## Studies on RA Derivatives. V. Synthesis and Antitumor Activity of Ala<sup>2</sup>-Modified RA-VII Derivatives<sup>1)</sup>

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A number of RA-VII derivatives having various amino acids including proline (6), pipecolic acid (11), norvaline (12), ornithine (14), aspartic acid (15) and methionine (20) in place of Ala<sup>2</sup> have been synthesized from RA-X methyl ester (3) and evaluated for cytotoxicity to P388 leukemia and KB cells *in vitro*. Comparison of the cytotoxicity of these compounds suggests that the polarity and the length of the 2nd amino acid residue affect the activity. An NMR study revealed that, in solution, 6 and 11 are locked in one conformational state, corresponding to conformer A of RA-VII.

**Keywords** RA-VII; cyclic hexapeptide; antitumor activity; cytotoxicity; RA-X methyl ester; conformationally locked derivative

RA-VII (1) is a bicyclic hexapeptide isolated from the roots of Rubia akane and R. cordifolia (Rubiaceae).2) It has attracted much attention because of its significant antileukemic and antitumor activities coupled with its characteristic structure incorporating the isodityrosine moiety. Recently RA-VII-related compounds which have serine (RA-III, 2),2) threonine (RA-VIII),3) pyroglutamic acid (RA-IX) or glutamic acid (RA-X)4) in place of alanine at the 2nd position have been isolated from the same plants. The cytotoxicity of these compounds is estimated to range from 0.0023 to 0.37  $\mu$ g/ml against P388 leukemia cells. These large differences in the activity are presumably due to the changes in the structure at the 2nd position. In order to obtain information about the structural effect of the 2nd amino acid (Aa<sup>2</sup>) side chain on the cytotoxic activity, we planned to prepare a number of Aa<sup>2</sup>-modified RA derivatives which have not been isolated from natural sources.

This paper describes the preparation of Ala<sup>2</sup>-modified RA-VII derivatives, some aspects of their conformations in solution, and the evaluation of their antitumor activity.

Chemistry The starting material was RA-X methyl ester (3).<sup>4)</sup> The manipulation of the ester group of 3 afforded proline (6), pipecolic acid (11), norvaline (12), ornithine (14), aspartic acid (15), homoserine (17), vinyl glycine (19) and methionine (20) derivatives. The synthetic scheme for these derivatives is depicted in Charts 1 and 2.

RA-X methyl ester (3) was reduced with lithium borohydride (LiBH<sub>4</sub>) in tetrahydrofuran (THF) at room temperature to provide the alcohol 4 in 74% yield. The alcohol 4 was treated with methanesulfonyl chloride (MsCl) and triethylamine (Et<sub>3</sub>N) at  $-78\,^{\circ}$ C in dichloromethane (CH<sub>2</sub>Cl<sub>2</sub>) in the presence of 4-dimethylaminopyridine (DMAP) to afford the mesylate 5 in 96% yield. Compound 5 was then cyclized in CH<sub>2</sub>Cl<sub>2</sub>–50% NaOH containing 0.5 eq of tetra-*n*-butylammonium bromide (*n*-Bu<sub>4</sub>NBr) to provide the proline derivative 6 in 82% yield.

Preparation of the pipecolic acid derivative 11 required C<sub>1</sub> elongation, which was conducted by means of the following procedures. The mesylate 5 was reacted with o-nitrophenyl selenocyanate (o-NO<sub>2</sub>PhSeCN) and sodium borohydride (NaBH<sub>4</sub>)<sup>5)</sup> in EtOH, and hydrogen peroxide oxidation of the resultant aryl selenide provided the allylglycine derivative 7 in 94% yield. Compound 7 was

subjected to Lemieux–Jonson oxidation with osmium tetroxide (OsO<sub>4</sub>) and sodium periodate (NaIO<sub>4</sub>) in dioxane– $H_2O$  to provide the aldehyde 8 in 76% yield. Condensation of 8 with ethyl diethylphosphonoacetate and sodium hydride (NaH) in the presence of 18-crown-6 in THF gave  $\alpha,\beta$ -unsaturated ester 9 (72%), which was subjected to catalytic hydrogenation, and successive reduction of the ester group with LiBH<sub>4</sub> provided the alcohol 10 in 31% yield. The alcohol 10 was converted to the pipecolic acid derivative 11 via the mesylate in a manner similar to that described for 6 in 56% yield from 10.

Catalytic hydrogenation of 7 afforded the norvaline derivative 12 in 87% yield. The ornithine derivative 14 was obtained by triphenylphosphine (Ph<sub>3</sub>P) reduction of the azide intermediate 13, which was prepared from the mesylate 5

Aspartic acid (15), vinylglycine (19) and methionine (20) derivatives were prepared from the aldehyde 8. Direct oxidation of the aldehyde 8 with silver(II) oxide (AgO) in THF- $H_2O$  afforded the aspartic acid derivative 15 in poor yield (14%). However, more satisfactory results were obtained by a two-step procedure involving hemiacetal oxidation followed by saponification of the resultant ester. Thus, the aldehyde 8 was reacted with *N*-iodosuccinimide (NIS) in methanol (MeOH)<sup>6)</sup> to afford the methyl ester 16, which was successively treated with lithium hydroxide (LiOH) in MeOH- $H_2O$  to provide 15 in 59% yield from 8.

The aldehyde 8 was reduced with NaBH<sub>4</sub> in 1,4-dioxane, and the resulting alcohol 17 (90%) was mesylated to afford 18 in 71% yield. The vinylglycine derivative 19 was prepared from 18 in the same manner as described for 7 in 17% yield. Substitution reaction of 18 with sodium thiomethoxide (NaSCH<sub>3</sub>) in chloroform (CHCl<sub>3</sub>) or THF gave the oxazine 21 as a major product with a small amount of the desired methionine derivative 20. However, after conversion of 18 into the corresponding iodide, treatment with NaSCH<sub>3</sub> in MeOH provided 20 in 91% yield.

All stereogenic centers of the obtained derivatives remained intact through the reactions, as indicated by comparisons of their chemical shifts and/or coupling constants in the proton (<sup>1</sup>H) and carbon 13 (<sup>13</sup>C) nuclear magnetic resonance (NMR) spectra with those of RA-VII (Table I).

Conformational Information Naturally occurring RAs

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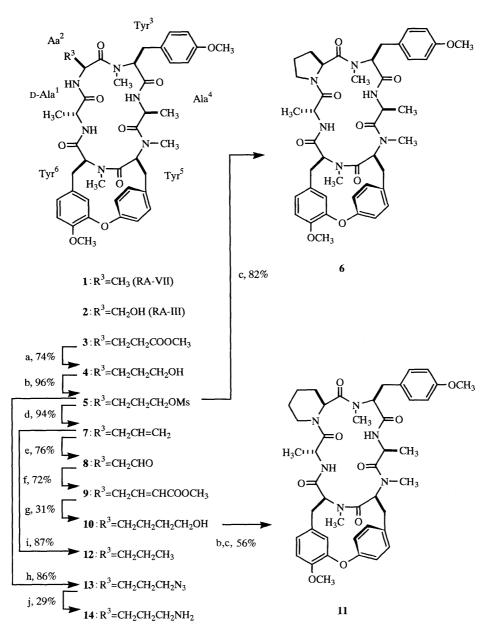


Chart 1. Structure and Synthetic Scheme of RA Derivatives

Ala = L-alanine, Tyr=N,O-dimethyl-L-tyrosine, D-Ala = D-alanine, Aa = L-amino acid. a) LiBH<sub>4</sub>, THF, r.t., 12 h. b) MsCl, DMAP, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h. c) n-Bu<sub>4</sub>Br, CH<sub>2</sub>Cl<sub>2</sub>-50% NaOH, r.t., 1 h. d) o-NO<sub>2</sub>PhSeCN, NaBH<sub>4</sub>, EtOH, r.t., 3 h; H<sub>2</sub>O<sub>2</sub>, THF-EtOH, r.t., 12 h; (CH<sub>3</sub>)<sub>2</sub>S, r.t., 12 h. e) OsO<sub>4</sub>, NaIO<sub>4</sub>, 1,4-dioxane-H<sub>2</sub>O, r.t., 18 h. f) Ethyl diethylphosphonoacetate, NaH, 18-crown-6, THF, r.t., 12 h. g) H<sub>2</sub>, 10% Pd-C, 1,4-dioxane, r.t., 24 h; LiBH<sub>4</sub>, THF, r.t., 24 h. h) NaN<sub>3</sub>, 18-crown-6, CH<sub>2</sub>Cl<sub>2</sub>, reflux, 9 h. i) H<sub>2</sub>, 10% Pd-C, 1,4-dioxane, r.t., 24 h. j) Ph<sub>3</sub>P, H<sub>2</sub>O, THF, r.t., 62 h.

adopt two or three conformational states in solution. Hand 13C-NMR spectroscopy, including two-dimensional (2D) NMR and molecular dynamics calculation studies for RA-VII revealed the two conformers, A and B, observed in CDCl<sub>3</sub> solution to be the trans and cis amide isomers with respect to the peptide bond between Ala<sup>2</sup> and Tyr<sup>3</sup>, respectively.7) The conformer ratio of natural RAs is influenced by structure, solvent and temperature, and the conformer A predominates (ca. 80-100%) in all cases. Similar conformational tendencies were observed in the derivatives prepared in this study. Among them, the proline (6) and pipecolic acid (11) derivatives showed a single conformer in various solvent(s), which suggested that these compounds are conformationally locked in one rigid structure (Table II). To elucidate the solution conformation of 6, conformational analysis was performed.

All proton and carbon resonances of **6** were unambiguously assigned by  $^1H^{-1}H$  correlated spectroscopy (H–H COSY),  $^{8)}$  heteronuclear multiple quantum coherence (HMQC)<sup>9)</sup> and heteronuclear multiple bond connectivity (HMBC)<sup>6)</sup> spectra. The  $^1H^-$  and  $^{13}C^-$ NMR parameters of **6** are quite similar to those of the major conformer of RA-VII. Phase-sensitive nuclear Overhauser effect spectroscopy (NOESYPH)<sup>10)</sup> correlations between  $\text{Pro}^2\text{-}C_\alpha H$  and  $\text{Tyr}^3\text{-}\text{NMe}$ ;  $\text{Tyr}^3\text{-}\text{NMe}$  and  $\text{Tyr}^3\text{-}C_\alpha H$  also suggest the presence of a *trans* peptide bond between  $\text{Pro}^2$  and  $\text{Tyr}^3$  (Fig. 1). The temperature coefficients  $(\Delta \delta/\Delta T)$  of the Ala<sup>4</sup>-NH shifts in dimethyl sulfoxide (DMSO)- $d_6^{-11}$  indicate internal hydrogen bonding to D-Ala<sup>1</sup> carbonyl oxygen, which can stabilize the backbone conformation (Table III).

Circular dichroism (CD) curve of 6 shows positive Cotton

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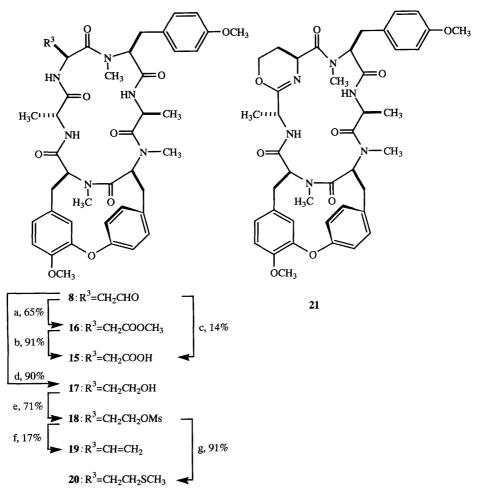


Chart 2 a) NIS,  $K_2CO_3$ ,  $CH_3OH$ , r.t., 7 h. b) LiOH,  $CH_3OH$ :  $H_2O=2:1$ , r.t., 4 h. c) AgO, THF:  $H_2O=9:1$ , reflux, 9 h. d) NaBH<sub>4</sub>, 1,4-dioxane, r.t., 3 h. e) MsCl, DMAP, Et<sub>3</sub>N,  $CH_2Cl_2$ , -78 °C, 2 h. f) o-NO<sub>2</sub>PhSeCN, NaBH<sub>4</sub>, EtOH, r.t., 24 h;  $H_2O_2$ , THF-EtOH, r.t., 12 h;  $(CH_3)_2S$ , r.t., 2 h. g) NaI,  $CH_3COCH_2CH_3$ , reflux, 10 h; NaSCH<sub>3</sub>,  $CH_3OH$ , r.t., 0.5 h.

