Protecting Group for Carboxyl Function: Mild and Facile Cleavage of 2-Cyanoethyl Ester under Non-hydrolytic Conditions

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The use of the 2-cyanoethyl group for carboxyl-protection is described. This group was readily introduced by esterification using ethylene cyanohydrin and the deprotection was carried out under mild conditions using tetrabutylammonium fluoride in dimethylformamide—tetrahydrofuran.

Keywords 2-cyanoethyl ester; carboxyl protecting group; tetrabutylammonium fluoride; deprotection; β -elimination; peptide synthesis

Easy protection of carboxylic acid and removal of the protecting group under mild non-hydrolytic conditions are important in organic synthesis. 1) Although carboxyl groups can be readily protected as the corresponding esters, regeneration of the parent acids is usually carried out in acidic or basic aqueous media. These conditions, however, sometimes cannot be used because of the instability or sensitivity of the given substrate. To overcome this disadvantage, a wide variety of methods have been developed for carboxyl protection based on β -elimination methods.²⁾ Recently, the 2-cyanoethyl group has been used in peptide synthesis.³⁾ However, the deblocking was achieved with K₂CO₃ in MeOH-H₂O, and is not suitable for the deprotection of 2-cyanoethyl esters bearing an alkali-sensitive group, such as acetate or 2-trimethylsilylethyl (TMSE) ester in the molecule.⁴⁾ We report here an alternative useful deblocking method of 2-cyanoethylester using tetrabutylammonium fluoride (TBAF) under non-hydrolytic conditions, allowing selective deprotection of the 2-cyanoethyl group.

Starting carboxylic acids (3c, d) were prepared from methyl 3-(4-hydroxyphenyl)propionate (1) by a standard method (Chart 1). The 2-cyanoethyl group was readily introduced³⁾ by using ethylene cyanohydrin in the presence of N,N'-dicyclohexylcarbodiimide (DCC) and a catalytic amount of 4-(dimethylamino)pyridine (DMAP) in dichloromethane, in high yields (Table I).

Deprotection of the 2-cyanoethyl group was achieved under mild non-hydrolytic conditions. Thus, treatment of 2-cyanoethyl 3-phenylpropionate (4a) in tetrahydrofuran (THF) with TBAF at room temperature for 1 h provided the corresponding ammonium carboxylate, from which 3-phenylpropionic acid (3a) was obtained in 95% yield. When the reaction was performed in N,N-dimethylform-

amide (DMF)-THF (5:1), it proceeded five times faster. Selective cleavage of the 2-cyanoethyl ester was accomplished in the presence of acetate, THP ether, methoxymethyl (MOM) ether, benzyl ester, TMSE ester, and benzyl carbamate. Thus, the 2-cyanoethyl esters (4b—g) were treated with TBAF in DMF-THF to give the parent carboxylic acids (3b—g) selectively in high yields. It is noteworthy that the cleavage of the 2-cyanoethyl group was faster than that of the TMSE group. These results are summarized in Table I.

Peptide synthesis⁵⁾ was carried out with Boc-Phe-OH (5). The 2-cyanoethyl ester (6) was prepared from 5 by the method mentioned above and treated with TBAF to afford crude 5. The degree of racemization calculated

Table I. Preparation of 2-Cyanoethyl Esters (4a—g) and Cleavage of 4a—g into 3a—g

RCO₂H
$$\xrightarrow{\text{HO}(\text{CH}_2)_2\text{CN}}$$
 RCO₂(CH₂)₂CN $\xrightarrow{\text{1) TBAF}}$ 3a—g

