

## ABSOLUTE STEREOSTRUCTURES OF PAEONISUFFRONE AND PAEONISUFFRAL, TWO NEW LABILE MONOTERPENES, FROM CHINESE MOUTAN CORTEX

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Two new labile monoterpenes named paeonisuffrone and paeonisuffral were isolated from Chinese Moutan Cortex, the dried root of *Paeonia suffruticosa* ANDREWS. The absolute stereostructures of paeonisuffrone and paeonisuffral were elucidated on the basis of chemical and physicochemical evidence which included the application of a modified Mosher's method and lipase catalyzed debenzoylation.

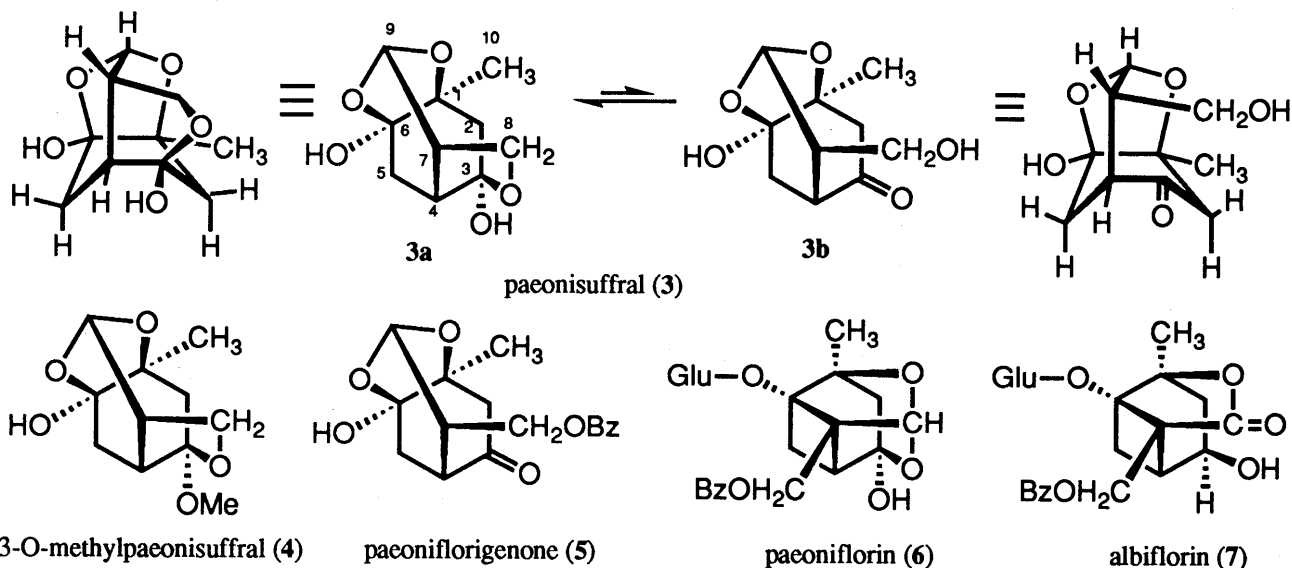
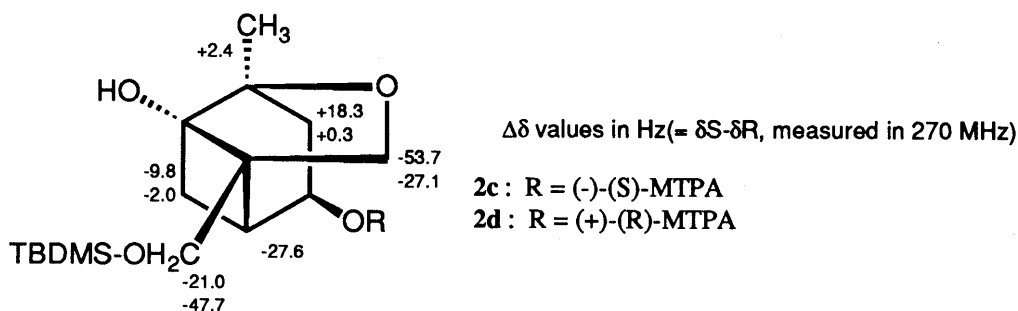
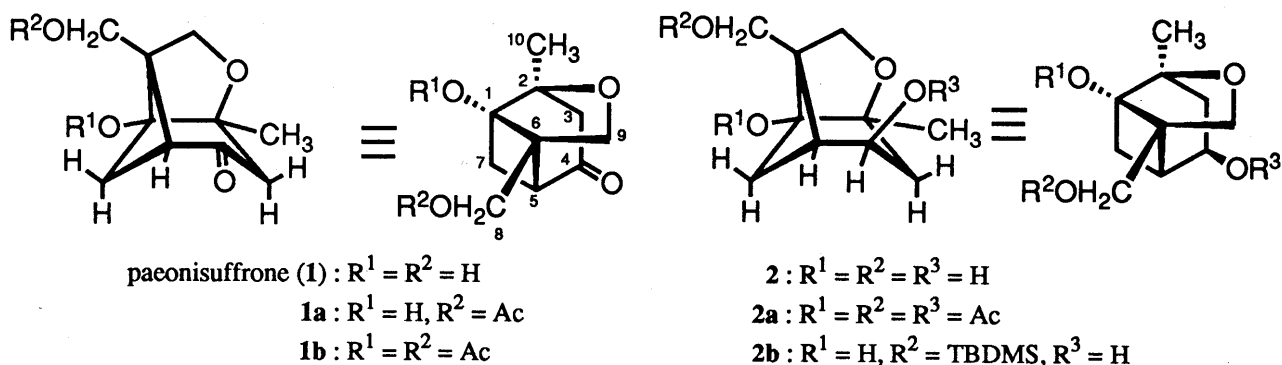
**KEYWORDS** Moutan Cortex; *Paeonia suffruticosa*; Paeoniaceae; paeonisuffrone, paeonisuffral; Mosher's method modified

