Pyridonecarboxylic Acids as Antibacterial Agents. IX.^{1a)} Synthesis and Structure—Activity Relationship of 3-Substituted 10-(1-Aminocyclopropyl)-9-fluoro-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylic Acids and Their 1-Thio and 1-Aza Analogues²⁾

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A series of the title compounds listed in Chart 1 have been synthesized to study the effects of 3-alkyl substituents on the antibacterial potency and *in vivo* efficacy of 10-(1-aminocyclopropyl)-9-fluoro-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-6-carboxylic acid and its 1-thio and 1-aza variants. Compound (S)-1, which proved most active in vitro against five representative gram-positive and gram-negative organisms, was assayed *in vivo* using Staphylococcus aureus and Pseudomonas aeruginosa mouse infection models. It exhibited an excellent *in vivo* efficacy, being superior to ofloxacin and ciprofloxacin, and was then assayed for convulsion-inducing activity, mammalian cell cytotoxicity, and topoisomerase II inhibition. The biological results showed that (S)-1 displayed antibacterial and toxicological advantages over ofloxacin and ciprofloxacin. Compound (S)-1 and its methanesulfonate showed high serum concentrations after oral and intravenous administrations to mice.

Keywords antibacterial agent; fluoroquinolone; structure-activity relationship; aminocyclopropyl substituent; toxicological property; pharmacokinetic property

In the preceding paper, ^{1a)} we disclosed that compound 1, a 10-(1-aminocyclopropyl) analogue of ofloxacin (OFLX),3) exhibits potent in vitro antibacterial activity against both gram-positive and gram-negative bacteria. comparable to that of OFLX or ciprofloxacin (CPFX),4) and we showed that 1 is considerably less toxic than OFLX and CPFX in terms of acute toxicity, cytotoxicity, and convulsion-inducing activity. This encouraging discovery prompted us to synthesize both enantiomers of 1 and a series of its 3-alkyl variants (2a-g), as well as 1-thio and 1-aza analogues of 1 (2h, i) (Chart 1). This paper describes the synthesis of these new quinolones and the results of their biological evaluation. Among the compounds synthe sized in this study, the (S) enantiomer of 1 has been found to be the best compound, being superior to OFLX and CPFX, in terms of in vivo antibacterial activity and

toxicological and pharmacokinetic properties.

Chemistry

All of the designed molecules were synthesized from the N-benzyloxycarbonyl (Z) derivative 3^{1a} of ethyl 4-(1-aminocyclopropyl)-2,3,5-trifluorobenzoylacetate utilizing the pyridone annulation methodology⁵ reported in the synthesis of OFLX and its analogues (Chart 2).

Reaction of the β -keto ester 3 with N,N-dimethylform-amide dimethyl acetal followed by treatment of the resulting (α -dimethylamino)methylene derivative with 2-aminoalcohols (4a—h) afforded the corresponding enamines 5a—h. These compounds were subjected to a double cyclization by heating in N,N-dimethylformamide (DMF) in the presence of potassium carbonate to give pyridobenzoxazines 6a—h6) in 61—89% overall yield,

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Chart 3

except for the fluoromethyl compound 6e (15%). These tricyclic compounds were converted to 1 and 2a—f by ester hydrolysis with ethanolic NaOH and subsequent removal of the Z-protecting group by catalytic hydrogenolysis, except for base-sensitive 6e, where acidic conditions were used for this step: treatment with HBr in acetic acid for N-deprotection and ester hydrolysis by heating with 6 n HCl. The 3-methylene-1,4-oxazine compound 2g was obtained from the alcohol 6f by treatment of its O-methanesulfonate with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), then subjecting the resultant olefin 7 to removal of O- and N-protecting

The 1,4-thiazine and 1,4-diazine analogues **2h** and **2i** were prepared from **5a** *via* the common intermediate, *N*-1-[(1-bromomethyl)ethyl]quinolone **9** obtained by bromi-

groups.

nation of the pyridone annulation product 8 (26% overall yield) (Chart 3). To obtain 2h, the bromide 9 was first subjected to a thiazine-ring formation by treatment with potassium ethyl xanthate in DMF, then the remaining ester and Z-amide groups were hydrolyzed under basic and acidic conditions, respectively, to give 2h in 54% overall yield. On the other hand, 2i was derived from 9 by carrying out a three-step sequence of reactions: acid-catalyzed ester hydrolysis to 10; cyclization to 12 with methylamine; and catalytic hydrogenolysis of the Z-group (32% overall yield). Characterization data for the new quinolones obtained above are recorded in Table I.

