Double Bond Migration on the $22(17\rightarrow28)abeo$ -Lupane Skeleton

Hiroyuki Fuchino, Osamu Nozawa, and Nobutoshi Tanaka*

Faculty of Pharmaceutical Sciences, Science University of Tokyo, Funakawara-machi, Ichigaya, Shinjuku-ku, Tokyo 162, Japan. Received February 23, 1994; accepted April 23, 1994

Double bond migration on the $22(17 \rightarrow 28)abeo$ -lupane skeleton was investigated using 28-p-toluenesulfonyloxy-lupane (4) and its 20(29)-ene derivative (7) as starting materials. In the case of 4, the double bond, formed between C-17 and C-28 after elimination of the p-toluenesulfonyloxy group followed by E-ring expansion, migrated to C-13 and C-18 under acidic conditions. In the case of 7, the double bond, formed after the elimination and the E-ring expansion, migrated in response to migration of the other double bond, 20(29)-ene, under acidic conditions. First, 20(29)-ene migrated to 19-ene (9), and then 17(28)-ene migrated to 13(18)-ene to form a conjugated 13(18),19-diene (10). Further migration proceeded to give an equilibrium mixture of 17α -H-11,13(18)-diene (11), 12,17-diene (12) and 17β -H-11,13(18)-diene (13) in a ratio of 3:3:5.

Keywords double bond migration; $22(17\rightarrow28)abeo$ -lupane; ring expansion; 28-p-toluenesulfonyloxylupane; 28-p-toluenesulfonyloxylup-20(29)-ene; betulin

Betulin (1) is a naturally occurring triterpene, abundantly available mainly from birch bark.¹⁾ Despite considerable effort, no notable chemical, industrial and pharmacological application has been established so far.

The chemical reactions of the functional groups, a primary and a secondary hydroxyl groups and a double bond, have been investigated thoroughly.²⁾ Two types of E-ring expansion have been reported. One is the acid-catalyzed biogenetic type reaction in which the bond from C-21 to C-19 shifts to C-20, affording a germanicane-type compound, allobetulin (2).³⁾ The other occurs through dehydration of the 28-hydroxyl group⁴⁾ or solvolysis of the 28-p-toluenesulfonyloxy group,⁵⁾ in which the bond from C-22 to C-17 shifts to C-28 affording $22(17\rightarrow28)abeo$ -lupene derivatives (3).

As the compounds formed in the latter reaction have a new skeleton, further chemical modification might generate new pharmacological activities. The most important factor in designing chemical modification is the position of the double bonds. Therefore, the migration of the double bonds following the E-ring expansion was examined in detail. In this study, we prepared 28-p-toluenesulfonyloxylupane (4) and its 20(29)-ene derivative (7) as starting materials to avoid undesirable side reaction of the C-3 function and to make its easy to monitor the reaction by gas-liquid chromatography (GC).

Reaction of 28-p-Toluenesulfonyloxylupane (4) The reaction of **4** in N,N-dimethylaniline⁵⁾ has been reported to yield a single product, $22(17 \rightarrow 28)abeo$ -lup-17(28)-ene (5). Solvolysis of **4** in acetic acid containing 20% sodium

acetate also gave a single product, 5. The structure of 5 was confirmed by proton (¹H) and carbon-13 (¹³C) nuclear magnetic resonance (NMR) spectroscopy (Table I and Experimental) and chemical conversion of 8 to 5 (Experimental). On the other hand, solvolysis in acetic acid gave 6 as the main product. The ¹³C-NMR spectral data for 6 (Table I) showed the presence of a tetrasubstituted double bond in the molecule. In the ¹H-NMR spectrum of 6 in CDCl₃, proton signals of allylic methines at $\delta 2.02$ (1H, ddd, J=12.0, 8.1, 4.4 Hz) and 2.36 (1H, ddd, J=10.3, 4.8, 2.2 Hz) and of an allylic methylene at δ 1.79 (1H, ddd, J=15.0, 5.5, 2.9 Hz) and 2.68 (1H, ddd, J=15.0, 4.8, 2.2 Hz) were observed. The signal at 2.36 is coupled with the signal of C_{20} -H at 1.90 (1H, d-septet, J = 10.3, 6.6 Hz), and the signal at 2.68 showed the nuclear Overhauser effect (NOE) with the signals of the isopropyl group at 0.68 (3H, d, $J = 6.6 \,\text{Hz}$), 0.91 (3H, d, $J = 6.6 \,\text{Hz}$) and 1.90 (1H, d-septet, J = 10.3, 6.6 Hz) in NOE correlation spectroscopy (NOESY). These data indicated the position of the double bond to be C-13 and C-18. Confirmation of the structure, including the stereochemistry at C-17 (17 α -H), was obtained by chemical conversion of compound 11, which will be mentioned later, into 6 by catalytic hydrogenation. Thus, the structure of 6 was determined as (17S)-22 $(17\rightarrow28)$ abeo-lup-13(18)-ene. The result indicates that the double bond formed after the ring expansion migrated to a more stable position under acidic conditions.