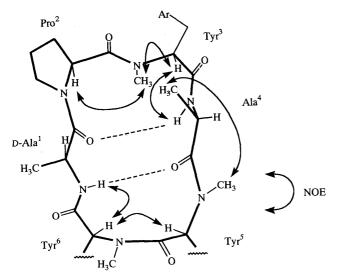


Fig. 1. Selected NOE Correlations of 6 in DMSO-d<sub>6</sub> at 303 K

effects at 287 and 215 nm, and negative ones at 233 and 206 nm. The strong band indicates rigid backbone geometry (Fig. 2).

Biological Results and Discussion Thirteen RA derivatives including intermediates prepared in this study were evaluated for *in vitro* cytotoxicity against P388 leukemia

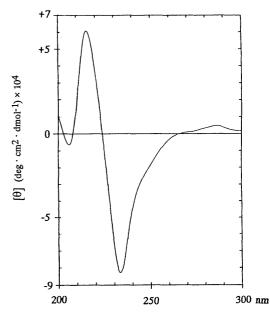


Fig. 2. The CD Spectrum of 6 in CH<sub>3</sub>OH

and KB cells, and RA-VII (1) and III (2) were re-evaluated for comparison (Table IV).

Compounds which possess a polar functionality at the side chain showed reduced activity, especially in the case

TABLE I. 13C-NMR Spectral Data for RA Derivatives

Amino acid ca	arbon	4	6	7	8	9	10	11	12	13	14	15 <sup>a)</sup>	16	17	19	20
D-Ala <sup>1</sup>	$C_{\alpha}$	47.93	46.93	47.96	46.18	47.59	48.00	46.23	47.90	48.06	47.81	46.31	46.23	46.93	47.94	47.79
	$C_{\beta}$	20.70	18.17	20.87	21.05	20.94	20.84	18.18	20.84	20.94	20.89	20.79	20.81	20.82	20.70	20.85
Aa <sup>2</sup>	$C_{\alpha}$	48.94	55.51	48.20	47.95	47.94	49.06	47.84	48.67	48.41	49.25	47.79	47.92	48.09	51.88	47.82
	$\mathbf{C}_{\boldsymbol{\beta}}$	27.76	25.26	35.50	45.87	32.79	30.96	17.97	33.33	25.02	23.68	35.25	35.93	33.47	131.52	30.40
	$\mathbf{C}_{\gamma}$	28.26	28.16	132.06	197.96	142.40	22.04	24.46	18.84	28.48	27.35		52.08	58.70	120.16	30.05
	$C_{\delta}$	61.94	46.78	119.58		125.49	32.11	25.11	13.80	50.84	39.36					15.36
$C_{\varepsilon}$	$C_{\varepsilon}$					60.60	62.17	42.75								
	$\mathbf{C}_{\zeta}$ $\mathbf{C}_{\alpha}$					14.22										
Tyr <sup>3</sup>		68.41	68.28	68.69	69.21	68.84	68.51	67.96	68.56	68.52	68.62	68.92	68.95	68.58	68.57	68.38
	$C_{\beta}$	32.99	32.96	32.92	32.75	32.79	32.97	32.89	32.92	32.92	32.96	32.63	32.82	32.97	32.75	32.84
	$\mathbf{C}_{\gamma}$	130.67		130.86	130.93	130.70	130.78			130.61	130.98		131.03	130.78	130.61	130.57
	$C_{\delta}$		130.20	130.29	130.29	130.12				130.19					130.32	
	$\mathbf{C}_{\varepsilon}$		114.03		114.06	114.18	114.08	114.02	114.00				114.06	114.09	114.04	
	$\mathbf{C}_{\zeta}$		158.30	158.42	158.36	158.45	158.43	158.38	158.39			158.32		158.44		
	$C_N$	39.95	39.91	39.84	40.25	33.93	39.91	39.46	39.81	39.88	40.16	40.21	40.09	39.86	39.68	39.98
	$C_{o}$	55.25	55.24	55.28	55.26	55.23	55.29	55.23	55.23	55.28	55.23	55.23	55.23	55.29	55.25	55.20
Ala <sup>4</sup>	$C_{\alpha}$	46.36	46.27	46.37	43.97	46.30	46.39	45.58	46.32	46.39	46.23	45.52	45.37	46.47	46.51	46.22
	$C_{\beta}$	18.54	18.15	18.60	16.80	18.60	18.60	17.97	18.50	18.58	18.67	18.44	18.67	18.51	18.41	18.51
Tyr <sup>5</sup>	$C_{\alpha}$	54.28	54.86	54.22	55.26	54.19	54.22	54.69	54.19	54.13	54.59	54.26	54.31	54.11	54.21	54.15
	$C_{\beta}$	36.98	36.82	37.04	37.00	37.02	37.03	36.87	36.97	37.05	36.99	36.90	36.99	36.97	36.99	36.91
	$\mathbf{C}_{y}$	135.08	135.72	135.11	135.06		135.09		135.09	135.03	135.33	134.92	135.12		135.10	
	$C_{\delta a}$		132.66	132.79	132.73	132.77	132.78		132.75	132.81	132.69	132.72	132.72	132.91	132.81	132.67
	$C_{\delta b}$		130.87	131.00		130.96	131.00				130.98		130.94	130.91	130.96	
	$C_{\epsilon a}$		124.04					124.06							124.26	
	$C_{\epsilon b}$		125.91		125.95			125.89					125.91	125.90		
	$\mathbf{C}_{\zeta}$		158.18			158.25						158.17			158.24	
	$C_N$	30.51	30.66	30.50	30.47	30.47	30.52	30.52	30.46	30.50	30.58	30.48	30.42	30.56	30.53	30.45
Tyr <sup>6</sup>	$C_{\alpha}$	57.41	56.98	57.44	57.45	57.45	57.45	57.11	57.41	57.47	57.37	57.37	57.49	57.45	57.41	57.30
	$C_{\beta}$	35.44	35.75	35.40	35.29	35.33	35.40	35.72	35.38	35.35	35.46	35.60	35.36	35.30	35.46	35.31
	$\mathbf{C}_{\scriptscriptstyle{y}}$		128.31	128.15	128.05	128.09				128.10	128.34	128.01	128.23	128.10		128.02
	$C_{\delta a}$		121.16							120.89						
	$C_{\delta b}$		113.34							113.42						
•	$C_{\epsilon a}$		112.32							112.32						
	$C_{\varepsilon b}$		153.12							153.14			153.22	153.14		
	$C_{\zeta}$		146.57							146.54			146.58		146.52	
	$C_N$	29.28	29.61	29.24	29.21	29.20	29.26	29.51	29.21	28.48	29.37	29.27	29.19	29.23	29.28	29.18
(	$C_{o}$	56.17	56.15	56.19	56.15	56.16	56.18	56.15	56.17	56.18	56.20	56.09	56.23	56.13	56.18	56.07
$C_{C=0}$			168.59 (T	* _′	167.73					167.71		167.94	167.78		167.87	
			168.67 (T		169.50	167.57				169.48		169.40	169.47	169.46		169.31
				-Ala <sup>1</sup> ) 170.62											170.42	
			170.94 (T		170.78		171.77			171.62					170.62	
			171.71 (A							171.76						
		172.33	172.01 (P	ro <sup>2</sup> ) 172.07	171.97		172.28	172.81	172.24	172.51	172.93			172.55	172.19	172.31
						172.33						172.21	171.95			

In CDCl<sub>3</sub>, major conformer,  $100 \, \text{MHz}$ ,  $\delta$ -values. a) 75 MHz.

TABLE II. Conformer Ratio (A:B) of RA Derivatives

Commound	CDCl <sub>3</sub>			DMSO- $d_6$			$CD_3OD$		
Compound	Α		В	A		В	Α		В
RA-VII <sup>6)</sup>	89	:	11						
6	100	:	0	100	:	0	100	:	C
11	100	:	0						

Table III. Effect of Temperature on the NH Chemical Shift of  ${\bf 6}$  in DMSO- $d_6$ 

	D-Ala <sup>1</sup>	Ala <sup>4</sup>
$\Delta\delta/\Delta T$ (ppm/K)	$4.66 \times 10^{-3}$	$-3.16 \times 10^{-3}$

of ornithine (14) and aspartic acid (15) derivatives. In this connection, it is noteworthy that  $\varepsilon$ -hydroxynorleucine (10) and homoserine (17) derivatives are less toxic than

TABLE IV. Cytotoxicity of RA Derivatives toward P388 Leukemia and KB Cells

Camanad	$IC_{50} (\mu g/ml)$				
Compound	P388	KB			
1 (RA-VII)	0.0013	0.0023			
2 (RA-III)	0.011	0.024			
4	0.14	0.084			
6	0.079	0.074			
7	0.031	0.059			
8	0.027	0.031			
10	0.18	0.14			
11	0.079	0.21			
12	0.011	0.019			
13	0.030	0.013			
14	>10	>10			
15	0.99	1.88			
17	0.048	0.052			
19	0.020	0.042			
20	0.017	0.040			

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Table V. In Vivo Antitumor Activity of RA Derivatives toward P388 Leukemia

Compound	T/C (%) Dose (mg/kg)								
Compound	0.4	1.6	3.13	6.25	12.5	25.0			
1 (RA-VII)	144	152	163	Toxic					
2 (RA-III)	149	156		160	159				
6	101	107		121	128	132			
15	96	103		126					
20	116	146		158					

P388 cells ( $10^6$ ) were transplanted intraperitoneally (i.p.) into CDF<sub>1</sub> mice on day 0 and the compound were administered i.p. on days 1—5. T/C (%)=(mean survival time of tested mice)/(mean survival time of control mice) × 100.

methionine (20) and norvaline (12) derivatives, respectively, having a non-polar residue with similar length.

Another clear relationship exists between the length of the residue and cytotoxicity. In a series of compounds having a hydroxyl group at the end of the side chain, cytotoxicity decreases with increase in the length of the carbon chain (2>17>4>10). The same tendencies are observed among other homologues (e.g. 19>7:1>12). However, the observation that the azido intermediate 13 having a rather long residue shows high cytotoxicity suggests that a lengthy side chain can be compatible with activity in the case of a less polar functionality.

The conformationally locked derivatives, 6 and 11, show potent cytotoxicity, suggesting that this conformation is an active one for expressing the activity.

The proline (6), aspartic acid (15) and methionine (20) derivatives were evaluated for *in vivo* antitumor activity against P388 leukemia (Table V). The observed antitumor activities of these compounds approximately paralleled the cytotoxicities.

## Experimental

Melting points were determined on a Yanagimoto micromelting point apparatus and were not corrected. Infrared (IR) spectra were taken on a Perkin Elmer 1710 spectrophotometer. Optical rotations were measured with a JASCO DIP-360 polarimeter, and  $\lceil \alpha \rceil_D$  values are given in  $10^{-1}$  deg cm<sup>2</sup> g<sup>-1</sup>. The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were recorded on Bruker AM-400, AM-500 and Varian Gemini 300 spectrometers. Chemical shifts were expressed in ppm with tetramethylsilane as an internal standard. The mass spectra (MS) were taken with Hitachi M-80 and VG AutoSpecE spectrometers. The ultraviolet (UV) and visible absorption spectra were recorded on a Shimadzu UV-240 spectrometer. CD spectra were recorded on a JASCO J-702 spectrometer. Silicagel column chromatography was performed with a CIG column system (22 mm i.d. × 100 mm, Kusano Scientific Co., Tokyo) prepacked with  $10\,\mu$  silica gel.

