Isolation and Structure Elucidation of a Novel Alkaloid, Incartine, a Supposed Biosynthetic Intermediate, from Flowers of *Lycoris incarnata*

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A novel alkaloid incartine (1), a supposed biosynthetic intermediate from galanthine (2) to narcissidine (3), was isolated from flowers of Lycoris incarnata (Amaryllidaceae) together with the known alkaloids galanthine (2), ungiminorine (4), ungiminorine N-oxide (5), galanthamide (6), galanthamine N-oxide (7), lycoramine (8), sanguinine (9), lycorine (10), and O-demethyllycoramine (11). 1-Palmitoyl-2-linoleoylphosphatidylethanolamine (12) and 1-palmitoyl-2-linoleoylphosphatidylmethanol sodium salt (13) were also identified in the flower.

Keywords incartine; Amaryllidaceae alkaloid; biosynthetic intermediate; Lycoris incarnata; flower; phospholipid

The Amaryllidaceae alkaloids have been studied extensively because of the variety of their structures and biological activities and also from the biosynthetic viewpoint. However, flowers of Amaryllidaceae plants have not attracted much attention from phytochemists. In a previous paper, we reported the isolation of a new alkaloid, hippeastrine N-oxide, together with known alkaloids from flowers of Lycoris radiata Herb. Recently, we have isolated a novel alkaloid, incartine (1), which was supposed to be a biosynthetic intermediate from galanthine (2) to narcissidine (3), from flowers of Lycoris incarnata. This paper describes in detail the isolation and the structural elucidation of incartine (1), together with nine known alkaloids, galanthine (2), ungiminorine (4), ungiminorine N-oxide (5), galanthamine (6), galanth-

amine N-oxide (7), lycoramine (8), sanguinine (9), lycorine (10), and O-demethyllycoramine (11), from fresh flowers of this plant. 1-Palmitoyl-2-linoleoylphosphatidylethanolamine (12) and 1-palmitoyl-2-linoleoylphosphatidylmethanol sodium salt (13) were also isolated.

Crude extract of fresh flowers of *Lycoris incarnata* obtained by the modified method of Ghosal *et al.*²⁾ was subjected to column and preparative thin layer chromatographies (PTLC), as described in Experimental to give compounds 1, 2, 4—12 and 13.

The new compound, incartine, $C_{18}H_{23}NO_5$, was isolated as colorless prisms, mp 183—185 °C. The infrared (IR) spectrum showed hydroxy group absorption at $3433\,\mathrm{cm}^{-1}$, but no absorption due to a carbonyl group. The proton nuclear magnetic resonance (1H -NMR)

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spectrum revealed the presence of two para-oriented aromatic protons (δ 6.97 and 6.95), two aromatic methoxy groups (δ 3.92 and 3.90), one aliphatic methoxy group $(\delta 3.57)$, and benzyl protons ($\delta 4.71$ and 4.31, each doublet, J=13.0 Hz), but no N-methyl signal. These data suggested that the compound has a lycorine-type skeleton. This suggestion was supported by a two-dimensional ¹H-¹H shift correlation spectroscopy (2D-COSY) experiment, which indicated the presence of the sequence H_{11c}-H_{11b}-H₁-H₂-H₃ (see also below). These findings and the molecular formula (C₁₈H₂₃NO₅) indicate that the structure of incartine should be very similar to that of narcissidine (3), ⁶⁾ galanthine N-oxide (14), ⁷⁾ or the 3,3a-epoxy derivative of galanthine (2). Since the ¹H-NMR spectrum of incartine did not show an olefinic proton signal, the compound was considered to be an α - or β -3,3a-epoxy derivative (1 or 15) of galanthine (2).

In order to elucidate the structure and the stereochemistry of incartine, a decoupling experiment and the nuclear magnetic double resonance (NMDR) analysis were carried out. The completely assigned chemical shifts, the coupling patterns (see Experimental), and the nuclear Overhauser effect (NOE) enhancements (Fig. 1) suggested the relative configuration of the 3,3a-epoxy ring of incartine to be α . The conformation of α -3,3a-epoxygalanthine (1) inspected from a Dreiding model was compared with that of the 3α , 3α -glycol derivative (16) reported by Toda et al.8 (see Fig. 2). It was reported8) that the C-ring of 16 took a distorted boat conformation with trans diaxial orientation of H-2 and H-3 ($J=8.3\,\mathrm{Hz}$). On the contrary, a small coupling $(J=1.2 \,\mathrm{Hz})$ of the corresponding hydrogens in incartine suggested the dihedral angle between them to be ca. 90°. A long-range coupling $(J=0.8 \,\mathrm{Hz})$ between H-1 and H-3 was observed in this case. These observations show that the C-ring takes a distorted chair conformation in the α -3,3a-epoxy compound. The β -orientation of the epoxy ring, such as in 15, would give a different coupling pattern. From these findings, incartine was concluded to be galanthine α -3,3a-epoxide (1).