Biological Assays and Discussion

In Vitro and in Vivo Antibacterial Activity All of the tricyclic (1-aminocyclopropyl)quinolones (Chart 1) syn-

Table I. Physical Data for 1,4-Oxazino-, 1,4-Thiazino-, and 1,4-Diazinoquinolones

Compd. mp (dec. °C)		Recryst.	Yield ^{a)} (%)	Formula	Analysis (%) Calcd (Found)		IR (KBr)	1 H-NMR (CF $_{3}$ COOD) δ (ppm)	
			(74)		C	Н	N	CIII	
(R) -1 $^{b)}$	269—273	CHCl ₃ -MeOH	74	C ₁₆ H ₁₅ FN ₂ O ₄	60.38	4.75	8.80	1711	1.3—2.3 (7H, m), 4.6—5.5 (3H, m), 8.05 (1H,
$(S)-1^{c)}$	269—272	CHCl ₃ -MeOH	63	$C_{16}H_{15}FN_{2}O_{4}$	(60.40 60.38	4.82 4.75	8.86) 8.80	1711	d, $J = 9.5 \text{ Hz}$), 9.43 (1H, s) 1.3—2.2 (7H, m), 4.5—5.6 (3H, m), 8.06 (1H,
2a	261—265	CHCl ₃ -EtOH	53	$C_{17}H_{17}FN_2O_4$	(60.30 61.44	4.67 5.16	9.06) 8.43	1713	d, $J=9.5$ Hz), 9.42 (1H, s)
2b		J		 	(61.42	5.22	8.49)		0.9—2.4 (9H, m), 4.5—5.3 (3H, m), 8.05 (1H, d, <i>J</i> =9.5 Hz), 9.37 (1H, s)
20	223—226	EIOH	67	$C_{18}H_{19}FN_2O_4$ ·1/2H ₂ O	60.84 (60.62	5.67 5.54	7.88 7.92)	1718	0.9-2.4(11H, m), 4.7-5.3(3H, m), 8.05(1H, d, J=9.5 Hz), 9.36(1H, s)
2c	240245	6 n HCl–EtOH	12	C ₁₆ H ₁₄ F ₂ N ₂ O ₄ ·HCl·3/4H ₂ O		4.31	7.25	1718	1.3—2.3 (4H, m), 4.4—5.8 (5H, m), 8.08 (1H,
2 d	255260	6 N HCl–EtOH	23	$C_{16}H_{15}FN_2O_5$	50.60	4.25 4.51	7.24) 7.38	1711	d, J=9.5 Hz), 9.39 (1H, s) 1.0—1.8 (4H, m), 4.3—5.2 (5H, m), 7.64 (1H,
2 e	269—272	CHCl ₃ -EtOH	67	\cdot HCl·1/2H ₂ O C ₁₇ H ₁₇ FN ₂ O ₄	(50.55 60.62	4.30 5.24	7.45) 8.32		d, J=10 Hz), 8.84 (1H, s) 1.3—2.3 (10H, m), 4.71 (2H, s), 8.08 (1H, d,
2f	255—260	CHCl ₃ –EtOH	49	· 1/4H ₂ O C ₁₇ H ₁₅ FN ₂ O ₄	(60.90 60.17	5.21 4.75	8.56) 8.26		J = 10 Hz), 9.48 (1H, s)
2-				$\cdot 1/2 H_2 O$	(60.44	4.51	8.19)		1.2—2.4 (8H, m), 4.81 (2H, s), 8.06 (1H, d, J=9Hz), 9.07 (1H, s)
2 g	275—280	EtOH-MeOH	19	$C_{16}H_{13}FN_2O_4$ ·HBr	48.38 (48.37	3.55 3.59	7.05 7.05)	1715	1.3—2.3 (4H, m), 5.34 (2H, s), 5.95 (1H, d, J=4.5 Hz), 6.28 (1H, d, J=4.5 Hz), 8.04 (1H,
2h	276—280	FtOH.	14	C H ENOS	`		Í		d, $J=9.5$ Hz), 9.55 (1H, s)
			14	$C_{16}H_{15}FN_2O_3S$	46.28 (46.24	3.88 4.00	6.75 6.63)	1701	1.4—2.4 (7H, m), 3.2—4.3 (2H, m), 5.2—5.8 (1H, m), 8.24 (1H, d, <i>J</i> =9.5 Hz), 9.51 (1H, s)
2i	256—260	6 n HCl–EtOH	8	C ₁₇ H ₁₈ FN ₃ O ₃ ·HCl	55.52 (55.47	5.21 5.44	11.42 11.28)	1687	1.2—2.4 (7H, m), 3.3—4.2 (5H, m), 4.8—5.4 (1H, m), 7.89 (1H, d, $J = 10$ Hz), 9.29 (1H, s)

a) Overall yield from 3. b) $[\alpha]_D^{25} + 80.3^{\circ} (c = 1.0, 1.0 \text{ N NaOH})$. c) $[\alpha]_D^{25} - 80.0^{\circ} (c = 1.0, 1.0 \text{ N NaOH})$.

thesized in this study and the two reference drugs (OFLX, CPFX) were tested for in vitro antibacterial activity against five selected gram-positive and gram-negative bacteria. Their minimum inhibitory concentrations (MICs, µg/ml) determined by conventional agar dilution procedures 1c,7) are recorded in Table II. Of the three 3-methyl compounds (1, 2h, 2i; all racemates) which differ only in the heteroatom at the 1-position, the oxazine compound 1 is evidently more potent than the other two, the activity order being 1>2h>2i. Among the 3-substituted 1,4-oxazinoquinolones other than 1, the 3-fluoromethyl compound 2c displayed the best antibacterial profile, but its MICs against the tested organisms are equal to those of 2h. The best overall antimicrobial activity (equal to CPFX) was obtained with (S)-1, which was more active than (R,S)-1against Staphylococcus aureus (two-fold MIC). It is interesting that (R)-1 is much less active than (S)-1, particularly against S. aureus and P. aeruginosa.