Next, the reaction was examined in diethylene glycol, being monitored by thin-layer chromatography (TLC) and

© 1994 Pharmaceutical Society of Japan

TABLE I. 13C-NMR Data in CDCl₃

С	5	6	8	9	10	11	13
1	40.4	40.5	40.4	40.4	40.5	39.6	39.7
2	18.6	18.7	18.6	18.6	18.7	18.5	18.4
3	42.1	42.1	42.1	42.1	42.1	42.3	42.3
4	33.3	33.3	33.3	33.3	33.2	33.3	33.3
5	56.4	56.4	56.4	56.5	56.4	56.1	56.0
6	18.7	18.8	18.7	18.7	18.7	18.7	18.7
7	34.5	34.9	34.2	$34.5^{a)}$	35.8	32.5	32.6
8	41.1	41.2	41.1	41.3	41.1	40.5	40.6
9	50.1	50.8	50.5	51.1	50.8	54.2	54.0
10	37.5	37.6	37.4	37.5	37.7	37.1	37.0
11	20.8	21.6	21.2	21.2	21.4	125.5	125.3
12	25.8	25.3	26.6	25.8	26.2	125.4	125.0
13	43.3	133.0	46.1	42.3	131.6	132.1	134.0
14	42.6	43.9	42.3	42.3	43.1	41.6	41.7
15	34.1	26.2	33.8	34.1 ^{a)}	26.2	24.0	26.3
16	33.3	25.7	32.8	32.4	25.3	25.4	28.1
17	141.8	33.1	141.7	143.6	40.1	32.9	33.7
18	40.3	135.3	41.0	44.8	135.8	140.3	139.3
19	38.5	45.5	43.9	130.7	136.0	44.4	42.8
20	28.1	27.3	150.8	123.4	120.5	26.8	27.6
21	21.2^{a}	30.4	27.7	27.3	32.7	30.4	29.0
22	21.0^{a}	22.3	23.5	23.9	28.5	22.0	20.5
23	33.4	33.4	33.4	33.4	33.5	33.2	33.2
24	21.5	21.7	21.6	21.6	21.7	21.2	21.2
25	16.2	16.4	16.3	16.3	16.3	17.9	17.9
26	15.9	17.9	15.9	16.0	18.6	16.6	17.1
27	14.9	20.8	14.9	15.0	21.2	19.6	20.3
28	117.4	34.7	118.3	. 117.3	34.9	36.2	37.6
29	20.7^{b}	21.0	108.9	$19.9^{b)}$	18.7 ^{a)}	21.0	21.3^{a}
30	22.1 b)	21.0	22.4	21.0^{b}	21.6°	21.0	21.7 ^{a)}

a, b) Assignments may be interchanged in each column.

GC. Below $120\,^{\circ}$ C, the E-ring expansion did not occur. At about $125\,^{\circ}$ C, the reaction proceeded slowly, giving a mixture of 5 and 6 in a ratio of 5:1 after 15h. The ratio of 6 increased in proportion to the reaction temperature. At boiling point (245 °C), the product was practically all 6. In this reaction, p-toluenesulfonic acid, which was produced by the reaction is considered to catalyze the migration of the double bond. In fact, compound 5 in diethylene glycol without acid did not generate 6 even at boiling temperature. During these reactions, a putative intermediate, $22(17 \rightarrow 28)abeo$ -lup-17-ene, was not detected by TLC or GC.

In conclusion, elimination of the 28-p-toluenesulfonyloxy group causes E-ring expansion and forms a double bond between C-17 and C-28. Under acidic conditions, the double bond migrates to between C-13 and C-18.

Reaction of 28-p-Toluenesulfonyloxylup-20(29)-ene (7) Solvolysis of 7 has been reported to cause E-ring expansion

with a 17(28)-double bond, too. When the reaction was carried out in *N*,*N*-dimethylaniline, isomerization of the isopropenyl side chain to the isopropylidene chain was observed,⁵⁾ while in acetic acid containing sodium acetate, the isopropenyl side chain remained intact.⁶⁾ Under similar reaction conditions to the latter (acetic acid containing 20% sodium acetate), 7 gave a single product, 8. The structure of 8 was confirmed by ¹H- and ¹³C-NMR spectroscopic studies including ¹H-¹H shift correlation spectroscopy (COSY), ¹³C-¹H COSY and long-range ¹³C-¹H COSY.

On the other hand, the reaction in acetic acid caused migration of the double bonds after the E-ring expansion, giving many products. The reaction was monitored by GC (Fig. 3). In the first stage, the main product was compound 9, which was replaced by four compounds, 10, 11, 12 and 13, as the reaction proceeded. After 10 h, an equilibrium mixture was obtained which consisted mainly of 11, 12 and 13 in a ratio of 3:3:5.

