Synthesis and Antibacterial Activities of 2-Oxaisocephems

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A series of 2-oxaisocephems with a thio-substituted methyl group at the 3-position and a 2-aminothiazol-4-yl moiety at the 7-position was synthesized via benzyl 3-acetyloxymethyl-7-azido-8-oxo-1-aza-4-oxabicyclo[4.2.0]oct-2-ene-2-carboxylate (2), derived from benzyl acetoacetate (1). The new 2-oxaisocephems were tested for antibacterial activities. Among them, the derivatives having a [2-(2-aminothiazol-4-yl)-2-(Z)-cyclopentyloxyimino]acetamido group at the 7-position characteristically showed potent activities against gram-positive bacteria including methicillin-resistant Staphylococcus aureus (MRSA) and Enterococcus faecalis as compared with cefuzonam and cefmenoxime, which are third-generation cephalosporins.

Keywords 2-oxaisocephem; methicillin-resistant *Staphylococcus aureus*; antibacterial activity; *Enterococcus faecalis*; [2-(2-aminothiazol-4-yl)-2-(Z)-cyclopentyloxyimino]acetamido group; thio-substituted methyl group

Several investigators have examined 2-oxaisocephem derivatives, 1-3) but Doyle *et al.* 1) reported that 2-oxaisocephems lacked comprehensive antibacterial activity, and 2-oxaisocephem and cephalosporin nuclei with the same side-chain possessed about the same inherent activity. 1a) Later, Mastalerz *et al.* proposed the synthesis of orally absorbable 2-oxaisocephems which would be effective against gram-positive organisms. 2) However, these compounds mostly have the side-chains of the first-generation cephalosporins at the 7-position. We have been searching for compounds with more potent and broad-spectrum antibacterial activity.

The cephalosporin class of antibacterial agents continues to be of clinical importance for the treatment of infection. A significant advance in this area was the introduction of the [2-(2-aminothiazol-4-yl)-2-(Z)-methoxyimino]acetamido side-chain at the 7-position.⁴⁾ Extended modification of this side-chain in combination

with alteration of the substituent at the 3-position of the cephalosporin nucleus has led to the preparation of some potent antibiotics. Cephalosporins with an aminothiazolyloxyiminoacetamido group at the 7-position have been introduced as third-generation antibacterial agents. We speculated that the introduction of side-chains of the third-generation cephalosporins into the 7-position of 2-oxaisocephems and alteration of the 3-substituents might enhance the activity and broaden the antibacterial spectrum. To discover agents which display a better gram-positive spectrum while maintaining gramnegative activity, we have prepared a series of novel 2-oxaisocephems. In particular, we wished to find compounds with improved antibacterial activity against methicillin-resistant Staphylococcus aureus (MRSA) and Enterococcus faecalis.

We report herein the synthesis and antibacterial activity of new 2-oxaisocephem derivatives having a thio-

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substituted methyl group at the 3-position and the 2-aminothiazol-4-yl moiety at the 7-position.

Synthesis

First, the 3-thio-substituted compounds 5, which are key intermediates in the synthesis of new 2-oxaisocephems (8a—x and 10a—d), were prepared. The starting material 2 was synthesized in 10 steps from benzyl acetoacetate 1 according to Doyle's method. 1a,e) To introduce the thio-substituted methyl group at the 3-postition of 2-oxaisocephems, the use of trimethylsilyl iodide (TMSI)⁵⁾ was found to be efficient. Typical procedures to obtain 5 are as follows. A solution of the 3-acetyloxymethyl derivative 2 (10 g, 26.88 mmol) in anhydrous CH₂Cl₂ (150 ml) was treated with 2.2 eq of TMSI (11.82 g, 59.14 mmol) at 0 °C for 1 h. A solution of 2 eq of thiol compound 3 and triethylamine in dry CH₂Cl₂ (100 ml) was added, and the reaction mixture was stirred at room temperature for 2 h to give 4. The crude 4, without purification, was allowed to react with 2 eq of diphenyldiazomethane at room temperature for 2h to afford 5 (65–76% yields). Compounds 3, 4, and 5 are shown in Table I.

