Fern Constituents: Triterpenoids Isolated from the Leaves of *Adiantum monochlamys*. Filicenol A, Filicenol B, Isoadiantol B, Hakonanediol and Epihakonanediol

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Five new triterpenoids, filicenol A (1), filicenol B (2), isoadiantol B (3), hakonanediol (4) and epihakonanediol (5), were isolated from the leaves of *Adiantum monochlamys* EATON, and their structures were elucidated on the basis of spectral data and chemical correlations with known compounds. Additional data are presented for previously reported compounds (6—21) from the same source.

Keywords fern; Adiantum monochlamys; triterpenoid; filicenol; isoadiantol; hakonanediol

Adiantum monochlamys Eaton (Hakone-shida in Japanese, Pteridaceae) is a rather common fern distributed in southern Honshū, Shikoku and Kyūshū, Japan, and has been used as a diuretic by suggestion of Kaempfer. In the previous communications, we have reported the structures of seventeen kinds of triterpenoids belonging to the hopane and migrated hopane groups isolated from the dried leaves. This paper deals with the isolation and structure elucidation of five new triterpenoids, filicenol A (1), filicenol B (2), isoadiantol B (3), hakonanediol (4) and epihakonanediol (5) (Chart 1) from the same source. Some additional data are also presented on previously reported compounds (6—21).

Results and Discussion

Air dried leaves were extracted with methanol. A suspension of the methanolic extract in water were extracted with ether. The ether extract was subjected to various kinds of chromatography (see Experimental) to obtain compounds 1-15 and 17-21; Table I gives physical constants and yields. Adianene ozonide (16) was only obtained from n-hexane extract of fresh leaves of the same fern. 2d

A new compound, filicenol A (1), was obtained as colorless needles. The infrared absorption (IR) spectrum indicated the presence of a hydroxyl group and a trisubstituted double bond. The mass spectrum (MS) of 1 showed a molecular ion, m/z 426.3850 ($C_{30}H_{50}O$) and many

significant fragment ions at m/z (rel. int.) 408 (46; M⁺ – CH₃ – H₂O), 383 (2; M⁺ – C₃H₇), 365 (7; M⁺ – C₃H₇ – H₂O), 341 (22; a), 323 (12; a – H₂O), 273 (26; b), 255 (5; b – H₂O), 205 (19; c) and 191 (100; d) (Chart 2). These

TABLE I. Triterpenoids Isolated from the Leaves of Adiantum monochlamys

	mp (°C)	[α] _D	Yield (%)	Lit.
Filicenol A (1)	222—225	+52.0	0.025	
Filicenol B (2)	218-221	+57.0	0.055	
Isoadiantol B (3)	213.5—215	+16.0	0.031	
Hakonanediol (4)	270-272	+21.0	0.0059	
Epihakonanediol (5)	299-301	+5.0	0.0066	
Fern-8-ene (6)	190—192	+30.6	0.027	2a
Fern-9(11)-ene (7)	171—172	-19.6	0.0021	2c
Ferna-7,9(11)-diene (8)	202-203	-177.9	0.0051	2c
Fern-7-ene (9)	212.5-214	-28.8	0.028	2a
Adian-5-ene (10)	193.5—195	+51.9	0.042	2a
Neohop-13(18)-ene (11)	199—201	+2.1	0.001	2c
Neohop-12-ene (12)	210-211	+41.6	0.0081	2c
Filic-3-ene (13)	232-234	+50.0	0.028	2a
Hop-22(29)-ene (14)	210-212	_	0.0007	2a
Neohopa-11,13(18)-diene (15)	214.5—215.5	+25.7	0.002	2c
Adianene ozonide (16)	154—157	+19.4	0.121	2d
Adiantone (17)	227-230	+80.1	0.111	2b
Isoadiantone (18)	236-237	+3.6	0.0011	2b
Tetrahymanol (19)	> 300		0.022	6
Ketohakonanol (20)	295-297	+8.0	0.0013	2b
Hydroxyadiantone (21)	270—275	+50.0	0.230	2b

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fragment ions suggested for 1 the filicane skeleton with a hydroxyl group in the left-hand part of the molecule.³⁾ The ¹H-nuclear magnetic resonance (¹H-NMR) spectrum

of 1 indicated the presence of five tertiary, two secondary and one olefinic methyl groups and one olefinic proton. The chemical shifts of methyl groups were similar to those of filic-3-ene (13) (Table II). The presence of a secondary hydroxyl group (equatorial) was suggested by a proton signal at δ 3.593 (1H, dd), and the signal of a trisubstituted double bond was observed at δ 5.199 (br s). In the ¹³C-NMR spectrum (Table III), signals of 1 were coincident with those of 13 except for the hydroxy methine carbon and its neighbors. Assignments of the 1H- and ¹³C-NMR spectra of the compounds shown in Tables II and III were confirmed as necessary by proton-proton and ¹³C-proton correlated spectroscopy (¹H-¹H and ¹³C-¹H COSY), ¹H-detected heteronuclear multiple bond correlation (HMBC) spectrum and distortionless enhancement by polarization transfer (DEPT) spectrum methods. Acetylation of 1 in a usual manner afforded filicenol A acetate (1a), the ¹H-NMR data for which are given in Table II. Compound 1 was oxidized to give filicenone A (22), and