Compound (S)-1, which displayed excellent *in vitro* antibacterial activity, was tested for *in vivo* efficacy against acute, lethal systemic infections due to gram-positive Staphylococcus aureus and gram-negative Pseudomonas aeruginosa in male ICR-strain mice. The effective dose (ED₅₀, mg/mouse) for (S)-1 and those of the reference drugs (OFLX, CPFX) are listed in Table III, in which MICs against the corresponding organisms are included for reference. Comparison of the oral (p.o.) efficacy data indicates that (S)-1 is 1.5—2.5 times more active orally than the reference quinolones in the S. aureus and P. aeruginosa infection models. On subcutaneous (s.c.) administration, on the other hand, (S)-1 is 2.1—3.9 times more active than OFLX, but shows the same order of activity as CPFX. A noteworthy difference between *in vitro*

TABLE II. In Vitro Antibacterial Activity (MICs, μg/ml)^{a)}

	Microorganism ^{b)}							
Compd.	Gram-positive	Gram-negative						
	S. au.	E. c.	K. pn.	Pr. v.	P. ae			
(R)-1	50	1.56	1.56	1.56	100			
(S)-1	0.2	≤ 0.05	≤ 0.05	≤0.05	0.39			
(R,S)-1	0.39	≤ 0.05	≤0.05	≤0.05	0.39			
2a	1.56	≤ 0.05	≤0.05	≤0.05	0.78			
2b	6.25	0.1	0.2	0.1	3.13			
2c	0.78	≤ 0.05	≤ 0.05	≤0.05	1.56			
2d	12.5	0.39	0.39	0.39	12.5			
2e	3.13	0.1	0.1	0.2	12.5			
2f	3.13	0.1	0.2	0.2	3.13			
2 g	3.13	0.1	0.2	0.2	3.13			
2h	0.78	≤ 0.05	≤0.05	≤0.05	1.56			
2i	6.25	0.2	0.39	0.39	12.5			
OFLX	0.39	≤ 0.05	≤0.05	≤0.05	0.78			
CPFX	0.2	≤0.05	_ ≤0.05	< 0.05	0.39			

a) Inoculation was performed with one loopful of 10° cells/ml. b) S. au. = Staphylococcus aureus FDA 209P, E. c. = Escherichia coli NIHJ, K. pn. = Klebsiella pneumoniae Y-50, Pr. v. = Proteus vulgaris GN 3027, P. ae. = Pseudomonas aeruginosa IFO 3445.

and in vivo activity is seen in the oral efficacy data with P. aeruginosa: while (S)-1 is less active than CPFX in vitro, its in vivo activity is 1.6 times superior to that of CPFX.

Toxicological Properties of (S)-1 Compound (S)-1, which showed an excellent antibacterial activity, was evaluated for acute toxicity 1b and convulsion-inducing activity 1c,8 in male ICR-strain mice by intravenous and intracerebral administrations, respectively. The dosemortality ratios data are recorded in Table IV, together

TABLE III. In Vivo Efficacy against Systemic Infections in Mice^{a)}

	-	MIC (μg/ml)	ED ₅₀ mg/mouse (95% confidence limits)					
Organism	Compd.			p.o.		s.c.		
S. aureus Smith ^{b)}	(S)-1	0.20	0.056	(0.0330.095)	0.021	(0.0150.030)		
9,	OFLX	0.39	0.11	(0.05-0.23)	0.044	(0.029 - 0.067)		
	CPFX	0.39	0.14	(0.08-0.26)	0.026	(0.020 - 0.034)		
P. aeruginosa	(S)-1	0.39	0.16	(0.11 - 0.23)		(0.0350.089)		
S-68°)	OFLX	0.78	0.39	(0.26-0.58)		(0.150.33)		
	CPFX	0.20	0.26	(0.230.29)	0.044	(0.0310.063)		

a) Male ICR-strain mice (18—21 g body weight). b) Challenge dose (for oral dosing, 2.2×10^7 CFU/mouse; for subcutaneous dosing, 2.4×10^7 CFU/mouse). c) Challenge dose (for oral dosing, 4.9×10^6 CFU/mouse; for subcutaneous dosing, 1.6×10^7 CFU/mouse). CFU, colony forming unit.