3a—g

4a—g

| R | Yield (%) of 4 | Deprotection conditions (h) | Yield (%) of 3 |
|--|----------------|-----------------------------|--------------------|
| Ph(CH ₂) ₂ | 4a (94) | THF, 1 DMF–THF, 0.2 | 3a (95) 3a (98) |
| $4-AcOC_6H_4(CH_2)_2$ | 4b (92) | DMF-THF, 0.3 | 3b (84) |
| $4\text{-THPOC}_6\text{H}_4(\text{CH}_2)_2$ | 4c (95) | DMF-THF, 0.3 | 3c (88) |
| $4-MOMOC_6H_4(CH_2)_2$ | 4d (92) | DMF-THF, 0.3 | 3d (96) |
| PhCH ₂ OCO(CH ₂) ₂ | 4e (86) | DMF-THF, 0.2 | 3e (99) |
| $TMSEOCO(CH_2)_2$ | 4f (92) | DMF-THF, 0.2 | 3f $(64)^{a}$ |
| PhCH ₂ OCON | 4g (97) | DMF-THF, 0.3 | 3g (100) |

a) A small amount (4%) of 2-cyanoethyl hydrogen succinate was present.

 $\begin{array}{c} \text{10-camphorsulfonic acid,} \\ \text{4-HOC}_6\text{H}_4(\text{CH}_2)_2\text{CO}_2\text{Me} & \overbrace{\begin{matrix} & & \\ &$

Chart 1

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from the optical rotations through this operation $(5 \rightarrow \text{crude } 6 \rightarrow \text{crude } 5)$ was 0.4%. Crystallization of crude 5 gave enantiomeric pure 5. Next, the Boc group of 6 was deprotected by trifluoroacetic acid (TFA) followed by DCC coupling in the presence of 1-hydroxybenzotriazole (HOBt) with Z-Gly-OH to give the dipeptide (7). The 2-cyanoethyl ester was cleaved by TBAF to afford crude 8. The degree of racemization through this operation $(6 \rightarrow \text{crude } 7 \rightarrow \text{crude } 8)$ was less than 1.0%. Crystallization of crude 8 gave enantiomeric pure 8.

Experimental

All melting points are uncorrected. Optical rotations were measured with a Perkin-Elmer 241 polarimeter using a 10 cm cell. Infrared (IR) absorption spectra were recorded on a JASCO HPIR-102 spectrophotometer. Proton nuclear magnetic resonance (¹H-HMR) spectra were measured on Varian VXR-200 (200 MHz), Hitachi R-250HT (250 MHz), and JEOL JNM-EX270 (270 MHz) spectrometers with tetramethylsilane as an internal standard. Mass spectra (MS) were obtained on a JEOL JMS-D300 [for electron impact (EI)- and exact MS] or a JEOL HX-100 [for fast atom bombardment mass spectra (FAB-MS)] mass spectrometer. E. Merck Silica gel 60 (70—230 mesh ASTM) was used for column chromatography. The known carboxylic acids (1,6) 3b,7) 3e,8) 3f,9) and 3g¹⁰) were prepared by the reported methods

Methyl 3-(4-Tetrahydropyran-2-yloxyphenyl)propionate (2c) 10-Camphorsulfonic acid (65 mg, 0.28 mmol) was added to a solution of 1^6) (1.0 g, 5.55 mmol) and 3,4-dihydro-2*H*-pyran (2.34 mg, 27.8 mmol) in dry CH₂Cl₂ (10 ml) at 0°C, and the mixture was stirred at the same temperature for 30 min. After quenching of the reaction with saturated aqueous NaHCO₃ and CH₂Cl₂, the organic layer was separated. The extract was dried over MgSO₄ and concentrated under reduced pressure. The oily residue was chromatographed on silica gel (hexane–AcOEt, 19:1) to give 2c (1.40 g, 96%) as an oil, bp 165–175 °C (0.5 mmHg) (bath temperature). IR (CHCl₃) v: 1730 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.4—2.1 (m, 6H), 2.59 (t, 2H, J=7.7 Hz, CH₂CO₂), 2.89 (t, 2H, J=7.7 Hz, CH₂Ph), 3.5—3.7 (m, 1H), 3.66 (s, 3H, CO₂CH₃), 3.8—4.0 (m, 1H), 5.37 (t, 1H, J=3.1 Hz, OCHO), 6.97 (d, 2H, J=8.6 Hz, phenyl protons), 7.10 (d, 2H, J=8.6 Hz, phenyl protons). *Anal*. Calcd for C₁₅H₂₀O₄: C, 68.16; H, 7.63. Found: C, 68.15; H, 7.59.