Table II. ¹H-NMR Spectral Data for Triterpenoids of Filicane, Adianane and Hopane Groups (270/400/500 MHz, CDCl₃, δ)

Protons	1	1a	2	2a	13	22	23	24	25
H-23	1.824	1.574	1.589	1.584	1.575	1.856	1.130	1.162	1.600
25	(dd, 3.2, 1.5)	(brs)	(dd, 2.5, 1.5)	(d, 1.5)	(dd, 3.0, 2.0)	(dd, 2.5, 1.5)			(dd, 3.0, 1.7)
H-24	0.999	1.092	1.063	0.994	0.975	1.350	1.066	0.936	0.892
H-25	0.886	0.924	3.839	4.202	0.892	1.116	1.036	0.886	10.210
			(1H, d, 12.0)	(1H, d, 12.0)					(1H, s)
			3.915	4.388					
			(1H, d, 12.0)	(1H, d, 12.0)					
H-26	0.913	0.894	0.923	0.896	0.912	0.892	0.928	0.905	0.866
H-27	0.913	0.908	0.946	0.944	0.921	0.962	0.928	0.926	0.922
H-28	0.777	0.770	0.777	0.778	0.775	0.764	0.772	0.782	0.766
H-29 ^{a)}	0.886	0.876	0.884	0.884	0.886	0.880	0.876	0.884	0.880
	(d, 6.4)	(d, 6.4)	(d, 6.4)	(d, 6.6)	(d, 6.7)	(d, 6.6)	(d, 6.6)	(d, 6.7)	(d, 6.6)
H-30 ^{a)}	0.826	0.818	0.824	0.826	0.825	0.822	0.824	0.825	0.820
	(d, 6.4)	(d, 6.4)	(d, 6.4)	(d, 6.6)	(d, 6.7)	(d, 6.6)	(d, 6.6)	(d, 6.7)	(d, 6.6)
CH = C	5.202	5.172	5.188	5.202	5.161	5.340			5.164
	(1H, brs)	(1H, brs)	(1H, br s)	(1H, brs)	(1H, br s)	(1H, br s)			(1H, br s)
CH-O-	3.595	4.744		` _ ′				3.577	***************************************
	(1H, dd, 10.2,	(1H, dd, 9.8,						(1H, ddd, 10.4,	
	4.8)	4.9)						10.4, 5.5)	
CH ₃ -CO	_	2.032		2.052		_		***************************************	

Protons	26	27	3	4	4 a	5	28	29	30
H-23	0.882	0.908	0.846	0.844	0.846	0.846	0.845	0.847	0.848
H-24	0.960	1.044	0.792	0.790	0.794	0.795	0.792	0.794	0.792
H-25	3.500		0.816	0.810	0.817	0.816	0.814	0.816	0.816
	(1H, d, 8.3) 4.414 (1H, d, 8.3)								
H-26	0.912	0.916	0.950	0.958	0.964	0.976	0.951	0.970	0.970
H-27	0.920	1.014	0.968	0.968	0.973	0.976	0.951	0.942	0.952
H-28	0.770	0.774	0.682 (d, 1.0)	0.790	0.812	0.795	0.703	0.648	0.662 (d, 1.0)
H-29 ^{a)}	0.882 (d, 6.6)	0.878 (d, 6.6)	1.160 (d, 6.1)	1.072	1.102	1.115	0.927 (d, 6.1)	0.889 (d, 6.7)	1.182 (d, 6.4)
H-30 ^{a)}	0.824 (d, 6.6)	0.826 (d, 6.6)	· _				0.806 (d, 6.1)	0.788 (d, 6.7)	
CH = C	`- '	· —			_	-			
СН-О-		_	3.732 (1H, dddd, 6.4, 6.4, 6.4, 6.4)	3.416 (1H, dd, 12.1, 4.4)	4.636 (1H, dd, 11.0, 5.0)	3.546 (1H, ddd, 4.8, 2.4, 2.4)	_		3.874 (1H, dddd, 6.4 6.4, 6.4, 2.4)
CH ₃ -CO	_				2.086		_		

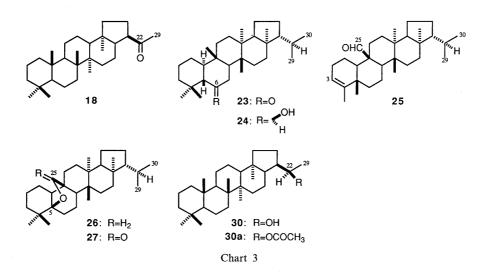
Signals unless otherwise stated are 3H, singlet. Multiplicity and coupling constants are shown in parentheses. a) Assignments of signals might be reversed, although the signals of H-29 and H-30 correspond to those of C-29 and C-30 in Table III, respectively.