TABLE IV. Acute Toxicity and Convulsion Induction in Male ICR-Strain Mice

	Acute tox	icity (i.v.) ^{a)}	Convulsion induction (i.c.) ^{b)}			
Compd.	Dose (mg/kg)	Mortality (after 7 d)	Clonic seizure	Tonic seizure	Mortality (after 24 h)	
(S)-1	500	0/5	0/8 10/10	0/8 8/10	0/8 8/10	
OFLX CPFX	250 250	3/5 1/5	10/10	10/10	10/10	

a) Intravenous dose to 18—24 g body weight mice. b) Intracerebral dose of $50\,\mu g$ to 20—25 g body weight mice.

Table V. Assays for Chinese Hamster V79 Cell Cytotoxicity and for Inhibition of DNA Gyrase and Topoisomerase II

_	$IC_{50} (\mu g/ml)$					
Compd.	V-79 cell	DNA gyrase ^{a)}	Topoisomerase II ^{b)}			
(S)-1	295	0.68	2010			
OFLX	120	0.70	1358			
CPFX	70	0.35	236			

a) Isolated and purified from E. coli KL-16.
 b) Isolated and purified from calf thymus.

with the data obtained with two reference drugs. The acute toxicity data indicate that (S)-1 is much less toxic ($LD_{50} = > 500 \,\mathrm{mg/kg}$) than OFLX and CPFX. Furthermore, convulsion-inducing action of (S)-1 proved to be significantly lower than that of the reference drugs in both clonic and tonic seizures, 100% survival being observed at the 50 μ g dose level.

The clonogenic cytotoxicity of (S)-1 was determined in Chinese hamster V-79 cells according to the method reported in the preceding paper, ^{1a)} and the 50%-inhibitory concentrations (IC₅₀, μ g/ml) obtained are recorded in Table V. The data reveal that (S)-1 is significantly less toxic than the two reference drugs against these mammalian cells.

Inhibitory activity against DNA gyrase and mammalian topoisomerase II was determined with the enzymes isolated and purified from *Escherichia coli* KL-16 and calf thymus cells, respectively, according to the procedures of Hoshino and his co-workers.⁹⁾ The IC₅₀ data included in Table V indicate that the DNA gyrase supercoiling inhibition activity of (S)-1 is less potent than that of CPFX but is at the same level as that of OFLX. An important finding

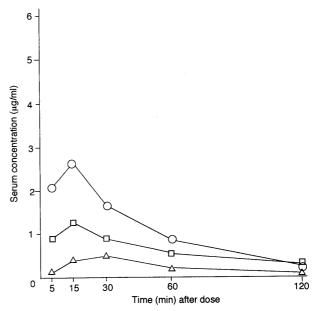


Fig. 1. Mean Serum Concentrations after Oral Administration of 5 mg/kg to Mice (n=5)

 \bigcirc , (S)-1; \square , OFLX; \triangle , CPFX.

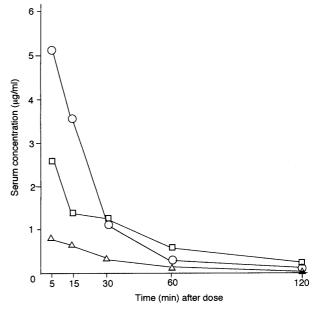


Fig. 2. Mean Serum Concentrations after Intravenous Administration of 5 mg/kg to Mice (n=5)

 \bigcirc , methanesulfonate of (S)-1; \square , OFLX; \triangle , CPFX.

is that the inhibitory activity of (S)-1 against mammalian topoisomerase II is 1.5 or 8.5 times weaker than that of OFLX or CPFX, respectively. Thus the selectivity index (the value obtained by division of IC₅₀ for topoisomerase II by IC₅₀ for DNA gyrase) is the best for (S)-1 (2956), being 1.5 or 4.4 times greater than those of OFLX (1940) and CPFX (674), respectively.

Pharmacokinetic Properties of (S)-1 and Its Methanesulfonate Serum concentrations of (S)-1 as well as of its methanesulfonate, which has a good water solubility (>200 mg/ml at 25 °C), were determined by oral and intravenous administration to mice. The results for oral administration of (S)-1 and the reference agents (Fig. 1) indicate that the maximum serum concentration ($C_{\rm max}$) of (S)-1 (obtained 15 min after dosing) is 2.1 and 5.5 times those of OFLX and CPFX, respectively. The half-life time of (S)-1 (30 min) was, however, about 50% shorter than those of the reference agents, and approximately 27% of the administered drug was recovered from urine over 24 h. On intravenous administration of (S)-1 methanesulfonate, on the other hand, the serum concentration five minutes after dosing (5.12 μ g/ml) was 2.0 and 6.3 times higher than those of OFLX and CPFX, respectively, (Fig. 2) and approximately 42% was recovered from the urine over 24 h.

In conclusion, we have obtained a new, highly potent quinolone antibacterial agent (S)-1 (T-3761) simply by replacing the 10-piperazinyl substituent of OFLX with the hitherto unexplored 1-aminocyclopropyl group. Our new drug is safer than most existing quinolones, yet has a broad antibacterial spectrum. Clinical trials of both T-3761 and its methanesulfonate (T-3762) as oral and intravenous drugs, respectively, are in progress.

