

THE SYNTHESIS OF *TRANS*-3,4-BIS-(3,4-DIHYDROXYPHENYL)PYRROLIDINE, A NOVEL DA₁ AGONIST

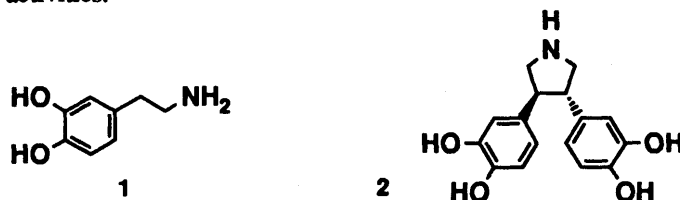
Tomonori HOSHIKO,* Hiroki ISHIHARA, Mitsumasa SHINO and Nobuyuki MORI
Eisai Tsukuba Research Laboratories, 5-1-3 Tokodai, Tsukuba, Ibaraki, 300-26, Japan

The synthesis of *trans*-3,4-bis-(3,4-dihydroxyphenyl)pyrrolidine by the Michael reaction of esterenolate with nitro olefin is described. This compound was expected to be the agonist toward dopamine (DA₁) receptor with equipotent affinity and to be more selective than dopamine itself.

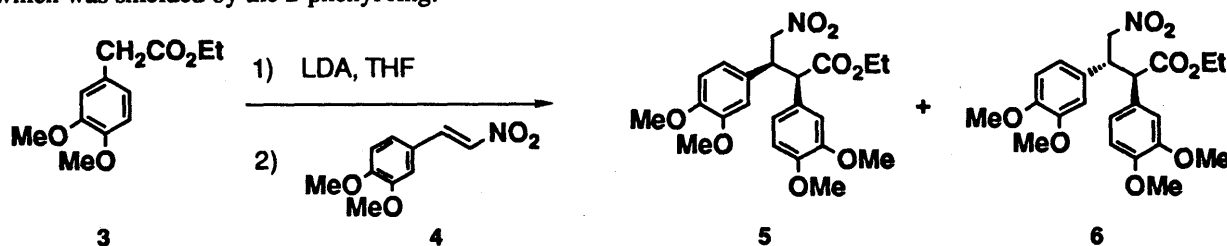
KEYWORDS dopamine; DA₁ agonist; *trans*-3,4-bis-(3,4-dihydroxyphenyl)pyrrolidine; Michael reaction; renal blood flow; mean arterial pressure

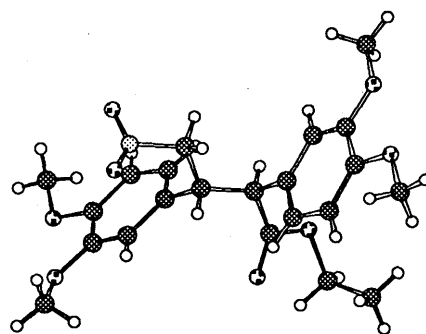
Dopamine (1) is a fundamental sympathomimetic amine and is implicated in a wide variety of both central and peripheral functions.¹⁾ An underproduction of dopamine, for example, causes Parkinson's disease,²⁾ while a hyperstimulation of its receptor associates with schizophrenia³⁾ in the brain. And it is suggested that 1 is linked with regulation of blood pressure or renal blood flow peripherally.⁴⁾ The development of both dopamine agonists and antagonists may therefore be of interest in various fields of therapy. As of now, two pharmacologically distinct dopamine receptor subtypes are known,⁵⁾ and they exist both in the central nervous system (D₁ and D₂ receptors) and in the cardiovascular system (DA₁ and DA₂ receptors).⁶⁾ Although a number of agonists and antagonists of D₂ / DA₂ receptors have already been found,⁷⁾ no agonist which acts on D₁ / DA₁ receptors is known except for benzazepine derivatives.⁸⁾

We have designed a novel DA₁ agonist, *trans*-3,4-bis-(3,4-dihydroxyphenyl)pyrrolidine (BDP, 2), whose skeleton is completely different from that of benzazepine derivatives. In this paper, we report an efficient synthesis of BDP(2) and its biological and pharmacological activities.

