

## Comparison of Singlet Oxygen Production Efficiency of C<sub>60</sub> with Other Photosensitizers, Based on 1268 nm Emission

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Singlet oxygen generation from laser-excited C<sub>60</sub> was measured directly by using a sensitive near-infrared emission spectrometer to monitor the O<sub>2</sub>(<sup>1</sup>A<sub>g</sub>)→O<sub>2</sub>(<sup>3</sup>Σ<sub>g</sub><sup>-</sup>) transition at 1268 nm. The emission intensity was proportional to both the laser power and the concentration of C<sub>60</sub>. When nitrogen gas was bubbled into the C<sub>60</sub> solution, the spectrum disappeared. Dose-dependent quenching of singlet oxygen by β-carotene was observed by monitoring the 1268 nm emission. Based on the quenching, the rate constant of the reaction of singlet oxygen with β-carotene was determined to be 1.5–2.3 × 10<sup>10</sup> M<sup>-1</sup> s<sup>-1</sup>, which is consistent with the reported value (1.5–2.5 × 10<sup>10</sup> M<sup>-1</sup> s<sup>-1</sup>). A comparison of the singlet oxygen-producing ability of C<sub>60</sub> as a photosensitizer with those of rose bengal, eosin and methylene blue, revealed C<sub>60</sub> to have considerably higher efficiency.

**Keywords** singlet oxygen; C<sub>60</sub>; fullerene; photosensitizer; near-infrared emission

Singlet oxygen (<sup>1</sup>A<sub>g</sub>), a highly reactive and toxic intermediate, plays an important role in many photochemical and photobiological processes<sup>1</sup> and has been postulated to be a major species mediating photochemical action in biological systems.<sup>2</sup> Erythropoietic protoporphyria, tetracycline phototoxicity and photodynamic therapy of cancer are known to involve singlet oxygen as a species responsible for the action processes.<sup>3</sup> Many attempts have been made to find evidence of singlet oxygen generation in various photosensitized reactions and enzymatic systems.<sup>4</sup> Most of the investigations were based upon monitoring weak visible chemiluminescence, chemical products, chemical scavenger techniques and deuterium kinetic effects. Compared with these nonspecific techniques for the detection of singlet oxygen, direct observation of the (0,0) <sup>1</sup>A<sub>g</sub>→<sup>3</sup>Σ<sub>g</sub><sup>-</sup> singlet oxygen emission at 1268 nm has proven to be the most reliable approach for singlet oxygen detection. Since the first observation of photosensitized singlet oxygen emission in solution,<sup>5</sup> emission measurement has been widely used for detection and investigation of singlet oxygen in photochemical and biological systems.<sup>6</sup>

The recent frenzy surrounding the fullerenes has encouraged investigators to elucidate fully the properties of C<sub>60</sub>. In particular, the electronic properties of this unique π-system have attracted the attention of both photochemists and theoreticians. Based on photoacoustic,<sup>7</sup> thermal lensing<sup>8</sup> and singlet oxygen luminescence studies,<sup>9</sup> C<sub>60</sub> has been postulated to produce singlet oxygen by energy transfer with unit efficiency. Foote and co-workers have conducted detailed studies on singlet oxygen generation in C<sub>60</sub> solution using a kinetic apparatus based on a Ge-detector monitoring 1270 nm emission through interference filters.<sup>9,10</sup>

We have recently constructed a sensitive near-infrared emission spectrometer with a Ge-detector, which can scan from 1000 to 1600 nm. We applied this apparatus for detection of singlet oxygen produced by laser-excited C<sub>60</sub>. Since the most difficult problem in time-resolved

measurement of 1270 nm emission using interference filters is to distinguish the singlet oxygen phosphorescence from the near-infrared tail of the dye fluorescence and from the near-infrared emission due to temperature change, a better approach for comparison of singlet oxygen production efficiency of photosensitizers should be to acquire the signal due to the O<sub>2</sub>(<sup>1</sup>A<sub>g</sub>)→O<sub>2</sub>(<sup>3</sup>Σ<sub>g</sub><sup>-</sup>) transition at 1268 nm by scanning the near-infrared wavelength region.