Methyl 3-[4-(Methoxymethoxy)phenyl]propionate (2d) A solution of 16 (3.0 g, 16.6 mmol) in dry THF (10 ml) was added to a suspension of NaH (60% dispersion in mineral oil, 731 mg, 18.3 mmol) in dry THF (10 ml) at 0 °C, and the mixture was stirred at room temperature for 30 min. To this mixture, a solution of chloromethyl methyl ether (1.26 ml, 16.6 mmol) in dry THF (15 ml) was added at 0 °C, and the whole was stirred at the same temperature for 1 h. After quenching of the reaction with saturated aqueous NH₄Cl and AcOEt, the organic layer was separated. The extract was dried over MgSO₄ and concentrated under reduced pressure. The oily residue was chromatographed on silica gel (hexane-AcOEt, 6:1) to give 2d (3.01 g, 81%) as an oil, bp 120—125°C (0.1 mmHg) (bath temperature). IR (CHCl₃) v: 1725 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.59 (t, 2H, J=7.5 Hz, CH₂CO₂), 2.90 (t, 2H, J=7.5 Hz, CH₂Ph), 3.46 (s, 3H, OCH₃), 3.66 (s, 3H, CO₂CH₃), 5.14 (s, 2H, OCH₂O), 6.96 (d, 2H, J=8.6 Hz, phenyl protons), 7.11 (d, 2H, J=8.6 Hz, phenyl protons) 8.6 Hz, phenyl protons). Anal. Calcd for C₁₂H₁₆O₄: C, 64.27; H, 7.19. Found: C, 64.05; H, 7.38.

3-(4-Tetrahydropyran-2-yloxyphenyl)propionic Acid (3c) A 1 N NaOH solution (5.3 ml, 5.3 mmol) was added to a solution of **2c** (693 mg,

2.62 mmol) in MeOH (10 ml) at 0°C, and the mixture was stirred at room temperature for 3 h. The MeOH was removed under reduced pressure and the aqueous solution was neutralized with solid citric acid, and then extracted with AcOEt. The extract was washed with water, dried over MgSO₄ and concentrated under reduced pressure. The resulting residue was crystallized from CH₂Cl₂-hexane to give 3c (640 mg, 98%) as colorless prisms, mp 85–86°C. IR (CHCl₃) v: 1715 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.5—2.1 (m, 6H), 2.64 (t, 2H, J=7.7 Hz, CH₂CO₂), 2.90 (t, 2H, J=7.7 Hz, CH₂Ph), 3.5–3.7 (m, 1H), 3.8—4.0 (m, 1H), 5.38 (t), H, J=3.1 Hz, OCHO), 6.97 (d, 2H, J=8.6 Hz, phenyl protons), 7.11 (d, 2H, J=8.6 Hz, phenyl protons). Exact MS Calcd for C₁₄H₁₈O₄: 250.1205. Found: 250.1210.

3-[4-(Methoxymethoxy)phenyl]propionic Acid (3d) A $0.5\,\mathrm{N}$ NaOH solution (12 ml, 6.0 mmol) was added to a solution of 2d (602 mg, 2.68 mmol) in MeOH (14 ml) at 0 °C, and the mixture was stirred at room temperature for 4 h. The MeOH was removed under reduced pressure and the aqueous solution was neutralized with solid NH₄Cl, and then extracted with AcOEt. The extract was dried over MgSO₄ and concentrated under reduced pressure. The resulting residues crystallized from CHCl₃-hexane to give 3d (390 mg, 70%) as colorless prisms, mp $58\,^{\circ}$ C. IR (CHCl₃) v: $1740\,\mathrm{cm}^{-1}$. 1 H-NMR (CDCl₃) δ : 2.64 (t, 2 H, 2 H- 2 6Hz, CH₂CO₂), 2 90 (t, 2 H, 2 H- 2 6Hz, CH₂Ph), 3 47 (s, 3 H, OCH₃), 5 .15 (s, 2 H, OCH₂O), 6 96 (d, 2 H, 2 H- 3 6Hz, phenyl protons), 7 12 (d, 2 H, 2 H- 3 6Hz, phenyl protons). Anal. Calcd for 2 C₁1H₁₄O₄: C, 6 2.85; H, 6 71. Found: C, 6 2.64; H, 6 76.