[2-(δ-Hydroxynorvaline)]-RA-VII (4) A solution of LiBH<sub>4</sub> in THF (2 M, 2.0 ml, 4.0 mmol) was added to a solution of 3 (842.4 mg, 1.0 mmol) in THF (30 ml). The mixture was stirred at room temperature for 12 h, then concentrated in vacuo, and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 4 (603 mg, 74%) as a colorless powder, mp 231-232 °C,  $[\alpha]_D - 93.3^\circ$  (c=0.15, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3404, 1673, 1636 cm<sup>-1</sup>. UV  $\lambda_{max}$  (EtOH) nm (log  $\epsilon$ ): 220 (4.48), 278 (3.63), 283 (3.59). High-resolution FAB-MS Calcd for  $C_{43}H_{55}N_6O_9$ : 815.3980 [M+H]<sup>+</sup>, Found: 815.3915. FAB-MS m/z (%): 815 (10,  $\lceil M+1 \rceil^+$ ), 121 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.10 (3H, d, J = 6.7 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.30 (3H, d, J = 6.8 Hz, Ala<sup>1</sup>- $H_{\beta}$ ), 1.55—1.61 (2H, m, Nva<sup>2</sup>- $H_{\beta}$ ), 1.79—1.88 (2H, m, Nva<sup>2</sup>- $H_{\gamma}$ ), 2.63 (1H, dd, J=11.4, 3.0 Hz, Tyr<sup>5</sup>-H<sub> $\beta a$ </sub>), 2.68 (3H, s, Tyr<sup>6</sup>-NMe), 2.93 (3H, s, Tyr<sup>3</sup>-NMe), 2.99 (1H, dd, J = 18.3, 4.2 Hz, Tyr<sup>6</sup>-H<sub> $\beta a$ </sub>), 3.09 (1H, dd,  $J = 18.3, 12.1 \text{ Hz}, \text{Tyr}^6 - \text{H}_{6b}$ , 3.11 (3H, s, Tyr<sup>5</sup>-NMe), 3.27 (1H, dd, J = 14.5,

11.3 Hz,  ${\rm Tyr^3}$ -H<sub>\$\theta\$\), 3.40 (1H, dd, \$J = 14.5, 4.0 Hz,  ${\rm Tyr^3}$ -H<sub>\$\theta\$\), 3.61 (1H, dd, \$J = 11.3, 4.0 Hz,  ${\rm Tyr^3}$ -H<sub>\$\theta\$\), 3.63—3.69 (3H, m,  ${\rm Nva^2}$ -H<sub>\$\theta\$\), 3.78 (3H, s,  ${\rm Tyr^3}$ -OMe), 3.92 (3H, s,  ${\rm Tyr^6}$ -OMe), 4.34 (1H, d, J = 1.8 Hz,  ${\rm Try^6}$ -H<sub>\$\theta\$\), 4.41 (1H, dq, J = 7.1, 6.8 Hz,  ${\rm Ala^1}$ -H<sub>\$\theta\$\), 4.55 (1H, dd, J = 12.0, 4.2 Hz,  ${\rm Tyr^6}$ -H<sub>\$\theta\$\), 4.71—4.76 (2H, m,  ${\rm Ala^1}$ -H<sub>\$\theta\$\, and  ${\rm Nva^2}$ -H<sub>\$\theta\$\), 5.40 (1H, dd, J = 11.4, 3.0 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\), 6.54 (1H, d, J = 7.1 Hz,  ${\rm Ala^1}$ -NH), 6.57 (1H, dd, J = 8.4, 1.8 Hz,  ${\rm Tyr^6}$ -H<sub>\$\theta\$\), 6.57 (1H, dd, J = 8.4, 1.8 Hz,  ${\rm Tyr^6}$ -H<sub>\$\theta\$\), 6.74 (1H, d, J = 8.4 Hz,  ${\rm Tyr^6}$ -H<sub>\$\theta\$\), 6.82 (2H, d, J = 8.6 Hz,  ${\rm Tyr^3}$ -H<sub>\$\theta\$\), 6.87 (1H, dd, J = 8.4, 2.4 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\), 7.05 (2H, d, J = 8.6 Hz,  ${\rm Tyr^6}$ -H<sub>\$\theta\$\), 7.19 (1H, dd, J = 8.4, 2.4 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\), 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\), 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\), 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\)}, 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\)}, 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\)}, 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\)}, 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\)}, 7.25 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz,  ${\rm Tyr^5}$ -H<sub>\$\theta\$\}</sub>), 7.40 (1H, dd, </sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub>

[2-(\delta-Hydroxynorvaline)]-RA-VII Mesylate (5) MsCl (0.43 ml, 5.6 mmol) was added dropwise to a stirred solution of 4 (760 mg, 0.93 mmol), DMAP (340 mg, 2.79 mmol) and Et<sub>3</sub>N (0.38 ml, 2.8 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 ml) at -78 °C. The mixture was stirred at -78 °C for 1 h, diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1) to give 5 (797 mg, 96%) as an amorphous powder. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.10 (3H, d, J=7.5 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.30 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub> $\beta$ </sub>), 1.50—1.90 (4H, m, Nva<sup>2</sup>-H<sub> $\beta$ </sub> and Nva<sup>2</sup>-H<sub> $\gamma$ </sub>), 2.63 (1H, dd, J=11.2, 3.1 Hz, Ala<sup>1</sup>-H<sub> $\beta$ a</sub>), 2.67  $(3H, s, Tyr^6-NMe), 2.92 (3H, s, Tyr^3-NMe), 2.92-3.09 (2H, m, Tyr^6-H<sub>B</sub>),$ 3.02 (3H, s, Nva<sup>2</sup>-OMs), 3.13 (3H, s, Tyr<sup>5</sup>-NMe), 3.31 (1H, dd, J = 14.6, 11.1 Hz,  $Tyr^3-H_{\beta a}$ ), 3.41 (1H, dd, J=14.6, 4.6 Hz,  $Tyr^3-H_{\beta b}$ ), 3.63 (1H, dd, J=11.1, 4.6 Hz, Tyr<sup>3</sup>-H<sub> $\alpha$ </sub>), 3.66 (1H, t, J=11.2 Hz, Tyr<sup>5</sup>-H<sub> $\beta$ b</sub>), 3.78 (3H, s,  $Tyr^3$ -OMe), 3.93 (3H, s,  $Tyr^6$ -OMe), 4.25 (2H, t, J=5.9 Hz, Nva<sup>2</sup>-H<sub> $\delta$ </sub>), 4.34 (1H, d, J=2.1 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.39 (1H, dq, J=8.9, 6.9 Hz, Ala<sup>1</sup>-H<sub>a</sub>), 4.53 (1H, dd, J = 12.0, 4.0 Hz, Tyr<sup>6</sup>-H<sub>a</sub>), 4.72 (1H, qd, J = 7.5, 7.2 Hz, Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 4.79—4.84 (1H, m, Nva<sup>2</sup>-H<sub> $\alpha$ </sub>), 5.38 (1H, dd, J=11.2, 3.1 Hz,  $Tyr^5$ -H<sub> $\alpha$ </sub>), 6.37 (1H, d, J=6.9 Hz,  $Nva^2$ -NH), 6.57 (1H, dd, J=8.4, 2.1 Hz, Tyr<sup>6</sup>-H<sub>δb</sub>), 6.60 (1H, d, J=8.9 Hz, Ala<sup>1</sup>-NH), 6.70 (1H, d, J=7.2Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J=8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.84 (2H, d, J=8.6 Hz,  $Tyr^3-H_{\epsilon}$ ), 6.87 (1H, dd, J=8.4, 2.4 Hz,  $Tyr^5-H_{\epsilon a}$ ), 7.05 (2H, d, J=8.6 Hz, Tyr<sup>3</sup>-H<sub> $\delta$ </sub>), 7.20 (1H, dd, J=8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub> $\epsilon$ b</sub>), 7.26 (1H, dd, J=8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.41 (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-Proline]-RA-VII (6) A mixture of 5 (35.8 mg, 0.040 mmol), n-Bu<sub>4</sub>NBr (6.5 mg, 0.020 mmol), CH<sub>2</sub>Cl<sub>2</sub> (4 ml) and 50% NaOH (1 ml) was stirred vigorously at room temperature for 1 h. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 6 (26.1 mg, 82%) as a colorless powder, mp 294—295°C,  $[\alpha]_D$  –148.1° (c=0.11, CHCl<sub>3</sub>). IR v (CHCl<sub>3</sub>): 3400, 1680, 1651 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$  (EtOH) nm (log  $\varepsilon$ ): 225 (4.52), 277 (3.63), 283 (3.56). High-resolution FAB-MS Calcd for  $C_{43}H_{53}N_6O_9$ : 797.3910  $[M+H]^+$ . Found: 797.3873. FAB-Ms m/z (%): 797 (20,  $[M+H]^+$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>,  $\delta$ ): 0.95 (3H, d, J = 6.7 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.26 (3H, d,  $J = 6.7 \,\text{Hz}$ , Ala<sup>1</sup>-H<sub>\beta</sub>), 1.95—2.05 (3H, m, Pro<sup>2</sup>-H<sub>\beta</sub> and Pro<sup>2</sup>-H<sub>\beta</sub>), 2.59—2.63 (2H, m,  $\text{Pro}^2\text{-H}_{\beta b}$  and  $\text{Tyr}^5\text{-H}_{\beta a}$  (pro-R)), 2.78 (3H, s,  $\text{Tyr}^6\text{-NMe}$ ), 2.85 (1H, dd, J=17.9, 3.7 Hz,  $\text{Tyr}^6\text{-H}_{\beta a}$  (pro-S)), 3.06 (3H, s, Tyr<sup>5</sup>-NMe), 3.17 (1H, dd, J=17.9, 12.0 Hz, Tyr<sup>6</sup>-H<sub>βb</sub> (pro-R)), 3.25 (1H, dd, J=14.1, 10.9 Hz, Tyr<sup>3</sup>-H<sub>fla</sub>), 3.38 (1H, dd, J=14.1, 4.5 Hz,  $Tyr^3-H_{\beta b}$ ), 3.50 (1H, dt, J=9.7, 7.3 Hz,  $Pro^2-H_{\delta a}$ ), 3.58 (1H, dd, J=10.9, 4.5 Hz,  $Tyr^3-H_{\alpha}$ ), 3.63 (1H, dd, J=9.7, 4.4 Hz,  $Pro^2-H_{\delta b}$ ), 3.71 (1H, t, J=11.1 Hz, Tyr<sup>5</sup>-H<sub>\textit{\beta}\text{b}} (pro-S)), 3.78 (3H, s, Tyr<sup>3</sup>-OMe), 3.92 (3H, s,</sub> Tyr<sup>6</sup>-OMe), 4.32 (1H, d, J=1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.50 (1H, dq, J=13.5,  $6.7 \,\mathrm{Hz}$ , Ala<sup>1</sup>-H<sub>a</sub>), 4.55 (1H, dd, J = 12.0, 3.7 Hz, Tyr<sup>6</sup>-H<sub>a</sub>), 4.72 (1H, dd,  $J=7.8, 2.9 \text{ Hz}, \text{Pro}^2-\text{H}_{\alpha}), 4.89 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{H}_{\alpha}), 5.49 \text{ (1H, td, } J=15.5, 6.7 \text{ Hz, Ala}^4-\text{Hz, } J=15.5, 6.7 \text{ Hz, } J=$ dd, J=11.1, 2.5 Hz,  $Tyr^5-H_a$ ), 6.56 (1H, dd, J=8.3, 1.8 Hz,  $Tyr^6-H_{\delta b}$ ),  $6.79 (1H, d, J = 8.3 Hz, Tyr^6 - H_e), 6.82 - 6.86 (2H, m, Ala^1 - NH, Ala^4 - NH),$ 6.83 (2H, d, J = 8.5 Hz, Tyr<sup>3</sup>-H<sub>e</sub>), 6.85 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>ea</sub>), 7.05 (2H, d,  $J = 8.5 \,\text{Hz}$ , Tyr<sup>3</sup>-H<sub>δ</sub>), 7.19 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>sh</sub>), 7.25 (1H, dd, J=8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub>8b</sub>), 7.40 (1H, dd, J=8.4, 2.1 Hz, Tyr5-Hab)

[2-Allylglycine]-RA-VII (7) A mixture of 5 (89.3 mg, 0.10 mmol), o-NO<sub>2</sub>PhSeCN (90.8 mg, 0.40 mmol), NaBH<sub>4</sub> (18.2 mg, 0.48 mmol) and EtOH (10 ml) was stirred at room temperature for 3 h. The mixture was concentrated *in vacuo*, and the résidue was dissolved in  $\text{CH}_2\text{Cl}_2$ . This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried. The solvent was evaporated off *in vacuo* to leave a residue, which was chromatographed on silica gel with  $\text{CH}_2\text{Cl}_2$ -AcOEt—MeOH (12:2:1) to give the selenide. 35% H<sub>2</sub>O<sub>2</sub> (2 ml) was added to the selenide dissolved in THF–EtOH (1:1, 20 ml), and the mixture was stirred