In this work, we firstly measured the near-infrared emission spectrum corresponding to the O<sub>2</sub>(<sup>1</sup>A<sub>g</sub>)→O<sub>2</sub>(<sup>3</sup>Σ<sub>g</sub><sup>-</sup>) transition of singlet oxygen in C<sub>60</sub> solution under laser excitation. The experiments were performed using a sensitive near-infrared spectrometer to monitor quantitatively the emission intensity due to the O<sub>2</sub>(<sup>1</sup>A<sub>g</sub>)→O<sub>2</sub>(<sup>3</sup>Σ<sub>g</sub><sup>-</sup>) transition at 1268 nm. The essential parameter of a photosensitizer is its photodynamic efficiency, which is usually associated with a high quantum yield of singlet oxygen generation. In this paper, we also report the singlet oxygen-generating ability of C<sub>60</sub> compared with that of other sensitizers.

### Experimental

**Materials** Benzene (Wako Pure Chemical Industries Ltd., Japan), and methanol (Wako Pure Chemical Industries Ltd., Japan) were used as solvents for photosensitizers. C<sub>60</sub> was purified by using HPLC with an ODS column (eluting solvents: toluene/methanol=55/45). Rose bengal (RB) was from Wako Pure Chemical Co. Methylene blue (MB) and eosin yellowish (tetrabromofluorescein sodium salt) (EO) were from Tokyo Kasei Kogyo Co., Ltd., Japan.

**Singlet Oxygen Emission Measurement** Our experimental set up (Fig. 1) consisted of an argon (Ar) laser (Innova 70-4; Coherent Inc., U.S.A.) and a near-infrared Ge-detector (mode 403HS; Applied Detector Co., U.S.A.), cooled by liquid nitrogen, connected to the exit slit of a monochromator (model CT10; JASCO, Japan) with a blaze wavelength at 1250 nm to minimize the grating loss. An IR-80 cut off filter with 0% transmittance at less than 750 nm and 35% transmittance at 800 nm was placed at the entrance slit of the monochromator. A collecting lens focused the monochromator output onto the detector crystal. The Ar laser output at 514.5 nm was chopped at 800 Hz by an acousto-optic modulator (A-160; Hoya, Japan) driven by a driver (110-DS; Hoya, Japan). The signal output from the Ge-detector was fed to a model 124A lock-in amplifier via a model 116 preamplifier (both from E. G. & G. Princeton

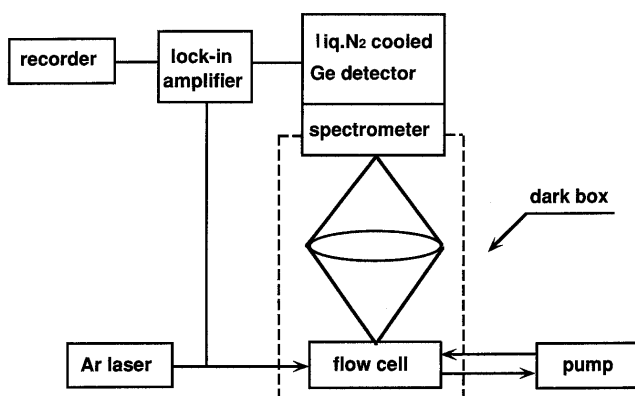


Fig. 1. Schematic Diagram of Our Near-Infrared Spectrometer

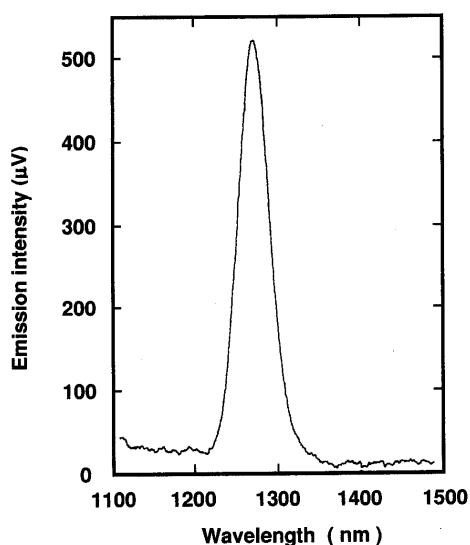


Fig. 2. Near-Infrared Singlet Oxygen Luminescence Emission Spectrum in  $C_{60}$  Benzene Solution ( $20 \mu M$ ) Excited by Ar Laser Light at 514.5 nm with 200 mW Output Power

Applied Research, U.S.A.) and synchronized with the internal reference signal of the lock-in amplifier. The signal output from the lock-in amplifier was fed to an XY recorder, and the emission spectrum was recorded by scanning the grating with a motor. In order to minimize photobleaching of the dye, the solution was circulated by a peristaltic pump through a quartz flow cell ( $3 \times 3$  mm).

