A New Triphenyl-Type Neolignan and a Biphenylneolignan from the Bark of *Illicium simonsii*

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A new triphenyl-type sesquineolignan, named simonsinol (1), was isolated from the bark of *Illicium simonsii* and its structure was elucidated based on the detailed analysis of its ¹H- and ¹³C-NMR spectra. A new biphenylneolignan, named isomagnolone (5), was also isolated and its structure was determined by the analysis of the two dimensional (2D) NMR spectra.

Keywords Illicium simonsii; triphenyl-type neolignan; simonsinol; Illiciaceae; isomagnolone; biphenylneolignan

A diverse series of sesquineolignans has been isolated from the *Illicium* plants; *i.e.* macranthol (2) from the pericarps of *Illicium macranthum*, 10 dunnianol (3) and isodunnianol (4) from the bark of *I. dunnianum*. 21 They are regarded as characteristic constituents of *Illicium* plants.

As a part of our continuing research aimed at the discovery of toxic sesquiterpenes and other sesquineolignans, this paper describes the isolation and structural elucidation of a new triphenyl- and a biphenyl-neolignan from the bark of *Illicium simonsii* MAXIM. The plant is found in western Szechuan and northern and eastern Assam at altitudes between 1800 and 3000 m.

The MeOH extract (362 g) of the bark (1 kg) of *Illicium simonsii* was dissolved in water, then extracted with *n*-hexane, EtOAc and *n*-BuOH, successively. The *n*-hexane soluble fraction produced a precipitate, which was purified by recrystallization from *n*-hexane, and proved to be identical to dunnianol (3) (2.4 g) following comparison with an authentic sample. 1,2) Chromatography of the mother liquor from 3 on silica-gel with *n*-hexane–EtOAc afforded a new sesquineolignan as a colorless oil (13.2 mg), which we named simonsinol (1).

The *n*-hexane soluble part was separated and purified by means of silica-gel and medium pressure liquid chromatography (MPLC) as described in the Experimental section to give dunnianol (3) (1.47 g), macranthol (2) (288.1 mg), isodunnianol (4) (1.90 g), simonsinol (1) (82.4 mg), together with a colorless oil (5 mg), named isomagnolone (5), and also β -eudesmol (7) (6.6 mg), and

 α -eudesmol (8) (4.0 mg).

Compounds 2, 3 and 4 were identified by direct comparison with authentic samples. The structure of compounds 7 and 8 were elucidated by comparisons of their spectral data with reported data.³⁾

Simonsinol (1) was obtained as a colorless oil and had a molecular formula C₂₇H₂₆O₃, identical with macranthol and dunnianol, being determined by electron-impact mass spectrometry (EI-MS) $(m/z: 398 \text{ [M}^+\text{]})$ and the number of carbon signals in the carbon-13 nuclear magnetic resonance (13C-NMR) spectrum of 1. Compound 1 gave a positive reaction to iron(III) chloride. The features of the infrared (IR) and proton nuclear magnetic resonance (¹H-NMR) spectra of 1 were very similar to those of 2 and 3, suggesting a triphenyl-type neolignan structure for 1. The signals of the methylenes of arylpropenyl moieties in the ¹H-NMR spectrum indicated a non-symmetrical structure like macranthol (see Table I). As shown in Table II, the carbon signals of rings A and B were very similar to those of dunnianol (3). On the other hand, the carbon signals of rings A and C were similar to those of honokiol. Thus, the structure of a triphenyl-type neolignan 1 was considered for simonsinol.

Simonsinol (1) afforded three O-methyl derivatives (1'a, 1'b and 1'c) on brief treatment with CH₃I and K₂CO₃ in acetone. The molecular ion peak of 1'a in the EI-MS (m/z: 440) suggested a tri-O-methyl derivative. In support of this, 1 afforded a tri-O-acetyl derivative (1") (m/z: 524) on treatment with a mixture of 4,4'-dimethylaminopyridine (DMAP), Ac₂O and pyridine. These results

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Table I. ¹H-NMR Spectral Data of Simonsinol (1) (400 MHz in CDCl₃)

