

## Two New Triterpenoidal Glycosides from *Medicago polymorpha* L.<sup>1)</sup>

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**Two new triterpenoid glycosides called medicago-saponins P<sub>1</sub> (1) and P<sub>2</sub> (2) were isolated together with five known glycosides from the aerial parts of *Medicago polymorpha* L. (Leguminosae). The structures of 1 and 2 were determined to be 3-O- $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 2)- $\alpha$ -L-arabinopyranosyl caulophyllogenin 28-O- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside and the desglucoside of 1.**

**Keywords** *Medicago polymorpha*; Leguminosae; triterpene saponin; oleanene glycoside; medicago-saponin; caulophyllogenin

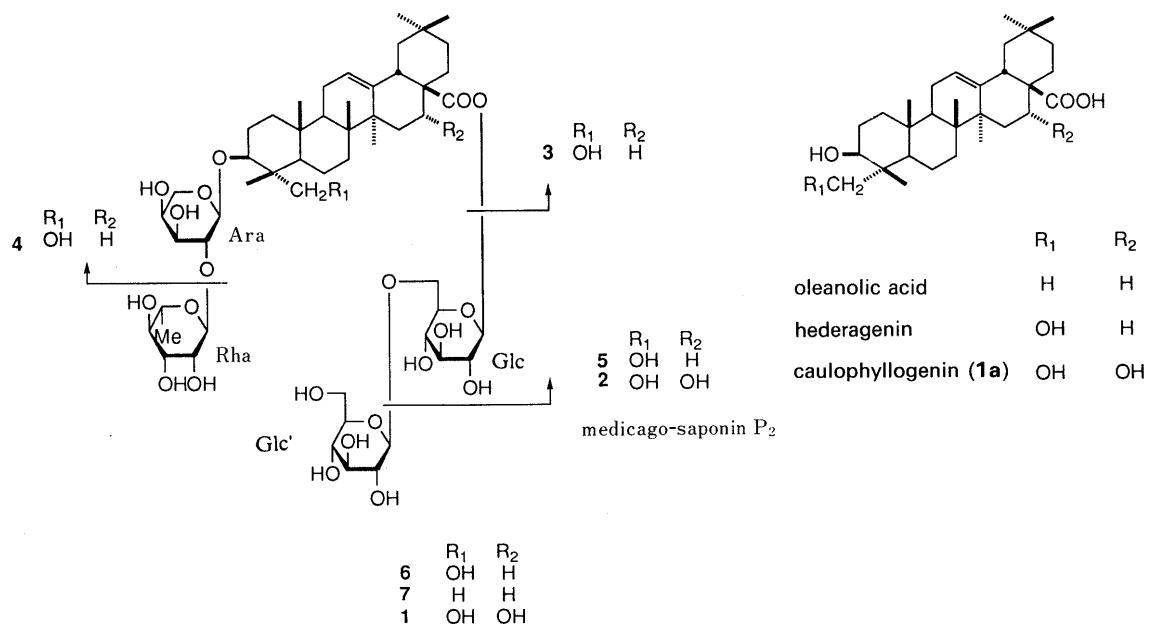
*Medicago polymorpha* L. (syn *M. denticulata* WILLD. and *M. hispida* GAERTN.) is used as an important fodder together with alfalfa (*M. sativa* L.) worldwide. Although earlier researchers found that alfalfa included many saponins and reported their structures,<sup>2)</sup> there have been few reports concerning the saponins of *M. polymorpha*.<sup>3)</sup> During the course of our study on leguminous plants,<sup>1)</sup> we have investigated the saponins of Japanese *M. polymorpha*. This paper deals with the structural elucidation and identification of these saponins.

A methanolic extract of the aerial parts of *M. polymorpha* was partitioned with 40% MeOH and ethyl acetate. A combination of various chromatographies of the former phase resulted in the isolation of seven saponins (1—7). Saponins 3—7 were identified as sapindoside A (3),<sup>4,5)</sup> compound 6 (4),<sup>6)</sup> compound 9 (5),<sup>5)</sup> compound 10 (6)<sup>5)</sup> and compound 11 (7)<sup>5)</sup> comparing the negative FAB-MS, and the <sup>1</sup>H- and <sup>13</sup>C-NMR spectra with those of the published data.