$Experimental^{10)}\\$

Ethyl 3-Alkyl-10-[(1-benzyloxycarbonylamino)cyclopropyl]-9-fluoro-3-methyl-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylates (6a—h) These compounds were prepared from 3 by the procedure reported in the previous paper. ^{1a})

6a: 89% yield. mp 183—185 °C (colorless prisms from AcOEt). $[\alpha]_D^{25}$ + 57.3° (c=1.0, CHCl₃). IR (KBr): 1722, 1696, 1677 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.1—1.8 (10H, m), 4.1—4.7 (5H, m), 5.02 (2H, s), 5.60 (1H, s), 7.27 (5H, s), 7.64 (1H, d, J=10.5 Hz), 8.31 (1H, s). *Anal.* Calcd for C₂₆H₂₅FN₂O₆: C, 64.99; H, 5.24; N, 5.83. Found: C, 64.85; H, 5.33; N, 5.91.

6b: 83% yield. mp 182—184 °C (colorless prisms from AcOEt). $[\alpha]_D^{25}$ -58.1° (c=1.0, CHCl₃). IR (KBr): 1722, 1695, 1677 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.2—1.8 (10H, m), 4.2—4.6 (5H, m), 5.03 (2H, s), 5.61 (1H, s), 7.27 (5H, s), 7.62 (1H, d, J=10.5 Hz), 8.30 (1H, s). *Anal.* Calcd for C₂₆H₂₅FN₂O₆: C, 64.99; H, 5.24; N, 5.83. Found: C, 65.06; H, 5.22; N, 5.67.

6c: 72% yield. mp 107—109 °C (colorless prisms from aqueous EtOH). IR (KBr): 1717 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.7—2.1 (12H, m), 3.8—4.7 (5H, m), 5.02 (2H, s), 5.60 (1H, s), 7.27 (5H, s), 7.62 (1H, d, J=10.5 Hz), 8.26 (1H, s). *Anal*. Calcd for C₂₇H₂₇FN₂O₆·1/2H₂O: C, 64.41; H, 5.61; N, 5.56. Found: C, 64.38; H, 5.89; N, 5.59.

6d: 89% yield. mp 148—149 °C (colorless prisms from aqueous EtOH). IR (KBr): 1716 cm⁻¹. ¹H-NMR (CDCl₃) δ : 0.7—2.1 (14H, m), 3.9—4.8 (5H, m), 5.02 (2H, s), 5.60 (1H, s), 7.27 (5H, s), 7.65 (1H, d, J=10.5 Hz), 8.26 (1H, s). *Anal*. Calcd for C₂₈H₂₉FN₂O₆: C, 66.13; H, 5.75; N, 5.51. Found: C, 66.20; H, 5.81; N, 5.60.

6e: 15% yield. mp 192—194 °C (colorless prisms from aqueous EtOH). IR (KBr): 1724, 1687 cm⁻¹. ¹H-NMR (CDCl₃) δ: 1.0—1.7 (7H, m), 4.2—5.3 (9H, m), 5.57 (1H, s), 7.27 (5H, s), 7.59 (1H, d, J=10.5 Hz), 8.27 (1H, s). *Anal*. Calcd for C₂₆H₂₄F₂N₂O₆: C, 62.65; H, 4.85; N, 5.62. Found: C, 62.61; H, 4.91; N, 5.69.

6f: 61% yield. mp 229—231°C (colorless needles from EtOH). IR (KBr): 1732, 1715 cm $^{-1}$. ¹H-NMR (CDCl₃) δ : 1.0—1.8 (7H, m), 3.8—4.8 (7H, m), 5.02 (2H, s), 5.59 (1H, s), 6.00 (1H, br s), 6.43 (1H, d, J=10.5 Hz), 7.26 (5H, s), 8.43 (1H, s). *Anal.* Calcd for $C_{26}H_{25}FN_{2}O_{7}$ ·1/4H₂O: C, 62.33; H, 5.13; N, 5.59. Found: C, 62.43; H, 5.35; N, 5.88.

6g: 81% yield. mp 194—196 °C (colorless needles from aqueous EtOH). IR (KBr): 1719 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.1—1.8 (13H, m), 4.14 (2H, s), 4.41 (2H, q, J=7.5 Hz), 5.03 (2H, s), 5.61 (1H, s), 7.27 (5H, s), 7.72 (1H, d, J=11 Hz), 8.56 (1H, s). *Anal.* Calcd for $C_{27}H_{27}FN_2O_6 \cdot 1/2H_2O$: C, 64.41; H, 5.61; N, 5.56. Found: C, 64.33; H, 5.88; N, 5.63.