Fuganti *et al.*⁵⁾ suggested that galanthine (2) is probably transformed to narcissidine (3) *via* the α -,3,3a-epoxide (1), since they found that galanthine (2) was converted to narcissidine (3) in *Sempre avanti* daffodil with loss of *pro*-S hydrogen from C-4 of the lycorane skeleton. Toda *et al.*⁸⁾ gave chemical support to this elimination step. However, the proposed intermediate, the α -epoxide (1), has never been isolated or synthesized. Therefore, this paper is the first to report isolation of the proposed epoxy intermediate.

Compound 5, $[\alpha]_D - 58.9^\circ$ (EtOH), was isolated as a pale yellow oil. The mass (MS) spectrum showed the molecular formula $C_{17}H_{19}NO_6$, suggesting the presence of one more oxygen atom than in ungiminorine (4). The ¹H-NMR spectrum of 5 showed a similarity to that of 4 except for deshielding of the protons at C-5, C-7 and C-11c. This alkaloid was identical with ungiminorine *N*-oxide (5) isolated from *Pancratium maritimum* (Amaryllidaceae).⁹⁾

Compounds 2, 4, 6—10 and 11 were identified as galanthine, ungiminorine, galanthamine, galanthamine *N*-oxide, lycoramine, sanguinine, lycorine, and *O*-de-

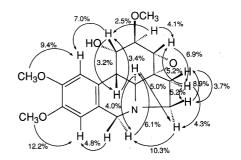


Fig. 1. NOE Enhancements for Incartine (1) in CDCl₃

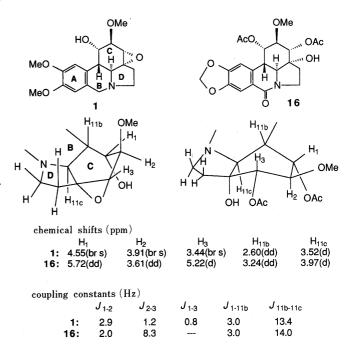


Fig. 2. Analysis of ¹H-NMR Spectra of 1 (at 400 MHz) and 16 (at 100 MHz, Ref. 8) in CDCl₃

methyllycoramine, respectively, by direct comparisons of their physical and spectral data and thin layer chromatographic (TLC) behavior with those of the authentic samples.

Compounds 12 and 13 gave a blue-violet color with molybdenum blue reagent. 100 Compound 12 showed a pale orange color with Dragendorff's reagent but compound 13 did not. The IR spectra of 12 and 13 revealed ester absorptions at 1741 and 1742 cm⁻¹, and phosphate absorptions at 1045 and 1050 cm⁻¹, respectively. ¹H-NMR spectra of 12 and 13 indicated the presence of palmitate and linoleate groups³⁾ in both compounds (see Experimental). The EI-MS spectra of 12 and 13 showed similar fragmentation patterns. The intensities (80 and 82%) of the fragments of m/z 313 generated by cleavage of a lineleate group from the fragments $(m/z 575)^{3}$ of 12 and 13 were higher than those (65 and 31%) of m/z 337 generated by cleavage of a palmitate group, respectively. These facts suggest that 12 and 13 are 1-palmitoyl-2linoleoylphosphatidyl derivatives.3,11) In addition, the ¹H-NMR spectrum of 12 showed the presence of an aminoethyl group and the FAB-MS revealed the molecular ion peak of m/z 716 (M+1) for $C_{39}H_{74}NO_8P$. From these data, compound 12 was concluded to be 1-palmitoyl-2linoleoylphosphatidylethanolamine. The FAB-MS of 13 showed the molecular ion peak of m/z 709 (M+1) for $C_{38}H_{70}NaO_8P$ and the H-NMR spectrum indicated the presence of a methoxy group. These findings suggested that compound 13 is 1-palmitoyl-2-linoleoylphosphatidylmethanol sodium salt. Compound 13 may be an artifact derived from a corresponding phosphatidylcholine or a phosphatidylethanolamine during the extraction of the flower with MeOH–CHCl₃.