Medicago-saponin P<sub>1</sub> (1) was obtained as a white amorphous powder,  $[\alpha]_D^{25} -20.1^\circ$  (pyridine). The IR spectrum of 1 featured absorptions of carbonyl (1735

cm<sup>-1</sup>) and a hydroxy group. In the negative FAB-MS, 1 showed an [M-H]<sup>-</sup> ion at *m/z* 1089. Fragment ion peaks at *m/z* 765 [M-hexose-hexose]<sup>-</sup> and 619 [M-hexose-hexose-methylpentose]<sup>-</sup> were also observed. These MS data appeared to a greater extent than those of 6 by a hydroxy group. By acid hydrolysis, 1 gave 1a as the sapogenol. The electron impact (EI)-MS of 1 showed a [M]<sup>+</sup> at *m/z* 488, indicating a hydroxy group located on 1a. By the <sup>1</sup>H- and <sup>13</sup>C-NMR spectral data (as listed in Tables I—III), the location of the hydroxy group was determined to be at C-16 $\alpha$ , which compares to those of hederagenin<sup>7)</sup> and acacic acid.<sup>8)</sup> Finally, 1a was identified with caulophyllogenin<sup>9a)</sup> because of the agreement of various data. In addition, the signals for the sugar region (Table II) were superimposable on those of 6 in the <sup>13</sup>C-NMR. Since the absolute configurations of the sugar species were identified according to the method reported by Hara *et al.*,<sup>10)</sup> the structure of 1 was established to be 3-O- $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 2)- $\alpha$ -L-arabinopyranosyl caulophyllogenin 28-O- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside.

Medicago-saponin P<sub>2</sub> (2) was obtained as a white



medicago-saponin P<sub>1</sub>

amorphous powder,  $[\alpha]_D -4.1^\circ$  (MeOH). An acidic hydrolysis of **2** gave caulophyllogenin (**1a**) as the sapogenol. In the FAB-MS of **2**, an  $[M + Na]^+$  ion at *m/z* 951 appeared to a lesser extent than that of **1** by a hexosyl moiety. Since the  $^{13}\text{C}$ -NMR signals of the sugar region (Table II) showed a good coincidence with those of **5**, **2** was concluded to be the desglucosyl compound of **1**, that is, 3-*O*- $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 2)- $\alpha$ -L-arabinopyranosyl caulophyllogenin 28-*O*- $\beta$ -D-glucopyranoside.

Saponins with caulophyllogenin as their aglycone are very rare. Only two examples are known in nature. They were isolated from *Caulophyllum robustum*<sup>9a)</sup> (Berberidaceae) and *Chrysanthellum procumbens*<sup>9b)</sup> (Compositae). Medicago-saponins P<sub>1</sub> and P<sub>2</sub> are the third and fourth saponins known to have caulophyllogenin, respectively, and are the first examples from Leguminosae.

TABLE I.  $^{13}\text{C}$ -NMR Data for Compounds **1**—**7** (Aglycone Moieties) and **1a** in Pyridine-*d*<sub>5</sub>