Next, we wished to convert the 3-thio-substituted compounds $\bf 5$ into the target compounds $\bf (8a-x)$. Reduction of $\bf 5$ using hydrogen sulfide $\bf (H_2S)$ in the presence of triethylamine in $\bf CH_2Cl_2$ at room temperature followed by in situ acylation with 1-hydroxybenzotriazole (HOBT) active esters $\bf 6$ gave $\bf 7$ having a 2-aminothiazol-4-yl moiety at the 7-position in good yields (70—75% yields). The HOBT active esters $\bf 6$ are listed in Table II. The

TABLE I. Compounds 3, 4, and 5

	a	b	c	d	e
R¹		-√s,'n		N-N CH ₃	— N

carboxylic acids used to prepare HOBT active esters were synthesized essentially according to the published method. (6) New 2-oxaisocephems 8 were easily derived from thus obtained 7. To obtain new compounds, it was required to cleave the benzhydryl ester of 7. When 7 was treated with trifluoroacetic acid (TFA) in the presence of anisole at 0 °C for 10 min, the benzhydryl ester was cleaved smoothly to afford 8 (60—65% yields). Treatment of 7 with TFA for a longer time often resulted in decomposition of products and difficulty of purification. The synthesized 7 and 8 are listed in Table III.

Furthermore, we synthesized 2-oxaisocephems having an (N-alkylpyridinium-4'-thio)methyl group at the 3-position from 8w and 8x. Compounds 8w and 8x were treated with 3 eq of N,O-bis(trimethylsilyl)acetamide (BSA) in N,N-dimethylformamide (DMF) at room temperature for 1 h, halides 9 were added, and the mixture was stirred at 0 °C-room temperature for 6 h to give 10a—d (55—60% yields). Compounds 10 were isolated as hydrogensulfates. The synthesized 10 are listed in Table IV.

In summary, we have established a convenient synthetic route to new 2-oxaisocephems from the 3-acetyloxymethyl derivative **2**, and this method allows us to introduce various substituents into the 3-position and the 7-position.

Biological Results

The compounds (8a—x and 10a—d) prepared in this investigation were tested for *in vitro* antibacterial activities against gram-positive (*Staphylococcus aureus* FDA 209P, MRSA 57, and *Enterococcus faecalis*) and gramnegative (*Escherichia coli* NIHJ JC-2 and *Pseudomonas aeruginosa* ATCC 10145) bacteria. The minimum inhibitory concentrations (MIC, μ g/ml; inoculum size, 10^6 cells/ml) against test organisms were determined by the twofold agar dilution method.⁷⁾ The results are summarized in Table VII. The antibacterial activity of cefuzonam and cefmenoxime as reference compounds is also presented.

Among the sixteen compounds (8a—p) with a (1,3,4-

TABLE II. Compounds 6

$$\begin{array}{c|c}
N & NOR^2 \\
 & CO_2 - N N \\
 & S \end{array}$$

	a	b	c	d	e	f		
R ²	CH ₃	CH ₂ CH ₃	CH ₂ SCH ₃	CH ₂ CH ₂ Cl	CH ₂ CH ₂ F	CH₂<		
	g	h	i	j	k	1		
R ²	CH ₂ CN	$CH_2CH = CH_2$	$CH_2CH = CHCH_3$	$CH_2CH_2CH = CH_2$	CH ₂ C≡CH	CH ₂ Ph		
	m	n	0	p				
\mathbb{R}^2	$-CH_2$ N		$\overline{}$	_				

TABLE III. Compounds 7 and 8

e III. Co	mpounds / and o					
	a	b	c	d	e	f
R¹	N-N —(°)	N-N (', '\)	N-N //_//		(_\%_\%	(_\%_\%
R ²	CH ₃	CH₂CH₃	CH₂SCH₃	CH ₂ CH ₂ Cl	CH ₂ CH ₂ F	CH ₂
	g	h	i	j	k	1
R ¹	N-N —(' _S ')	N-N (',')	N-N -// '\		N-N 	________________\
R ²	CH₂CN	$CH_2CH = CH_2$	CH ₂ CH=CHCH ₃	$CH_2CH_2CH = CH_2$	CH ₂ C≡CH	CH ₂ Ph
	m	n	0	p	q	r
\mathbb{R}^1					$ ^{N-N}_{S}$ $^{\sim}$ CH ₃	$-\sqrt{\sum_{N}^{N}}$
\mathbb{R}^2	$-CH_2 N$	_	-	—	→	CH ₃
	s	t .	u	V	W	x
\mathbb{R}^1	_√s,'n	–√s,'n			— N	$-\sqrt{\sum}N$
\mathbb{R}^2	CH₂<	$\overline{}$	CH ₃	$\overline{}$	CH_3	$\overline{}$
		-				