## Results

**Near-Infrared Emission Spectrum Due to Singlet Oxygen Generated with  $C_{60}$  as a Photosensitizer** Figure 2 shows a near-infrared emission spectrum of  $C_{60}$  benzene solution excited with Ar laser light at 514.5 nm, corresponding to the  $O_2(^1\Delta_g) \rightarrow O_2(^3\Sigma_g^-)$  transition, obtained by using the apparatus shown in Fig. 1. The scanning time for the spectrum was about 15 min with a 2 mm slit for both the entrance and exit slits of the monochromator with a grating blaze at 1250 nm. We can confidently assign the narrow band at 1268 nm to the (0,0) vibronic band of the  $O_2(^1\Delta_g) \rightarrow O_2(^3\Sigma_g^-)$  transition. When dioxygen gas bubbles were present in the solution, the spectrum had a peak at 1268 nm, but the spectrum disappeared when nitrogen gas was bubbled in. These results confirm that singlet oxygen is responsible for the emission at 1268 nm.

The  $C_{60}$ -sensitized singlet oxygen emission spectrum in

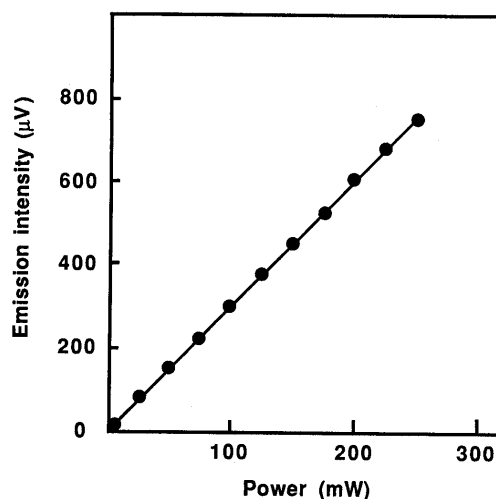


Fig. 3. Effect of Laser Power on the Singlet Oxygen Emission at 1268 nm from  $C_{60}$  Solution

Singlet oxygen generation in  $C_{60}$  benzene solution ( $25 \mu M$ ) excited by Ar laser light at 514.5 nm was monitored by measuring the emission intensity at 1268 nm.

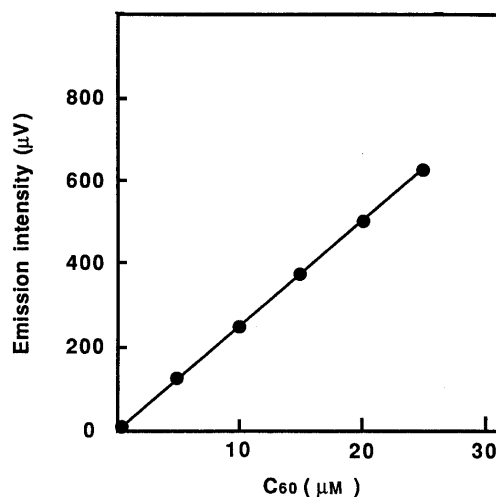


Fig. 4. Effect of  $C_{60}$  Concentration on the Singlet Oxygen Emission at 1268 nm from  $C_{60}$  Solution

Singlet oxygen generation in  $C_{60}$  benzene solution ( $5$ – $25 \mu M$ ) excited by Ar laser light at 514.5 nm with 200 mW output power was monitored by measuring the emission intensity at 1268 nm.

benzene solution did not show any overlap with fluorescence of  $C_{60}$ , in contrast to the cases of hematoporphyrin, MB and EO.

**Effect of Laser Power and  $C_{60}$  Concentration on 1268 nm Emission Intensity** Figure 3 shows the relationship between the emission intensity at 1268 nm in  $C_{60}$  solution and laser power. The emission intensity of singlet oxygen in laser-excited  $C_{60}$  solution increased with the increase of laser power in this range of laser power.