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at room temperature for 12 h. Dimethyl sulfide was added to the mixture at room temperature to decompose excess H2O2. The whole was stirred for 12 h, and concentrated in vacuo. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO3 and brine, and dried over MgSO4. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 7 (75.1 mg, 94%) as a colorless powder, mp 227—228 °C,  $[\alpha]_D$  –199.4° (c=0.10, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3413, 1674, 1636 cm $^{-1}$ . UV  $\lambda_{\rm max}$  (EtOH) nm (log  $\varepsilon$ ): 220 (4.60), 227 (3.78), 283 (3.71). High-resolution FAB-MS Calcd for C<sub>43</sub>H<sub>53</sub>N<sub>6</sub>O<sub>9</sub>: 797.3874  $[M+H]^+$ , Found: 797.3918. MS m/z (%): 796 (20,  $M^+$ ), 121 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.13 (3H, d, J = 6.7 Hz, Ala<sup>4</sup>-H<sub>g</sub>), 1.29 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub> $\beta$ </sub>), 2.42—2.54 (2H, m, Aa<sup>2</sup>-H<sub> $\beta$ </sub>), 2.63 (1H, dd, J=11.3, 2.9 Hz, Tyr<sup>5</sup>-H<sub>ga</sub>), 2.68 (3H, s, Tyr<sup>6</sup>-NMe), 2.92 (3H, s, Tyr<sup>3</sup>-NMe), 3.13 (3H, s, Tyr<sup>5</sup>-NMe), 3.29 (1H, dd, J=14.2, 11.1 Hz,  $Tyr^3-H_{\beta a}$ ), 3.45 (1H, dd, J = 14.2, 4.2 Hz,  $Tyr^3-H_{\beta b}$ ), 3.60 (1H, dd, J = 11.1, 4.2 Hz,  $Tyr^3-H_{\alpha}$ ), 3.67 (1H, t, J=11.3 Hz,  $Tyr^5-H_{\beta b}$ ), 3.79 (3H, s, Tyr<sup>3</sup>-OMe), 3.94 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J=2.1 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.37 (1H, t, J = 6.9 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.54 (1H, dd, J = 12.0, 3.9 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.74 (1H, dq, J=7.5, 6.7 Hz,  $Ala^4-H_a$ ), 4.84 (1H, m,  $Aa^2-H_a$ ), 5.18 (1H, dd, J = 10.3, 1.4 Hz, Aa<sup>2</sup>-H<sub> $\delta a$ </sub>), 5.22 (1H, dd, J = 17.1, 1.4 Hz, Aa<sup>2</sup>-H<sub> $\delta b$ </sub>), 5.40 (1H, dd, J = 11.4, 3.2 Hz, Tyr<sup>5</sup>-H<sub>a</sub>), 5.67—5.77 (1H, m, Aa<sup>2</sup>-H<sub>y</sub>), 6.16 (1H, d, J = 8.7 Hz, Aa<sup>2</sup>-NH), 6.38 (1H, d, J = 6.9 Hz, Ala<sup>1</sup>-NH), 6.58 (1H, dd, J = 8.3, 2.1 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.69 (1H, d, J = 7.5 Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J = 8.3 Hz,  $Tyr^6 - H_{\epsilon}$ ), 6.83 (2H, d, J = 8.6 Hz,  $Tyr^3 - H_{\epsilon}$ ), 6.87 (1H, dd,  $J=8.4, 2.4 \text{ Hz}, \text{ Tyr}^5 - H_{\epsilon a}$ , 7.08 (2H, d,  $J=8.6 \text{ Hz}, \text{ Tyr}^3 - H_{\delta}$ ), 7.21 (1H, dd, J=8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>εb</sub>), 7.26 (1H, dd, J=8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub>δa</sub>), 7.41 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-( $\beta$ -Formylalanine)]-RA-VII (8) OsO<sub>4</sub> (20.0 mg, 0.080 mmol) and NaIO<sub>4</sub> (160.4 mg, 0.75 mmol) were added to a solution of 7 (119.5 mg, 0.15 mmol) in 1,4-dioxane-H<sub>2</sub>O (4:1, 10 ml) and the mixture was stirred at room temperature for 18 h. The white precipitate was removed by filtration, then the filtrate was diluted with CH2Cl2, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 8 (91.8 mg, 76%) as a colorless powder, mp 322—323 °C,  $[\alpha]_D$  –129.8°  $(c = 0.13, \text{ CHCl}_3)$ . IR  $\nu$  (CHCl<sub>3</sub>): 3405, 1725, 1673, 1634 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$ (EtOH) nm ( $\log \varepsilon$ ): 224 (4.44), 277 (3.64), 283 (3.58). High-resolution FAB-MS Calcd for  $C_{42}H_{51}N_6O_{10}$ : 799.3667 [M+H]<sup>+</sup>. Found: 799.3722. MS m/z (%): 798 (100, M<sup>+</sup>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.12  $(3H, d, J=6.7 Hz, Ala^4-H_g)$ , 1.28  $(3H, d, J=7.0 Hz, Ala^4-H_g)$ , 2.63  $(1H, d, J=6.7 Hz, Ala^4-H_g)$ dd, J=11.4, 3.2 Hz, Tyr<sup>5</sup>-H<sub>βa</sub>), 2.66 (3H, s, Tyr<sup>6</sup>-NMe), 2.83 (1H, dd,  $J = 18.5, 5.0 \text{ Hz}, \text{Aa}^2 - \text{H}_{\beta a}$ , 2.92—2.95 (1H, m, Tyr<sup>6</sup>-H<sub>\theta a</sub>), 3.04—3.10 (1H, m,  $Tyr^6-H_{\beta b}$ ), 3.12 (3H, s,  $Tyr^3$ -NMe), 3.15 (1H, dd, J=14.2, 10.3 Hz, Tyr<sup>3</sup>-H<sub>8a</sub>), 3.17 (3H, s, Tyr<sup>5</sup>-NMe), 3.23 (1H, dd, J = 18.5, 9.2 Hz, Aa<sup>2</sup>-H<sub>8b</sub>), 3.42 (1H, dd, J = 14.2, 3.9 Hz, Tyr<sup>3</sup>-H<sub> $\beta$ b</sub>), 3.58 (1H, dd, J = 10.3, 3.9 Hz,  $\text{Tyr}^3 - \text{H}_{\alpha}$ ), 3.66 (1H, t, J = 11.4 Hz,  $\text{Tyr}^5 - \text{H}_{\beta b}$ ). 3.79 (3H, s,  $\text{Tyr}^3 - \text{OMe}$ ), 3.93 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J = 1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.36 (1H, dq, J = 7.1, 7.0 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.56 (1H, dd, J=11.9, 3.8 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.70 (1H, dq, J=7.3, 6.7 Hz, Ala<sup>4</sup>-H<sub>a</sub>), 5.24 (1H, ddd, J=9.2, 8.6, 5.9 Hz, Aa<sup>2</sup>-H<sub>a</sub>), 5.38 (1H, dd, J = 11.4, 3.2 Hz, Tyr<sup>3</sup>-H<sub>a</sub>), 6.40 (1H, d, J = 7.1 Hz, Ala<sup>1</sup>-NH), 6.58 (1H, dd, J = 8.4, 1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.77 (1H, d, J = 7.3 Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J = 8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.87 (2H, d, J = 8.6 Hz, Tyr<sup>3</sup>-H<sub>e</sub>), 6.87 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>za</sub>). 6.94 (1H, d, J = 8.6 Hz, Ala<sup>2</sup>-NH), 7.05  $(2H, d, J=8.6 Hz, Tyr^3-H_{\delta}), 7.21 (1H, dd, J=8.4, 2.4 Hz, Tyr^5-H_{sb}), 7.25$ (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.40 (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>), 9.74 (1H, s, Aa2-CHO).

[2-[(E)-2-Amino-4-hexene-1,6-dicarboxylic acid]]-RA-VII Ethyl Ester (9) Ethyl diethylphosphonoacetate (126  $\mu$ l, 0.63 mmol), NaH (60%, 25.2 mg, 0.63 mmol) and 18-crown-6 (40.4 mg, 0.15 mmol) were added to a solution of **8** (235.8 mg, 0.29 mmol) in THF (15 ml) at 0 °C. The mixture was stirred at room temperature for 12 h, then concentrated *in vacuo*. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off *in vacuo* to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1) to give **9** (185.7 mg, 72%) as an amorphous powder. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.12 (3H, d, J=6.6 Hz, Ala<sup>4</sup>-H<sub> $\rho$ </sub>), 1.27 (3H, t, J=7.1 Hz, Aa<sup>2</sup>-COOCH<sub>2</sub>CH<sub>3</sub>), 1.28 (3H, d, J=7.0 Hz, Ala<sup>1</sup>-H $_{\rho}$ ), 2.61—2.64 (3H, m, Aa<sup>2</sup>-H $_{\rho}$  and Tyr<sup>5</sup>-H $_{\rho a}$ ), 2.67 (3H, s, Tyr<sup>6</sup>-NMe), 2.91 (3H, s, Tyr<sup>3</sup>-NMe), 2.95 (1H, dd, J=20.3, 4.1 Hz, Tyr<sup>6</sup>-H $_{\rho a}$ ), 3.08 (1H, dd, J=20.3, 12.1 Hz, Tyr<sup>6</sup>-H $_{\rho b}$ ), 3.13 (3H, s, Tyr<sup>5</sup>-NMe), 3.26 (1H, dd, J=14.1, 11.2 Hz,

Tyr³-H<sub>βa</sub>), 3.42 (1H, dd, J=14.1, 4.1 Hz, Tyr³-H<sub>βb</sub>), 3.58 (1H, dd, J=11.2, 4.1 Hz, Tyr³-H<sub>α</sub>), 3.66 (1H, t, J=11.4 Hz, Tyr⁵-H<sub>βb</sub>), 3.78 (3H, s, Tyr³-OMe), 3.93 (3H, s, Tyr³-OMe), 4.20 (2H, q, J=7.1 Hz, Aa²-COOCH<sub>2</sub>CH<sub>3</sub>), 4.33 (1H, d, J=1.9 Hz, Tyr⁵-H<sub>α</sub>), 4.39 (1H, m, Ala¹-H<sub>α</sub>), 4.54 (1H, dd, J=12.1, 4.1 Hz, Tyr⁵-H<sub>α</sub>), 4.72 (1H, dq, J=7.4, 6.6 Hz, Ala⁴-H<sub>α</sub>), 4.92 (1H, m, Aa²-H<sub>α</sub>), 5.38 (1H, dd, J=11.4, 3.1 Hz, Tyr⁵-H<sub>α</sub>), 5.97 (1H, d, J=15.6 Hz, Aa²-H<sub>δ</sub>), 6.34 (1H, d, J=7.0 Hz, Ala¹-NH), 6.50 (1H, d, J=9.5 Hz, Aa²-NH), 6.57 (1H, dd, J=8.4, 1.8 Hz, Tyr⁵-H<sub>δ</sub>), 6.69 (1H, d, J=7.4 Hz, Ala⁴-NH), 6.78—6.89 (1H, m, Aa²-H<sub>γ</sub>), 6.79 (1H, dd, J=8.4, Hz, Tyr⁵-H<sub>δ</sub>), 6.83 (2H, d, J=8.6 Hz, Tyr³-H<sub>δ</sub>), 6.87 (1H, dd, J=8.4, 2.4 Hz, Tyr⁵-H<sub>δ</sub>), 7.03 (2H, d, J=8.6 Hz, Tyr³-H<sub>δ</sub>), 7.20 (1H, dd, J=8.4, 2.4 Hz, Tyr⁵-H<sub>δ</sub>), 7.26 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.26 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.42 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.26 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.42 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.42 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.40 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.40 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.40 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.40 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.40 (1H, dd, J=8.4, 2.2 Hz, Tyr⁵-H<sub>δ</sub>), 7.56 (1H, dd, J=8.