264

Table III. ¹³C-NMR Spectral Data for Triterpenoids of the Filicane and Hopane Groups (68/100/125 MHz, CDCl₃, δ)

	1	2	13	24	3	4a	5	28	29	30
C-1	16.9	18.0	17.5 ^{a)}	25.5	40.4	40.4	40.3	40.4	40.3	40.4
C-2	26.9	28.3	27.2	22.1	18.7	18.8	18.7	18.7	18.7	18.7
C-3	. 122.1	120.7	120.3	44.1	42.1	42.2	42.1	42.2	42.1	42.1
C-4	143.4	144.0	144.3	33.8	33.3	33.3	33.3	33.3	33.3	33.3
C-5	44.4	38.7	38.4	51.8	56.2	56.3	56.2	56.2	56.1	56.2
C-6	77.6	39.2	38.7	73.8	18.7	18.8	18.7	18.7	18.7	18.7
C-7	29.2	20.4	18.3^{a}	32.7	33.3	33.1	33.0	33.3	33.3	33.3
C-8	46.4	50.2	49.4	45.1	41.9	42.1	42.1	41.9^{b}	41.9	42.0
C-9	37.3	42.5	37.6	36.5	50.5	50.4	50.3	50.5	50.5	50.5
C-10	55.8	58.3	57.2	50.4	37.4	37.5	37.5	37.4	37.4	37.4
C-11	35.1	29.6	35.2	34.3	20.9	21.1	21.0	21.0	20.9	20.9
C-12	28.4	29.6	28.4	28.8	22.8	21.6	21.4	24.1	23.9	21.2
C-13	39.1	39.2	39.1	38.9	48.5	50.1	49.9	49.3	48.6	48.4
C-14	40.0	39.7	40.2	39.8	42.0	41.9	41.9	41.8 ^{b)}	42.3	42.3
C-15	29.1	29.3	29.1	29.0	32.7	32.5	32.4	33.7	32.7	32.6
C-16	35.6	35.8	35.7	35.6	23.9	17.3	17.4	22.6	21.6	23.9
C-17	42.8	42.7	42.8	42.8	55.0	56.3	49.8	54.7	53.2	51.6
C-18	51.7	51.6	51.8	51.8	45.1	37.4	37.3	44.4	44.5	44.6
C-19	19.9	19.9	19.9	19.9	39.7	38.1	32.4	41.7	39.8	39.7
C-20	28.4	28.4	28.4	28.4	24.3	25.1	25.4	27.6	22.7	21.2
C-21	60.1	60.1	60.1	60.1	47.5	81.9	74.0	47.9	45.5	46.8
C-22	30.7	30.8	30.8	30.8	72.8	74.4	73.3	32.0	28.8	67.5
C-23	22.1	18.0	17.9	34.3	33.4	33.5	33.4	33.4	33.4	33.4
C-24	15.3	20.1	20.7	21.0	21.6	21.7	21.6	21.6	21.6	21.6
C-25	20.1	64.8	20.6	17.5	15.9	16.0	15.9	15.9	15.9	15.9
C-26	16.0	15.5	16.1	16.2	16.8	16.6	$16.6^{c)}$	16.7°)	16.8	16.8
C-27	15.6	15.7	15.7	15.3	16.8	16.6	$16.5^{c)}$	16.8°)	16.7	16.8
C-28	16.3	16.3	16.3	16.3	15.2	15.5	14.8	15.8	15.2	15.9
C-29	22.0^{d}	22.0^{d}	22.0^{d}	22.0^{d}	21.8	17.8	22.0	22.8^{d}	22.1 ^d)	16.4
C-30	22.9^{d}	22.9^{d}	22.9^{d}	22.9^{d}	_			23.9^{d}	17.5^{d}	
CO		_			_	171.7	_			_
CH_3						21.4	_			

a-d) Assignments of signals might be reversed. Spectra of 13, 28 and 29 were run at 125 MHz, and assignments were confirmed by DEPT, $^{1}H^{-1}H$ and $^{13}C^{-1}H$ -COSY, and HMBC methods. Some of assignments reported 7) have been revised.



the chemical shifts of three methyl groups (H-23, H-24 and H-25) of 22 were shifted to lower fields compared with those of 13. Compound 22 was reduced by the Wolff–Kishner–Barton method to give 13′, which was identical with an authentic sample of 13.^{2a)} On the other hand, 1 was treated with BF₃-etherate/ether to give a saturated ketone, adianan-6-one (23), reduction of which afforded adianan- 6β -ol (24).⁴⁾ Compound 24 was dehydrated with POCl₃/pyridine to give adian-5-ene (10). Thus, the structure of filicenol A was established as filic-3-en- 6β -ol (1), the absolute

configuration of which was confirmed by the circular dichroism (CD) curve of 22.