During the course of our chemical studies on biologically active constituents of naturally occurring drug materials,<sup>1)</sup> we have investigated the chemical constituents of Moutan Cortex, the root cortex of *Paeonia suffruticosa* ANDREWS (Paeoniaceae)[Botanpi in Japanese].<sup>2)</sup> In our recent paper, we reported the isolation of five new glycosides of antioxidant activity, suffruticosides A, B, C, and D and galloyl-oxypaeoniflorin, together with a new paeonol glycoside, suffruticoside E, from the water-soluble portion of the Chinese Moutan Cortex, which is now in common use in Japan.<sup>3)</sup> In a continuing study, we have isolated two new labile monoterpenes named paeonisuffrone(1) and paeonisuffral(3) from the lipophilic portion of the same Chinese Moutan Cortex.<sup>4)</sup> This paper communicates the evidence consistent with the absolute stereostructures of paeonisuffrone(1) and paeonisuffral(3) as shown.<sup>5)</sup>

The MeOH extract (prepared below 25°C) of the Cortex was partitioned into an AcOEt-water mixture. Repeated silica gel column chromatography of the AcOEt-soluble portion furnished 1(0.0003%, from the Cortex), 3(0.0014%), and 3-O-methylpaeonisuffral(4, 0.0003%)<sup>6)</sup> together with paeonol(1.3%), resacetophenone(0.0030%), acetovanillone(0.0010%), 2, 5-dihydroxy-4-methylacetophenone(0.0020%), 2, 5-dihydroxy-4-methoxyacetophenone (0.0010%), acetoisovanillone(0.0030%), benzoyl-paeoniflorin(0.1000%), benzoyl-oxypaeoniflorin(0.0500%), and paeoniflorigenone(5, 0.0110%).<sup>7)</sup>