Table IV. Compounds 10

	a	b	c	d
\mathbb{R}^2	CH ₃		CH ₃	$\overline{}$
\mathbb{R}^3	CH ₃	CH_3	CH ₂ COCH ₃	CH ₂ COCH ₃

thiadiazol-2-yl)thiomethyl group at the 3-position, that with a [2-(2-aminothiazol-4-yl)-2-(Z)-cyclopentyloxyimino acetamido group at the 7-position (80) showed significantly increased activities against all the bacteria tested including MRSA and E. faecalis, which are resistant to most cephalosporins, as compared with cefuzonam and cefmenoxime. The replacement of the (1,3,4-thiadiazol-2yl)thiomethyl group at the 3-position of 80 by other thio-substituted methyl substituents (8q, 8t, 8v, and 8x) also increased the activities against both gram-positive and gram-negative bacteria as compared with reference compounds. In particular, 8v with a (1,2,4-thiadiazol-5yl)thiomethyl group at the 3-position showed wellbalanced and potent antibacterial activity against test organisms including MRSA and E. faecalis. But, substitution with a (2-methyl-1,3,4-thiadiazol-5-yl)thiomethyl or (4-pyridyl)thiomethyl group at the 3-position (8q and 8x) caused a decrease in the activity against MRSA as compared with 8t and 8v. In the four compounds (8a,

8r, 8u, and 8w) with a [2-(aminothiazol-4-yl)-2-(Z)-methoxyimino]acetamido group, which is often used as the side-chain of third-generation cephalosporins at the 7-position, a decrease of the activity against MRSA and E. faecalis was observed. Among the four 3-(N-alkyl-pryidinium-4'-thio)methyl compounds (10a—d), 10b and 10d derived from 8x also possessed broader spectra of antibacterial activity than cefuzonam and cefmenoxime, and 10b showed about the same in vitro activity as 8v. Substitution of the 7-position by a [2-(2-aminothiazol-4-yl)-2-(Z)-methoxyimino]acetamido group (10a and 10c) increased the activity against E. coli, but decreased the activity against MRSA and E. faecalis.

Among the above compounds, **8v** and **10b** showed potent and broad-spectrum activities against test organisms including MRSA and *E. faecalis*, which cause a serious clinical problem in antibacterial chemotherapy.

Experimental

All the melting points were determined on a Yanaco MP-500D apparatus and are uncorrected. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker AC250 instrument operating at 250 MHz. Chemical shifts are expressed in parts per million (ppm) on the δ scale from internal tetramethylsilane and coupling constants in Hz. Infrared (IR) spectra were measured for KBr pellets with a JASCO IR-810 infrared spectrophotometer.

Benzyl 3-Acetyloxymethyl-7-azido-8-oxo-1-aza-4-oxabicyclo[4.2.0]-oct-2-ene-2-carboxylate (2) This compound was prepared essentially as described by Doyle *et al.*^{1a,e)} in 10 steps from benzyl acetoacetate 1.

TABLE V. Benzhydryl 7-Azido-8-oxo-3-substituted-1-aza-4-oxabicyclo[4.2.0]oct-2-ene-2-carboxylates 5

		mp (°C) ^{b)}	Formula _	Analysis (%)					
Compound	Yield (%) ^{a)}			Calcd			Found		
				С	Н	N	С	Н	N
5b	74	208—209	C ₂₃ H ₁₈ N ₆ O ₄ S ₂	54.54	3.58	16.59	54.46	3.69	16.35
5c	70	205-206	$C_{23}H_{18}N_6O_4S_2$	54.54	3.58	16.59	54.33	3.56	16.41
5d	71	197—198	$C_{24}H_{20}N_6O_4S_2$	55.37	3.87	16.14	55.49	3.85	15.93
5e	65	192—193	$C_{26}H_{21}N_5O_4S$	62.51	4.24	14.02	62.41	4.22	13.80

a) Yield from 2. b) Decomposition.