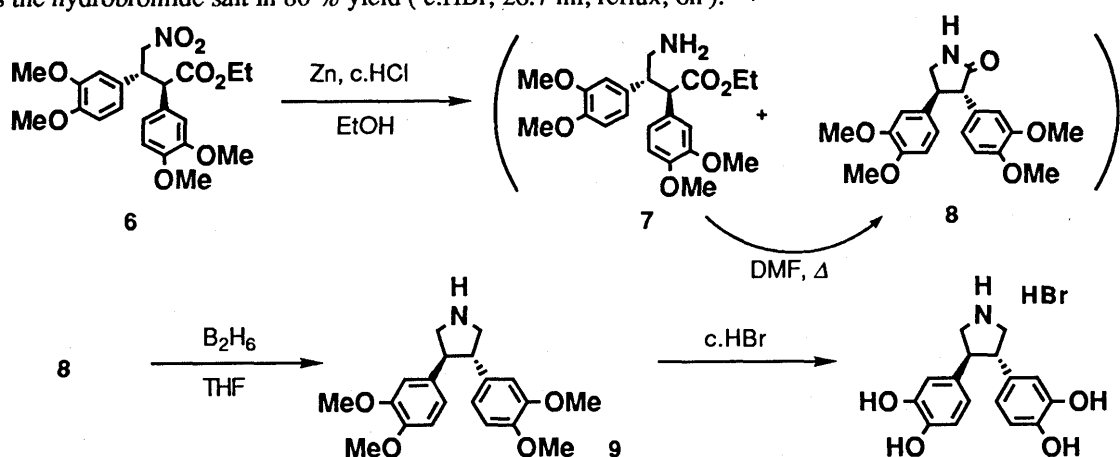


The first part of the synthesis of BDP(2) was the construction of the *trans*- γ -butyrolactam(8). The Michael reaction of ethyl 3,4-dimethoxyphenyl acetate(3)(20 mmol) with 2-(3,4-dimethoxyphenyl)-1-nitroethene(4)(20 mmol) prepared from 3,4-dimethoxybenzaldehyde and nitromethane⁹⁾ yielded ethyl 2,3-bis-(3,4-dimethoxyphenyl)-4-nitrobutyrate (1.2 eq of LDA, THF, -78 °C, 87%). After purification by silica-gel chromatography, the NMR spectra of the adduct indicated that it was a mixture of the two diastereomers(1 : 1.1). The methyl proton of ethyl ester appeared at δ 0.98 and δ 1.21, respectively. The diastereomers could not be separated by TLC, but were separable by means of recrystallization from EtOH. The obtained crystalline adduct was determined to be the *erythro* form (5) for stereochemistry by X-ray analysis.¹⁰⁾ The ORTEP drawing is shown below. This conformational analysis accounts for the chemical shift of the methyl proton on the ethyl ester ($\delta = 0.98$) of 5, which was shielded by the 2-phenyl ring.¹¹⁾



X-Ray Crystal Structure of 5 (*erythro* Form)

Ethyl *threo*-2,3-bis-(3,4-dimethoxyphenyl)-4-nitrobutyrate(6)(14.8mmol), which was obtained from mother liquor (*threo* / *erythro* = 11 / 1), was reduced with Zn(30mmol) and c.HCl(8.9ml) in EtOH(27ml) to afford a mixture of amino ester(7) and γ -butyrolactam(8). On heating the mixture in DMF, the amino ester(7) was cyclized to *trans*- γ -butyrolactam(8) and obtained in 61 % yield. The *trans*- γ -butyrolactam(8)(12.4 mmol) was treated with borane•THF complex (1M solution in THF, 50 ml) to afford the corresponding pyrrolidine(9) (60 %). The demethylation of the methoxyl group of 9 (5.0 mmol) gave BDP(2) as the hydrobromide salt in 80 % yield (c.HBr, 26.7 ml, reflux, 6h).¹²⁾