Figure 4 shows the dependence of the emission intensity at 1268 nm on the concentration of  $C_{60}$ . The absorbance spectrum of  $C_{60}$  at the same concentration (from  $5 \times 10^{-6}$  to  $25 \times 10^{-6} M$ ) changed dose-dependently, and the intensity of the absorbance at 514.5 nm was proportional to the concentration of  $C_{60}$  (Fig. 5). These results indicate that  $C_{60}$  exists as the monomer, and the interaction between molecules is essentially negligible in the benzene

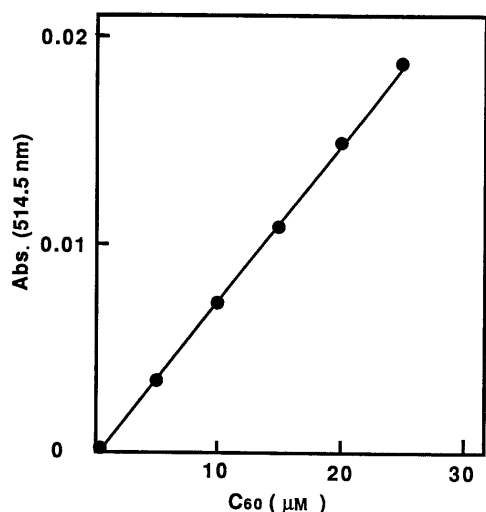


Fig. 5. Relationship between the Absorbance at 514.5 nm and the Concentration of  $C_{60}$

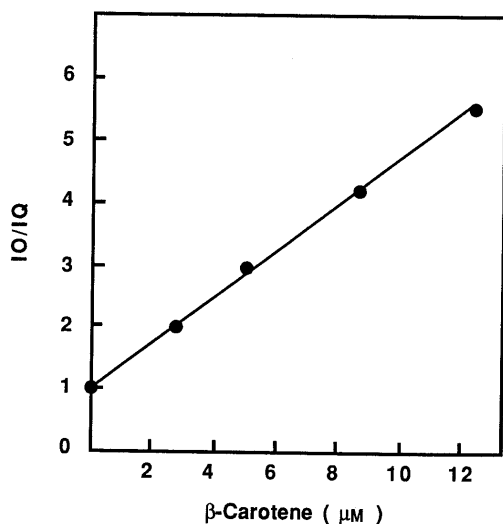


Fig. 6. Stern-Volmer Plots for Quenching of Singlet Oxygen with  $\beta$ -Carotene

Singlet oxygen was generated in  $C_{60}$  benzene solution excited by Ar laser light at 514.5 nm with 200 mW output power and monitoring was done by measuring the emission intensity at 1268 nm.

solution of  $C_{60}$  (from  $5 \times 10^{-6}$  to  $25 \times 10^{-6}$  M).

The emission at 1268 nm was also inhibited by the addition of a singlet oxygen quencher,  $\beta$ -carotene. Using  $C_{60}$ -excited singlet oxygen generation and quenching of singlet oxygen emission at 1268 nm with  $\beta$ -carotene, we determined the rate constant for  $[\beta\text{-carotene}][\text{singlet oxygen}]$ . The Stern-Volmer<sup>11)</sup> plot for the ratio of the emission intensity with (IQ) and without (IO) the quencher Q is  $IO/IQ = 1 + K_q\tau[Q]$ . A plot of  $IO/IQ$  vs.  $[Q]$  is shown in Fig. 6. From the singlet oxygen lifetime in benzene ( $\tau = 20\text{--}30 \mu\text{s}$ ),<sup>12)</sup> the value of  $K_q = 1.5\text{--}2.3 \times 10^{10} \text{ M}^{-1} \text{ s}^{-1}$  was obtained for  $\beta$ -carotene. This value is in good agreement with that in the literature.<sup>13)</sup>

**Comparison of Emission Intensity Due to  $C_{60}$  and Other Photosensitizers** Near-infrared spectroscopic studies in the wavelength region of 1268 nm were performed on other photosensitizer solutions in the same way as described

TABLE I. Comparison of Relative Emission Intensities at 1268 nm in Solutions of Various Photosensitizers

Photosensitizer	$A_{514.5}$	$\epsilon_{514.5}$	$O_2$ emission (I)	$I/\epsilon$	$I/\epsilon$ (rel.)
MB	0.0283	2830	485	0.1713	4.3
RB	0.2729	27290	2680	0.0982	2.5
EO	0.5471	54710	2160	0.0394	1.0
$C_{60}$	0.0074	740	350	0.4729	12.0

The emission intensity at 1268 nm was measured in the benzene/methanol solution of a photosensitizer at the concentration of  $10 \mu\text{M}$ , excited by Ar laser light at 514.5 nm with 200 mW output power.