Position	
3	7.12 d (J=2.2 Hz)
5	$7.10 \mathrm{d} (J = 2.2 \mathrm{Hz})$
7	3.37 br d (J = 7.0 Hz)
8	5.95 ddd (J = 7.0, 10.3, 16.9 Hz)
9	5.09 dd (J = 1.8, 16.9 Hz)
	5.05 dd (J = 1.8, 10.3 Hz)
3′	$7.26 \mathrm{d} (J = 2.2 \mathrm{Hz})$
5′	7.12 dd (J = 2.2, 8.8 Hz)
6'	$6.96 \mathrm{d} (J = 8.8 \mathrm{Hz})$
7′	$3.39 \mathrm{d} (J = 6.2 \mathrm{Hz})$
8'	6.00 ddd (J = 6.2, 9.9, 16.9 Hz)
9′	5.11 dd (J=1.8, 16.9 Hz)
	$5.07 \mathrm{dd} (J = 1.8, 9.9 \mathrm{Hz})$
1"	7.26 dd (J=2.3, 8.8 Hz)
3"	7.08 d (J=2.3)
6"	$6.89 \mathrm{d} (J = 8.8 \mathrm{Hz})$
7"	$3.45 \mathrm{d} (J = 7.0 \mathrm{Hz})$
8"	6.03 ddd (J = 7.0, 9.9, 16.9 Hz)
9"	5.18 dd (J=1.8, 16.9 Hz)
-	$5.15 \mathrm{dd} (J = 1.8, 9.9 \mathrm{Hz})$

Table II. ¹³C-NMR Data for the Aromatic Ring Moieties of Macranthol (2), Dunnianol (3), Simonsinol (1) (100 MHz in CDCl₃), with Magnolol and Honokiol

Carbon	2	3	Magnolol ^{a)}	Honokiol ^{a)}	1
1	130.0	147.7			147.4
2	124.7	125.4			128.9
3	150.9	131.3			131.2^{b}
4	128.3	134.0			132.8
5	130.9				130.8
6	130.2				124.5
1′	150.8	151.4	151.0	150.5	151.7
2'	127.5	124.4	124.5	127.8	124.9
3′	130.3	131.6	131.4	129.6	130.5
4′	132.4	133.2	133.3	132.3	133.1
5′	129.0	130.0	129.8	128.8	128.7
6′	115.8	117.2	116.8	116.4	117.7
1''	151.2			128.9	129.7
2"	123.3			130.3	129.4
3"	131.3			130.2	$131.3^{b)}$
4''	133.4			126.5	126.4
5''	130.3			153.6	154.1
6''	116.8			115.7	116.4

a) Quoted from reference 4. b) Assignments may be interchanged.

indicated that 1 has three phenolic hydroxyl groups. As was seen in the $^1\text{H-NMR}$ spectrum of the corresponding derivative of dunnianol, 2) one of the methoxy and acetoxy signals in 1'a and 1'' appeared at an unusually high field (δ 3.20 and 1.72, respectively), indicating that the position of the hydroxy group on ring A should be at C_1 . In the case of the methoxy and acetoxy derivatives of macranthol, which has the hydroxy group at the *meta* position to the arylpropenyl group on ring A, all acetoxy proton signals appeared in the normal field and one methoxy group appeared at a higher field.

When the methoxy methyl signals at δ 3.86 and 3.77 of 1'a were irradiated, the signals at δ 6.90 (1H, d, J=8.4 Hz) and 6.91 (1H, d, J=8.1 Hz) were both enhanced. Irradiation of the signal at δ 3.19 caused the enhancements

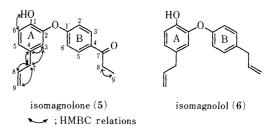


Fig. 2

Table III. ¹H-(400 MHz) and ¹³C-(100 MHz) NMR Spectral Data of Isomagnolone (5) and Isomagnolol (6) (CDCl₃)

	5	6a)	
_	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m C}$
1		146.0 s	145.9 s
2		141.8 s	143.6 s
3	$6.80 \mathrm{d} (J = 1.8 \mathrm{Hz})$	120.3 d	119.2 d
4		133.1 s	132.6 s
5	$6.94 \mathrm{dd} (J = 8.4, 1.8 \mathrm{Hz})$	126.1 s	124.8 d
6	$7.00 \mathrm{d} (J = 8.4 \mathrm{Hz})$	116.6 d	116.2 d
7	$3.29 \mathrm{d} (J = 6.6 \mathrm{Hz})$	39.3 t	
8	5.95 ddd (J = 15.8, 9.2, 6.6 Hz)	137.2 d	
9	5.03 dd (J = 15.8, 1.5 Hz)	116.0 t	
	5.04 dd (J = 9.2, 1.5 Hz)		
1'		161.0 s	155.4 s
2', 6'	$7.02 \mathrm{d} (J = 9.2 \mathrm{Hz})$	116.7 d	117.9 d
3', 5'	$7.96 \mathrm{d} (J = 9.2 \mathrm{Hz})$	130.4 d	129.9 d
4′		132.2 s	135.3 s
7′		193.5 s	
8′	$2.97 \mathrm{q} (J = 7.3 \mathrm{Hz})$	31.6 t	
9′	$1.22 \mathrm{t} (J = 7.3 \mathrm{Hz})$	8.3 q	
C_1 -OH	5.27 br s	•	

a) Quoted from reference 5.

of the signals at δ 7.39 (1H, d, J=2.2 Hz) and 7.42 (1H, dd, J=8.4, 2.2 Hz). In addition to these results, the ¹³C signals of rings A and B in 1 were compatible with those of dunnianol, indicating the position of the hydroxy group on ring C at 5".