The biological and pharmacological activities of BDP(2) were characterized by D₁ and D₂ binding assay and the test for the hemodynamics in anesthetised dogs. The binding affinities were determined by a replacement analysis in bindings of D₁(DA₁) antagonist ([³H]-Sch23390) and D₂ agonist ([³H]-spiperone) to membranes of the rat neurostriatum using the methods described previously.¹³⁾ Renal blood flow and mean arterial pressure were measured in anesthetized dogs by intravenous administration.

Table I. Binding and Pharmacological Data of BDP(2)

| Comp. | IC ₅₀ , ^{a)} μ M | | Ratio (D ₁ /D ₂) | ED ₁₅ , ^{b)} μ g/kg | |
|--------------|--|--------------------------------------|---|---|-----|
| | D ₁ binding ^{c)} | D ₂ binding ^{d)} | | MAP | RBF |
| BDP(2) | 7.2 \pm 0.1 | 2.35 | 3.1 | 300 | 100 |
| Dopamine (1) | 3.7 \pm 0.9 | 0.12 | 30.8 | Hypertensive ^{e)} | |

a) Concentration of compound that decreased specific binding of [³H]-Sch23390 or [³H]-spiperone to rat striatum by 50 %. b) The effective dose required to produce 15% of reduction of mean arterial pressure (MAP) or 15 % of increase in renal blood flow (RBF) after intravenous administration to anesthetized dog. Effective doses were determined from at least three doses (N=5). c) Each value was determined from six concentration by linear regression analysis (N=3) expressed as \pm SEM. d) Each value was determined from six concentration by linear regression analysis (N=2). e) Dopamine has α - and β - adrenergic activities in addition to DA₁ activity.

The data listed in the table show that BDP(2) has affinity toward both D₁ receptor and D₂ receptor. Since dopamine itself showed IC₅₀ = 3.7 μM toward D₁ receptor in this test, it was to be estimated that BDP(2) have the equal (or scarcely low) binding affinity toward the D₁ receptor to dopamine. Moreover, D₂ binding affinity of BDP(2) is lower than that of dopamine itself (IC₅₀ = 0.12 μM) and the D₁ / D₂ binding affinity ratio of BDP(2) is 3.1, whereas that of dopamine is 30.8. This result suggests that BDP(2) has more selective affinity toward D₁ receptor than dopamine. In order to clarify that BDP(2) was the agonist toward DA₁ receptor peripherally, we tried to examine the effect on renal blood flow and blood pressure. As shown in the table, BDP(2) increased RBF and reduced MAP at 300 μg/kg of intravenous administration, and these effects fully disappeared after administration of the specific DA₁ antagonist sch23390. These facts and the results of binding affinity indicated that BDP(2) is expected to be the agonist toward DA₁ receptor.

An extension of the present synthetic route and the synthesis of analogues of BDP(2) are now in progress.