above. The emission of singlet oxygen was detected in MB, RB, and EO benzene-methanol solutions as well. Table I compares the emission intensities of these solutions at 1268 nm. The values were calibrated on the basis of molar extinction coefficient and then normalized with respect to the value of the EO solution. The efficiencies ( $I/\epsilon$ ) of singlet oxygen generation were determined for MB, RB, EO and  $C_{60}$  as 0.1713, 0.0982, 0.0394 and 0.4729, respectively. The efficiency of singlet oxygen production by  $C_{60}$  is approximately 12-fold higher than that by EO in benzene-methanol solution under similar conditions. This difference in singlet oxygen generation may be ascribed to the differences in conversion efficiency from the excited singlet state of the dye to the triplet state and in the lifetime of the triplet state. The triplet state of  $C_{60}$  may be efficiently quenched by the ground state of dioxygen ( $^3O_2$ ).  $C_{60}$  was shown to generate singlet oxygen highly efficiently upon photoirradiation.

## Discussion

In this work, we firstly measured the direct near-infrared emission spectrum corresponding to the  $O_2(^1D_g) \rightarrow O_2(^3\Sigma_g^-)$  transition of singlet oxygen in  $C_{60}$  solution under laser excitation. The emission dependence on the intensity of laser power and on the concentration of the  $C_{60}$  suggests that  $C_{60}$  generates singlet oxygen quite efficiently upon photoirradiation, over a wide range of concentration.

We also compared the emission intensity of the  $C_{60}$  solution at 1268 nm with those of other dye solutions. The generation of singlet oxygen in  $C_{60}$  benzene-methanol solution was found to be 12-fold higher than that of EO benzene-methanol solution, and several-fold higher than with MB and RB (Table I). Although MB, RB and EO are the most frequently used sensitizers in photooxygenation and photodynamic studies,  $C_{60}$  had a much higher capacity of singlet oxygen generation than these sensitizers.

Because  $C_{60}$  possesses a high degree of symmetry, transitions between the singlet state ( $S_0$ ) and the singlet state ( $S_1$ ) are strongly forbidden. This forbiddenness, the small S-T splitting, the very low value of the fluorescence rate constant, and the large spin-orbital interaction in the spherical  $C_{60}$  molecule explain why intersystem crossing (ISC) is a dominant process. Therefore, the triplet state of  $C_{60}$  is formed in high yield and the triplet life time is very long ( $40 \pm 4 \mu\text{s}$ ).<sup>9)</sup> Generation of singlet oxygen may occur by energy transfer from the highly populated  $C_{60}$  triplet state to the dioxygen ground state ( $^3O_2$ ).

The intense absorption peak of  $C_{60}$  at 335 nm indicates

that C<sub>60</sub> generates large amounts of singlet oxygen under UV irradiation. Its very high singlet oxygen yield, especially under UV irradiation, and the inertness to photooxidative destruction suggest great potential for photodynamic action in biological systems.<sup>14)</sup>