The other O-methyl derivatives, 1'b and 1'c, were proved to be dimethyl derivatives by EI-MS (m/z: 426) and 1 H-NMR spectra, respectively. In the 1 H-NMR spectra, 1'b exhibited methoxy signals at δ 3.86 and 3.83. On the other hand, the two methoxy signals of 1'c appeared at δ 3.87 and 3.28. These observations indicated that 1'b is a 1',5"-di-O-methyl derivative and 1'c is a 1,1'- or 1,5'-di-O-methyl derivative. The nuclear Overhauser effect (NOE) experiments were performed on compound 1'b. When each of the two methoxy signal was irradiated, the aromatic proton, which coupled with the ortho proton, was enhanced in each case. This evidence supports the assigned structure of simonsinol (1).

Isomagnolone (5) was obtained as a colorless oil. The molecular formula of 5 was determined as $C_{18}H_{18}O_3$ by EI-MS (m/z: 282) and the number of signals in its 13 C-NMR spectrum. The IR spectrum indicated the absorption of a carbonyl group (1730 cm $^{-1}$) along with that of an aromatic ring. In the 1 H-NMR spectrum of 5, there was an ABC system due to aromatic protons at δ 7.00 (d, J=8.4 Hz), 6.94 (dd, J=8.4, 1.8 Hz), and 6.80 (d,

 $J=1.8\,\mathrm{Hz}$), together with signals of a p-substituted aromatic ring at δ 7.96 and 7.02 (each 2H, d, J=9.2 Hz). In addition, the proton signals of the α -properly moiety and ethyl group were observed as shown in Table III. As a biphenylneolignan, with a p-substituted benzene ring, only a type of isomagnolol (6) has been characterized so far. The ¹³C signals of isomagnolol were similar to those of 5, except for the signals of C-1, C-1', and C-4', as shown in Table III. Moreover, the presence of the isolated ethyl group and a carbonyl group in the IR (1730 cm⁻¹) and $^{13}\text{C-NMR}$ spectra (δ 193.5) suggested the presence of a 1-propanone moiety in 5. Protons on the p-substituted benzene ring (B-ring) appeared with a separation ca. 1 ppm, indicating that one of the substituents is ketonic. In contrast, the difference between the proton signals of the C-ring of isodunnianol (4), is only 0.2 ppm.²⁾ Figure 2 shows the observed heteronuclear multiple bond correlation (HMBC) of 5, which supports the location of the α -properly group. The carbon signal at δ 116.6 was ascribed to C-6 by heteronuclear multiple quantum coherence (HMQC), corresponding to the proton signal at δ 7.00 (d, J = 8.4 Hz). The observation of HMBC correlation between the C-6 signal and the 1-hydroxyl proton signal supported the location of 1-hydroxy group. Thus, the structure of 5 was assigned to isomagnolone.

Although β -eudesmol (7) and α -eudesmol (8) are well known sesquiterpenes, this is the first time that these compounds have been isolated from *Illicium* plants.

Experimental

 1 H- and 13 C-NMR spectra were recorded using JEOL GX-400 spectrometers. NOE and two dimensional correlation spectroscopy (2D COSY) experiments were also performed using this equipment. Chemical shifts are expressed in δ (ppm) values with tetramethylsilane as an internal standard. EI-MS are recorded on a JEOL JMX-DX-303 spectrometer. IR spectra were recorded on a Shimadzu IR-408 spectrometer.