6h: 64% yield. mp 175—177 °C (a white powder from aqueous EtOH). IR (KBr): 1722, 1694, 1683 cm $^{-1}$. 1 H-NMR (CDCl $_{3}$) δ : 1.0—1.8 (11H, m), 4.1—4.6 (4H, m), 5.03 (2H, s), 5.62 (1H, s), 7.29 (5H, s), 7.69 (1H, d, J=10.5 Hz), 8.14 (1H, s). *Anal*. Calcd for $C_{27}H_{25}FN_{2}O_{6} \cdot 1/4H_{2}O$: C, 65.25; H, 5.17; N, 5.64. Found: C, 65.48; H, 5.19; N, 5.74.

(S)-10-(1-Aminocyclopropyl)-9-fluoro-3-methyl-7-oxo-2,3-dihydro-7Hpyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylic Acid ((S)-1) A solution of the ethyl ester 6b (7.00 g, 14.6 mmol) in a mixture of EtOH (70 ml) and dioxane (70 ml) was stirred at room temperature for 2 h after addition of 1 N NaOH (70 ml). The mixture was poured into a two-layer mixture of H₂O (200 ml) and CHCl₃ (200 ml), and the whole was acidified to pH 2 by addition of 6 N HCl. The aqueous layer was separated, and the organic layer was washed successively with water and saturated brine, dried, and concentrated. The solid residue was washed with Et₂O to give the corresponding carboxylic acid (6.34 g, 96%). An analytical sample was obtained by recrystallization from EtOH as colorless prisms, mp 181—183 °C. $[\alpha]_D^{25}$ –45.8° (c=1.0, CHCl₃). IR (KBr): 1731, 1715 cm⁻ ¹H-NMR (CDCl₃) δ : 1.0—1.9 (7H, m), 4.1—4.7 (3H, m), 5.01 (2H, s), 5.63 (1H, s), 7.28 (5H, s), 7.70 (1H, d, $J=10\,\text{Hz}$), 8.66 (1H, s), 14.62 (1H, brs). Anal. Calcd for C₂₄H₂₁FN₂O₆: C, 63.71; H, 4.68; N, 6.19. Found: C, 63.79; H, 4.67; N, 6.04.

A solution of this material (6.30 g, 13.9 mmol) in AcOH (126 ml) was stirred under atmospheric pressure of hydrogen after addition of 5% Pd–C (1.26 g). After 2 h, the catalyst was filtered off, and the filtrate was concentrated under reduced pressure. The resulting amino compound was isolated as its hydrochloride by concentration of a solution in 6 N HCl (21 ml) followed by crystallization of the residue from EtOH. This material was dissolved in 40% aqueous EtOH (100 ml) containing KOH (1.80 g, 32.1 mmol), and the solution was saturated with CO₂. The resulting precipitate was collected by filtration to give (S)-1 (3.85 g, 87%).

By subjecting 6a, c, d, f—h to the same two-step deprotection procedure, (R)-1 and 2a, b, d—f were obtained. Base-sensitive 2c was obtained from 6e by successive treatment with 30% HBr-AcOH at room temperature for 1 h and with 6 N HCl at reflux temperature.

10-(1-Aminocyclopropyl)-9-fluoro-3-methylene-7-oxo-2,3-dihydro-7Hpyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylic Acid (2g) Methanesulfonyl chloride (260 mg, 2.27 mmol) was added dropwise to a stirred suspension of 6f (380 mg, 0.77 mmol) and triethylamine (230 mg, 2.27 mmol) in CH₂Cl₂ (4 ml) at 5 to 10 °C. After the addition was completed, the mixture was stirred at room temperature for 30 min, then poured into a two-layer mixture of ice-water (10 ml) and CH₂Cl₂ (10 ml), and the whole was acidified to pH 1 with 2 N HCl. The organic layer was separated, washed successively with water and saturated brine, dried, and concentrated. The resultant crude O-mesylate was dissolved in benzene (4.4 ml) containing DBU (175 mg, 1.15 mmol), and the solution was heated under reflux for 1 h. The reaction mixture was cooled to room temperature, and poured into a two-layer mixture of H₂O (20 ml) and AcOEt (20 ml), and the whole was acidified to pH 1 with 2 N HCl. The organic layer was washed successively with water and saturated brine, dried, and concentrated. The residue was purified by chromatography (silica gel 10 g, toluene: AcOEt = 3:2) to give 7 (260 mg, 71%). An analytical sample was obtained by recrystallization from EtOH as colorless prisms, mp 195—197 °C. IR (KBr): 1724, 1700, 1687 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.2—1.7 (7H, m), 4.41 (2H, q, J=7.5 Hz), 4.78 (2H, s), 5.02 (2H, s), 5.12 (1H, d, J=2.5 Hz), 5.45 (1H, d, J=2.5 Hz), 5.60 (1H, s), 7.27 (5H, s), 7.66 (1H, d, J = 10.5 Hz), 8.64 (1H, s). Anal. Calcd for C₂₆H₂₃FN₂O₆: C, 65.27; H, 4.85; N, 5.85. Found: C, 65.24; H, 4.89; N, 5.93.