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#### References

- 1) A. P. Schaap, "Singlet Molecular Oxygen Benchmark Papers in Organic Chemistry," Vol. 5, ed. by A. P. Schaap, Dowder, Hutchison and Ross, Stroudsburg, PA, 1976; A. A. Frimer, "Singlet O<sub>2</sub>," Vol. 1—4, ed. by A. A. Frimer, CRC Press, Inc., Boca Raton, FL, 1985; K. Gollnick, *Adv. Photochem.*, **6**, 1 (1968); J. D. Spikes, R. Livingston, *Adv. Radiation Biol.*, **3**, 29 (1969).
- 2) C. S. Foote, *Science*, **162**, 963 (1968); *idem*, "Light-Activated Pesticides, ACS Symposium Series 339," ed. by J. R. Heitz, K. R. Downum, American Chemical Society, Washington DC, pp. 22—38; T. Ito, *Photochem. Photobiol.*, **28**, 493 (1978).
- 3) T. Hasan, A. U. Khan, *Proc. Natl. Acad. Sci. U.S.A.*, **83**, 4604 (1986); A. S. W. Li, C. F. Chignell, R. D. Hall, *Photochem. Photobiol.*, **46**, 379 (1987); M. B. Poh-Fitzpatrick, *Liver Dis.*, **2**, 164 (1982); K. R. Weishaupt, C. J. Gomer, T. J. Dougherty, *Cancer Res.*, **36**, 2326 (1976); T. J. Dougherty, J. E. Kaufman, A. Goldfarb, K. R. Weishaupt, D. Boyle, A. Mittleman, *ibid.*, **38**, 2628 (1978); J. P. Keene, D. Kessel, E. J. Land, R. W. Redmond, T. G. Truscott, *Photochem. Photobiol.*, **43**, 117 (1986); D. Branult, C. Vever-Bizet, M. Rougee, R. Bensasson, *ibid.*, **47**, 151 (1988).
- 4) H. G. Krishnamurty, F. J. Simpson, *J. Biol. Chem.*, **245**, 1467 (1970); T. Matsuura, H. Matsushima, H. Sakamoto, *J. Am. Chem. Soc.*, **89**, 6370 (1967); R. C. Allen, R. L. Stjernholm, R. H. Steele, *Biochem. Biophys. Res. Commun.*, **47**, 679 (1972); H. Rosen, S. J. Klebanoff, *J. Biol. Chem.*, **252**, 4803 (1977); K. Takayama, T. Noguchi, M. Nakano, T. Migita, *Biochem. Biophys. Res. Commun.*, **75**, 1052 (1977); C. S. Foote, R. B. Abakerli, R. L. Clough, R. I. Lehrer, "Bioluminescence and Chemiluminescence," ed. by M. A. DeLuca, W. D. McElroy, Academic Press, Inc., New York, 1981, pp. 81—88.
- 5) A. A. Krasnovsky Jr., *Biofisika (Sov. Biophys.)*, **21**, 748 (1976); *idem*, *ibid.*, **22**, 927 (1977); A. U. Khan, M. Kasha, *Proc. Natl. Acad. Sci. U.S.A.*, **76**, 6047 (1979); A. U. Khan, *Chem. Phys. Lett.*, **72**, 112 (1980).
- 6) I. M. Byteva, G. P. Guvinovitch, *Z. Prikl. Spekt.*, **29**, 154 (1978); A. A. Krasnovsky, Jr., *Photochem. Photobiol.*, **29**, 29 (1979); A. U. Khan, *J. Am. Chem. Soc.*, **103**, 6516 (1981); A. U. Khan, P. Gebauer, L. P. Hager, *Proc. Natl. Acad. Sci. U.S.A.*, **80**, 5195 (1983); J. R. Kanofsky, *J. Biol. Chem.*, **258**, 5991 (1983); A. U. Khan, *J. Am. Chem. Soc.*, **105**, 7195 (1983); J. R. Kanofsky, J. Wright, G. E. Miles-Richardson, A. I. Tauber, *J. Clin. Invest.*, **74**, 1489 (1984); J. R. Kanofsky, *J. Biol. Chem.*, **259**, 5596 (1984); A. U. Khan, *Biochem. Biophys. Res. Commun.*, **122**, 668 (1984); *idem* *J. Photochem.*, **25**, 327 (1984); J. R. Kanofsky, *ibid.*, **25**, 105 (1984); J. R. Kanofsky, B. Axelrod, *J. Biol. Chem.*, **261**, 1099 (1986); A. U. Khan, *J. Biolumin. Chemilumin.*, **4**, 200 (1989); A. A. Krasnovsky, Jr., "Light in Biology and Medicine," Vol. 2, ed. by R. H. Douglas, J. Moan, G. Ronto, Plenum Press, New York, 1991, pp. 437—452; A. Baker, J. R. Kanofsky, *Arch. Biochem. Biophys.*, **286**, 70 (1991).
- 7) R. R. Hung, J. J. Grabowski, *J. Phys. Chem.*, **95**, 6073 (1991).
- 8) M. Terazima, N. Hirota, H. Shinohara, Y. Saito, *J. Phys. Chem.*, **95**, 9080 (1991).
- 9) J. W. Arbogast, A. P. Darmanyan, C. S. Foote, Y. Rubin, F. N. Diederich, M. M. Alvarez, S. J. Anz, R. L. Whetten, *J. Phys. Chem.*, **95**, 11 (1991).
- 10) A. A. Krasnovsky, Jr, C. S. Foote, *J. Am. Chem. Soc.*, **115**, 6013 (1993).
- 11) O. Stern, M. Volmer, *Phys. Z.*, **20**, 183 (1919).
- 12) P. B. Merkel, D. R. Kearns, *J. Am. Chem. Soc.*, **94**, 1029 (1972).
- 13) P. B. Merkel, D. R. Kearns, *J. Am. Chem. Soc.*, **94**, 7244 (1972).
- 14) H. Tokuyama, S. Yamago, E. Nakamura, T. Shiraki, Y. Sugiura, *J. Am. Chem. Soc.*, **115**, 7918 (1993).