The second new compound, filicenol B (2) was obtained as colorless needles. The MS of 2 showed a molecular ion, m/z 426.3870 ($C_{30}H_{50}O$), and many significant fragment ions at m/z (rel. int.) 411 (10; M^+-CH_3), 395 (51; M^+-CH_2OH), 383 (4; $M^+-C_3H_7$), 341 (4, a'), 273 (7, b'), 255 (8, b'- H_2O), 205 (54; d), 191 (31; c) and 177 (100; e) (Chart 2). These fragment ions suggested that 2 has a filicane skeleton with a hydroxyl group on the left-hand

part of molecule.3) The 1H-NMR spectrum of 2 indicated the presence of four tertiary, two secondary, and one olefinic methyl groups, one hydroxymethyl group and one trisubstituted double bond. The chemical shifts of methyl groups were similar to those of 13, as shown in Table II. Thirty carbon signals were observed in the 13C-NMR spectrum of 2 and their chemical shift values were similar to those of 13 except for C-25, as shown in Table III. Compound 2 was acetylated in a usual manner to give filicenol B acetate (2a), the ¹H-NMR spetrum of which is shown in Table II. Oxidation of 2 with CrO₃-pyridine afforded an aldehyde (25), and 25 was reduced by the Wolff-Kishner-Barton method to give a hydrocarbon (13'), which was identified as filic-3-ene by comparison with an authentic sample.^{2a)} Furthermore, treatment of 2 with 1 N H₂SO₄/AcOH gave an epoxide (26), oxidation of which with K₂Cr₂O₇ afforded a five membered lactone (27). Compound 27 was assumed to be adianan- 5β ,25olide and 26 to be adianan-5 β ,25-epoxide on the basis of their ¹H-NMR spectra (Table II). Thus, the structure of 2 was concluded to be 25-hydroxyfilic-3-ene.

The third new compound, isoadiantol B (3) was obtained as colorless needles. The MS of 3 showed a molecular ion, m/z 414.3876 (C₂₉H₅₀O) and many significant fragments at m/z (rel. int.) 399 (9; M^+-CH_3), 381 (7; $M^+-CH_3 H_2O$), 367 (8; $M^+ - C_2H_5O$), 193 (73; d), 191 (100; e) and 175 (63; d-H₂O) (Chart 2). These fragment ions suggested the 30-norhopane skeleton for 3.3) The ¹H-NMR spectrum of 3 indicated the presence of six tertiary and one secondary methyl groups (Table II). The secondary methyl proton of H-29 (δ 1.16, d, J=6.1 Hz) was observed at lower field than that of hopane (28) or isohopane (29). As shown in Table III, the ¹³C-NMR spectrum of 3 revealed carbon signals that were very similar to those of 28 except for the C-22 hydroxymethine signal. Therefore, 3 was supposed to be 30-norisohopan-22-ol, and the structure was finally confirmed by direct comparison with the products prepared from isoadiantone (18)^{2b)} by LiAlH₄ reduction. The products were two diastereomeric 30-norisohopan-22-ols, isoadiantol A (30, less polar) and isoadiantol B (3', more polar), of which the latter was proved to be identical with 3. The establish the absolute configuration at C-22 of 30 and 3, the differences between the molecular rotations $[M]_D$ of the alcohols (30 and 3) and their respective

TABLE IV. Comparison of Molecular Rotation of Pregnane Derivatives and Isoadiantol A and B

		Alcohol (A) $[M]_D$	Acetate (B) $[M]_D$	$B-A$ $\Delta[M]_D$
Isoadiantol A (30)	22R	+21	+73	+ 52
Isoadiantol B (3)	22S	+66	+46	-20
Adiantol A	22R	+161	+160	— 1
Adiantol B ^{a)}	22S	+306	+251	-55
Pregn-5-ene-3β,20-diol	20R	-204	 149	+55
	20S	– 179	— 197	-18
3-Oxopregn-4-en-20-ol	20R	+272	+483	+211
	20S	+319	+323	+4
3,11-Dioxopregn-20-ol	20R	+175	+280	+105
	20S	+230	+200	-30

a) The absolute configuration at C-22 of the bromoacetate was determined by X-ray analysis. $^{8)}$

acetates were compared with those of paired adiantols³⁾ and pregnan-20-ol derivatives⁵⁾ (Table IV). As the $[M]_D$ increment for conversion of 3 to its acetate (3a) was negative, as found for the 20S compounds of adiantols and pregnan-20-ol derivatives, compound 3 was assumed to have 22S configuration. Thus, isoadiantol B (3) was established as (22S)-30-norisohopan-22-ol.