TABLE VI. Substituted 8-Oxo-1-aza-4-oxabicyclo[4.2.0]oct-2-ene-2-carboxylic Acids 8 and 10

Compound	Yield (%) ^{a)}	1 H-NMR δ (250 MHz, DMSO- d_{6})
8b	62	1.24 (3H, t, $J = 6.3$ Hz), 3.90—4.30 (4H, m), 4.42—4.63 (3H, m), 5.70 (1H, dd, $J = 4.8$, 8.4 Hz), 6.78 (1H, s), 7.25 (2H, s), 9.20 (1H, d, $J = 8.4$ Hz), 9.57 (1H, s)
8c	60	2.20 (3H, s), 3.70—4.10 (2H, m), 4.40—4.70 (3H, m), 5.20 (2H, s), 5.74 (1H, dd, <i>J</i> =4.8, 8.4 Hz), 6.82 (1H, s), 7.30 (2H, s), 9.25 (1H, d, <i>J</i> =8.4 Hz), 9.51 (1H, s)
8d	61	3.71 - 4.13 (4H, m), 4.30 (2H, t, $J = 6.3$ Hz), $4.45 - 4.65$ (3H, m), 5.71 (1H, dd, $J = 4.8$, 8.5 Hz), 6.83 (1H, s), 7.31 (2H, s), 9.25 (1H, d, $J = 8.5$ Hz), 9.56 (1H, s)
8e	61	3.84 - 4.25 (2H, m), $4.50 - 4.94$ (7H, m), 5.71 (1H, dd, $J = 4.7$, 8.5 Hz), 6.84 (1H, s), 7.32 (2H, s), 9.28 (1H, d, $J = 8.5$ Hz), 9.56 (1H, s)
8f	64	0.2—0.38 (2H, m), 0.50—0.63 (2H, m), 1.00—1.23 (1H, m), 3.80—4.08 (4H, m), 4.48—4.65 (3H, m), 5.67 (1H, dd, <i>J</i> =4.8, 8.5 Hz), 6.77 (1H, s), 7.22 (2H, s), 9.24 (1H, d, <i>J</i> =8.5 Hz), 9.57 (1H, s)
8g	62	3.77—4.20 (2H, m), 4.47 —4.67 (3H, m), 5.05 (2H, s), 5.75 (1H, dd, J =4.8, 8.4 Hz), 6.95 (1H, s), 7.25 (2H, s), 9.44 (1H, d, J =8.4 Hz), 9.56 (1H, s)
8h	63	3.83 - 4.05 (2H, m), $4.43 - 4.70$ (3H, m), $5.15 - 5.40$ (3H, m), 5.70 (1H, dd, $J = 4.7$, 8.4 Hz), $5.83 - 6.05$ (2H, m), 6.79 (1H, s), 7.22 (2H, s), 9.23 (1H, d, $J = 8.4$ Hz), 9.57 (1H, s)
8i	61	1.73—1.92 (3H, m), 3.73—4.17 (2H, m), 4.47—5.00 (5H, m), 5.43—5.60 (1H, m), 5.64—5.93 (2H, m), 6.81 (1H, s), 7.23 (2H, s), 9.19 (1H, d, J=8.5 Hz), 9.51 (1H, s)
8 j	60	2.25—2.48 (2H, m), 3.51—4.51 (4H, m), 4.33—4.75 (3H, m), 4.92—5.27 (2H, m), 5.66—6.05 (2H, m), 6.85 (1H, s), 7.22 (2H, s), 9.27 (1H, d, <i>J</i> =8.5 Hz), 9.56 (1H, s)
8k	61	3.45 - 3.58 (1H, m), $3.70 - 4.10$ (2H, m), $4.28 - 4.88$ (5H, m), 5.69 (1H, dd, $J = 4.8$, 8.4 Hz), 6.84 (1H, s), 7.23 (2H, s), 9.27 (1H, d, $J = 8.4$ Hz), 9.52 (1H, s)
81	65	3.71— 4.55 (5H, m), 5.14 (2H, s), 5.69 (1H, dd, J = 4.8 , 8.5 Hz), 6.82 (1H, s), 7.16 — 7.44 (7H, m), 9.28 (1H, d, J = 8.5 Hz), 9.57 (1H, s)
8m	62	3.78—4.17 (2H, m), 4.31–4.82 (3H, m), 5.38 (2H, s), 5.73 (1H, dd, <i>J</i> =4.7, 8.3 Hz), 6.79 (1H, s), 7.24 (2H, s), 7.71 (2H, d, <i>J</i> =6.3 Hz), 8.69 (2H, d, <i>J</i> =6.3 Hz), 9.33 (1H, d, <i>J</i> =8.3 Hz), 9.44 (1H, s)
8n	65	0.75 - 1.00 (6H, m), $1.45 - 1.78$ (4H, m), $3.83 - 4.15$ (3H, m), $4.43 - 4.65$ (3H, m), 5.68 (1H, dd, $J = 4.7$, 8.4 Hz), 6.77 (1H, s), 7.22 (2H, s), 9.21 (1H, d, $J = 8.4$ Hz), 9.57 (1H, s)
80	65	1.38—1.86 (8H, m), 3.84—4.06 (2H, m), 4.45—4.73 (4H, m), 5.66 (1H, dd, J =4.8, 8.5 Hz), 6.75 (1H, s), 7.22 (2H, s), 9.16 (1H, d, J =8.5 Hz), 9.57 (1H, s)