Typical Example of the Preparations of 2-Cyanoethyl Esters: 2-Cyanoethyl 3-Phenylpropionate (4a) A solution of DCC (604 mg, 2.93 mmol) in dry CH₂Cl₂ (4 ml) was added to a solution of 3a (400 mg, 2.67 mmol), ethylene cyanohydrin (189 mg, 2.66 mmol), and DMAP (16.3 mg, 0.13 mmol) in dry CH₂Cl₂ (4 ml) at 0 °C, and the whole was stirred at room temperature for 5 h, and then filtered to remove N,N'dicyclohexylurea (DCU). The filtrate was washed with 10% aqueous HCl, saturated aqueous NaHCO₃. The organic layer was dried over MgSO₄ and the solution was concentrated under reduced pressure. The oily residue was chromatographed on silica gel (hexane-Et₂O, 3:1) to give 4a (509 mg, 94%) as an oil, bp 139—141 °C (0.3 mmHg). IR (CHCl₃) v: 2250, 1740 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.64 (t, 2H, J= 6.3 Hz, CH₂CN), 2.69 (t, 2H, J = 8.0 Hz, CH₂CO₂), 2.97 (t, 2H, J =8.0 Hz, CH₂Ph), 4.26 (t, 2H, J=6.3 Hz, CO₂CH₂), 7.1—7.4 (m, 5H, phenyl protons). Anal. Calcd for C₁₂H₁₃NO₂: C, 70.92; H, 6.45; N, 6.89. Found: C, 70.82; H, 6.44; N, 6.87.

2-Cyanoethyl 3-(4-Acetoxyphenyl)propionate (4b) This compound was prepared from $3b^{7}$ (1.0 g, 4.80 mmol). Purification by chromatography on silica gel (hexane–Et₂O, 1:2) gave **4b** (1.16 g, 92%) as an oil, bp 165—175 °C (0.4 mmHg) (bath temperature). IR (CHCl₃) v: 2260, 1750 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.28 (s, 3H, CH₃CO), 2.64 (t, 2H, J=6.3 Hz, CH₂CN), 2.68 (t, 2H, J=7.6 Hz, CH₂CO₂), 2.96 (t, 2H, J=7.6 Hz, CH₂Ph), 4.25 (d, 2H, J=6.3 Hz, CO₂CH₂), 7.01 (d, 2H, J=8.4 Hz, phenyl protons), 7.21 (d, 2H, J=8.4 Hz, phenyl protons). Exact MS Calcd for C₁₄H₁₅NO₄: 261.1001. Found: 261.1007.

2-Cyanoethyl 3-(4-Tetrahydropyran-2-yloxyphenyl)propionate (4c) This compound was prepared from **3c** (400 mg, 1.60 mmol). Purification by chromatography on silica gel (hexane–Et₂O, 1:1) gave **4d** (460 mg, 95%) as an oil. IR (CHCl₃) v: 2260, 1740 cm $^{-1}$. 1 H-NMR (CDCl₃) δ : 1.5—2.1 (m, 6H), 2.6—2.7 (m, 4H, CH₂CO₂ and CH₂CN), 2.91 (t, 2H, J=7.6 Hz, CH₂Ph), 3.5—3.7 (m, 1H), 3.8—4.0 (m, 1H), 4.26 (t, 2H, J=6.3 Hz, CO₂CH₂), 5.38 (t, 1H, J=3.3 Hz, OCHO), 6.98 (d, 2H, J=8.6 Hz, phenyl protons). Anal. Calcd for C₁₇H₂₁NO₄: C, 67.31; H, 6.98; N, 4.62. Found: C, 67.03; H, 7.02; N, 4.65.