The other two new compounds, hakonanediol (4) and epihakonanediol (5) were both obtained as colorless needles. The MS of 4 and 5 showed the molecular ions, m/z430.3833 and 430.3815 ($C_{29}H_{50}O_2$), respectively, and significant fragment ions at m/z (rel. int.) 412 (7, 6; M^+ H_2O), 397 (5, 4; $M^+-CH_3-H_2O$) and 191 (100, 100; g) (Chart 2). These fragment ions suggested that both 4 and 5 have the hopane-like skeleton on the left-hand part, and a hydroxyl group on the right-hand part of the molecules.3) The 1H-NMR spectra of both 4 and 5 indicated the presence of seven tertiary methyl groups, and the chemical shifts of five methyl groups on rings A, B and C closely resembled to those of hopane (28) (Table II). Two hydroxyl groups of 4 and 5 were secondary and tertiary. The coupling pattern $[\delta 3.416 \text{ (dd, } J=12.1, 4.4 \text{ Hz})]$ of a geminal proton of the secondary hydroxyl group in 4 suggests that the proton is axial, while the coupling pattern $[\delta \ 3.546 \ (ddd, J=4.8, 2.4, 2.4 \ Hz)]$ of that in 5 suggests that the proton is equatorial. Acetylation of 4 and 5 in a usual way afforded hakonanediol monoacetate (4a) and epihakonanediol monoacetate (5a). The ¹³C-NMR data for 4a and 5 are given in Table III, and support the above conclusion. The structures of 4 and 5 were finally established by the identity of the IR and ¹H-NMR spectra with those of two diols, 4' and 5', respectively, obtained from ketohakonanol (20)^{2b)} by LiAlH₄ reduction.

It is noteworthy that all twenty-one triterpenoids obtained from the leaves of *Adiantum monochlamys* (Table I) are pentacyclic and belong to the hopane and migrated hopane or closely related groups. The isolation of the many nor-compounds, 3, 4, 5, 17, 18, 20 and 21, is also a characteristic feature of this *Adiantum* fern.

Experimental

Melting points were measured on a Yanagimoto micro apparatus and were corrected. Specific rotation was observed in CHCl₃ solutions (c=0.5-1.2) at 22-24°C. CD was measured with a JASCO J-600 apparatus. ¹H- and ¹³C-NMR spectra were taken at 270/400/500 and 68/100/125 MHz by the Fourier transform (FT) method with tetramethylsilane as an internal standard. MS were recorded (direct inlet) at 30 eV and relative intensities of peaks were reported with reference to the most intense peak higher than m/z 100. Gas liquid chromatography (GLC) was performed on a 1 m glass column containing Chromosorb G HP with 1.4% SE-30 at 260 °C under N₂ using cholestane as an internal standard (its retention time was set at 3.0 min). Silica gel 60, 230-400 mesh (Merck), Al₂O₃ (Woelm or Wako, neutral or basic, grade 1 or 3) and 20%-AgNO₃ impregnated silica gel were used for column chromatography (CC). Precoated Silica gel 60 plates (Merck) were used for thin layer chromatography (TLC), and spots were detected by spraying with concentrated H₂SO₄ followed by heating.

Plant Materials The leaves of Adiantum monochlamys were collected in August, 1965, and September 1976, at Mariko, Shizuoka, Shizuoka Prefecture. Voucher specimens have been deposited in the Herbarium of Showa College of Pharmaceutical Sciences, Tokyo.

Extraction of Dried Leaves and Separation The drid leaves (1.3 kg) were extracted three times with MeOH (40 l each). The extract was concentrated to 10 l in total, and was allowed to stand overnight. Waxy substances were filtered off, and the solution was evaporated to dryness. The resultant solid was extracted with Et_2O and water, and the Et_2O

solution was dried and evaporated to give a residue $(48.5 \,\mathrm{g})$. The residue was refluxed with *n*-hexane $(2 \,\mathrm{l})$. After cooling, the less soluble material $(9.0 \,\mathrm{g})$ was filtered off (fraction A), and the solution was evaporated to dryness. The residue was chromatographed on silica gel with *n*-hexane (fr. B), *n*-hexane-benzene (8:2) (fr. C), *n*-hexane-benzene (1:1) (fr. D), benzene (fr. E), benzene-Et₂O (9:1) (fr. F), and Et₂O (fr. G) to give six fractions.

Fern-8-ene (6), Fern-9(11)-ene (7), Ferna-7,9(11)-diene (8), Fern-7-ene (9), Adian-5-ene (10), Neohop-13(18)-ene (11), Neohop-12-ene (12), Filic-3-ene (13), Hop-22(29)-ene (14) and Neohopa-11,13(18)-diene (15) Fraction B was repeatedly chromatographed on Al_2O_3 and 20% AgNO₃-impregnated silica gel to give the following crystalline solids (weight) in order of elution (recrystallized from acetone to obtain pure specimens): 6 (310 mg), 7 (23 mg), 8 (58 mg), 9 (318 mg), 10 (478 mg), 11 (12 mg), 12 (91 mg), 13 (313 mg), 14 (8 mg) and 15 (23 mg). 2a,c