[2-(\varepsilon-Hydroxynorleucine)]-RA-VII (10) Palladium (10%) on activated carbon (100 mg) was added to a solution of 9 (185.7 mg, 0.21 mmol) in 1,4-dioxane (20 ml), and the mixture was stirred vigorously at room temperature for 24 h under hydrogen. The mixture was filtered, and the filtrate was concentrated in vacuo. The residue was dissolved in THF (30 ml), and LiBH<sub>4</sub> (2 m in THF, 1.05 ml, 2.3 mmol) was added to the solution at 0 °C. Stirring was continued at room temperature for 24 h, then the mixture was concentrated in vacuo. The residue was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 10 (75.4 mg, 31%) as a colorless powder, mp 227—228 °C,  $[\alpha]_D$  –176.3°  $(c = 0.14, \text{CHCl}_3)$ . IR  $\nu$  (CHCl<sub>3</sub>): 3403, 1674, 1636 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$  (EtOH) nm (log ε): 221 (4.49), 276 (3.62), 282 (3.56). High-resolution FAB-MS Calcd for  $C_{44}H_{57}N_6O_{10}$ : 829.4136 [M+H]<sup>+</sup>. Found: 829.4054. FAB-MS m/z (%): 829 (10, [M+H]<sup>+</sup>). 121 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.13 (3H, d, J = 6.7 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.31 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub> $\beta$ </sub>), 1.33—1.46 (2H, m, Nle<sup>2</sup>-H<sub> $\beta$ </sub>), 1.54—1.78 (4H, m, Nle<sup>2</sup>-H, and Nle<sup>2</sup>-H<sub> $\delta$ </sub>), 2.63 (1H, dd, J = 11.4, 3.2 Hz, Tyr<sup>5</sup>-H<sub> $\beta$ </sub>), 2.68 (3H, s, Tyr<sup>6</sup>-NMe), 2.93 (3H, s, Tyr<sup>3</sup>-NMe), 2.95 (1H, dd, J=18.1, 3.9 Hz, Tyr<sup>6</sup>-H<sub>ga</sub>), 3.10 (1H, dd, J = 18.1, 12.0 Hz, Tyr<sup>6</sup>-H<sub> $\beta$ b</sub>), 3.13 (3H, s, Tyr<sup>5</sup>-NMe), 3.31 (1H, dd, J = 14.2, 11.1 Hz, Tyr<sup>3</sup>-H<sub> $\beta a$ </sub>), 3.44 (1H, dd, J = 14.2, 4.3 Hz, Tyr<sup>3</sup>-H<sub> $\beta b$ </sub>), 3.62 (1H, dd, J = 11.1, 4.3 Hz, Tyr<sup>3</sup>-H<sub>a</sub>), 3.64—3.70 (3H, m, Nle<sup>2</sup>-H<sub>e</sub> and  $Tyr^{5}-H_{gb}$ ), 3.79 (3H, s,  $Tyr^{3}$ -OMe), 3.94 (3H, s,  $Tyr^{6}$ -OMe), 4.34 (1H, d, J=1.9 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.37 (1H, dq, J=6.9, 6.8 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.54 (1H, dd, J = 12.0, 3.9 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.70—4.79 (2H, m, Nle<sup>2</sup>-H<sub> $\alpha$ </sub> and Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 5.41 (1H, dd, J = 11.4, 3.2 Hz, Tyr<sup>5</sup>-H<sub> $\alpha$ </sub>), 6.42 (1H, d, J = 6.8 Hz, Ala<sup>1</sup>-NH), 6.45 (1H, d, J = 8.6 Hz, Nle<sup>2</sup>-NH), 6.58 (1H, dd, J = 8.4, 1.9 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.72 (1H, d, J = 7.5 Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J = 8.4 Hz, Tyr<sup>6</sup>-H<sub>8</sub>), 6.84  $(2H, d, J=8.6 Hz, Tyr^3-H_c)$ , 6.88 (1H, dd, J=8.4, 2.4 Hz,  $Tyr^5-H_{ca}$ ), 7.08  $(2H, d, J=8.6 Hz, Tyr^3-H_{\delta}), 7.21 (1H, dd, J=8.4, 2.4 Hz, Tyr^5-H_{\epsilon b}), 7.26$ (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.42 (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-Pipecolic acid]-RA-VII (11) MsCl (42  $\mu$ l, 0.54 mmol) was added dropwise to a solution of 10 (75.3 mg, 0.090 mmol), DMAP (33.0 mg, 0.27 mmol) and Et<sub>3</sub>N (38  $\mu$ l, 0.27 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (8 ml) at  $-78\,^{\circ}$ C, and stirring was continued for 1 h. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1) to afford the mesylate (58.1 mg) as an amorphous powder. A mixture of the mesylate (58.1 mg, 0.064 mmol), n-Bu<sub>4</sub>NBr (10.3 mg, 0.032 mmol), CH<sub>2</sub>Cl<sub>2</sub> (4 ml) and 50% NaOH (1 ml) was stirred vigorously at room temperature for 1 h. The mixture was diluted with CH2Cl2, washed successively with H2O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 11 (41.6 mg, 56% from 10) as a colorless powder, mp 250—251 °C,  $[\alpha]_D$  —210.2° (c=0.11, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3392, 1681, 1646 cm<sup>-1</sup>. UV  $\lambda_{max}$  (EtOH) nm ( $\log \varepsilon$ ): 220 (4.52), 277 (3.59), 282 (3.54). High-resolution FAB-MS Calcd for  $C_{44}H_{55}N_6O_9$ : 811.4031  $[M+H]^+$ . Found: 811.4089. FAB-MS m/z (%): 811 (5,  $[M+H]^{+}$ ).  $^{1}H-NMR$  (CDCl<sub>3</sub>,  $\delta$ ): 0.97 (3H, d, J=6.7 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.24 (3H, d, J = 6.9 Hz, Ala-<sup>1</sup>H<sub> $\beta$ </sub>), 1.45—1.52 (1H, m, Pip<sup>2</sup>-H<sub> $\delta a$ </sub>), 1.63—1.72 (2H, m,  $Pip^2-H_{\beta}$ ), 1.77—1.81 (1H, m,  $Pip^2-H_{\gamma a}$ ), 1.85—1.88 (1H, m,  $Pip^2-H_{\delta b}$ ), 2.05—2.10 (1H, m,  $Pip^2-H_{\gamma b}$ ), 2.60 (1H, dd, J=11.2, 2.7 Hz,  $Tyr^5-H_{\beta a}$ ), 2.74 (3H, s,  $Tyr^6-NMe$ ), 2.83 (3H, s,  $Tyr^3-NMe$ ), 2.89 (1H, dd,  $J = 18.0, 3.7 \text{ Hz}, \text{Tyr}^6 - \text{H}_{\beta a}$ ), 3.06 (3H, s, Tyr<sup>5</sup>-NMe), 3.13 (1H, dd, J = 18.0, 12.0 Hz, Tyr<sup>6</sup>-H<sub> $\beta$ b</sub>), 3.32 (1H, dd, J = 14.4, 10.3 Hz, Tyr<sup>3</sup>-H<sub> $\beta$ a</sub>), 3.35 (1H, dd, J=14.4, 5.3 Hz, Tyr<sup>3</sup>-H<sub>βb</sub>), 3.53 (1H, m, Pip<sup>2</sup>-H<sub>εa</sub>), 3.53 (1H, dd, J=10.3, 5.3 Hz, Tyr<sup>3</sup>-H<sub>a</sub>), 3.68 (1H, t, J=11.2 Hz, Tyr<sup>5</sup>-H<sub>βb</sub>), 3.92 (1H,

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dd, J = 22.6, 2.3 Hz, Pip²-H<sub>εb</sub>), 4.33 (1H, d, J = 1.8 Hz, Tyr²-H<sub>δa</sub>), 4.54 (1H, dd, J = 12.0, 3.7 Hz, Tyr²-H<sub>α</sub>), 4.64 (1H, qd, J = 6.9, 6.7 Hz, Ala¹-H<sub>α</sub>), 4.79 (1H, dq, J = 8.2, 6.7 Hz, Ala⁴-H<sub>α</sub>), 5.08 (1H, dd, J = 6.3, 1.8 Hz, Pip²-H<sub>α</sub>), 5.45 (1H, dd, J = 11.2, 2.7 Hz, Tyr²-H<sub>α</sub>), 6.55 (1H, dd, J = 8.6, 1.8 Hz, Tyr²-H<sub>δb</sub>), 6.67 (1H, d, J = 8.2 Hz, Ala⁴-NH), 6.73 (1H, d, J = 8.6 Hz, Tyr²-H<sub>ε</sub>), 6.82 (2H, d, J = 8.5 Hz, Tyr³-H<sub>ε</sub>), 6.86 (1H, dd, J = 8.4, 2.4 Hz, Tyr²-H<sub>ε</sub>), 7.04 (2H, d, J = 8.5 Hz, Tyr³-H<sub>δ</sub>), 7.19 (1H, dd, J = 8.4, 2.4 Hz, Tyr²-H<sub>ε</sub>), 7.25 (1H, dd, J = 8.4, 2.1 Hz, Tyr²-H<sub>δb</sub>).

[2-Norvaline]-RA-VII (12) Palladium (10%) on activated carbon (20 mg) was added to a solution of 7 (39.8 mg, 0.050 mmol) in 1,4-dioxane (5 ml), and the mixture was stirred vigorously at room temperature for 24h under hydrogen. The mixture was filtered, and the filtrate was concentrated in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH to give 12 (34.8 mg, 87%) as a colorless powder, mp 227—228 °C,  $[\alpha]_D$  –204.5° (c=0.16, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3410, 1674, 1636 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$  (EtOH) nm (log  $\varepsilon$ ): 223 (4.73), 277 (3.85), 282 (3.80). High-resolution FAB-MS Calcd for C<sub>43</sub>H<sub>55</sub>N<sub>6</sub>O<sub>9</sub>: 799.4031 [M+H]<sup>+</sup> Found: 799.4043. FAB-MS m/z (%): 799 (20, [M+H]<sup>+</sup>). 164 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 0.93 (3H, t, J=7.3 Hz, Nva<sup>2</sup>-H<sub> $\beta$ </sub>), 1.11 (3H, d, J=6.7 Hz, Ala<sup>4</sup>-H<sub>8</sub>), 1.28 (3H, d, J=6.9 Hz, Ala<sup>1</sup>-H<sub>8</sub>), 1.27—1.36 (2H, m, Nva<sup>2</sup>-H<sub>g</sub>), 2.63 (1H, dd, J=11.7, 3.1 Hz, Tyr<sup>5</sup>-H<sub>ga</sub>), 2.67 (3H, s, Tyr<sup>6</sup>-NMe), 2.91 (3H, s, Tyr<sup>3</sup>-NMe), 2.96—3.08 (2H, m,  $Tyr^6-H_6$ ), 3.11 (3H, s,  $Tyr^5-NMe$ ), 3.29 (1H, dd, J=14.2, 11.0 Hz,  $Tyr^3-H_{\beta a}$ ), 3.43 (1H, dd, J=14.2, 4.3 Hz,  $Tyr^5-H_{\beta b}$ ), 3.61 (1H, dd, J=11.0, 4.3 Hz,  $Tyr^3-H_{\alpha}$ ), 3.65 (1H, t, J=11.7 Hz,  $Tyr^5-H_{\beta b}$ ), 3.78 (3H, s,  $Tyr^3$ -OMe), 3.92 (3H, s,  $Tyr^6$ -OMe), 4.34 (1H, d, J=2.0 Hz,  $Tyr^6$ -H<sub> $\delta a$ </sub>), 4.37 (1H, t, J = 6.9 Hz, Ala<sup>1</sup>-H<sub>a</sub>), 4.53 (1H, dd, J = 12.0, 3.9 Hz, Tyr<sup>6</sup>-H<sub>a</sub>), 4.71—4.79 (2H, m, Nva<sup>2</sup>-H<sub> $\alpha$ </sub> and Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 5.39 (1H, dd, J=11.7, 3.1 Hz,  $Tyr^5-H_a$ ), 6.42 (1H, d, J=6.9 Hz, Ala<sup>1</sup>-NH), 6.56 (1H, d, J=8.4 Hz, Nva<sup>2</sup>-NH), 6.57 (1H, dd, J = 8.4, 2.0 Hz, Tyr<sup>6</sup>-H<sub> $\delta$ b</sub>), 6.69 (1H, d, J = 7.5 Hz, Ala<sup>4</sup>-NH), 6.79 (1H, d, J=8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.82 (2H, d, J=8.6 Hz,  $Tyr^3-H_e$ , 6.86 (1H, dd, J=8.4, 2.4 Hz,  $Tyr^5-H_{ea}$ ), 7.07 (2H, d, J=8.6 Hz,  $\text{Tyr}^3$ - $\text{H}_{\delta}$ ), 7.20 (1H, dd, 8.4, 2.4 Hz,  $\text{Tyr}^5$ - $\text{H}_{\epsilon b}$ ), 7.25 (1H, dd, J=8.4, 2.3 Hz,  $\text{Tyr}^6\text{-H}_{\delta a}$ ), 7.41 (1H, dd, J = 8.4, 2.3 Hz,  $\text{Tyr}^6\text{-H}_{\delta b}$ ).