2-Cyanoethyl 3-[4-(Methoxymethoxy)phenyl]propionate (4d) This compound was prepared from **3d** (1.4 g, 6.7 mmol). Purification by chromatography on silica gel (hexane–Et₂O, 2:1) gave **4d** (1.61 g, 92%) as an oil, bp 160-165 °C (0.3 mmHg) (bath temperature). IR (CHCl₃) v: 2250, 1735 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.6—2.7 (m, 4H, CH₂CO₂, CH₂CN), 2.91 (t, 2H, J=7.7 Hz, CH₂Ph), 3.46 (s, 3H, OCH₃), 4.25 (t, 2H, J=6.3 Hz, CO₂CH₂), 5.14 (s, 2H, OCH₂O), 6.96 (d, 2H, J=8.8 Hz, phenyl protons), 7.12 (d, 2H, J=8.8 Hz, phenyl protons). *Anal.* Calcd for C₁₄H₁₇NO₄: C, 63.86; H, 6.51; N, 5.32. Found: C, 63.66; H, 6.71; N. 5.30.

Benzyl 2-Cyanoethyl Succinate (4e) This compound was prepared from $3e^{8)}$ (3.0 g, 14.4 mmol). Purification by chromatography on silica gel (hexane–AcOEt, 6:1) gave 4e (3.24 g, 86%) as an oil, bp 160-170 °C (0.4 mmHg) (bath temperature). IR (CHCl₃) v: 2250, 1725 cm⁻¹.

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¹H-NMR (CDCl₃) δ : 2.64 (t, 2H, J=6.3 Hz, CH₂CN), 2.70 (s, 4H, CH₂CH₂), 4.27 (t, 2H, J=6.3 Hz, OCH₂), 5.14 (s, 2H, CH₂Ph), 7.2—7.5 (m, 5H, phenyl protons). Exact MS Calcd for C₁₄H₁₅NO₄: 261.0998. Found: 261.0995.

2-Cyanoethyl 2-(Trimethylsilyl)ethyl Succinate (4f) This compound was prepared from **3f** ⁹⁾ (127 mg, 0.58 mmol). Purification by chromatography on silica gel (hexane–Et₂O, 2:1) gave **4f** (145 mg, 92%) as an oil, 125—135 °C (0.4 mmHg) (bath temperature). IR (CHCl₃) v: 2260, 1730 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.04 (s, 9H, CH₃×3), 0.9—1.1 (m, 2H, CH₂Si), 2.6—2.7 (m, 4H, CH₂CH₂), 2.72 (t, 2H, J=6.3 Hz, CH₂CN), 4.1—4.3 (m, 2H, OCH₂), 4.31 (t, 2H, J=6.3 Hz, OCH₂). *Anal*. Calcd for C₁₂H₂₁NO₄Si: C, 53.10; H, 7.80; N, 5.16. Found: C, 52.83; H, 7.71; N, 5.14.

2-Cyanoethyl (1-Benzyloxycarbonyl-4-piperidine)carboxylate (4g) This compound was prepared from $3g^{10}$ (3.0 g, 11.4 mmol). Purification by recrystallization gave 4g (3.50 g, 97%) as colorless prisms, mp 78 °C (Et₂O-hexane). IR (CHCl₃) v: 1735, 1685 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.5—1.8 (m, 2H), 1.8—2.0 (m, 2H); 2.45—2.65 (m, 1H, CHCO), 2.71 (t, 2H, J=6.2 Hz, CH₂CN), 2.8—3.05 (m, 2H), 4.0—4.2 (m, 2H), 4.29 (t, 2H, J=6.2 Hz, CO₂CH₂), 5.13 (s, 2H, CH₂Ph), 7.3—7.4 (m, 5H, phenyl protons). *Anal.* Calcd for C₁₇H₂₀N₂O₄: C, 64.54; H, 6.37; N, 8.86. Found: C, 64.41; H, 6.44; N, 8.84.