Adiantone (17) and Isoadiantone (18) Fraction C was chromatographed several times on silica gel. The crystalline product (3250 mg) from the *n*-hexane-benzene (8:2) eluate was recrystallized from MeOH to give 17. IR $y_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1705. The product (15 mg) from the second eluate with the same solvent was recrystallized from MeOH to give 18. IR $y_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1705.^{2b}

Filicenol A Acetate (1a) and Filicenol B Acetate (2a) Fraction D was treated with pyridine–Ac₂O overnight at room temperature, and the reaction mixture was repeatedly chromatographed on Al₂O₃. The crystalline product (350 mg) from the first eluate with *n*-hexane was recrystallized from acetone to give 1a, mp 249—252 °C, $[\alpha]_D$ +17.0°. IRv^{KBr}_{max} cm⁻¹: 1720, 1246, 1023. The crystalline product (750 mg) from the second eluate with the same solvent was recrystallized from acetone to give 2a, mp 214—216 °C, $[\alpha]_D$ +75.0°. IR v^{KBr}_{max} cm⁻¹: 1737, 1245, 1035.

Filicenol A (1) 1a (250 mg) was treated with LiAlH₄, and the product was chromatographed on Al₂O₃. The crystalline product from the *n*-hexane-benzene (1:1) eluate was recrystallized from acetone to give 1. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3550, 1013. *Anal.* Calcd for C₃₀H₅₀O: C, 84.44; H, 11.81. Found; C, 84.61; H, 12.06.

Filicenol B (2) 2a (700 mg) was treated with LiAlH₄, and the product was chromatographed on Al_2O_3 . The crystalline product from the *n*-hexane-benzene (1:1) eluate was recrystallized from acetone to give 2. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 3525, 1032. *Anal.* Calcd for $C_{30}H_{50}O$: C, 84.44; H, 11.81. Found; C, 84.68; H, 11.82.

Isoadiantol B Acetate (3a) and Tetrahymanol Acetate (19a) Fraction E was treated with pyridine–Ac₂O overnight at room temperature, and the reaction product was recrystallized repeatedly from CHCl₃–MeOH to give less soluble crystals, 19a (200 mg), mp > 300 °C, $[\alpha]_D$ +32.1°. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1730, 1250, and more soluble crystals, 3a (250 mg), mp 217.5—220 °C, $[\alpha]_D$ +13.2°. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1730, 1242. 19a were identical (IR and TLC comparisons) with an authentic sample.⁶)

Isoadiantol B (3) 3a (100 mg) was treated with LiAlH₄, and the product was chromatographed on Al_2O_3 . The crystalline product from the *n*-hexane–henzene (1:1) eluate was recrystallized from Et₂O–MeOH to give 3. IR ν_{max}^{KBr} cm⁻¹: 3470, 1111, 1072.

Tetrahymanol (19) 19a (150 mg) was treated with LiAlH₄, and the product was chromatographed on Al_2O_3 . The crystalline product from the *n*-hexane-benzene (1:1) eluate was recrystallized from Et₂O-MeOH to give 19. IR v_{max}^{KBF} cm⁻¹: 3310, 1045, 1033.

Ketohakonanol (20) Fraction F was chromatographed on Al_2O_3 , and the crystalline product (18 mg) from the benzene–Et₂O (9:1) eluate was recrystallized from CHCl₃–MeOH to give **20**. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3480, 1067, 1713.^{2b)}

Hakonanediol (4) and Epihakonanediol (5) Fraction G was chromatographed on Al₂O₃. The crystalline product (90 mg) from the Et₂O eluate was recrystallized from CHCl₃–MeOH to give **5**. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3300, 1080, 1070, 1040. *Anal.* Calcd for C₂₉H₅₀O₂: C, 80.87; H, 11.70. Found: C, 80.86; H, 11.68. The crystalline product (80 mg) from the methanol eluate was recrystallized from CHCl₃–MeOH to give **4**. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3400, 1069, 1044. *Anal.* Calcd for C₂₉H₅₀O₂: C, 80.87; H, 11.70; H, 11.70. Found: C, 80.88; H, 11.68.

Hakonanediol Monoacetate (4a) 4 (20 mg) are treated with pyridine– Ac_2O overnight at room temperature, and the product was recrystallized from acetone to give **4a**, mp 265—268 °C. IRv_{max}^{KBr} cm⁻¹: 3550, 1075, 1039, 1713, 1256.

Epihakonanediol Monoacetate (5a) 5 (20 mg) was treated with pyridine– Ac_2O overnight at room temperature, and the product was chromatographed on Al_2O_3 . The crystalline product from the benzene eluate was recrystallized from acetone to give **5a**, mp 255–258 °C. IR v_{max}^{KBr} cm⁻¹: 3530, 1070, 1054, 1028, 1728, 1250.