[2-(\delta-Azidonorvaline)]-RA-VII (13) NaN<sub>3</sub> (588.7 mg, 9.0 mmol), 18crown-6 (158.6 mg, 0.60 mmol) and 5 (268.9 mg, 0.30 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 ml) and the solution was refluxed for 9 h. The mixture was concentrated in vacuo. The residue was dissolved in CH2Cl2. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 13 (218.5 mg, 86%) as a colorless powder, mp 211-212 °C, [ $\alpha$ ]<sub>D</sub>  $-209.5^{\circ}$ C (c=0.11, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3420, 2103, 1677,  $1634\,\mathrm{cm^{-1}}$ . UV  $\lambda_{\mathrm{max}}$  (EtOH) nm (log ε): 225 (4.53), 277 (3.66), 285 (3.60). High-resolution FAB-MS Calcd for  $C_{43}H_{53}N_6O_9$ : 840.4043 [M+H]<sup>+</sup> Found: 840.4043. FAB-MS m/z(%): 840 (70, [M+H]<sup>+</sup>), 164 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.14 (3H, d, J = 6.6 Hz, Ala<sup>4</sup>-H<sub>8</sub>), 1.31 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub>B</sub>), 1.54—1.63 (2H, m, Nva<sup>2</sup>-H<sub>B</sub>), 1.76—1.82 (2H, m, Nva<sup>2</sup>-H<sub>y</sub>), 2.64 (1H, dd, J=11.2, 3.0 Hz, Tyr<sup>5</sup>-H<sub>βa</sub>), 2.67 (3H, s,  $Tyr^6$ -NMe), 2.91 (3H, s,  $Tyr^3$ -NMe), 2.96 (1H, dd, J=17.7, 3.7 Hz,  $Tyr^6-H_{ga}$ ), 3.10 (1H, dd, J=17.7, 11.8 Hz,  $Tyr^6-H_{gb}$ ), 3.14 (3H, s, Tyr<sup>5</sup>-NMe), 3.33 (1H, dd, J = 14.2, 11.1 Hz, Tyr<sup>3</sup>-H<sub>ga</sub>), 3.33—3.37 (2H, m, Nva<sup>2</sup>-H<sub> $\delta$ </sub>), 3.44 (1H, dd, J=14.2, 4.4Hz, Tyr<sup>3</sup>-H<sub> $\beta$ b</sub>), 3.63 (1H, dd, J=11.1, 4.4Hz, Tyr<sup>3</sup>-H<sub> $\alpha$ </sub>), 3.67 (1H, t, J=11.2Hz, Tyr<sup>5</sup>-H<sub> $\beta$ b</sub>), 3.79 (3H, s,  $Tyr^3$ -OMe), 3.94 (3H, s,  $Tyr^6$ -OMe), 4.34 (1H, d, J=1.8 Hz,  $Tyr^6$ -H<sub>ôa</sub>), 4.37 (1H, dq, J=8.8, 6.6 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.53 (1H, dd, J=11.8, 3.7 Hz,  $Tyr^6-H_{\alpha}$ ), 4.70 (1H, dq, J=7.3, 6.9 Hz,  $Ala^4-H_{\alpha}$ ), 4.81 (1H, m,  $Nva^2-H_{\alpha}$ ), 5.39 (1H, dd, J = 11.2, 3.0 Hz, Tyr<sup>5</sup>-H<sub> $\alpha$ </sub>), 6.24 (1H, d, J = 8.8 Hz, Ala<sup>1</sup>-NH), 6.35 (1H, d, J = 6.8 Hz, Ala<sup>2</sup>-NH), 6.58 (1H, dd, J = 8.4, 1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.69 (1H, d, J=7.3 Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J=8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.84 (2H, d, J = 8.6 Hz, Tyr<sup>3</sup>-H<sub>e</sub>), 6.88 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>ea</sub>), 7.07  $(2H, d, J=8.6 \text{ Hz}, \text{Tyr}^3-H_{\delta}), 7.21 \text{ (1H, dd, } J=8.4, 2.4 \text{ Hz}, \text{Tyr}^5-H_{\epsilon b}), 7.27$ (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.42 (1H, dd, J = 8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-Ornithine]-RA-VII (14) A mixture of 13 (153.5 mg, 0.18 mmol),  $Ph_3P$  (94.4 mg, 0.36 mmol),  $H_2O$  (50  $\mu$ l) and THF (10 ml) was stirred at room temperature for 62 h. The mixture was concentrated *in vacuo*, and the residue was dissolved in  $CH_2Cl_2$ . This solution was washed successively with  $H_2O$ , 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off *in vacuo* to leave a residue, which was chromatographed on silica gel with  $CH_2Cl_2$ -AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 14 (43.8 mg, 29%) as a colorless powder, mp 195—197 °C,  $[\alpha]_D$  – 129.8°

 $(c = 0.13, \text{ CHCl}_3)$ . IR  $\nu$  (CHCl<sub>3</sub>): 3392, 1662 cm<sup>-1</sup>. UN  $\lambda_{\text{max}}$  (EtOH) nm (log  $\varepsilon$ ): 220 (4.35), 277 (3.45), 282 (3.39). High-resolution FAB-MS Calcd for C<sub>43</sub>H<sub>56</sub>N<sub>7</sub>O<sub>9</sub>: 814.4140 [M+H]<sup>+</sup>. Found: 814.4089. FAB-MS m/z(%): 814 (20,  $[M+H]^+$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.06  $(3H, d, J = 6.6 \text{ Hz}, \text{Ala}^4 - \text{H}_{\beta}), 1.35 (3H, d, J = 6.6 \text{ Hz}, \text{Ala}^4 - \text{H}_{\beta}), 1.77 - 1.79$  $(2H, m, Orn^2-H_y)$ , 1.97—2.06  $(2H, m, Orn^2-H_\delta)$ , 2.19—2.27  $(2H, m, Orn^2-H_\delta)$ Orn<sup>2</sup>-H<sub> $\beta$ </sub>), 2.61 (1H, dd, J=11.2, 2.5 Hz, Tyr<sup>5</sup>-H<sub> $\beta a$ </sub>), 2.68 (3H, s, Tyr<sup>6</sup>-NMe), 2.89—2.94 (1H, m, Tyr<sup>6</sup>-H<sub> $\beta$ a</sub>), 2.96 (3H, s, Tyr<sup>3</sup>-NMe), 3.06-3.10 (1H, m,  $Tyr^6-H_{\beta b}$ ), 3.08 (3H, s,  $Tyr^3-NMe$ ), 3.20 (1H, dd,  $J = 13.5, 10.8 \text{ Hz}, \text{Tyr}^3 - \text{H}_{\beta a}), 3.34 (1\text{H}, \text{dd}, J = 13.5, 4.4 \text{ Hz}, \text{Tyr}^3 - \text{H}_{\beta b}), 3.61$  $(1H, dd, J = 10.8, 4.4 Hz, Tyr^3 - H_{\alpha}), 3.71 (4H, m, Tyr^3 - OMe and Tyr^5 - H_{\beta b}),$ 3.93 (3H, s, Tyr<sup>6</sup>-OMe), 4.35 (1H, d, J=1.4 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.49—4.56 (2H, m, Ala<sup>1</sup>-H<sub> $\alpha$ </sub> and Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.61 (1H, m, Orn<sup>2</sup>-H<sub> $\alpha$ </sub>), 4.73 (1H, dq, J=8.5, 6.6 Hz,  $Ala^4-H_a$ ), 5.42 (1H, dd, J=11.2, 2.5 Hz,  $Tyr^5-H_a$ ), 6.43  $(1H, d, J = 6.9 \text{ Hz}, \text{Ala}^{1} - \text{NH}), 6.57 (1H, dd, J = 8.2, 1.4 \text{ Hz}, \text{Tyr}^{6} - \text{H}_{\delta b}), 6.79$ (1H, d, J=8.2 Hz,  $Tyr^6-H_e$ ), 6.81 (2H, d, J=8.3 Hz,  $Tyr^3-H_e$ ), 6.86 (1H, d, J = 8.5 Hz, Ala<sup>4</sup>-NH), 6.87 (1H, dd, J = 8.4, 2.3 Hz, Tyr<sup>5</sup>-H<sub>εa</sub>), 7.09 (2H, d, J = 8.3 Hz,  $Tyr^3 - H_{\delta h}$ , 7.20 (1H, dd, J = 8.5, 2.3 Hz,  $Tyr^5 - H_{\delta h}$ ), 7.25 (1H, dd, J = 8.4, 2.0 Hz,  $Tyr^5 - H_{\delta h}$ ), 7.40 (1H, dd, J = 8.5, 2.0 Hz,  $Tyr^5 - H_{\delta h}$ ), 8.76 (1H, d, J = 6.2 Hz, Orn<sup>2</sup>-NH).

[2-Aspartic Acid]-RA-VII (15). a) Oxidation of 8 with AgO AgO (24.8 mg, 0.20 mmol) was added to a solution of 8 (39.9 mg, 0.50 mmol) in THF-H<sub>2</sub>O (9:1, 2ml) and the mixture was refluxed for 9 h. The mixture filtered, and the filtrate was concentrated in vacuo. The residue was dissolved in CH2Cl2. This solution was washed successively with H2O, 1 N HCl, saturated NaHCO3 and brine, and dried over MgSO4. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 15 (5.7 mg, 14%) as a colorless powder, mp 245—246 °C,  $[\alpha]_D$  -187.0° (c=0.11,CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3395, 1673, 1636 cm<sup>-1</sup>. UV  $\lambda_{max}$  (EtOH) nm (log  $\varepsilon$ ): 220 (4.48), 277 (3.58), 282 (3.52). High-resolution FAB-MS Calcd for  $C_{42}H_{55}N_6O_{11}$ : 815.3616 [M+H]<sup>+</sup>. Found: 815.3644. FAB-MS m/z (%): 815 (50,  $[M+H]^+$ ). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.08 (3H, d,  $J = 6.6 \,\text{Hz}$ , Ala<sup>4</sup>-H<sub>g</sub>), 1.28 (3H, d,  $J = 6.9 \,\text{Hz}$ , Ala<sup>1</sup>-H<sub>g</sub>), 2.62 (1H, dd, J = 11.3, 2.8 Hz,  $Tyr^5 - H_{\beta a}$ ), 2.69—2.90 (4H, m,  $Asp^2 - H_{\beta}$  and  $Tyr^6 - H_{\beta}$ ), 2.65 (3H, s, Tyr<sup>6</sup>-NMe), 3.05 (3H, s, Tyr<sup>3</sup>-NMe), 3.09 (3H, s, Tyr<sup>5</sup> -NMe), 3.16 (1H, dd, J=14.0, 10.4 Hz, Tyr<sup>3</sup>-H<sub>βa</sub>), 3.35 (1H, dd, J=14.0, 4.2 Hz,  $\text{Tyr}^3$ - $\text{H}_{\beta b}$ ), 3.59 (1H, dd, J=10.4, 4.2 Hz,  $\text{Tyr}^3$ - $\text{H}_{\alpha}$ ), 3.65 (1H, t, J=11.3 Hz,  $\text{Tyr}^5$ - $\text{H}_{\beta b}$ ), 3.78 (3H, s,  $\text{Tyr}^3$ -OMe), 3.92 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J = 1.5 Hz, Tyr<sup>6</sup>-H<sub>ha</sub>), 4.47 (1H, dq, J = 7.4, 6.9 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.57 (1H, dd, J=10.8, 4.5 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.69 (1H, dq, J=8.5,  $6.6 \,\mathrm{Hz}$ ,  $\mathrm{Ala^4-H_a}$ ), 5.10 (1H, dd, J=13.7, 7.6 Hz,  $\mathrm{Asp^2-H_a}$ ), 5.38 (1H, dd, J = 11.3, 2.8 Hz, Tyr<sup>5</sup>-H<sub> $\alpha$ </sub>), 6.53 (1H, d, J = 7.4 Hz, Ala<sup>1</sup>-NH), 6.56 (1H, d, J=8.5 Hz, Ala<sup>4</sup>-NH), 6.56 (1H, dd, J=8.5, 1.5 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.79 (1H, d,  $J = 8.5 \,\text{Hz}$ ,  $\text{Tyr}^6 - \text{H}_{\epsilon}$ ), 6.84 (2H, d,  $J = 8.4 \,\text{Hz}$ ,  $\text{Tyr}^3 - \text{H}_{\epsilon}$ ), 6.86 (1H, dd,  $J = 8.6, 2.4 \text{ Hz}, \text{Tyr}^5 - \text{H}_{\epsilon a}$ , 7.07 (2H, d,  $J = 8.4 \text{ Hz}, \text{Tyr}^3 - \text{H}_{\delta}$ ), 7.20 (1H, dd, J=8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>εb</sub>), 7.24 (1H, dd, J=8.6, 2.1 Hz, Tyr<sup>5</sup>-H<sub>δa</sub>), 7.40 (1H, dd, J = 8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>), 7.80 (1H, d, J = 7.6 Hz, Asp<sup>2</sup>-NH).