Typical Example of the Cleavage of 2-Cyanoethyl Esters with a Base: 3-Phenylpropionic Acid (3a) A solution of $1.0\,\mathrm{m}$ TBAF¹¹ ($1.0\,\mathrm{m}$ l, $1.0\,\mathrm{mmol}$) in THF was added to a solution of 4a ($200\,\mathrm{mg}$, $0.99\,\mathrm{mmol}$) in DMF (5 ml) at room temperature, and the mixture was stirred for $10\,\mathrm{min}$. The mixture was added to saturated aqueous NaHCO3 and the solution was washed with AcOEt. The aqueous layer was acidified with 10% aqueous HCl and extracted with AcOEt. The extract was washed with water, dried over MgSO4, and concentrated under reduced pressure. The residue was solidified to give 3a ($145\,\mathrm{mg}$, 98%) as colorless prisms, mp $47-48\,^{\circ}\mathrm{C}$ (hexane), lit, $^{12}\mathrm{mp}$ $48.5\,^{\circ}\mathrm{C}$. IR (CHCl3) v: $1713\,\mathrm{cm}^{-1}$. $^{1}\mathrm{H}$ -NMR (CDCl3) δ : 2.68 (t, $2\mathrm{H}$, $J=7.6\,\mathrm{Hz}$, $2\mathrm{Hz}$), 2.96 (t, $2\mathrm{H}$, $J=7.6\,\mathrm{Hz}$, $2\mathrm{Hz}$), 2.96 (t, $2\mathrm{H}$, $J=7.6\,\mathrm{Hz}$, $2\mathrm{Hz}$), 2.96 (t, $2\mathrm{Hz}$), 2.96 (t), $2\mathrm{Hz}$), $2\mathrm{Hz}$

3-(4-Acetoxyphenyl)propionic Acid (3b) By the same procedure as described for the cleavage of **4a**, this compound was recovered from **4b** (200 mg, 0.77 mmol) as colorless plates (134 mg, 84%), mp 95—96 °C (benzene–Et₂O), lit., ⁷⁾ mp 93.5—95.5 °C. IR (CHCl₃) v: 1755, 1715 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.29 (s, 3H, CH₃CO), 2.68 (t, 2H, J=7.7 Hz, CH₂CO₂), 2.95 (t, 2H, J=7.7 Hz, CH₂Ph), 7.01 (d, 2H, J=8.6 Hz, phenyl protons), 7.22 (d, 2H, J=8.6 Hz, phenyl protons).

3-(4-Tetrahydropyran-2-yloxyphenyl)propionic Acid (3c) By the same procedure as described for the cleavage of 4a, this compound was recovered from 4c (200 mg, 0.66 mmol) as colorless prisms (145 mg, 88%), mp 84-85 °C (CH₂Cl₂-hexane).

3-[4-(Methoxymethoxy)phenyl]propionic Acid (3d) By the same procedure as described for the cleavage of **4a**, this compound was recovered from **4d** (200 mg, 0.76 mmol) as colorless prisms (153 mg, 96%), mp 57—58 °C (CHCl₃-hexane).

Benzyl Hydrogen Succinate (3e) By the same procedure as described for the cleavage of **4a**, this compound was recovered from **4e** (200 mg, 0.77 mmol) as colorless plates (158 mg, 99%), mp $56-57\,^{\circ}\text{C}$ (CHCl₃-hexane), lit., ⁸⁾ mp $58-59\,^{\circ}\text{C}$. IR (CHCl₃) ν : $1710\,\text{cm}^{-1}$. $^{1}\text{H-NMR}$ (CDCl₃) δ : 2.67 (s, 4H, CH₂CH₂), 5.13 (s, 2H, CH₂Ph), 7.2—7.4 (m, 5H, phenyl protons), 10.5-11.0 (br, 1H, CO₂H).