Experimental

All melting points are given as uncorrected values. The spectrophotometers used were a Perkin–Elmer 1720 infrared Fourier-transform spectrophotometer for IR spectra, a JEOL JMS-D 300 for MS, a Union PM-201 for optical rotations, and JEOL JNM-FX 200, JEOL JNM-GSX 400 and Bruker AM-400 spectrometers for ¹H-NMR spectra with tetramethylsilane as an internal standard. The plates used for PTLC were coated with silica gel (Kieselgel PF₂₅₄, Merck) and aluminum oxide (PF₂₅₄, Merck). The following solvent systems were used: 1) CHCl₃–MeOH (5:1); 2) CHCl₃–MeOH–H₂O (60:35:10); 3) CHCl₃–AcOEt–MeOH–H₂O (70:30:10:2); 4) CHCl₃–MeOH–NH₄OH–H₂O (70:26:2:2); 5) CHCl₃–MeOH (10:1). UV light, I₂ vapor, Dragendorff's reagent and molybdenum blue reagent¹⁰⁾ were used for location of compounds.

Extraction Following the modified method of Ghosal *et al.*,²⁾ fresh flowers (5.3 kg) of *Lycoris incarnata* collected in our Faculty plot were ground in 8.741 of CHCl₃–MeOH (2:1) in a mixer. The extract was warmed at 60 °C for 1 h, then 0.1 m EDTA (87 ml) was added to retard the phospholipase activity and the mixture was kept at room temperature overnight, then filtered to give two layers. The CHCl₃ and MeOH–H₂O layers were concentrated *in vacuo* to afford sticky extracts, 18.5 and 98 g, respectively.

Treatment of the CHCl₃ Extract The CHCl₃ extract was subjected to column chromatography using HCl-washed Florisil (25×5.5 cm). Elution was carried out successively with benzene (1.85 l), CHCl₃ (6 l), CHCl₃–MeOH (95:5, 2.85 l, fraction (fr.) I, 1.627 g), CHCl₃–MeOH (9:1, 1.75 l, fr. II, 488 mg), CHCl₃–MeOH (3:1, 3.45 l, fr. III, 305 mg), and CHCl₃–MeOH (1:1, 3.49 l, fr. IV, 360 mg).

Fraction I (1.627 g) was subjected to column chromatography on SiO₂. Elution was carried out successively with CHCl₃–MeOH (10:1), CHCl₃–MeOH (5:1), and MeOH. The MeOH fraction gave an oil, which was subjected to PTLC (SiO₂, solvent 2) to afford incartine (1) (Rf 0.53–0.62, 14 mg).

Fraction II (488 mg) was subjected to PTLC (SiO_2 , solvent 1) to afford an oil (Rf 0.28—0.36, 45.4 mg). This crude material was further purified by PTLC (SiO_2 , solvent 4) to give 1-palmitoyl-2-linoleoylphosphatidylethanolamine (12) (Rf 0.68—0.74, 7.7 mg).

Fraction III (305 mg) was subjected to PTLC (SiO₂, solvent 1) to give two fractions (Rf 0.38—0.41, fr. III-A, 17.2 mg; Rf 0.13—0.23, fr. III-B, 52.9 mg). Purification of fr. III-A and fr. III-B by PTLC (Al₂O₃, solvent 3 and 1) gave ungiminorine (4) (Rf 0.65—0.75, 7.2 mg) and ungiminorine N-oxide (5) (Rf 0.61—0.65, 2.9 mg), respectively.

Fraction IV (360 mg) was subjected to PTLC (SiO₂, solvent 4) to afford two fractions (Rf 0.80—0.92, fr. IV-A, 33 mg, Rf 0.45—0.55, fr. IV-B, 139.4 mg). Fraction IV-A was subjected to PTLC (SiO₂, solvent 1) to give galanthamine (6) (Rf 0.19—0.27, 5 mg) and lycoramine (8), (Rf 0.37—0.44, 8.5 mg). Fraction IV-B was subjected to PTLC (SiO₂, solvent 4) to afford an oil (Rf 0.59—0.69, 75.1 mg), which was further separated by PTLC (Al₂O₃, solvent I) to give galanthamine N-oxide (7) (Rf 0.76—0.81, 1.6 mg) and 1-palmitoyl-2-linoleoylphosphatidylmethanol sodium salt (13) (Rf 0.02—0.08, 47 mg).

Treatment of the MeOH- H_2O Extract The MeOH- H_2O extract (98 g) was successively triturated with hot hexane, benzene, CHCl₃, and CHCl₃-MeOH (1:1) (fr. V, 6.454 g). The insoluble material was an oil (fr. VI, 55 g), which was soluble in MeOH.