b) Oxidation of 8 with NIS NIS (42.2 mg, 0.19 mmol) and K<sub>2</sub>CO<sub>3</sub> (25.9 mg, 0.19 mmol) were added to a solution of 8 (59.8 mg, 0.075 mmol) in CH<sub>3</sub>OH (2 ml) and the mixture was stirred at room temperature for 7h. Then 5% Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> was added to the mixture, and the whole was concentrated in vacuo. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1) to give [2-aspartic acid]-RA-VII methyl ester (16) (40.6 mg, 65%) as an amorphous powder. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.12 (3H, d, J = 6.7 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.29 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub>6</sub>), 2.63 (1H, dd, J=11.3, 3.0 Hz, Tyr<sup>5</sup>-H<sub>6a</sub>), 2.67 (1H, dd, J=16.6, 6.0 Hz,  $Asp^2-H_{\beta a}$ ), 2.67 (3H, s,  $Tyr^6$ -NMe), 2.92 (1H, dd, J=16.5, 8.4 Hz, Asp<sup>2</sup>-H<sub> $\beta$ b</sub>), 2.98—3.08 (2H, m, Tyr<sup>6</sup>-H<sub> $\beta$ </sub>), 3.10 (3H, s, Tyr<sup>3</sup>-NMe), 3.12 (3H, s, Tyr<sup>5</sup>-NMe), 3.24 (1H, dd, J=14.2, 10.5 Hz, Tyr<sup>3</sup>-H<sub> $\beta a$ </sub>), 3.44 (1H, dd, J = 14.2, 4.2 Hz,  $\text{Tyr}^3 - \text{H}_{\beta b}$ ),  $3.59 \text{ (1H, dd, } J = 10.5, 4.2 \text{ Hz, Tyr}^3 - \text{H}_{\alpha}$ ), 3.67 (1H, t, J = 11.3 Hz, Tyr<sup>5</sup>-H<sub>βb</sub>), 3.73 (3H, s, Asp<sup>2</sup>-OMe), 3.79 (3H, s, Tyr<sup>3</sup>-OMe), 3.93 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J=1.7 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.38 (1H, t, J = 6.9 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.56 (1H, dd, J = 12.4, 3.8 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.71 (1H, dq, J=8.7, 6.7 Hz, Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 5.19 (1H, td, J=8.4, 6.0 Hz,  $Asp^2-H_{\alpha}$ ), 5.39 (1H, dd, J=11.3, 3.0 Hz,  $Tyr^5-H_{\alpha}$ ), 6.38 (1H, d, J=6.9 Hz, Ala<sup>1</sup>-NH), 6.58 (1H, dd, J = 8.4, 1.7 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.70 (1H, d, J = 8.7 Hz, Ala<sup>4</sup>-NH), 6.80 (2H, d, J=8.4 Hz,  $Tyr^6$ -H<sub> $\epsilon$ </sub>,  $Asp^2$ -NH), 6.83 (2H, d,  $J = 8.6 \text{ Hz}, \text{ Tyr}^3 - \text{H}_{\epsilon}$ , 6.88 (1H, dd, J = 8.6, 2.4 Hz,  $\text{Tyr}^5 - \text{H}_{\epsilon a}$ ), 7.07 (2H, d,  $J = 8.6 \,\mathrm{Hz}$ , Tyr<sup>3</sup>-H<sub>6</sub>), 7.21 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>6</sub>), 7.26 (1H, dd, J=8.6, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.41 (1H, dd, J=8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

LiOH· $\rm H_2O$  (3.3 mg, 0.079 mmol) was added to a solution of 16 (40.5 mg, 0.048 mmol) in MeOH– $\rm H_2O$  (2:1, 2 ml) and the mixture was stirred at room temperature for 4 h, then acidified with 1 n HCl and concentrated *in vacuo*. The residue was dissolved in  $\rm CH_2Cl_2$ . This solution was washed successively with  $\rm H_2O$ , 1 n HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off *in vacuo* to leave a residue, which was chromatographed on silica gel with  $\rm CH_2Cl_2$ –AcOEt–MeOH (12:2:1), followed by recrystallization from MeOH–isopropyl ether to give 15 (36.3 mg, 91%) as a colorless powder.

[2-Homoserine]-RA-VII (17) NaBH<sub>4</sub> (51.5 mg, 1.36 mmol) was added to a solution of 8 (272.0 mg, 0.34 mmol) in 1,4-dioxane (20 ml), and the mixture was stirred at room temperature for 3 h, then concentrated in vacuo. The residue was dissolved in CH2Cl2. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 17 (246.6 mg, 90%) as a colorless powder, mp 261—262 °C,  $[\alpha]_D$  – 202.7° (c=0.09, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3401, 1669, 1636 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$  (EtOH) nm (log  $\varepsilon$ ): 223 (4.55), 277 (3.64), 282 (3.60). High-resolution FAB-MS Calcd for  $C_{42}H_{53}N_6O_{10}$ : 801.3823 [M+H]<sup>+</sup>. Found: 801.3763. FAB-MS m/z (%): 801 (15, [M+H]+), 337 (100). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.15 (3H, d,  $J = 6.6 \,\text{Hz}$ ,  $A \ln^4 - H_{\theta}$ ), 1.31 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub>B</sub>), 1.93 (2H, m, Hse<sup>2</sup>-H<sub>B</sub>), 2.64 (1H, dd, J = 11.4, 2.7 Hz,  $Tyr^5-H_{\beta a}$ ), 2.66 (3H, s,  $Tyr^6-NMe$ ), 2.93 (3H, s,  $Tyr^3-NMe$ ), 3.00 (1H, dd,  $J = 18.3, 3.7 \text{ Hz}, \text{Tyr}^6 - \text{H}_{\beta a}$ ), 3.08 (1H, dd,  $J = 18.3, 12.0 \text{ Hz}, \text{Tyr}^6 - \text{H}_{\beta b}$ ), 3.14 (3H, s, Tyr<sup>5</sup>-NMe), 3.23 (1H, dd, J = 14.2, 10.9 Hz, Tyr<sup>3</sup>-H<sub>ga</sub>), 3.42 (1H, dd, J = 14.2, 4.3 Hz,  $Tyr^3 - H_{\beta b}$ ), 3.59 (1H, dd, J = 10.9, 4.3 Hz,  $Tyr^3 - H_{\alpha}$ ), 3.65 (1H, t, J = 11.4 Hz, Tyr<sup>2</sup>-H<sub>Bb</sub>), 3.70—3.79 (2H, m, Hse<sup>2</sup>-H<sub>y</sub>), 3.80 (3H, s, Tyr<sup>3</sup>-OMe), 3.90 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J = 2.0 Hz, Tyr<sup>6</sup>-H<sub>2a</sub>), 4.41 (1H, dq, J=7.2, 6.9 Hz, Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 4.52 (1H, dd, J=12.0, 3.7 Hz,  $Tyr^6-H_\alpha$ ), 4.73 (1H, dq, J=7.4, 6.6 Hz,  $Ala^4-H_\alpha$ ), 5.01 (1H, td, J=7.7, 7.0 Hz, Hse<sup>2</sup>-H<sub>a</sub>), 5.38 (1H, dd, J=11.4, 2.7 Hz, Tyr<sup>5</sup>-H<sub>a</sub>), 6.50 (1H, d,  $J = 7.2 \,\text{Hz}$ , Ala<sup>1</sup>-NH), 6.57 (1H, dd, J = 8.4, 2.0 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.73 (1H, d, J = 7.4 Hz, Ala<sup>4</sup>-NH), 6.76 (1H, d, J = 8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.81—6.85 (1H, m, Hse<sup>2</sup>-NH), 6.83 (2H, d, J=8.6 Hz, Tyr<sup>3</sup>-H<sub>e</sub>), 6.88 (1H, dd, J=8.4, 2.4 Hz,  $Tyr^5-H_{ea}$ ), 7.03 (2H, d, J=8.6 Hz,  $Tyr^3-H_{\delta}$ ), 7.21 (1H, dd, J=8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>sb</sub>), 7.27 (1H, dd, J=8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub>δa</sub>), 7.43 (1H, dd, J = 8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-Homoserine]-RA-VII Mesylate (18) MsCl (30  $\mu$ l, 0.34 mmol) was added dropwise to a solution of 17 (46.4 mg, 0.058 mmol), DMAP (21.2 mg, 0.17 mmol) and Et<sub>3</sub>N (24  $\mu$ l, 0.17 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 ml) at -78 °C. The mixture was stirred at -78 °C for 2 h, then diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1) to give **18** (36.1 mg, 71%) as an amorphous powder. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.08 (3H, d, J = 6.6 Hz, Ala<sup>4</sup>-H<sub> $\beta$ </sub>), 1.28 (3H, d, J = 7.0 Hz, Ala<sup>1</sup>-H<sub>B</sub>), 2.08—2.14 (2H, m, Hse<sup>2</sup>-H<sub>B</sub>), 2.60—2.62 (1H, m,  $Tyr^5-H_{6a}$ ), 2.64 (3H, s,  $Tyr^6-NMe$ ), 2.93 (3H, s,  $Tyr^3-NMe$ ), 2.96 (3H, s, Hse<sup>2</sup>-OMs), 2.98—3.07 (2H, m,  $Tyr^6$ -H<sub>g</sub>), 3.10 (3H, s,  $Tyr^5$ -NMe), 3.26 (1H, dd, J=14.1, 11.0 Hz, Tyr<sup>3</sup>-H<sub>βa</sub>), 3.38 (1H, dd, J=14.1, 4.8 Hz,  $\text{Tyr}^3$ - $\text{H}_{8b}$ ), 3.61 (1H, dd, J = 11.0, 4.8 Hz,  $\text{Tyr}^3$ - $\text{H}_{\alpha}$ ), 3.63 (1H, t, J = 11.2 Hz,  $Tyr^5-H_{gb}$ ), 3.76 (3H, s,  $Tyr^3$ -OMe), 3.90 (3H, s,  $Tyr^6$ -OMe), 4.14—4.20  $(2H, m, Hse^2-H_y)$ , 4.31 (1H, d, J=1.9 Hz,  $Tyr^6-H_{\delta a}$ ), 4.40 (1H, m, Ala<sup>1</sup>-H<sub>\alpha</sub>), 4.51 (1H, dd, J=11.7, 3.8 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.70 (1H, dq, J=7.3, 6.6 Hz, Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 4.95 (1H, dd, J=11.3, 3.2 Hz, Hse<sup>2</sup>-H<sub> $\alpha$ </sub>), 5.36 (1H, dd, J=11.2, 3.2 Hz,  $Tyr^5-H_a$ ), 6.40 (1H, d, J=7.0 Hz,  $Hse^2-NH$ ), 6.54—6.57 (1H, m, Ala<sup>1</sup>-NH), 6.55 (1H, dd, J = 8.3, 1.9 Hz, Tyr<sup>6</sup>-H<sub>δb</sub>), 6.70 (1H, d, J = 7.3 Hz, Ala<sup>4</sup>-NH), 6.77 (1H, d, J=8.3 Hz,  $Tyr^6-H_e$ ), 6.82 (2H, d, J=8.5 Hz,  $Tyr^3-H_e$ ), 6.84 (1H, dd, J=8.4, 2.1 Hz,  $Tyr^5-H_{ea}$ ), 7.14 (1H, dd, J=8.4, 2.1 Hz,  $Tyr^5-H_{\varepsilon b}$ ), 7.23 (1H, dd, J=8.4, 2.1 Hz,  $Tyr^5-H_{\delta a}$ ), 7.38 (1H, dd, J = 8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub>δh</sub>).