REFERENCES AND NOTES

- 1) L. I. Goldberg, *Pharm. Rev.*, **24**, 1 (1972). L. I. Goldberg, *New Engl. J. Med.*, **291**, 707 (1974). T. L. Sourkes, *Psychoneuroendocrinology*, **1**, 69 (1975).
- 2) H. Corrodi, K. Fuxe, T. Hokfelt, P. Lidbrink, and U. Ungerstedt, *J. Pharm. Pharmacol.*, **25**, 409 (1979).
- 3) For example: O. Hornykiewicz, *Nature*, **299**, 484 (1982).
- 4) R. A. Hahn, J. R. Wardell, H. M. Sarau, and P. T. Ridley, *J. Pharm. Exp. Ther.*, **233**, 305 (1982).
- 5) J. W. Keabian and D. B. Calne, *Nature(London)*, **277**, 93 (1979).
- 6) L. I. Goldberg and J. D. Kohli, *Trens Pharmacol. Sci.*, **2**, 64 (1983).
- 7) For example, bromocriptine, lisuride, and pergolide are known as D₂ agonists and (-)-sulpiride and domperidone are known as D₂ antagonist. See J. W. Stoof and J. W. Keabian, *Life Sciences*, **35**, 2281 (1984).
- 8) J. Weinstock, D. L. Ladd, J. W. Wilson, C. K. Brush, N. C. F. Yim, G. Gallagher, Jr., M. E. McCarthy, J. Silvestri, H. M. Sarau, K. E. Flaim, D. M. Ackerman, P. E. Setler, A. J. Tobia, and R. A. Hahn, *J. Med. Chem.*, **29**, 2315 (1986). C. Kaiser and T. Jain, *Med. Res. Rev.*, **5**, 145 (1985). S. K. Kulkarni and A. K. Mehta, *Drugs of Today*, **22**, 29 (1986).
- 9) J. R. McCarthy, J. McCowan, M. B. Zimmerman, M. A. Wenger, and L. W. Emmert, *J. Med. Chem.*, **29**, 2586 (1986).
- 10) Crystal Data. **5**: C₂₂H₂₇NO₈; M = 433.36; monoclinic; P2₁/n(No.14); a=16.029(5)Å; b=11.574(4)Å; c=12.169(5)Å and β=101.93°(3); V=2208.9Å³; z=4; μ(CuKα)=7.9cm⁻¹; no. of reflections with I > 3σ(I)= 2214; no. of refinement parameter=280; final R values, R=0.109; R_w=0.091.
- 11) The J_{2H-3H} values of both adducts could not be measured directly from the signals of ²H because the signals of ²H overlapped with those of OMe proton. **5**: (NMR,400 MHz, CDCl₃) δ 0.98 (3H, t, J= 7.1 Hz), 3.84-3.99(3H, m), 3.87(3H, s), 3.897(3H, s), 3.902(3H, s), 3.94(3H, s),4.04-4.16(1H, m), 4.32(1H, dd, J= 4.3 Hz, 12.4 Hz), 4.42(1H, dd, J= 10.1 Hz, 12.4 Hz), 6.80-6.89(4H, m), 6.99-7.03(2H, m). Mp 144-146 °C. **6**: (NMR,400 MHz, CDCl₃) δ 1.21(3H, t, J= 7.1 Hz), 3.73(3H, s), 3.76(3H, s), 3.78(3H, s), 3.71-3.95(3H, m), 4.06-4.21(1H, m), 4.78(1H, dd, J= 4.9 Hz, 12.4 Hz), 4.86(1H, dd, J= 10.1 Hz, 12.4 Hz), 6.52(1H, m), 6.60-6.68(5H, m).
- 12) **2**: (NMR(400 MHz, D₂O) δ 3.50 (2H, t, J= 11 Hz), 3.50 - 3.62 (2H, m), 3.96(2H, dd, J= 7 Hz, J= 11 Hz), 6.82(2H, dd, J= 2 Hz, J= 8 Hz), 6.91(2H, d, J= 2 Hz), 6.93(2H, d, J= 8 Hz) Hz. Mp 208 - 210 °C. Calcd for C₁₆H₁₇NO₄.HBr C, 52.19; H 4.93; N, 3.80 %. Found C, 52.20; H, 5.17; N, 3.26 %. High resolution mass (FAB) calcd for C₁₆H₁₈NO₄; MH⁺ 288.1236. Found MH⁺ 288.1238.
- 13) W. Billard, V. Ruperto, G. Crosby, L. C. Iorio and A. Barnett, *Life Science*, **35**, 1885 (1984).

(Received January 12, 1993)