8 p	65	1.12—1.98 (10H, m), 3.85—4.13 (3H, m), 4.43—4.63 (3H, m), 5.67 (1H, dd, <i>J</i> =4.8, 8.5 Hz), 6.74 (1H, s), 7.21 (2H, s), 9.17 (1H, d, <i>J</i> =8.5 Hz), 9.57 (1H, s)
8q	60	1.40—1.87 (8H, m), 2.69 (3H, s), 3.83—4.12 (2H, m), 4.43—4.75 (4H, m), 5.65 (1H, dd, <i>J</i> =4.8, 8.5 Hz), 6.75 (1H, s), 7.22 (2H, s), 9.16 (1H, d, <i>J</i> =8.5 Hz)
8r	62	3.75— 4.02 (5H, m), 4.30 (1H, d, $J=14$ Hz), 4.45 (1H, d, $J=14$ Hz), 4.61 (1H, dd, $J=3$, 11.1 Hz), 5.73 (1H, dd, $J=4.8$, 9 Hz), 6.81 (1H, s), 7.25 (2H, s), 8.90 (1H, s), 9.24 (1H, d, $J=9$ Hz)
8s	63	0.18—0.39 (2H, m), 0.51—0.64 (2H, m), 0.92—1.26 (1H, m), 3.63—4.08 (4H, m), 4.25 (1H, d, <i>J</i> =14Hz), 4.53 (1H, d, <i>J</i> =14Hz), 4.62 (1H, dd, <i>J</i> =3.1, 11.2Hz), 5.67 (1H, dd, <i>J</i> =4.7, 8.4Hz), 6.75 (1H, s), 7.17 (2H, s), 8.85 (1H, s), 9.18 (1H, d, <i>J</i> =8.4Hz)
8t	65	1.35— 1.90 (8H, m), 3.85 — 4.10 (2H, m), 4.25 (1H, d, $J = 14.2$ Hz), 4.45 — 4.72 (3H, m), 5.66 (1H, dd, $J = 4.8$, 8.4 Hz), 6.75 (1H, s), 7.24 (2H, s), 8.90 (1H, s), 9.16 (1H, d, $J = 8.4$ Hz)
8u	61	3.80 - 4.03 (5H, m), 4.44 - 4.70 (3H, m), 5.72 (1H, dd, J = 4.8, 8.4 Hz), 6.79 (1H, s), 7.21 (2H, s), 8.73 (1H, s), 9.23 (1H, d, J = 8.4 Hz)
8v	65	1.38—1.90 (8H, m), 3.80—4.05 (2H, m), 4.48—4.70 (4H, m), 5.66 (1H, dd, <i>J</i> =4.8, 8.4 Hz), 6.74 (1H, s), 7.23 (2H, s), 8.73 (1H, s), 9.14 (1H, d, <i>J</i> =8.4 Hz)
8w	60	3.85—4.19 (5H, m), 4.25 (1H, d, <i>J</i> =14 Hz), 4.40 (1H, d, <i>J</i> =14 Hz), 4.48 (1H, dd, <i>J</i> =3, 10.3 Hz), 5.85 (1H, dd, <i>J</i> =4.6, 8.3 Hz), 6.69 (1H, s), 7.32 (2H, d, <i>J</i> =6.2 Hz), 8.36 (2H, d, <i>J</i> =6.2 Hz), 9.14 (1H, d, <i>J</i> =8.3 Hz)
8x	60	1.40—1.88 (8H, m), 3.83—4.08 (2H, m), 4.28 (1H, d, J =14.1 Hz), 4.41 (1H, d, J =14.1 Hz), 4.49 (1H, dd, J =2.8, 10 Hz), 4.59—4.76 (1H, m), 5.63 (1H, dd, J =4.4, 8.3 Hz), 6.74 (1H, s), 7.38 (2H, d, J =6.2 Hz), 8.38 (2H, d, J =6.2 Hz), 9.12 (1H, d, J =8.3 Hz)
10b	60	1.40—1.85 (8H, m), 3.85—4.05 (2H, m), 4.19 (3H, s), 4.45—4.75 (4H, m), 5.65 (1H, dd, <i>J</i> =4.4, 8.2 Hz), 6.78 (1H, s), 8.02 (2H, d, <i>J</i> =7.1 Hz), 8.70 (2H, d, <i>J</i> =7.1 Hz), 9.20 (1H, d, <i>J</i> =8.2 Hz)
10c	55	2.27 (3H, s), 3.78—4.21 (4H, m), 4.45—4.75 (4H, m), 5.50 (2H, s), 5.64 (1H, dd, <i>J</i> =4.5, 8.3 Hz), 6.75 (1H, s), 8.09 (2H, d, <i>J</i> =7.1 Hz), 8.52 (2H, d, <i>J</i> =7.1 Hz), 9.15 (1H, d, <i>J</i> =8.3 Hz)
10d	58	1.40—1.83 (8H, m), 2.29 (3H, s), 3.94—4.10 (2H, m), 4.41—4.77 (4H, m), 5.56 (2H, s), 5.66 (1H, dd, <i>J</i> =4.5, 8.3 Hz), 6.76 (1H, s), 8.10 (2H, d, <i>J</i> =7.1 Hz), 8.52 (2H, d, <i>J</i> =7.1 Hz), 9.14 (1H, d, <i>J</i> =8.3 Hz)