Paeonisuffrone(1), white powder,  $[\alpha]_D -16.8^\circ$  (c=1.5, MeOH), showed absorption bands due to hydroxyl(3426  $\text{cm}^{-1}$ ) and ketone(1725  $\text{cm}^{-1}$ ) functions in its IR spectrum. The positive FAB-MS of 1 showed the quasimolecular ion peaks at m/z 237(M+K)<sup>+</sup> and m/z 221(M+Na)<sup>+</sup>, while the negative FAB-MS showed the quasimolecular ion peak at m/z 197(M-H)<sup>-</sup>. The high resolution MS measurement of 1 revealed the molecular formula C<sub>10</sub>H<sub>14</sub>O<sub>4</sub>. The <sup>1</sup>H(270 MHz, CD<sub>3</sub>OD,  $\delta$ ) and <sup>13</sup>C NMR(Table I) spectra of 1 showed signals ascribable to a tert.-methyl[ $\delta$  1.31,  $\delta$ c 19.2(10-H<sub>3</sub>)], two methylenes [ $\delta$  2.31, 2.91(ABq, J=18Hz, 3-H<sub>2</sub>);  $\delta$  2.21(d, J=11Hz), 2.47(dd, J=7, 11Hz) (7-H<sub>2</sub>)], two methylenes bearing an oxygen function [ $\delta$  3.57, 3.88(ABq, J=10Hz, 9-H<sub>2</sub>);  $\delta$  3.84, 3.89(ABq, J=12Hz, 8-H<sub>2</sub>)], a ketone carbonyl( $\delta$ c 212.9), two quart. carbons bearing an oxygen function [ $\delta$ c 82.2, 87.6(1, 2-C)] and another one quart. carbon[ $\delta$ c 63.0(6-C)].

Acetylation of 1 with Ac<sub>2</sub>O-pyridine afforded the monoacetate(1a), white powder,  $[\alpha]_D -48.9^\circ$  (c=0.6, EtOH), C<sub>12</sub>H<sub>16</sub>O<sub>5</sub>, IR(KBr,  $\text{cm}^{-1}$ ): 3566, 2975, 1732, 1240, 1042, <sup>1</sup>H NMR(CDCl<sub>3</sub>,  $\delta$ ): 1.42(s, 10-H<sub>3</sub>), 2.10(s, OAc), 2.21(d, J=11Hz), 2.50(dd, J=7, 11Hz) (7-H<sub>2</sub>), 2.54, 2.78(ABq, J=18Hz, 3-H<sub>2</sub>), 2.84(d, J=7Hz, 5-H), 3.70, 3.92(2H, ABq, J=11Hz, 9-H<sub>2</sub>), 4.38, 4.42(ABq, J=12Hz, 8-H<sub>2</sub>), while acetylation of 1 with Ac<sub>2</sub>O-pyridine in the presence of dimethylaminopyridine(DMAP) yielded the diacetate(1b), white powder,  $[\alpha]_D -9.0^\circ$  (c=0.5, EtOH), C<sub>10</sub>H<sub>16</sub>O<sub>4</sub>, IR(KBr,  $\text{cm}^{-1}$ ): 2930, 1790, 1744, 1372, 1242, 1134, 1046, 1022, <sup>1</sup>H NMR(CDCl<sub>3</sub>,  $\delta$ ): 1.36(s, 10-H<sub>3</sub>), 2.09, 2.12(both s, OAcx2), 2.54, 2.92(ABq, J=17Hz, 3-H<sub>2</sub>), 2.76(2H, s, 7-H<sub>2</sub>), 3.00(m, 5-H), 3.72, 3.92(ABq, J=10Hz, 9-H<sub>2</sub>), 4.36, 4.40(ABq, J=12Hz, 8-H<sub>2</sub>). Reduction of 1 with NaBH<sub>4</sub> in MeOH furnished 2, white powder,  $[\alpha]_D -6.2^\circ$  (c=0.3, EtOH), C<sub>16</sub>H<sub>22</sub>O<sub>7</sub>, IR(KBr,  $\text{cm}^{-1}$ ): 3400, 2930, <sup>1</sup>H NMR(CD<sub>3</sub>OD,  $\delta$ ): 1.22(s, 10-H<sub>3</sub>), 1.54(d, J=10Hz), 2.03(dd, J=8, 10Hz) (7-H<sub>2</sub>), 1.78(d, J=16Hz), 2.29(dd, J=8, 16Hz) (3-H<sub>2</sub>), 2.42(dd, J=4, 8Hz, 5-H), 3.68, 4.62(ABq, J=8Hz, 9-H<sub>2</sub>), 3.80, 3.83(2H, ABq, J=12Hz, 8-H<sub>2</sub>), 4.11(dd, J=4, 8Hz, 4-H). Acetylation of 2 with Ac<sub>2</sub>O-pyridine-DMAP gave the triacetate(2a), white powder,  $[\alpha]_D -1.5^\circ$  (c=0.3, EtOH), IR(KBr,  $\text{cm}^{-1}$ ): 2924, 1740, 1250, <sup>1</sup>H NMR(CDCl<sub>3</sub>,  $\delta$ ): 1.27(s, 10-H<sub>3</sub>), 1.98(d, J=16Hz), 2.51(dd, J=8, 16Hz) (3-H<sub>2</sub>), 2.06, 2.07, 2.09(3H