The Cleavage of 4f with TBAF By the same procedure as described for the cleavage of 4a, a mixture of 3f and 5b (14:1, 103 mg, 64%) was obtained. This ratio was determined by comparison of the methylene proton signals in the ¹H-NMR spectra. The methylene protons (CH₂O) of 3f appeared as a multiplet (4.14—4.26 ppm). The methylene protons (CH₂O) of 2-cyanoethyl hydrogen succinate ¹³⁾ appeared as a triplet (4.32 ppm, J=6.3 Hz). The spectral data of 3f ⁹⁾ were abstracted from those of the mixture. IR (CHCl₃) v: 1710 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.04 (s, 9H, CH₃×3), 0.95—1.15 (m, 2H, CH₂Si), 2.5—2.8 (m, 4H, CH₂CH₂), 4.14—4.26 (m, 2H, CH₂O).

(1-Benzyloxycarbonyl-4-piperidine)carboxylic Acid (3g) By the same procedure as described for the cleavage of 4a, this compound was recovered from 4g (200 mg, 0.63 mmol) as an oil (166 mg, 100%). 10 IR (CHCl₃) ν : 1685 cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.5—1.8 (m, 2H), 1.8—2.05 (m, 2H), 2.4—2.6 (m, 1H, CHCO), 2.8—3.1 (m, 2H), 4.0—4.2 (m, 2H), 5.13 (s, 2H, CH₂Ph), 7.2—7.4 (m, 5H, phenyl protons).

Boc-Phe-OH (5) A solution of $1.0\,\mathrm{m}$ TBAF ($1.3\,\mathrm{ml}$, $1.3\,\mathrm{mmol}$) in THF was added to a solution of crude 6 ($200\,\mathrm{mg}$, $0.63\,\mathrm{mmol}$) in DMF ($3\,\mathrm{ml}$) at $0\,\mathrm{^{\circ}C}$, and the stirring was continued for $40\,\mathrm{min}$ at room tem-

perature. The mixture was added to saturated aqueous NaHCO₃ and the solution was washed with AcOEt. The aqueous layer was acidified with 5% aqueous KHSO₄ and extracted with AcOEt. The extract was washed with water, drid over MgSO₄, and concentrated to give crude 5, $[\alpha]_D^{20}$ +24.5° (c=2, EtOH). Recrystallization from AcOEt–hexane gave pure 5 (145 mg, 87%) as colorless prisms, mp 84—86°C, $[\alpha]_D^{20}$ +24.7° (c=2, EtOH), lit., ¹⁴ mp 85—87°C, $[\alpha]_D^{20}$ +24.7±0.5° (c=1.5, EtOH). IR (CHCl₃) v: 1700 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.29 (s, 3H, CH₃), 1.42 (s, 6H, CH₃×2), 2.8—3.3 (m, 2H, CH₂Ph), 4.5—4.7 (br, 1H, CH), 4.8—5.0 (br, 1H, NH), 7.1—7.4 (m, 5H, phenyl protons).