a) 8b-x: yields from 7b-x respectively, 10b and 10d: yields from 8x, 10c: yield from 8w.

TABLE VII. In Vitro Antibacterial Activity (MIC, µg/ml)

Compound	S. aureus FDA 209P	MRSA 57	E. faecalis	E. coli NIHJ JC-2	P. aeruginosa ATCC 10145
8a	0.78	>100	50	0.2	12 5
8b	1.56	> 100	50	0.78	25
8c	1.56	>100	25	1.56	25
8d	0.78	>100	12.5	0.78	12.5
8e	1.56	>100	25	0.2	12.5
8f	0.78	>100	25	0.39	12.5
8g	1.56	> 100	50	0.2	12.5
8h	0.78	100	12.5	0.39	12.5
8i	0.39	100	12.5	0.78	12.5
8j	0.39	100	12.5	1.56	12.5
8k	1.56	>100	25	0.39	12.5
81	0.39	100	12.5	0.78	12.5
8m	0.78	100	25	0.78	12.5
8n	0.39	25	12.5	0.78	12.5
8o	0.39	12.5	6.25	0.39	12.5
8p	0.39	12.5	6.25	1.56	12.5
8q	0.39	25	6.25	0.78	12.5
8r	0.2	>100	25	0.2	25
8s	0.39	50	6.25	0.39	6.25
8t	0.39	6.25	3.13	0.78	12.5
8u	0.39	50	25	0.2	6.25
8v	0.2	3.13	1.56	0.39	6.25
8w	0.39	> 100	100	0.2	>100
8x	0.2	25	3.13	0.78	12.5
10a	0.2	100	25	< 0.025	12.5
10b	0.1	6.25	1.56	0.39	6.25
10c	0.39	100	50	< 0.025	25
10d	0.39	12.5	3.13	0.39	12.5
Cefuzonam	0.39	100	100	0.1	25
Cefmenoxime	1.56	>100	>100	0.1	25