Adiantone (17) and Hydroxyadiantone (21) Fraction A was refluxed with benzene (200 ml) for 1 h. After cooling, the less soluble material was filtered off (3.1 g, fr. H), and the solution was evaporated to dryness (4.5 g, fr. I). Fraction I was chromatographed on silica gel, and the crystalline product (1.5 g) from the n-hexane-benzene (8:2) eluate was recrystallized from CHCl₃-MeOH to give 17. 2b Fraction H was repeatedly recrystallized from tetrahydrofuran (THF)-MeOH to give 21. IR v_{max}^{KBr} cm $^{-1}$: 3430, 1082, 1695. 3

Extraction of Fresh Leaves and Separation The fresh leaves $(1.4 \, \text{kg})$ were extracted twice with n-hexane $(45 \, 1 \, \text{each})$ to give extracts $(20 \, \text{g})$ and azeotropic H_2O (840 ml). The extracts were refluxed with n-hexane (2.0 l). After cooling, the less soluble material was filtered off (fr. J), and the solution was evaporated to dryness (fr. K). Fraction K was chromatographed on silica gel with n-hexane (fr. L), n-hexane-benzene (9:1) (fr. M), n-hexane-benzene (8:2) (fr. N), n-hexane-benzene (1:1) (fr. O), benzene (fr. P), benzene-Et₂O (9:1) (fr. Q) and Et₂O (fr. R) to give seven fractions.

Adianene Ozonide (16) Fraction M was rechromatographed on silica gel, and the product (700 mg) from the *n*-hexane-benzene (9:1) eluate was recrystallized from *n*-hexane to give $\mathbf{16}$.

Oxidation of Filicenol A (1) A solution of 1 (250 mg) in pyridine (20 ml) was combined with CrO_3 (200 mg) in pyridine (6 ml), and the mixture was stirred at room temperature for 2 h. The reaction mixture was chromatographed on Al_2O_3 . The product (205 mg) from the *n*-hexane eluate was recrystallized from *n*-hexane to give filic-3-en-6-one (22), mp 280—281 °C. IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 1699. CD (c=0.11, dioxane) [θ] (nm): +9914 (298.5). Further elution with benzene gave the starting material.

Reduction of 22 22 (20 mg) was reduced by the Wolff-Kishner-Barton method. The reaction product was chromatographed on Al_2O_3 , and the crystalline product (16 mg) from the *n*-hexane eluate was recrystallized from acetone to give a hydrocarbon, mp 232—234 °C, which was identified by comparison (IR, GLC) with an authentic sample of filic-3-ene (12). 2a

Acid Induced Rearrangement of Filicenol A (1) 1 (200 mg) was treated with BF₃-etherate (25 ml) in absolute Et₂O (120 ml) at room temperature for 2 d. The reaction mixture was chromatographed on Al₂O₃. The product (60 mg) from the *n*-hexane eluate was recrystallized from methanol to give adianan-6-one (23), mp 260—261 °C, $[\alpha]_D$ +16.5°. IR ν_{max}^{RBr} cm⁻¹: 1698.

Reduction of 23 23 (50 mg) was treated with LiAlH₄, and the product was chromatographed on Al₂O₃. The crystalline product from the *n*-hexane eluate was recrystallized from Et₂O–MeOH to give adianan-6 β -ol (24), mp 213.5—216 °C; 227—229 °C, [α]_D +20.5°. IR ν ^{KBr}_{max} cm⁻¹: 3495, 1065, 1044, 986.

Dehydration of 24 24 (15 mg) was treated with POCl₃ (1 ml) in pyridine (7 ml) at 100 °C for 1 h. The reaction mixture was chromatographed on Al_2O_3 . The crystalline product (10 mg) from the *n*-hexane eluate was recrystallized from acetone to give a hydrocarbon, mp 187—189 °C, which was identical (IR and GLC comparisons) with an authentic sample to adian-5-ene (10).^{2a)}

Oxidation of Filicenol B (2) A solution of 2 (150 mg) in pyridine (15 ml) was combined with CrO_3 (200 mg) in pyridine (6 ml) and the mixture was stirred at -5 °C for 2 h. The product was chromatographed on Al_2O_3 . The first product (100 mg) from the *n*-hexane-benzene (9.5:0.5) eluate was recrystallized from $CHCl_3$ -MeOH to give filic-3-en-25-al (25), mp 248—252 °C, $[\alpha]_D$ +9.5°. IR ν_{max}^{KBr} cm⁻¹: 1690. *Anal.* Calcd for $C_{30}H_{48}O$: C, 84.84; H, 11.39. Found: C, 84.68; H, 11.43. Further elution with benzene gave the starting material.