[2-Vinylglycine]-RA-VII (19) A mixture of 18 (105.0 mg, 0.12 mmol), o-NO<sub>2</sub>PhSeCN (54.5 mg, 0.24 mmol), NaBH<sub>4</sub> (9.1 mg, 0.24 mmol) and EtOH (10 ml) was stirred at room temperature for 24 h. The mixture was concentrated *in vacuo*, and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. This solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off *in vacuo* to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1) to give the selenide. 35% H<sub>2</sub>O<sub>2</sub> (2 ml) was added to the selenide dissolved in THF-EtOH (1:1, 20 ml), and the whole was stirred at room temperature for 12 h. Dimethyl sulfide was added to decompose excess H<sub>2</sub>O<sub>2</sub>, and the mixture was stirred at room temperature for 2 h. Then H<sub>2</sub>O was added and the whole was concentrated

in vacuo. The residue was dissolved in CH2Cl2, and this solution was washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH2Cl2-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 19 (16.3 mg, 17%) as a colorless powder, mp 261—262 °C,  $[\alpha]_D$  -202.3° (c=0.12, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3400, 1675, 1636 cm<sup>-1</sup>. UV  $\lambda_{\text{max}}$  (EtOH) nm (log  $\varepsilon$ ): 219 (4.48), 277 (3.61), 282 (3.56). High-resolution FAB-MS Calcd for  $C_{42}H_{51}N_6O_9$ : 783.3718 [M+H]<sup>+</sup>. Found: 783.3720. FAB-MS m/z (%): 783 (75, [M+H]<sup>+</sup>). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.12 (3H, d, J = 6.6 Hz, Ala<sup>4</sup>-H<sub>8</sub>), 1.32 (3H, d, J = 6.9 Hz, Ala<sup>1</sup>-H<sub>B</sub>), 2.60 (1H, dd, J = 11.9, 3.9 Hz, Tyr<sup>5</sup>-H<sub>Ba</sub>), 2.68 (3H, s, Tyr<sup>6</sup>-NMe), 2.87 (3H, s, Tyr<sup>3</sup>-NMe), 2.91—3.10 (2H, m, Tyr<sup>6</sup>-H<sub> $\beta$ </sub>), 3.12 (3H, s, Tyr<sup>5</sup>-NMe), 3.29 (1H, dd, J = 14.2, 11.0 Hz, Tyr<sup>3</sup>-H<sub>βa</sub>), 3.40 (1H, dd, J = 14.2, 4.4 Hz,  $\text{Tyr}^3 - \text{H}_{\beta b}$ ), 3.57 (1H, dd, J = 11.0, 4.4 Hz,  $\text{Tyr}^3 - \text{H}_{\alpha}$ ), 3.66 (1H, t, J = 11.9 Hz, Tyr<sup>5</sup>-H<sub>βb</sub>), 3.78 (3H, s, Tyr<sup>3</sup>-OMe), 3.93 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J = 1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.42 (1H, qd, J = 6.9, 6.8 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.54 (1H, dd, J=11.9, 3.9 Hz, Tyr<sup>6</sup>-H<sub> $\alpha$ </sub>), 4.74 (1H, dq, J=7.5, 6.6 Hz,  $Ala^4-H_{\alpha}$ ), 5.33—5.42 (4H, m,  $Aa^2-H_{\alpha}$ ,  $Aa^2-H_{\gamma}$  and  $Tyr^5-H_{\alpha}$ ), 5.86—5.95 (1H, m,  $Aa^2$ -H<sub> $\theta$ </sub>), 6.46 (1H, d, J=6.8 Hz,  $Ala^1$ -NH), 6.58 (1H, dd, J = 8.4, 1.8 Hz, Tyr<sup>6</sup>-H<sub> $\delta b$ </sub>), 6.71 (1H, d, J = 7.5 Hz, Ala<sup>4</sup>-NH), 6.76 (1H, d, J=7.5 Hz, Aa<sup>2</sup>-NH), 6.79 (1H, d, J=8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.80 (2H, d,  $J = 8.6 \,\mathrm{Hz}$ , Tyr<sup>3</sup>-H<sub>s</sub>), 6.87 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>s</sub>), 7.02 (2H, d,  $J = 8.6 \,\mathrm{Hz}, \,\mathrm{Tyr^3 - H_\delta}), \,7.19 \,\,(1\mathrm{H}, \,\mathrm{dd}, \,J = 8.3, \,2.4 \,\mathrm{Hz}, \,\mathrm{Tyr^5 - H_{eb}}), \,7.26 \,\,(1\mathrm{H}, \,\mathrm{dd}, \,J = 8.3, \,2.4 \,\mathrm{Hz})$ dd, J=8.4, 2.1 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.41 (1H, dd, J=8.3, 2.1 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

[2-Methionine]-RA-VII (20) NaI (21.4 mg, 0.14 mmol) was added to a solution of 18 (50.0 mg, 0.057 mmol) in methyl ethyl ketone (3 ml), and the mixture was refluxed for 10 h, then concentrated in vacuo to leave a residue. A mixture of the residue, NaSCH<sub>3</sub> (19.9 mg, 0.28 mmol) and MeOH (4 ml) was stirred at room temperature for 0.5 h, then diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed successively with H<sub>2</sub>O, 1 N HCl, saturated NaHCO<sub>3</sub> and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated off in vacuo to leave a residue, which was chromatographed on silica gel with CH<sub>2</sub>Cl<sub>2</sub>-AcOEt-MeOH (12:2:1), followed by recrystallization from MeOH-isopropyl ether to give 20 (43.0 mg, 91%) as a colorless powder, mp 204—205 °C,  $[\alpha]_D$  —188.2° (c=0.13, CHCl<sub>3</sub>). IR  $\nu$  (CHCl<sub>3</sub>): 3407, 1676, 1636 cm<sup>-1</sup>. UV  $\lambda_{max}$  (EtOH) nm ( $\log \varepsilon$ ): 221 (4.47), 277 (3.59), 282 (3.34). High-resolution FAB-MS Calcd for C<sub>43</sub>H<sub>55</sub>N<sub>6</sub>O<sub>9</sub>S: 831.3751  $[M+H]^+$ . Found: 831.3754. FAB-MS m/z (%): 831 (70,  $[M+H]^+$ ).  $^1H$ -NMR (CDCl<sub>3</sub>, major conformer,  $\delta$ ): 1.12 (3H, d, J=6.6 Hz, Ala<sup>4</sup>-H<sub>g</sub>), 1.30 (3H, d,  $J = 7.0 \,\text{Hz}$ , Ala<sup>1</sup>-H<sub>\theta</sub>), 1.95—2.05 (2H, m, Met<sup>2</sup>-H<sub>\theta</sub>), 2.08 (3H, s, Met<sup>2</sup>-SMe), 2.43—2.53 (2H, m, Met<sup>2</sup>-H<sub>y</sub>), 2.64 (1H, dd, J=11.3, 3.1 Hz, Tyr<sup>5</sup>-H<sub> $\beta a$ </sub>), 2.67 (3H, s, Tyr<sup>6</sup>-NMe), 2.92—2.98 (1H, m, Tyr<sup>6</sup>-H<sub> $\beta a$ </sub>), 2.98 (3H, s, Tyr<sup>3</sup>-NMe), 3.01—3.10 (1H, m, Tyr<sup>6</sup>-H<sub> $\beta$ b</sub>), 3.12 (3H, s, Tyr<sup>5</sup>-NMe), 3.30 (1H, dd, J = 14.3, 11.0 Hz, Tyr<sup>3</sup>-H<sub>gg</sub>), 3.44 (1H, dd, J = 14.3, 4.4 Hz, Tyr<sup>3</sup>-H<sub> $\beta$ b</sub>), 3.63 (1H, dd, J=11.0, 4.4 Hz, Tyr<sup>3</sup>-H<sub> $\alpha$ </sub>), 3.66 (1H, t, J = 11.3 Hz, Tyr<sup>5</sup>-H<sub>βb</sub>), 3.78 (3H, s, Tyr<sup>3</sup>-OMe), 3.93 (3H, s, Tyr<sup>6</sup>-OMe), 4.34 (1H, d, J = 1.9 Hz, Tyr<sup>6</sup>-H<sub> $\delta a$ </sub>), 4.38 (1H, dd, J = 7.0, 6.8 Hz, Ala<sup>1</sup>-H<sub> $\alpha$ </sub>), 4.54 (1H, dd, J=12.3, 4.2 Hz,  $Tyr^6-H_\alpha$ ), 4.73 (1H, dq, J=7.4, 6.6 Hz, Ala<sup>4</sup>-H<sub> $\alpha$ </sub>), 4.97 (1H, dd, J=15.9, 7.5 Hz, Met<sup>2</sup>-H<sub> $\alpha$ </sub>), 5.39 (1H, dd, J=11.3, 3.1 Hz,  $Tyr^5$ -H<sub>a</sub>), 6.41 (1H, d, J = 6.8 Hz,  $Ala^1$ -NH), 6.52 (1H, d,  $J=8.7 \text{ Hz}, \text{ Met}^2-\text{NH}), 6.57 (1\text{H}, \text{dd}, J=8.4, 1.9 \text{ Hz}, \text{Tyr}^6-\text{H}_{\delta b}), 6.71 (1\text{H}, \text{dd}, J=8.4, 1.9 \text{ Hz}, \text{Tyr}^6-\text{H}_{\delta b})$ d, J = 7.4 Hz, Ala<sup>4</sup>-NH), 6.80 (1H, d, J = 8.4 Hz, Tyr<sup>6</sup>-H<sub>e</sub>), 6.83 (2H, d,  $J = 8.6 \,\mathrm{Hz}$ , Tyr<sup>3</sup>-H<sub>e</sub>), 6.87 (1H, dd, J = 8.4, 2.4 Hz, Tyr<sup>5</sup>-H<sub>ea</sub>), 7.06 (2H, d,  $J = 8.6 \text{ Hz}, \text{ Tyr}^3 - \text{H}_{\delta}), 7.21 \text{ (1H, dd, } J = 8.4, 2.4 \text{ Hz, Tyr}^5 - \text{H}_{\epsilon b}), 7.26 \text{ (1H, dd)}$ dd, J=8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta a$ </sub>), 7.42 (1H, dd, J=8.4, 2.2 Hz, Tyr<sup>5</sup>-H<sub> $\delta b$ </sub>).

Cell Survival by MTT [3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl Tetrazolium Bromide] Assay MTT colorimetric assay was performed in a 96-well plate. 12) The assay is dependent on the reduction of MTT by the mitochondrial dehydrogenase of viable cells to a blue formazan product which can be measured spectrophotometrically. Human KB oral epidermoid carcinoma cells (1 × 10<sup>4</sup> cells/ml) or mouse P388 leukemia cells  $(2 \times 10^4 \text{ cells/ml})$  were inoculated in each well with  $100 \,\mu\text{l}$  of RPMI1640 medium (Gibco, Grand Island, NY) supplemented with 10% fetal calf serum (Flow Laboratories, U.K.), 100 units/ml of penicillin and 100 µg/ml of streptomycin. After overnight incubation (37 °C, 5% CO<sub>2</sub>), 100 µl of sample solution was added to each well and the plates were incubated for 3 d (KB Cell) or 2 d (P388). Then 50  $\mu$ l of MTT (200  $\mu$ g/ml.PBS) was added to each well and the plates were incubated for a further 4 h. The resulting formazan was dissolved in 150  $\mu$ l of DMSO. The plates was placed on a plate shaker for 5 min and read immediately at 540 nm. The  $IC_{50}$  ( $\mu g/ml$ ) value was defined as that concentration of sample which caused 50% reduction of growth in sample-treated cells, with respect to the controls. The IC<sub>50</sub> was calculated by using the probit test.

In Vivo Antitumor Activity P388 murine leukemia cells  $(1 \times 10^6 \text{ cells})$ 

were inoculated i.p. into female CDF<sub>1</sub> mice (6—7 weeks old, control: n=16; test: n=8) on day 0. Samples, suspended in 0.5% gum arabic-saline solution, were administered i.p. on days 1-5. The antitumor activity was estimated according to the NCI tumor panel screening method. 13)

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## References

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