Benzhydryl 7-Azido-8-oxo-3-[(1,3,4-thiadiazol-2-yl)thiomethyl]-1-aza-4-oxabicyclo[4.2.0]oct-2-ene-2-carboxylate (5a) Trimethylsilyl iodide (11.82 g, 59.14 mmol) was added dropwise to a stirred solution of 2 (10 g, 26.86 mmol) in CH₂Cl₂ (150 ml) at 0 °C. The mixture was stirred for 1 h, then a solution of 2-mercapto-1,3,4-thiadiazole 3a (6.35 g, 53.72 mmol) and triethylamine (5.46 g, 53.72 mmol) in CH₂Cl₂ (100 ml) was added dropwise over a period of 30 min. The reaction mixture was stirred at room temperature for 2h, and MeOH (50 ml) was added. The mixture was evaporated under reduced pressure. After treatment of the residue with Et_2O , the resultant crude 4a ($R^1 = (1,3,4-thiadiazol-2-yl)thiomethyl)$ was collected by filtration. Diphenyldiazomethane (10.43 g, 53.72 mmol) was added to a solution of this crude 4a in tetrahydrofuran (THF) (100 ml) and MeOH (100 ml), and the mixture was stirred at room temperature for 2h. The solvent was evaporated off under reduced pressure. The residue was purified by silica gel column chromatography (eluent, CH₂Cl₂/AcOEt = 6/1) and recrystallized from AcOEt/n-hexane to give 5a ($10.\overline{3}4$ g, 76%) as colorless needles, mp 196—197 °C (dec.). NMR (CDCl₃) δ : 3.72—3.85 (1H, m), 4.00 (1H, dd, J=9.5, 11.1 Hz), 4.47 (1H, d, J = 13.7 Hz), 4.59 (1H, dd, J = 3.7, 11.1 Hz), 4.76 (1H, d, J=13.7 Hz), 5.24 (1H, d, J=5.1 Hz), 6.89 (1H, s), 7.24—7.60 (10H, m), 8.95 (1H, s). IR cm⁻¹: 2090, 1780, 1710, 1610. *Anal.* Calcd for $C_{23}H_{18}N_6O_4S_2$: C, 54.54: H, 3.58; N, 16.59. Found: C, 54.83; H, 3.52; N, 16.29.

Compounds 5b—e were obtained by the same procedure as described for 5a; yields, melting points, and elemental analysis data are given in Table V.

Benzhydryl 7-[2-(2-Aminothiazol-4-yl)-2-[(Z)-methoxyimino]acetamido]-8-oxo-3-[(1,3,4-thiadiazol-2-yl)thiomethyl]-1-aza-4-oxabicyclo-[4.2.0]oct-2-ene-2-carboxylate (7a) A stirred mixture of 5a (5g, 9.87 mmol) and triethylamine (2g, 19.74 mmol) in CH₂Cl₂ (100 ml) was cooled in an ice bath and saturated with H₂S. While stirring, gas evolution occurred. The solution was stirred at room temperature for 1 h, washed with 5% aqueous NaHCO₃ solution (50 ml × 3) and brine (50 ml), dried over Na₂SO₄, and filtered. To this filtrate was added the HOBT active ester 6a prepared from the corresponding carboxylic acid (1.99 g, 9.87 mmol), N, N'-dicyclohexylcarbodiimide (DCC) (2.04 g, 9.87 mmol), and HOBT (1.33 g, 9.87 mmol) in CH₂Cl₂ (50 ml), and this mixture was

stirred at room temperature for 10 h. The solvent was removed under reduced pressure and the residue was purified by silica gel column chromatography (eluent, $\text{CH}_2\text{Cl}_2/\text{AcOEt}=6/1$) to give **7a** (4.72 g, 72%) as a pale yellow powder. NMR (CDCl₃) δ : 3.80—4.20 (5H, m), 4.23 (1H, d, J=13.7 Hz), 4.31 (1H, d, J=13.7 Hz), 4.68 (1H, dd, J=3.7, 11.1 Hz), 5.65 (1H, dd, J=4.8, 6.5 Hz), 6.84 (1H, s), 6.90 (1H, s), 7.20—7.60 (10H, m), 9.01 (1H, s). IR cm⁻¹: 3340, 1780, 1700, 1670, 1610, 1530.