Reduction of 25 25 ($\overline{20}$ mg) was reduced by the Wolff–Kishner–Barton method. The product was chromatographed on Al₂O₃, and the crystalline product (15 mg) from the *n*-hexane eluate was recrystallized from acetone to give a hydrocarbon, mp 232—234 °C, which was identical (IR and GLC comparisons) with an authentic sample of filic-3-ene (13').^{2a)}

Acid Treatment of Filicenol B (2) A solution of 2 (200 mg) in benzene (14 ml) was treated with concentrated $\rm H_2SO_4$ (1.06 ml) in acetic acid (24 ml) at room temperature for 1 d in a nitrogen atmosphere. The reaction mixture was chromatographed on $\rm Al_2O_3$. The crystalline product (90 mg) from the *n*-hexane-benzene (9:1) eluate was recrystallized from acetone to give adian-5(25)-epoxide (26), mp 201–204 °C, $[\alpha]_D+16.5^\circ$. IR $\nu_{\rm MS}^{\rm RBr}$ cm⁻¹: 1122, 1102. *Anal.* Calcd for $\rm C_{30}H_{50}O$: C, 84.44; H, 11.81. Found: C, 84.50; H, 11.83.

Oxidation of 26 A solution of 26 (75 mg) in acetic acid (40 ml) was treated with $K_2Cr_2O_7$ (130 mg) at 100 °C for 8 h. The reaction mixture was chromatographed on Al_2O_3 . Elution with *n*-hexane gave the starting material, and the product (40 mg) from the benzene eluate was recrystallized from MeOH to give adian-5 β ,25-olide (27), mp > 300 °C, $[\alpha]_D$ +18.5°.

IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1751.

Reduction of Isoadiantone (18) 18 (500 mg) was treated with LiAlH₄, and the mixture was repeatedly chromatographed on Al₂O₃. The product from the initial *n*-hexane-benzene (9:1) eluate was recrystallized from Et₂O-MeOH to give isoadiantol A (30), mp 194—196 °C. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3345, 1132, 1050. The product from the second eluate with the same solvent was recrystallized from Et₂O-MeOH to give isoadiantol B (3'), mp 211.5—212.5 °C. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3470, 1111, 1072.

Isoadiantol A Acetate (30a) 30 was treated with pyridine–Ac₂O overnight at room temperature, and the product was chromatographed on Al₂O₃ with *n*-hexane to give a crystalline material, which was recrystallized from Et₂O–MeOH to afford **30a**, mp 197–199 °C, $[\alpha]_D$ + 16.0°, IR ν_{max}^{RBr} cm⁻¹: 1726, 1250.

Reduction of Ketohakonanol (20) 20 (400 mg) in absolute THF (150 ml) was treated with LiAlH₄ at 40—45 °C for 5 h. The product was chromatographed on Al₂O₃. The product from the Et₂O eluate was recrystallized from CHCl₃—MeOH to give epihakonanediol (5'), mp 282.5—285 °C. IR $\nu_{\rm max}^{\rm KBr}$ cm $^{-1}$: 3300, 1080, 1070, 1040. The product from the methanol eluate was recrystallized from CHCl₃—MeOH to give hakonanediol (4'), mp 264—269 °C. IR $\nu_{\rm max}^{\rm KBr}$ cm $^{-1}$: 3400, 1069, 1044.

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References and Notes

- T. Makino, "New Illustrated Flora of Japan," Hokuryukan, Tokyo, 1961, p. 20.
- a) H. Ageta, K. Iwata and S. Natori, Tetrahedron Lett., 1964, 3413; b) H. Ageta, K. Iwata, Y. Arai, Y. Tsuda, K. Isobe and S. Fukushima, ibid., 1966, 5679; c) H. Ageta, K. Shiojima and Y. Arai, Chem. Commun., 1968, 1105; d) H. Ageta, K. Shiojima, R. Kamaya and K. Masuda, Tetrahedron Lett., 1978, 899.
- K. Shiojima, Y. Arai, K. Masuda, Y. Takase, T. Ageta and H. Ageta, Chem. Pharm. Bull., 40, 1683 (1992).
- Although the reaction product was assumed to be 5αH compound, the product was in fact the 5βH isomer (23). The former has a boat conformation at ring B (calculated by Chem 3 D plus), and the C-25 methyl signal would appear at very high field in its ¹H-NMR spectrum owing to the effect of the carbonyl group at C-6, while the latter has all chair conformations at rings A-D and no marked effect of the carbonyl on the C-25 methyl group is expected. As the steric energy of the former is 90.8 and that of the latter is 84.9, the reaction product was obtained as the more stable isomer, 23, When the four isomers of 24 (5αH, 6αH; 5αH, 6βH; 5βH, 6αH; 5βH, 6βH) were simulated by using Chem 3 D plus, only the 5βH, 6αH isomer could account for the observed splitting pattern of 6α-H (Table II).
- 5) L. Fieser and M. Fieser, "Steroids," Reinhold, New York, 1959, p. 612.
- H. Ageta, K. Shiojima and K. Masuda, *Chem. Pharm. Bull.*, 30, 2272 (1982).
- A. L. Wilkins, P. M. Bird and P. M. Jeger, Magnet. Reson. Chem., 25, 503 (1987).
- 8) H. Koyama and H. Nakai, J. Chem. Soc., B, 1970, 546.