each, all s, OAcx3), 2.23(d,  $J=12$ Hz), 2.43(dd,  $J=8, 12$ Hz) (7-H<sub>2</sub>), 2.67(dd,  $J=4, 8$ Hz, 5-H), 3.83, 4.44(2H, ABq,  $J=9$ Hz, 9-H<sub>2</sub>), 4.27, 4.39(ABq,  $J=12$ Hz, 8-H<sub>2</sub>), 5.12(dd,  $J=4, 8$ Hz, 4-H). Comparison of <sup>1</sup>H NMR data for 2 and 2a with those for 1, 1a, and 1b showed the presence of a partial structure (-CH<sub>2</sub>-CO-CH-CH<sub>2</sub>-) from C-3 to C-7.

The connectivities of the quart. carbons(C-1, 2, 6) were clarified by HMBC experiment with 1a. Namely, the long-range correlations were observed between the following carbons and protons of 1a(2-C: 3-H<sub>2</sub>, 10-H<sub>3</sub>; 1-C: 3-H<sub>2</sub>, 5-H, 8-H<sub>2</sub>; 6-C: 7-H<sub>2</sub>, 8-H<sub>2</sub>). Furthermore, the NOE correlations were observed in the pairs of protons in 2a[3-H<sub>2</sub>&10-H<sub>3</sub>; 3 $\alpha$ -H&5-H; 7 $\beta$ -H&8-H<sub>2</sub>] and comparison of the <sup>1</sup>H-<sup>1</sup>H coupling constants for 1 with those for paeoniflorin(6) and albiflorin(7)<sup>8</sup> has finally led us to formulate the stereostructure of 1 as shown.

The absolute configuration of 1 has been determined by application of a modified Mosher's method.<sup>9</sup>) Thus, silylation of 1 with t-butyltrimethylsilyl chloride(TBDMS-Cl) and imidazole in DMF(r.t., 1h) and subsequent reduction with NaBH<sub>4</sub> provided 2b, which was treated with (-)-(S)- and (+)-(R)- $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetic acid(MTPA) and dicyclohexylcarbodiimide(DCC) in CH<sub>2</sub>Cl<sub>2</sub> in the presence of DMAP to afford the (-)-(S)-MTPA ester(2c) and (+)-(R)-MTPA ester(2d), respectively. The signals due to protons on C-3 and C-10 in the (+)-(R)-MTPA ester(2d) appeared at higher fields than those of the (-)-(S)-MTPA ester(2c)[ $\Delta\delta$ : positive], while the signals due to protons attached to C-5 and C-7 of 2d were observed at lower fields as compared to those of (2c)[ $\Delta\delta$ : negative]. Consequently, the absolute configuration at C-4 has been

Table I.  $^{13}\text{C}$  NMR Data for 1, 1a, 1b, 2, 3a, 3b, and 4

Carbons	1 <sup>a)</sup>	1 <sup>b)</sup>	1a <sup>a)</sup>	1b <sup>a)</sup>	2 <sup>b)</sup>	3a <sup>b)</sup>	3b <sup>b)</sup>	4 <sup>b)</sup>
1	81.7	82.2	81.4	85.6	82.6	79.7	79.7	79.7
2	86.1	87.6	86.1	86.0	90.5	45.2	48.1	43.9
3	48.6	49.8	48.5	48.9	45.4	107.6	213.0	110.9
4	209.9	212.9	209.1	208.2	70.4	45.6	47.9	43.9
5	48.5	49.5	48.6	51.0	40.1	32.0	35.7	32.0
6	60.4	63.0	59.8	59.0	59.1	102.7	97.3	102.9
7	31.2	31.7	31.2	31.9	30.8	44.0	47.3	40.5
8	61.2	62.7	69.8	70.5	64.1	67.2	61.2	68.3
9	68.8	71.7	63.0	63.1	70.3	101.1	101.0	101.1
10	18.9	19.2	19.0	19.8	20.4	22.6	21.9	22.7
COCH <sub>3</sub>			171.7	171.0				
				169.7				
COCH <sub>3</sub>			21.0	20.9 <sup>c)</sup>				
				21.1 <sup>c)</sup>				
OCH <sub>3</sub>								49.3