Carbon No.	<b>1a</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>
1	38.9	38.9	38.9	38.9	38.8	38.9	38.8	38.8
2	27.7	26.0	26.1	26.2	26.1	26.0	26.0	26.4
3	73.6	80.9	81.2	81.0	81.9	80.9	80.9	88.7
4	42.2	43.3	43.3	43.5	43.5	43.3	43.3	39.4
5	47.2	47.1	47.1	47.7	47.6	47.5	47.5	55.8
6	18.7	18.0	18.1	18.2	18.2	18.2	18.0	18.4
7	33.3	32.9	32.9	33.2	32.8	32.6	32.6	33.0
8	40.0	39.9	40.0	39.7	40.0	39.8	39.7	39.8
9	48.9	47.6	47.6	48.1	48.2	48.0	48.0	47.9
10	37.3	36.8	36.8	36.9	36.9	36.7	36.7	36.9
11	23.9	23.7	23.7	23.8	23.9	23.7	23.7	23.7
12	122.5	122.5	122.6	122.6	122.9	122.8	122.7	122.7
13	145.1	144.2	144.5	144.8	144.2	143.9	143.9	144.0
14	42.9	41.8	42.0	42.1	42.9	42.0	42.0	42.0
15	36.2	35.9	35.7	28.3	28.3	28.1	28.1	28.2
16	74.7	74.1	73.4	23.6	23.4	23.2	23.2	23.3
17	48.6	48.9	49.2	46.6	47.0	46.8	46.9	46.9
18	41.5	41.0	41.2	42.0	41.7	41.6	41.5	41.6
19	47.3	46.9	47.1	46.4	46.2	46.0	46.0	46.1
20	31.1	30.5	30.6	30.9	30.8	30.6	30.5	30.6
21	36.2	35.7	35.7	34.2	34.0	33.8	33.8	33.9
22	32.9	31.9	31.8	32.8	32.5	32.4	32.3	32.4
23	68.2	63.7	63.8	64.0	64.5	63.8	63.8	15.5
24	13.1	13.7	13.7	14.0	13.6	13.8	13.7	28.0
25	16.1	16.1	16.2	16.1	16.2	16.0	16.0	16.8
26	17.6	17.4	17.5	17.5	17.6	17.4	17.4	17.4
27	27.2	27.0	27.1	26.2	26.0	25.9	25.8	25.9
28	178.0	175.8	176.4	180.2	176.6	176.3	176.4	176.4
29	33.3	33.0	33.0	33.2	33.1	33.0	32.9	33.0
30	24.8	24.5	24.6	23.8	23.7	23.5	23.5	23.5

TABLE II.  $^{13}\text{C}$ -NMR Data for Compounds **1**—**7** (Sugar Moieties) in Pyridine-*d*<sub>5</sub>

Carbon No.	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>
Ara							
1	104.0	103.8	104.0	106.6	104.1	104.0	104.6
2	75.6	76.0	75.8	73.1	75.7	75.7	75.8
3	74.2	73.6	74.7	74.7	74.4	74.3	73.9
4	69.5	70.4	69.7	69.6	69.5	69.5	69.8
5	65.3	64.8	65.6	67.0	65.3	65.3	64.3
Rha							
1	101.4	101.4	101.6		101.5	101.5	101.6
2	72.1 <sup>a)</sup>	71.6 <sup>a)</sup>	72.3 <sup>a)</sup>		72.1 <sup>a)</sup>	72.1 <sup>a)</sup>	72.2 <sup>a)</sup>
3	72.3 <sup>a)</sup>	71.9 <sup>a)</sup>	72.5 <sup>a)</sup>		72.4 <sup>a)</sup>	72.3 <sup>a)</sup>	72.4 <sup>a)</sup>
4	73.7	73.4	74.1		73.9	73.7	73.4
5	69.0	68.6	69.3		69.1	69.0	68.4
6	18.3	18.2	18.5		18.3	18.3	18.4
Glc							
1	95.6	95.6		95.7	95.6	95.5	95.5
2	73.8	74.0		73.9	74.0	73.9	73.8
3	78.2 <sup>b)</sup>	78.8 <sup>b)</sup>		78.4 <sup>a)</sup>	79.1 <sup>b)</sup>	78.2 <sup>b)</sup>	78.3 <sup>b)</sup>
4	70.6	70.7		70.9	71.0	70.7	70.7
5	77.7	78.0 <sup>b)</sup>		78.0	78.7 <sup>b)</sup>	77.7	77.8
6	69.1	61.8		69.4	62.0	69.1	69.2
Glc							
1	105.0			105.3		105.0	105.1
2	74.9			75.2		74.9	75.0
3	78.1 <sup>b)</sup>			78.4 <sup>a)</sup>		78.2 <sup>b)</sup>	78.3 <sup>b)</sup>
4	71.2			71.5		71.3	71.4
5	78.4 <sup>b)</sup>			78.7 <sup>a)</sup>		78.5 <sup>b)</sup>	78.5 <sup>b)</sup>
6	62.3			62.6		62.4	62.5

a, b) In each vertical column may be interchanged.