Fraction V (6.454 g) was subjected to flash chromatography on SiO_2 with solvent 4 to give two fractions (550 ml, fr. V-A, 165.8 mg; 500 ml, fr. V-B, 119.8 mg). Fraction V-A was subjected to PTLC (SiO_2 , solvent 2) to give three fractions (Rf 0.55—0.63, fr. V-A-1, 34.7 mg; Rf 0.66—0.74, fr. V-A-2, 76 mg; Rf 0.86—0.89, fr. V-A-3, 2.1 mg of galanthine (2)). Fraction V-A-1 was purified by PTLC (Al_2O_3 , solvent 3) to give sanguinine (9) (Rf 0.50—0.59, 3.9 mg). Fraction V-A-2 was

triturated with MeOH to afford lycorine (10) (53.3 mg). Fraction V-B was subjected to PTLC (SiO₂, solvent 2) to give an oil (21.3 mg), which was further purified by PTLC (Al₂O₃, solvent 1) to afford *O*-demethyllycoramine (11) (*Rf* 0.13—0.44, 1.9 mg).

Fraction VI (55 g) was subjected to flash chromatography on SiO_2 with solvent 4. The solid obtained from the first fraction (1000 ml) was washed with MeOH to afford lycorine (10) (114 mg). The MeOH solution gave an oil (208 mg), which was subjected to PTLC (SiO₂, solvent 2) to give an amorphous solid (68.6 mg). This was further purified by PTLC (Al₂O₃, solvent 5) to afford ungiminorine (4) (Rf 0.38—0.44, 2.6 mg) and lycoramine (8) (Rf 0.78—0.82, 5.6 mg).

Incartine (1) Colorless prisms (from MeOH), mp 183—185 °C. IR (KBr): 3433 (OH), 1615, 1516, 1456, 1326, 1290, 1085 cm $^{-1}$. High MS m/z [M $^+$]: Calcd for C₁₈H₂₃NO₅: 333.1573. Found: 333.1538. EI-MS m/z (%): 333 (M $^+$, 50), 332 (100), 296 (25), 295 (22), 294 (20), 266 (17), 259 (74), 258 (65), 244 (30), 242 (23). $^1\mathrm{H}\text{-NMR}$ (CDCl₃, 400 MHz) δ: 6.97 (1H, s, H-8), 6.95 (1H, s, H-11), 4.71 (1H, d, $J_{7\beta^-7\alpha}=13.0$ Hz, H-7β), 4.55 (1H, br s, $J_{1-2}=2.9$ Hz, $J_{1-11b}=3.0$ Hz, $J_{1-3}=0.8$ Hz, H-1), 4.31 (1H, d, $J_{7\alpha^-7\beta}=13.0$ Hz, H-7α), 4.08 (1H, dd, $J_{5\beta^-5\alpha}=10.8$ Hz, $J_{5\beta^-4\beta}=7.2$ Hz, H-5β), 3.92 (3H, s, OCH₃-10), 3.90 (3H, s, OCH₃-9), 3.91 (1H, br s, $J_{2-1}=2.9$ Hz, $J_{2-3}=1.2$ Hz, H-2), 3.76 (1H, ddd, $J_{5\alpha^-5\beta}=10.8$ Hz, $J_{5\alpha^-4\beta}=14.2$ Hz, $J_{5\alpha^-4\alpha}=5.3$ Hz, H-5α), 3.57 (3H, s, OCH₃-2), 3.52 (1H, d, $J_{11c-11b}=13.4$ Hz, H-11c), 3.44 (1H, br s, $J_{3-2}=1.2$ Hz, $J_{3-1}=0.8$ Hz, H-3), 3.38 (1H, ddd, $J_{4\beta^-4\alpha}=13.1$ Hz, $J_{4\beta^-5\alpha}=14.2$ Hz, $J_{4\beta^-5\beta}=7.2$ Hz, H-4β), 2.60 (1H, dd, $J_{11b-11c}=13.4$ Hz, $J_{11b-1}=3.0$ Hz, H-11b), 2.04 (1H, dd, $J_{4\alpha^-4\beta}=13.1$ Hz, $J_{4\alpha^-5\alpha}=5.3$ Hz, H-4α).