The spectra were measured  
a) in CDCl<sub>3</sub>, or b) in CD<sub>3</sub>OD,  
and c) assignments may be  
interchangeable.

elucidated to be R and the absolute structure of paeonisuffrone(1) has been determined as shown.

Paeonisuffral(3), white powder,  $[\alpha]_{\text{D}} +39.7^{\circ}$  (c=0.6, MeOH), C<sub>10</sub>H<sub>14</sub>O<sub>5</sub>, IR(KBr, cm<sup>-1</sup>): 3400, 2936, 1719, 1356, 1071, positive FAB-MS(m/z): 215(M+H)<sup>+</sup>, was obtained as a tautomeric mixture of the ketal(3a) and the ketone(3b) forms.<sup>10)</sup> The confirmative assignments for <sup>1</sup>H and <sup>13</sup>C NMR spectra of 3a and 3b were obtained by using <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C COSY and HMBC, and the <sup>1</sup>H and <sup>13</sup>C NMR data for 3b were found similar to those data for paeoniflorigenone(5), except for some signals due to the benzoyl group of the latter. Treatment of 3 with silica gel in methanol yielded 3-O-methylpaeonisuffral(4), white powder, C<sub>11</sub>H<sub>16</sub>O<sub>5</sub>, IR(KBr, cm<sup>-1</sup>): 3419, 2936, 1356, 1073,  $[\alpha]_{\text{D}} +21.1^{\circ}$  (c=0.4, MeOH), <sup>1</sup>H NMR(CD<sub>3</sub>OD,  $\delta$ ): 1.15(s, 10-H<sub>3</sub>), 1.76, 2.01(d, J=14Hz, 2-H<sub>2</sub>), 1.76(d, J=13Hz), 2.07(dd, J=5, 13Hz) (5-H<sub>2</sub>), 2.34(m, 7-H), 2.70(m, 4-H), 3.25(s, OMe), 3.71(d, J=9Hz), 3.92(dd, J=4, 9Hz) (8-H<sub>2</sub>), 5.21(d, J=3Hz, 9-H). Finally, debenzoylation of 5 with lipase(from *Candida cylindracea*) furnished 3[a mixture (ca. 3:1) of 3a and 3b]. Based on this evidence, the absolute structure of paeonisuffral(3) has been clarified.

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- 10) Two tautomers(3a and 3b) were distributed in CD<sub>3</sub>OD in an approximate ratio of 3:1 as shown by <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD,  $\delta$ ). 3a : 1.14(s, 10-H<sub>3</sub>), 1.77(d, J=14Hz), 2.01(dd, J=1, 14Hz) (2-H<sub>2</sub>), 1.85(dd, J=1, 13Hz), 2.13(dd, J=4, 13Hz) (5-H<sub>2</sub>), 2.33(m, 7-H), 2.46(m, 4-H), 3.64(d, J=8Hz), 3.96(dd, J=5, 8Hz) (8-H<sub>2</sub>), 5.19(d, J=3Hz, 9-H); 3b : 1.22(s, 10-H<sub>3</sub>), 2.01(m, 7-H), 2.18(dd, J=2, 13Hz), 2.28(dd, J=3, 13Hz) (5-H<sub>2</sub>), 2.48(d, J=17Hz), 2.72(d, J=17Hz) (2-H<sub>2</sub>), 2.71(m, 4-H), 3.29(d, J=9, 11Hz), 3.47(dd, J=6, 11Hz) (8-H<sub>2</sub>), 5.37(br s, 9-H).

(Received January 5, 1993)