TABLE III.  $^1\text{H}$ -NMR Data for Compounds **1**—**7** and **1a** in Pyridine-*d*<sub>5</sub>

	<b>1a</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>
<i>tert</i> -Me	1.02 (s)	0.96 (s)	0.95 (s)	0.94 (s)	0.87 (s)	0.88 (s)	0.86 (s)	0.89 (s)
	1.05 (s)	1.03 (s)	0.96 (s)	0.94 (s)	0.88 (s)	0.89 (s)	0.87 (s)	0.89 (s)
	1.08 (s)	1.03 (s)	0.99 (s)	1.00 (s)	0.94 (s)	0.96 (s)	0.99 (s)	0.89 (s)
	1.10 (s)	1.07 (s)	1.01 (s)	1.02 (s)	0.99 (s)	1.06 (s)	1.08 (s)	1.07 (s)
	1.19 (s)	1.17 (s)	1.12 (s)	1.06 (s)	1.14 (s)	1.12 (s)	1.14 (s)	1.11 (s)
	1.81 (s)	1.76 (s)	1.76 (s)	1.23 (s)	1.19 (s)	1.19 (s)	1.18 (s)	1.17 (s)
H-18	3.64 (br d, <i>J</i> =13.9)	3.51 (br d, <i>J</i> =13.9)	3.47 (br d, <i>J</i> =13.0)	3.28 (br d, <i>J</i> =9.5)	3.20 (br d, <i>J</i> =13.2)	3.18 (br d, <i>J</i> =10.3)	3.19 (br d, <i>J</i> =13.9)	3.21 (br d, <i>J</i> =13.9)
H-3	4.22 (dd, <i>J</i> =11.0, 5.1)	— <sup>a)</sup>	— <sup>a)</sup>	4.12 (m)	— <sup>a)</sup>	4.12 (m)	— <sup>a)</sup>	3.24 (dd, <i>J</i> =12.1, 4.0)
H-16	5.26 (br s)	5.27 (br s)	5.29 (br s)					
H-12	5.68 (br s)	5.60 (br s)	5.61 (br s)	5.47 (br s)	5.45 (br s)	5.42 (br s)	5.42 (br s)	5.43 (br s)
Glc H-1		6.26 (d, <i>J</i> =8.1)	6.2 (d, <i>J</i> =8.8)		6.26 (d, <i>J</i> =8.1)	6.31 (d, <i>J</i> =8.1)	6.27 (d, <i>J</i> =8.1)	6.27 (d, <i>J</i> =8.1)
Glc' H-1		5.02 (d, <i>J</i> =8.1)			4.98 (d, <i>J</i> =7.4)		5.05 (d, <i>J</i> =8.1)	5.04 (d, <i>J</i> =7.7)
Ara H-1		5.11 (d, <i>J</i> =5.9)	5.13 (d, <i>J</i> =5.9)	5.11 (d, <i>J</i> =6.6)	5.04 (d, <i>J</i> =7.3)	5.11 (d, <i>J</i> =5.9)	5.12 (d, <i>J</i> =5.9)	4.91 (d, <i>J</i> =5.1)
Rha H-1		6.23 (s)	6.05 (s)	6.24 (s)		6.21 (s)	6.25 (s)	6.12 (s)
Rha H-6		1.64 (d, <i>J</i> =6.6)	1.65 (d, <i>J</i> =6.6)	1.63 (d, <i>J</i> =5.9)		1.63 (d, <i>J</i> =5.9)	1.65 (d, <i>J</i> =5.9)	1.64 (d, <i>J</i> =6.2)

a) Overlapped with the other signals.