Galanthine (2) Pale yellow oil, $[\alpha]_{D}^{29}$ – 65.9° $(c=0.046, \text{CHCl}_3)$ (lit.¹³⁾ $[\alpha]_{D}^{22}$ – 94.7° $(c=0.686, \text{CHCl}_3)$). High MS m/z [M⁺]: Calcd for $C_{18}H_{23}NO_4$: 317.1628. Found: 317.1603. ¹H-NMR (CDCl₃, 200 MHz) δ : 6.68 (1H, s, H-11), 6.63 (1H, s, H-8), 5.61 (1H, br s, H-3), 4.67 (1H, s, H-1), 4.17 and 3.50 (each 1H, d, J=14 Hz, CH_2-7), 3.90 (3H, s, OCH₃-9), 3.87 (3H, s, OCH₃-10), 3.53 (3H, s, OCH₃-2).

Ungiminorine (4) Colorless prisms (from acetone), mp 207—208 °C (lit. 14) mp 208—211 °C), $[\alpha]_D^{24}$ –42° (c=0.46, EtOH) (lit. 14) $[\alpha]_D^{24}$ –49° (c=0.67, EtOH)). High MS m/z [M $^+$]: Calcd for C₁₇H₁₉NO₅: 317.1264. Found: 317.1236. 1 H-NMR (CDCl₃, 200 MHz) δ: 6.68 (1H, s, H-1l), 6.63 (1H, s, H-8), 5.88 (2H, s, OCH₂O), 5.56 (1H, br s, H-4), 4.65 (2H, m, H-1 and 3), 4.01 and 3.50 (each 1H, d, J=13 Hz, CH₂-7), 3.84 (1H, br s, H-1lc), 3.43 (3H, s, OCH₃), 2.67 (1H, d, J=11.2 Hz, H-11b).

Ungiminorine *N*-Oxide (5) Pale yellow oil, $[\alpha]_D^{28} - 58.9^{\circ}$ (c = 0.11, EtOH). IR (KBr): 3425, 2900, 1485 cm⁻¹. High MS m/z [M⁺]: Calcd for C₁₇H₁₉NO₆: 333.1213. Found: 333.1219. ¹H-NMR (CDCl₃, 200 MHz) δ: 6.98 (1H, s, H-11), 6.84 (1H, s, H-8), 5.97 and 5.95 (each 1H, d, J = 1.7 Hz, OCH₂O), 5.62 (1H, br s, H-4), 4.65 and 4.53 (each 1H, br s, CH₂-5), 4.50 and 4.16 (each 1H, d, J = 13 Hz, CH₂-7), 4.20 (1H, br s, H-11c), 3.34 (3H, s, OCH₃), 2.65 (1H, d, J = 12.5 Hz, H-11b).

Galanthamine (6) Colorless prisms (from acetone), mp 126—127 °C (lit. ¹⁵⁾ mp 127—128 °C), $[\alpha]_D^{25}$ –91.6° (c=0.19, EtOH) (lit. ¹⁵⁾ $[\alpha]_D^{23}$ –109.2° (c=0.85, EtOH)). High MS m/z [M⁺]: Calcd for C₁₇H₂₁NO₃: 287.1522. Found: 287.1495. ¹H-NMR (CDCl₃, 200 MHz) δ: 6.67 and 6.62 (each 1H, d, J=8 Hz, H-11 and 12), 6.01—6.05 (2H, m, H-3 and 4), 4.62 (1H, br s, H-17), 4.16 (1H, m, H-2), 4.08 and 3.68 (each 1H, d, J=15 Hz, H-9), 3.84 (3H, s, OCH₃), 2.40 (3H, s, NCH₃).

Galanthamine *N*-Oxide (7) Colorless oil, $[\alpha]_D^{24} - 133.8^\circ$ (c = 0.05, MeOH) (lit. ¹⁵) $[\alpha]_D^{26} - 122.9^\circ$ (c = 0.38, MeOH)). IR (KBr): 3437, 2922, 1632, 1512 cm⁻¹. High MS m/z [M⁺]: Calcd for C₁₇H₂₁NO₄: 303.1469. Found: 303.1450. ¹H-NMR (CDCl₃, 200 MHz) δ: 6.77 (2H, s, H-11 and 12), 6.08—6.03 (2H, m, H-3 and 4), 4.65 (1H, br s, H-16), 4.70 and 4.40 (each 1H, d, J = 15 Hz, CH₂-9), 4.20 (1H, br s, H-2), 3.88 (3H, s, OCH₃), 3.10 (3H, s, NCH₃).

