 2,3-dihydro-3 β -methoxy-4 β -hydroxywithanolide E monoacetate⁹) (21.3 mg). During this separation procedure, the free compound of 2 and 2,3-dihydro-3 β -methoxy-4 β -hydroxywithanolide E monoacetate were difficult to separate and were converted to acetyl derivatives to facilitate separation.

Physapruin A (1) Colorless needles from CHCl₃-MeOH, mp 207—209 °C, $[\alpha]_D^{25} +159.1^\circ$ ($c=0.46$, MeOH). Positive FAB-MS (m/z): 509 [M+Na]⁺, 469 [M-H₂O+H]⁺, 451 [M-2H₂O+H]⁺, 433 [M-3H₂O+H]⁺, 415 [M-4H₂O+H]⁺, 283, 265, 169, 125. IR (KBr): 3408, 1696, 1664 cm⁻¹. CD ($c=0.60$, MeOH) $[\theta]$ (nm): -28000 (340) (negative max.), +83000 (256) (positive max.).

Physapruin B (2) Amorphous powder, $[\alpha]_D^{25} +47.0^\circ$ ($c=0.47$, MeOH). Positive FAB-MS (m/z): 625 [M+Na]⁺, 603 [M+H]⁺, 585 [M-H₂O+H]⁺, 549 [M-3H₂O+H]⁺, 417, 357, 283, 265, 169, 125. IR (KBr): 3452, 1740, 1714, 1690 cm⁻¹. CD ($c=0.047$, MeOH) $[\theta]$ (nm): -15000 (334)

(negative max.), +51000 (255) (positive max.).

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References and Notes

- 1) This work is Part XXVII in the series of studies on the constituents of solanaceous plants.
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