Compounds 7b-x were obtained by the same procedures as described for 7a.

7-[2-(2-Aminothiazol-4-yl)-2-[(Z)-methoxyimino]acetamido]-8-oxo-3-[(1,3,4-thiadiazol-2-yl)thiomethyl]-1-aza-4-oxabicyclo[4.2.0]oct-2-ene-2-carboxylic Acid (8a) A stirred suspension of 7a (2 g, 3.01 mmol) in anisole (4 ml) was treated with trifluoroacetic acid (10 ml) at 0 °C. Vigorous stirring was maintained for 10 min, after which time $\rm Et_2O$ (80 ml) was added to the solution. The resulting precipitates were collected by filtration, and washed with AcOEt. The precipitates were dissolved in 5% aqueous NaHCO₃ solution (100 ml) and insoluble substances were filtered off. The filtrate was adjusted to pH 4 with 10% HCl, and subjected to chromatography on Diaion HP-20 with $\rm CH_3CN-H_2O$ mixtures. After combining the appropriate fractions and evaporation under reduced pressure to remove $\rm CH_3CN$, freeze-drying gave 8a (944 mg, 63%) as a white powder. NMR (DMSO- d_6) δ : 3.75—4.08 (5H, m), 4.45—4.65 (3H, m), 5.72 (1H, dd, J=4.7, 9.1 Hz), 6.83 (1H, s), 7.31 (2H, s), 9.25 (1H, d, J=9.1 Hz), 9.58 (1H, s). IR cm $^{-1}$: 3400, 1760, 1750, 1700, 1670.

Compounds 8b—x were obtained by the same procedure as described for 8a; yields and NMR data are given in Table VI.

7-[2-(2-Aminothiazol-4-yl)-2-[(Z)-methoxyimino] acetamido]-3-[(1-yl)-2-[(Z)-methoxyimino]] $methyl pyridinium \hbox{-} 4-yl) thiomethyl] \hbox{-} 8-oxo-1-aza-4-oxabicyclo \hbox{$[4.2.0]$ octages a positive of the property of the$ 2-ene-2-carboxylic Acid Hydrogensulfate (10a) N,O-Bis(trimethylsilyl)acetamide (1.24 g, 6.12 mmol) was added dropwise to a solution of 8x (1 g, 2.04 mmol) in DMF (4 ml) at 0 °C, and the mixture was stirred at room temperature for 1 h. Then methyl iodide (868 mg, 6.12 mmol) was added. Stirring was continued at room temperature for 6 h, and iso-PrOH (20 ml) and Et₂O (20 ml) were added to the solution. The resulting precipitates were collected by filtration, washed with iso-PrOH and Et₂O, and dissolved in 5% aqueous NaHCO3 solution (100 ml), and insoluble substances were filtered off. The filtrate was adjusted to pH 6 with 10% HCl, and subjected to chromatography on Diaion HP-20 using CH₃CN-H₂O mixtures. The appropriate fractions were combined and evaporated under reduced pressure to remove CH3CN. Then 4N H2SO4 (2 ml) was added to the residual aqueous solution in an ice bath, and the mixture was stirred for 30 min. The resultant precipitates were collected by filtration to give 10a (700 mg, 57%) as a white powder. NMR (DMSO- d_6) δ : 3.85—4.13 (5H, m), 4.21 (3H, m), 4.45—4.73 (3H, m), 5.64 (1H, dd, J = 4.5, 8.4 Hz), 6.78 (1H, s), 8.02 (2H, d, J = 7.1 Hz), 8.71 (2H, d, J = 7.1 Hz), 9.21 (1H, d, J = 8.4 Hz). IR cm⁻¹: 3300, 1780, 1750, 1700, 1680.

Compounds 10b—d were obtained by the same procedure as described for 10a; yields and NMR data are given in Table VI.

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