The ester 7 (220 mg, 0.46 mmol) was saponified by the same procedure as described for **6b**, and the resultant carboxylic acid was treated with 30% HBr–AcOH (8 ml) at 5 to 10 °C for 1 h. The whole was concentrated under reduced pressure, and the residue was treated with Et₂O to induce crystallization. The crude hydrobromide of **2g** obtained by filtration was recrystallized from MeOH–EtOH.

Ethyl 7-[(1-Benzyloxycarbonylamino)cyclopropyl]-1-[(2-bromo-1-methyl)ethyl]-6,8-difluoro-1,4-dihydro-4-oxoquinoline-3-carboxylate (9) A solution of (R,S)-5a $(5.60\,\mathrm{g},\ 10.8\,\mathrm{mmol})$ in DMF $(56\,\mathrm{ml})$ was stirred at $70-80\,^\circ\mathrm{C}$ for 12 h after addition of NaHCO $_3$ $(0.99\,\mathrm{g},\ 11.8\,\mathrm{mmol})$. The mixture was then poured into a two-layer mixture of ice-water $(200\,\mathrm{ml})$ and AcOEt $(150\,\mathrm{ml})$, and the whole was adjusted to pH 2 with $6\,\mathrm{N}$ HCl. The organic layer was separated, washed successively with water and saturated brine, dried, and concentrated. The residue was subjected to chromatography (silica gel $100\,\mathrm{g}$, toluene: AcOEt=1:3) to give 8 $(1.73\,\mathrm{g},\ 32\%)$. An analytical sample was obtained by recrystallization from AcOEt as colorless prisms, mp $157-159\,^\circ\mathrm{C}$. IR (KBr): $1725\,\mathrm{cm}^{-1}$. 14-NMR (CDCl $_3$) δ : 1.0-1.9 $(104,\,\mathrm{m})$, 3.6-4.6 $(54,\,\mathrm{m})$, 5.04 $(24,\,\mathrm{s})$, 5.2-5.7 $(24,\,\mathrm{m})$, 7.0-7.5 $(64,\,\mathrm{m})$, 8.64 $(14,\,\mathrm{s})$. Anal. Calcd for $C_{26}H_{26}F_{2}N_{2}O_{6}$: C, 62.40; H, 5.24; N, 5.60. Found: C, 62.50; H, 5.28; N, 5.65.

Carbon tetrabromide (500 mg, 1.51 mmol) and triphenylphosphine (390 mg, 1.49 mmol) were added to a stirred suspension of **8** (500 mg, 1.00 mmol) in CH₂Cl₂ (5 ml). After being stirred at room temperature for 5 h, the mixture was concentrated under reduced pressure, and the residue was subjected to chromatography (silica gel 20 g, toluene: AcOEt=1:1) to give **9** (460 mg, 82%). An analytical sample was obtained by recrystallization from iso-Pr₂O–AcOEt as colorless needles, mp 148—149 °C. IR (KBr): 1730, 1692 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.1—2.0 (10H, m), 3.68 (2H, d, J=4 Hz), 4.40 (2H, q, J=7 Hz), 5.04 (2H, s), 5.2—5.8 (2H, m), 7.28 (5H, s), 8.02 (1H, dd, J=10, 2 Hz), 8.55 (1H, s). *Anal.* Calcd for C₂₆H₂₅BrF₂N₂O₅: C, 55.43; H, 4.47; N, 4.97. Found: C, 55.41; H, 4.54; N, 5.00.

10-(1-Aminocyclopropyl)-9-fluoro-3-methyl-7-oxo-2,3-dihydro-7*H***-pyrido[1,2,3-***de***]-1,4-benzothiazine-6-carboxylic Acid (2h)** Potassium ethyl xanthate (290 mg, 1.81 mmol) was added to a stirred solution of **9** (250 mg, 0.44 mmol) in DMF (5 ml) at room temperature. After 12 h, the reaction mixture was poured into a two-layer mixture of ice-water (40 ml) and AcOEt (30 ml), and the whole was acidified to pH 2 with 2 N HCl. The organic layer was separated, washed successively with water and saturated brine, dried, and concentrated. The residue was purified by chromatography (silica gel 5 g, CHCl₃: EtOH = 5:1) to give **11** (120 mg, 54%). An analytical sample was obtained by recrystallization from EtOH as pale yellow needles, mp 209—211 °C. IR (KBr): 1719, 1695, 1679 cm⁻¹.
¹H-NMR (CDCl₃) δ: 1.0—1.8 (10H, m), 2.7—3.7 (2H, m), 4.1—4.8 (3H, m), 5.05 (2H, s), 5.81 (1H, s), 7.29 (5H, s), 7.90 (1H, d, *J* = 10 Hz), 8.37 (1H, s). *Anal.* Calcd for C₂₆H₂₅FN₂O₅S: C, 62.89; H, 5.07; N, 5.64. Found: C, 62.52; H, 5.05; N, 5.76.

The hydrobromide of 2h was obtained from 11 by two-step deprotection as performed with 7.