Lycoramine (8) Colorless prisms (from acetone), mp 122—124 °C (lit. $^{16)}$ mp 122—124 °C, $[\alpha]_D^{22} - 100.0^\circ$ (c = 0.05, EtOH) (lit. $^{16)}$ $[\alpha]_D^{22} - 96.0^\circ$ (c = 0.71, EtOH)). High MS m/z [M $^+$]: Calcd for $C_{17}H_{23}NO_3$: 289.1679. Found: 289.1663. 1 H-NMR (CDCl $_3$, 200 MHz) δ : 6.666 and 6.60 (each 1H, d, J = 8 Hz, H-11 and 12), 4.37 (1H, m, H-17), 4.05 (1H, m, H-2), 4.01 and 3.62 (each 1H, d, J = 15 Hz, CH $_2$ -9), 3.86 (3H, s, OCH $_3$), 2.37 (3H, s, NCH $_3$).

Sanguinine (9) Pale yellow oil, $[\alpha]_D^{27} - 100.0^\circ$ (c = 0.08, EtOH) (lit.¹⁷⁾ $[\alpha]_D^{27} - 133.0^\circ$ (c = 0.23, EtOH)). High MS m/z [M⁺]: Calcd for $C_{16}H_{19}NO_3$: 273.1366. Found: 273.1350. ¹H-NMR (CDCl₃, 200 MHz) δ : 6.67 and 6.43 (each 1H, d, J = 8 Hz, H-11 and 12), 4.43 (1H, m, H-17), 4.09 (1H, m, H-2), 4.05 and 3.63 (each 1H, d, J = 15 Hz, CH₂-9), 2.38 (3H, s, NCH₃).

Lycorine (10) Colorless pillars (from MeOH), mp 258—259 °C (lit. 3)

mp 257—258 °C).

O-Demethyllycoramine (11) Pale yellow oil, $[\alpha]_D^{24} - 103.1^\circ$ (c = 0.21, EtOH) (lit.¹⁶⁾ $[\alpha]_D^{23} - 111.9^\circ$ (c = 0.59, EtOH)). High MS m/z [M⁺]: Calcd for C₁₆H₂₁NO₃: 275.1522. Found: 275.1513. ¹H-NMR (CDCl₃, 200 MHz) δ: 6.64 and 6.52 (each 1H, d, J = 8 Hz, H-11 and 12), 4.36 (1H, br s, H-17), 4.11 (1H, m, H-2), 3.98 and 3.60 (each 1H, d, J = 15 Hz, CH₂-9), 2.37 (3H, s, NCH₃).

1-Palmitoyl-2-linoleoylphosphatidylethanolamine (12) Pale yellow oil. EI-MS m/z (%): 575 (12), 337 (65), 313 (80), 263 (48), 239 (25). FAB-MS m/z: 716 [M+1] (thioglycerol). IR (KBr): 2925, 2854, 1741, 1652, 1446, 1385, 1074, 1045 cm⁻¹. ¹H-NMR (CDCl₃, 200 MHz), δ: 5.35 (4H, m, -CH=CH-×2), 5.22 (1H, m, -CH-), 4.48—3.78 (4H, m, OCH₂CH-×2), 3.88—3.04 (4H, m, OCH₂CH₂N), 3.48 (3H, s, NH₃), 2.76 (2H, t, J=6 Hz, = C-CH₂-C=), 2.28 (4H, m, COCH₂CH₂-×2), 2.03 (4H, m, = C-CH₂-×2), 1.58 (4H, br s, COCH₂CH₂-×2), 0.88 (6H, m, CH₃).

1-Palmitoyl-2-linoleoylphosphatidylmethanol Sodium Salt (13) Pale yellow oil. EI-MS m/z (%): 575 (58), 337 (31), 313 (82), 263 (39), 239 (32). FAB-MS m/z: 709 [M+1] (thioglycerol); 731 [M+Na] (thioglycerol+NaI). IR (KBr): 2926, 2855, 1742, 1466, 1234, 1073, 1050 cm⁻¹. ¹H-NMR (CDCl₃, 200 MHz) δ: 5.34 (4H, m, -CH=CH-×2), 5.25 (1H, m, -CH-), 4.37—3.88 (4H, m, OCH₂CH-×2), 3.57 and 3.51 (3H, each s, OCH₃), 2.77 (2H, t, J=6 Hz, =C-CH₂-C=), 2.30 (4H, m, COCH₂CH₂-×2), 2.05 (4H, m, =C-CH₂-×2), 1.59 (4H, br s, COCH₂CH₂-×2), 0.89 (6H, m, CH₃).

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