Extraction and Isolation The bark (1 kg) of I. simonsii, collected in Sichuan province, China, in the summer season of 1989, was extracted three times with methanol (61) to give a MeOH extract (362g). This was suspended in water and extracted successively with n-hexane, EtOAc, and n-BuOH. The n-hexane extract was separated into a precipitate (3.45 g) and a soluble fraction (14.9 g) when concentrated. The former gave dunnianol (3) (2.4 g) by crystallization from *n*-hexane. The mother liquor from 3 afforded simonsinol (1) (13.2 mg) after separation by silica-gel chromatography with n-hexane-EtOAc (9:1). The n-hexane soluble part was chromatographed on silica-gel to give four fractions (I-IV). Repeated silica-gel column chromatography of fraction II afforded isodunnianol (4) (1.90 g), β -eudesmol (7) (6.6 mg) and α-eudesmol (8) (4.0 mg). Fraction III was subjected to silica-gel chromatography and then MPLC using a Kusano Si-5 column, to give simonsinol (1) (82.4 mg). Similar separation of fraction IV, gave dunnianol (3) (1.47 g), macranthol (2) (288.6 mg) and isomagnolone (5) (5 mg). Compounds 2, 3 and 4 were identified by direct comparison with authentic samples. Compounds 7 and 8 were identified as β -eudesmol and α -eudesmol, respectively, by comparison with reference data.³⁷

Simonsinol (1) A colorless oil, MS m/z: 398 [M⁺], $C_{27}H_{26}O_3$. IR $v_{max}^{CHCI_3}$ cm⁻¹: 3510, 3020, 1638, 1598, 1210.

Methylation of Simonsinol (1) A mixture of 1 (22.8 mg), K_2CO_3 (1 g), CH_3I (1.3 ml) in dry acetone (20 ml), was refluxed for 8 h. After the solution had cooled, K_2CO_3 was removed by filtration. The filtrate was concentrated to dryness under reduced pressure, and the residual oil was chromatographed on silica-gel [n-hexane-EtOAc (1:9)] to give three O-methylated compounds (1'a (12.1 mg), 1'b (5.2 mg) and 1'c (7.2 mg)). 1'a: colorless oil, EI-MS m/z: 440 (M⁺), 426 (M⁺-14). ¹H-NMR

(CDCl₃) δ : 3.20 (3H, s, OMe), 3.37 (2H, d, J = 6.6 Hz), 3.40 (2H, d, $J = 6.6 \,\mathrm{Hz}$), 3.42 (2H, d, $J = 6.6 \,\mathrm{Hz}$), 3.78 (6H, s, OMe), 5.02 (1H, dd, J=9.9, 1.5 Hz), 5.05 (1H, dd, J=17.2, 1.8 Hz), 5.06 (1H, dd, J=9.9, 1.8 Hz), 5.07 (1H, dd, J = 17.6, 1.5 Hz), 5.09 (1H, dd, J = 9.9, 1.8 Hz), 5.12 (1H, dd, J=17.2, 1.8 Hz), 5.97—6.03 (3H, m), 6.90 (1H, d, J=8.4 Hz), 6.90 (1H, d, J=8.1 Hz), 6.91 (1H, d, J=8.1 Hz), 7.03 (1H, d, J=2.2 Hz), 7.12 (2H, d, J=2.2 Hz), 7.13 (1H, dd, J=8.1, 2.2 Hz), 7.39 (1H, d, J = 2.2 Hz), 7.42 (1H, dd, J = 8.4, 2.2 Hz). ¹³C-NMR (CDCl₃) δ: 34.2 (t), 39.2 (t), 39.5 (t), 55.3 (q), 55.7 (q), 60.2 (q), 110.0 (d), 110.9 (d), 115.2 (t), 115.3 (t), 115.7 (t), 127.9 (d), 128.0 (s), 128.1 (s), 128.3 (d), 130.2 (d), 130.4 (d), 130.7 (d), 131.2 (s), 131.5 (d), 131.7 (s), 132.4 (s), 134.3 (s), 134.7 (s), 137.0 (d), 137.4 (d), 137.8 (d), 153.7 (s), 155.2 (s), 156.4 (s); 1'b: colorless oil, EI-MS m/z: 426 (M⁺). ¹H-NMR (CDCl₃) δ : 3.395 (2H, d, $J=7.0\,\text{Hz}$), 3.398 (2H, d, $J=7.0\,\text{Hz}$), 3.43 (2H, d, J = 7.0 Hz), 3.83 (3H, s, OMe), 3.86 (3H, s, OMe), 5.03 (1H, dd, J = 10.3, 1.5 Hz), 5.05 (1H, dd, J = 10.2, 1.8 Hz), 5.06 (1H, dd, J = 10.3, 1.5 Hz), 5.08 (1H, dd, J = 15.0, 1.5 Hz), 5.10 (1H, dd, J = 15.0, 1.5 Hz), 5.12 (1H, dd, J = 15.0, 1.8 Hz), 5.90—6.10 (3H, m), 6.92 (1H, d, J = 8.4 Hz), 7.00 (1H, d, J=7.7 Hz), 7.12 (1H, d, J=2.2 Hz), 7.18 (2H, d, J=2.2 Hz), 7.19(1H, dd, J=7.7, 2.2 Hz), 7.36 (1H, d, J=2.2 Hz), 7.42 (1H, dd, J=8.4,2.2 Hz). 13 C-NMR (CDCl₃) δ : 34.3 (t), 39.2 (t), 39.4 (t), 55.4 (q), 56.0 (q), 110.1 (d), 111.4 (d), 115.3 (t), 115.5 (t), 115.7 (t), 126.6 (s), 127.2 (s), 128.3 (d), 129.0 (d), 129.7 (s), 130.1 (s), 130.2 (2d), 130.7 (s), 131.0 (d), 131.9 (s), 132.5 (d), 133.3 (s), 136.9 (d), 137.4 (s), 137.7 (d), 148.7 (s), 154.2 (s), 156.4 (s); 1'c: colorless oil, EI-MS m/z: 426 (M⁺). ¹H-NMR (CDCl₃) δ : 3.28 (3H, s, OMe), 3.39 (2H, d, J = 6.6 Hz), 3.44 (4H, d, $J = 6.6 \,\mathrm{Hz}$), 3.88 (3H, s, OMe), 5.04 (1H, dd, J = 9.9, 1.8 Hz), 5.05 (1H, dd, J = 16.9, 1.5 Hz), 5.07 (1H, dd, J = 9.9, 1.5 Hz), 5.09 (1H, dd, J = 15.0, 1.8 Hz), 5.10 (1H, dd, J=9.5, 1.5 Hz), 5.13 (1H, dd, J=16.9, 1.5 Hz), 5.98-6.04 (3H, m), 6.92 (1H, d, J=8.1 Hz), 7.00 (1H, d, J=8.1 Hz), 7.12 (1H, d, J=2.2 Hz), 7.13 (1H, dd, J=8.1, 2.2 Hz), 7.15 (1H, d, J=2.2 Hz), 7.18 (1H, d, J=2.2 Hz), 7.37 (1H, d, J=2.2 Hz), 7.41 (1H, dd, J = 8.1, 2.2 Hz). ¹³C-NMR (CDCl₃) δ : 34.3 (t), 39.4 (t), 39.7 (t), 55.5 (q), 61.2 (q), 110.2 (d), 115.5 (2t), 116.2 (t), 118.8 (d), 126.5 (s), 128.2 (d), 128.5 (s), 129.5 (d), 130.1 (s), 130.8 (d), 130.9 (d), 131.1 (d), 131.2 (d), 132.2 (s), 132.5 (s), 135.2 (s), 136.9 (d), 137.1 (d), 137.2 (s), 137.9 (d), 151.7 (s), 152.1 (s), 156.8 (s).