## Experimental

The instruments and reagents used in this study were the same as those described in the previous paper.<sup>8)</sup>

**Extraction and Isolation** The fresh aerial parts(3.0 kg) of *Medicago polymorpha* collected in the medicinal garden of our Faculty were extracted with MeOH, and the extract (160 g) was partitioned with 40% MeOH and EtOAc. The aqueous extract (130 g) was subjected to Diaion HP-20 column chromatography using 0%→100% MeOH to give fractions 1 to 4. Each fraction was separated by Wako gel LP60 (0%→100% MeOH), Bondapak C<sub>18</sub> (0%→100% MeOH) and silica gel (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=9:1:0.1→6:4:1) to provide compounds **1** (0.029%), **2** (0.0019%), **3** (0.0012%), **4** (0.0005%), **5** (0.011%), **6** (0.049%) and **7** (0.0006%).

**Compound 1 (Medicago-Saponin P<sub>1</sub>)** A white amorphous powder, TLC *Rf*, 0.11 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>28</sup> -20.1° (*c*=0.30, pyridine). IR (KBr): 3390 (ν<sub>O-H</sub>), 1735 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Negative FAB-MS: *m/z* 1089 [M-H]<sup>-</sup>, 765 [M-H-Glc-Glc]<sup>-</sup>, 619 [M-H-Glc-Glc-Rha]<sup>-</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Characterization of Sapogenol and Sugars for 1** Compound **1** (12 mg) was dissolved in 2 N HCl/H<sub>2</sub>O (2 ml) and heated at 90°C for 2 h. After filtration of the mixture, the precipitate was recrystallized with MeOH to provide a sapogenol (**1a**), Colorless fine needles, mp >300°C, [α]<sub>D</sub><sup>28</sup> +24.4° (*c*=0.4, pyridine). IR (KBr): 3410 (ν<sub>O-H</sub>), 1735 (ν<sub>C=O</sub>) cm<sup>-1</sup>. EI-MS: *m/z* 488 [M<sup>+</sup>]. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Table I. The filtrate was neutralized by Amberlite MB-3 column chromatography. After evaporation, the residue (5 mg) was dissolved in pyridine (0.7 ml), then the mixture was added to a pyridine solution (2 ml) of L-cysteine methyl ester hydrochloride (0.06 mol/l) and warmed at 60°C for 1 h. The mixture was evaporated under N<sub>2</sub> stream and dried *in vacuo*. The obtained syrup was trimethylsilylated with *N*-trimethylsilylimidazole (0.4 ml) at 60°C for 1 h. After the addition of *n*-hexane (2 ml) and H<sub>2</sub>O (2 ml), the *n*-hexane layer was taken off and checked by GC. The retention time (*t*<sub>R</sub>) of the peaks was at 14.71 min (D-glucose, 14.75 min), 10.06 min (L-rhamnose, 10.11 min), and 8.34 min (L-arabinose, 8.36 min).

**Compound 2 (Medicago-Saponin P<sub>2</sub>)** A white amorphous powder, TLC *Rf*, 0.30 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>30</sup> -4.1° (*c*=0.54, MeOH). IR (KBr): 3410 (ν<sub>O-H</sub>), 1735 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Positive FAB-MS: *m/z* 951 [M+Na]<sup>+</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Characterization of Sapogenol** A sample of **2** was hydrolyzed in the same manner as above. The precipitate was identified with caulophyllogein (**1a**) by TLC *Rf*, 0.23 (CHCl<sub>3</sub>:MeOH=20:1).

**Compound 3** A white amorphous powder, TLC *Rf*, 0.61 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>30</sup> +19.9° (*c*=0.48, MeOH). IR (KBr): 3410 (ν<sub>O-H</sub>), 1700 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Positive FAB-MS: *m/z* 751 [M+H]<sup>+</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Compound 4** A white amorphous powder, TLC *Rf*, 0.30 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>30</sup> +17.7° (*c*=0.40, MeOH). IR (KBr): 3415 (ν<sub>O-H</sub>), 1725 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Positive FAB-MS: *m/z* 951 [M+Na]<sup>+</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Compound 5** A white amorphous powder, TLC *Rf*, 0.39 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>30</sup> +7.0° (*c*=0.49, MeOH). IR (KBr): 3415 (ν<sub>O-H</sub>), 1735 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Positive FAB-MS: *m/z* 935 [M+Na]<sup>+</sup>.

<sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Compound 6** A white amorphous powder, TLC *Rf*, 0.18 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>29</sup> +30.9° (*c*=0.33, MeOH). IR (KBr): 3395 (ν<sub>O-H</sub>), 1715 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Negative FAB-MS: *m/z* 1073 [M-H]<sup>-</sup>, 749 [M-H-Glc-Glc]<sup>-</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

**Compound 7** A white amorphous powder, TLC *Rf*, 0.26 (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O=7:3:0.5). [α]<sub>D</sub><sup>30</sup> -11.1° (*c*=0.41, MeOH). IR (KBr): 340 (ν<sub>O-H</sub>), 1735 (ν<sub>C=O</sub>) cm<sup>-1</sup>. Positive FAB-MS: *m/z* 1081 [M+Na]<sup>+</sup>. <sup>1</sup>H-NMR: Table III. <sup>13</sup>C-NMR: Tables I and II.

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## References and Notes

- 1) Part XL in a series of studies on leguminous plants.
- 2) I. Kitagawa, T. Taniyama, T. Murakami, M. Yoshihara, M. Yoshikawa, *Yakugaku Zasshi*, **108**, 547 (1988); G. Massiot, C. Lavaud, D. Guillaume, L. L. Men-Olivier, *J. Agric. Food Chem.*, **36**, 902 (1988); G. Massiot, C. Lavaud, L. L. Men-Olivier, G. V. Binst, S. P. F. Miller, H. M. Fales, *J. Chem. Soc., Perkin Trans 1*, **1988**, 3071; G. Massiot, C. Lavaud, V. Besson, L. L. Men-Olivier, G. V. Binst, *J. Agric. Food Chem.*, **39**, 78 (1991); G. Massiot, C. Lavaud, M. Benkhaled, L. L. Men-Olivier, *J. Nat. Prod.*, **55**, 1339 (1992); W. Oleszak, K. R. Price, I. J. Colquhoun, M. Jurzysta, M. Ploszynski, C. R. Fenwick, *J. Agric. Food Chem.*, **38**, 1810 (1990); W. Oleszak, M. Jurzysta, K. R. Price, C. R. Fenwick, *J. Chromatogr.*, **519**, 109 (1990); W. Oleszak, M. Jurzysta, M. Ploszynski, I. J. Colquhoun, K. R. Price, C. R. Fenwick, *J. Agric. Food Chem.*, **40**, 191 (1992).
- 3) S. B. Mahato, *Phytochemistry*, **30**, 3389 (1991).
- 4) H. Kimata, T. Nakashima, S. Kokubun, K. Nakayama, Y. Mitoma, T. Kitahara, N. Yata, O. Tanaka, *Chem. Pharm. Bull.*, **31**, 1998 (1983).
- 5) H. Kawai, M. Kuroyanagi, K. Umehara, A. Ueno, M. Satake, *Chem. Pharm. Bull.*, **36**, 4769 (1988).
- 6) B. Domon, K. Hostettman, *Helv. Chim. Acta*, **66**, 422 (1983).
- 7) J. Kinjo, K. Matsumoto, M. Inoue, T. Takeshita, T. Nohara, *Chem. Pharm. Bull.*, **39**, 116 (1991).
- 8) J. Kinjo, K. Araki, K. Fukui, H. Higuchi, T. Ikeda, T. Nohara, Y. Ida, N. Takemoto, M. Miyakoshi, J. Shoji, *Chem. Pharm. Bull.*, **40**, 3269 (1992).
- 9) a) L. I. Strigina, N. S. Chetyrina, V. V. Isakov, A. K. Dzisenko, G. B. Elyakov, *Phytochemistry*, **13**, 479 (1974); b) M. Becchi, M. Bruneteau, M. Trouilloud, H. Combier, H. Pontanier, G. Michel, *Eur. J. Biochem.*, **108**, 271 (1980).
- 10) S. Hara, H. Okabe, K. Mihashi, *Chem. Pharm. Bull.*, **35**, 501 (1987).