Z-Gly-Phe-OCet (7) A solution of TFA (5.4 ml) in dry CH₂Cl₂ (25 ml) was added to a solution of 6 (1.5 g, 4.7 mmol) in dry CH₂Cl₂ (5 ml) at 0 °C, and the stirring was continued for 3 h at room temperature. The mixture was evaporated under reduced pressure to remove TFA and CH₂Cl₂. To a solution of the residue in DMF (15 ml), a solution of triethylamine (1.6 ml, 11.5 mmol) in DMF (5 ml) was added at 0 °C. Z-Gly-OH (910 mg, 4.7 mmol), HOBt (680 mg, 5.2 mmol), and DCC (1.04 g, 5.2 mmol) were added to the above solution at 0 °C. The mixture was stirred for 23 h at the same temperature, then DCU precipitated was removed by suction. The filtrate was concentrated under reduced pressure. The residue was dissolved in AcOEt and the solution was washed with 5% aqueous KHSO4. The organic layer was dried over MgSO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (CH₂Cl₂-Et₂O, 3:1) to give crude 7, $[\alpha]_D^{20}$ +15.4° (c=2, CH₂Cl₂). Recrystallization from AcOEt–Et₂O gave pure sample of 7 (1.19 g, 62%) as colorless prisms, mp 93—94 °C, $[\alpha]_D^{20}$ +15.8° (c=2.9, CH₂Cl₂). IR (CHCl₃) v: 2250, 1720, 1670 cm⁻¹. ¹H-NMR (CDCl₃) δ : 2.61 (t, 2H, J = 6.4 Hz, CH₂CN), 3.11 (d, 2H, $J=6.0 \text{ Hz}, \text{ CH}_2\text{Ph}), 3.86 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28 \text{ (t, 2H, } J=4.4 \text{ Hz}, \text{ NCH}_2\text{CO)}, 4.28$ $J = 6.4 \text{ Hz}, \text{CO}_2\text{CH}_2$, 4.82 (q, 1H, J = 6.0 Hz, CH), 5.12 (s, 2H, OCH₂Ph), 5.3—5.5 (br, 1H, amide NH), 6.4—6.6 (br d, 1H, urethane NH), 7.1—7.5 (m, 10H, phenyl protons). Anal. Calcd for C₂₂H₂₃N₃O₅: C, 64.53; H, 5.66; N, 10.26. Found: C, 64.54; H, 5.71; N, 10.31.

Z-Gly-Phe-OH (8) A solution of 1.0 m TBAF (1.1 ml, 1.1 mmol) in THF was added to a solution of crude **7** (300 mg, 0.73 mmol) in DMF (5 ml) at 0 °C, and stirring was continued for 2 h at room temperature. The mixture was added to saturated aqueous NaHCO₃ and the solution was washed with AcOEt. The aqueous layer was acidified with 5% aqueous KHSO₄ and extracted with AcOEt. The extract was washed with water, dried over MgSO₄, and concentrated to give crude **8**, $[\alpha]_D^{24} + 37.7^{\circ}$ (c = 2, EtOH). Recrystallization from AcOEt–Et₂O gave pure **8** (217 mg, 83%) as colorless prisms, mp 126 °C, $[\alpha]_D^{24} + 38.3^{\circ}$ (c = 2.4, EtOH), lit., ¹⁵⁾ mp 125—126 °C, $[\alpha]_D^{24} + 38.5^{\circ}$ (c = 5, EtOH). IR (KBr) $v: 3275, 1730, 1680, 1650 \, \text{cm}^{-1}$. ¹H-NMR (acetone- d_6) δ: 3.0—3.3 (m, 2H, NCH₂CO), 3.83 (d, 2H, $J = 6.0 \, \text{Hz}$, CH₂Ph), 4.75 (q, 1H, $J = 6.0 \, \text{Hz}$, CH), 5.10 (s, 2H, OCH₂Ph), 6.5—6.7 (br, 1H, NH), 7.0—7.7 (m, 10H, phenyl protons).

References and Notes

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- 5) Amino acids and their derivatives are all of L form. Symbols and nomenclature follow the recommendations published by the IUPAC-IUB Commission on Biochemical Nomenclature for amino acids and peptides: Eur. J. Biochem., 138, 9 (1984). The following abbreviations for protecting groups in peptide synthesis are used: Boc, tert-butoxycarbonyl, Cet, 2-cyanoethyl; Z, benzyloxycarbonyl.
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- 13) This compound was easily prepared by catalytic hydrogenation of **4e** (72%) as colorless prisms, mp 45—45.5 °C (Et₂O-hexane). IR (CHCl₃) v: 2250, 1740, 1720 cm⁻¹. ¹H-NMR (CDCl₃) δ: 2.65—2.8
- (m, 6H), 4.32 (t, 2H, J=6.3 Hz, CO₂CH₂). *Anal.* Calcd for C₇H₉NO₄: C, 49.12; H, 5.30; N, 8.18. Found: C, 48.97; H, 5.34; N, 8.11.
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