Permethylation of Simonsinol (1) A mixture of 1 (23.2 mg), K_2CO_3 (3.6 g), CH_3I (3 ml), in dry acetone (20 ml) was refluxed for 24 h. After a work-up as described above, an oily product (20.7 mg) was obtained, identical with 1'a on thin-layer chromatography (TLC) and in terms of its $^1\text{H-}$ and $^{13}\text{C-NMR}$ spectra.

Acetylation of Simonsinol (1) A mixture of **1** (15.2 mg) and DMAP (2 mg) in dry pyridine (1 ml) and Ac₂O (0.5 ml) was left 2h at room temperature, then concentrated to dryness under reduced pressure. The residue was chromatographed on silica-gel [n-hexane–EtOAc (1:9)] to give an oily acetylated compound (1") (16.8 mg). EI-MS m/z: 524 (M⁺), 482, 440, 396. ¹H-NMR (CDCl₃) δ : 1.72 (3H, s, OAc), 2.11 (6H, s, OAc), 3.32 (4H, d, J=7.0 Hz), 3.40 (2H, d, J=6.6 Hz), 5.06—5.14 (6H, m), 5.91—6.03 (3H, m), 7.08 (1H, d, J=7.0 Hz), 7.09 (1H, d, J=6.6 Hz), 7.09 (1H, dd, J=6.6, 1.8 Hz), 7.15 (1H, d, J=1.8 Hz), 7.19 (1H, d, J=2.2 Hz), 7.21 (1H, d, J=2.2 Hz), 7.29 (1H, dd, J=7.0, 2.2 Hz), 7.30 (1H, d, J=2.2 Hz).

Isomagnolone (5) Colorless oil, MS m/z: 282 [M⁺], $C_{18}H_{18}O_3$. IR $v_{\max}^{CHC_1}$ cm⁻¹: 3510, 1730, 1495, 1462.

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