10-(1-Aminocyclopropyl)-9-fluoro-1,3-dimethyl-7-oxo-2,3-dihydro-1H,7H-pyrido[1,2,3-de]quinoxaline-6-carboxylic Acid (2i) The ester 9 (400 mg, 0.71 mmol) was added to a mixture of dioxane (4 ml) and 6 N HCl (4 ml), and the mixture was heated under reflux for 30 min. The reaction mixture was poured into a two-layer mixture of H_2O (50 ml) and CHCl₃ (50 ml), and the organic layer was separated, washed successively with water and saturated brine, dried, and concentrated. The solid residue was treated with Et_2O and collected by filtration to give 10 (180 mg, 47%). An analytical sample was obtained by recrystallization from iso-Pr₂O-AcOEt as colorless prisms, mp 167—169 °C. IR (KBr): 1737 cm⁻¹. 1 H-NMR (CDCl₃) δ : 1.0—2.0 (7H, m), 3.70 (2H, d, J=4.5 Hz), 5.03 (2H, s), 5.2—5.8 (2H, m), 7.28 (5H, s), 8.03 (1H, dd, J=9.5, 2 Hz), 8.77 (1H, s), 14.13 (1H, br s). *Anal*. Calcd for $C_{24}H_{21}BrF_{2}N_{2}O_{5}$: C, 53.85; H, 3.95; N, 5.23. Found: C, 54.02; H, 4.00: N, 5.45.

A solution of **10** (150 mg, 0.28 mmol) in 30% MeNH₂–EtOH (7.5 ml) was stirred at 45—55 °C for 12 h, then concentrated under reduced pressure. The residue was taken up in H₂O (50 ml) and CHCl₃ (30 ml), the whole was acidified to pH 2 with 6 n HCl, and the aqueous layer was separated. The organic layer was washed successively with water and saturated brine, dried, and concentrated. The solid residue was treated with Et₂O and collected by filtration to give **12** (90 mg, 69%). An analytical sample was obtained by recrystallization from EtOH as pale yellow needles, mp 218—220 °C. IR (KBr): 1711 cm⁻¹. ¹H-NMR (CDCl₃) δ : 1.1—1.8 (7H, m), 3.2—3.7 (5H, m), 4.2—4.6 (1H, m), 5.03 (2H, s), 5.80 (1H, s), 7.31 (5H, s), 7.70 (1H, d, J=11 Hz), 8.66 (1H, s), 14.75 (1H, br s). *Anal*. Calcd for C₂₅H₂₄FN₃O₅: C, 64.51; H, 5.20; N, 9.03. Found: C, 64.60; H, 5.16; N, 9.11.

Catalytic hydrogenolysis of 12 by the same procedure as described for 6b afforded 2i.

Methanesulfonate of (S)-1 Methanesulfonic acid (0.31 g, 3.22 mmol) was added to a stirred suspension of (S)-1 (1.00 g, 3.14 mmol) in EtOH (10 ml) at 50 °C, then the mixture was allowed to cool to room temperature. Precipitates were collected by filtration to give the methanesulfonate of (S)-1 (1.22 g, 94%), mp 258—259 °C (dec.), colorless prisms. [α]_D²⁰ -64.2° (c=1.0, 1.0 N NaOH). IR (KBr): 1704 cm⁻¹. ¹H-NMR (CF₃COOD) δ: 1.3—2.2 (7H, m), 3.09 (1H, s), 4.6—5.5 (3H,

m), 8.05 (1H, d, $J=9.5\,\text{Hz}$), 9.43 (1H, s). Anal. Calcd for $C_{16}H_{15}FN_2O_4\cdot CH_4O_3S$: C, 49.27; H, 4.62; N, 6.76. Found: C, 49.32; H, 4.66; N, 6.75.

In Vivo Efficacy The in vivo assay was carried out according to the method reported in the previous paper. $^{11)}$ Male ICR mice (18—21 g body weight, 10 mice per group) were used in this assay. Each mouse received 0.5-ml volume of bacterial suspension in 5% gastric mucin intraperitioneally. Test compounds, which were suspended in 0.5% aqueous methylcellulose or dissolved in dilute NaOH (ca. 1 eq), were administered orally or subcutaneously at 1 h after infection. Fifty percent effective dose values (ED₅₀) were calculated from the cumulative mortalities on the seventh day after infection.

Determination of Serum Concentrations and Urinary Recovery Male ICR mice (18—21 g body weight, 5 mice per group) which had been starved for 18 h were used in this assay. For the oral dosing test, each sample (10 mg) was suspended in 20 ml of 0.5% aqueous methylcellulose. For the intravenous dosing test, each sample (10 mg) was dissolved in 20 ml of water or in 20 ml of dilute NaOH (ca. 1 eq). These preparations (corresponding to 5 mg/kg) were administered orally or intravenously to mice. Serum samples were obtained at 5, 15, 30, 60, 120, and 240 min after dosing. Urine was collected over the period of 24h after dosing. Serum concentrations and urinary excretion were determined by microbiological assay with *E. coli* Kp.

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