Compounds 9—13 were isolated by column chromatography on silica gel impregnated with silver nitrate and preparative TLC. They are isomers with the molecular formula $C_{30}H_{48}$. As the physical properties and spectral data were identical with those reported,⁵⁾ the structure of 9 was determined to be 17(28),19-diene.

Compound 10 has a conjugated diene system [UV $\lambda_{\max}^{n-\text{hexane}}$ nm (log ε): 214 (4.43)] which consists of two tetrasubstituted double bonds [$^{13}\text{C-NMR}$ (CDCl₃) δ : 136.0, 135.8, 131.6, 120.5]. As one of them was assigned as an isopropylidene double bond [$^{1}\text{H-NMR}$ (CDCl₃) δ : 1.44 (3H, d, $J=2.0\,\text{Hz}$), 1.64 (3H, s)] and the other was correlated to the 14 α -methyl group in long-range $^{1}\text{H-}^{13}\text{C}$ COSY, the structure was determined as the 13(18),19-diene.

Compounds 11 and 13 are epimers having conjugated transoid dienes [UV $\lambda_{\max}^{\text{h-hexane}}$ nm (log ε): 243 (4.45), 252 (4.52), 261 (4.33) for 11 and 245 (4.17), 252 (4.22), 262 (4.04) for 13] consisting of *cis*-disubstituted and tetrasubstituted double bonds [1 H-NMR (CDCl₃) δ : 5.55 (1H, dd, J=10.5, 2.0 Hz) and 6.39 (1H, dd, J=10.5, 3.0 Hz) for 11 and 5.54 (1H, dd, J=10.5, 1.5 Hz) and 6.44 (1H, dd, J=10.5, 3.0 Hz) for 13]. The UV data are identical with those of oleane-11,13(18)-diene⁷⁾ and 11 and 13 were identified as 11,13(18)-diene compounds with the help of 1 H- 1 H COSY and long-range 1 H- 13 C COSY (Fig. 5). Either 11 or 13 gave the same equilibrium mixture of 11, 12 and 13 in boiling acetic acid containing 0.1% p-toluenesulfonic acid.

Compound 12 showed the presence of a conjugated

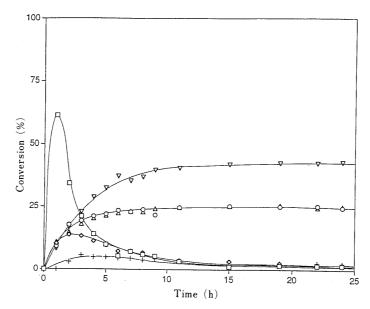


Fig. 3. Time Course of Each Product Formed from 7 in Boiling Acetic Acid

☐, compound 9; ⋄, compound 10; ○, compound 11; △, compound 12; ▽, compound 13; +, not determined.

diene [UV $\lambda_{\text{max}}^{n-\text{hexane}}$ nm (log ε): 239 (4.14), 246 (4.19), 254 (4.04)] consisting of a trisubstituted double bond and a tetrasubstituted double bond [$^1\text{H-NMR}$ (CDCl₃) δ : 5.53 (1H, br dd, J=5.3, 2.5 Hz); $^{13}\text{C-NMR}$ (CDCl₃) δ : 137.5 (C), 131.0 (C), 130.4 (C), 118.4 (CH)]. The UV data are the same as those of an ursa-12,17-diene derivative. Considering the above-mentioned interconversion reaction, the structures were determined as the 12,17-diene for 12 and 17-epimers of the 11,13(18)-diene for 11 and 13.

As each proton signal at C-12 of 11, 12 and 13 showed NOE with the proton signal at C-19, they were considered

to possess the same configuration at C-19, having the isopropyl side chain in a pseudoaxial position. On hydrogenation (H_2/Pt), compound 11 gave 6. Therefore the configuration at C-19 was determined as S (19 β -H). In NOESY, the proton signal at C-17 of 11 showed NOE with the methyl signals of the isopropyl side chain. Therefore, the configuration at C-17 was determined as S (17 α -H) for 11, and so R (17 β -H) for 13. This result was supported by 13 C-NMR data. In the case of the 17 α -H form, C-15 and C-28 are in a γ -gauche relationship which is expected to induce upfield shifts of their 13 C signals. 9

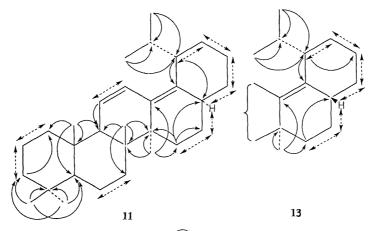


Fig. 5. ¹H-¹H COSY (←--→) and Long-Range ¹³C-¹H COSY (¹H → ¹³C) Connections for Compounds 11 and 13

In fact, the signals of C-15 and C-28 of 11 resonated at higher field than those of 13 by 2.3 and 1.4 ppm, respectively (Table I).

The reaction of 7 was also examined in diethylene glycol. Under reflux, 7 gave the equilibrium mixture of 11,12 and 13 after 1 h. At 125 °C, the reaction proceeded slowly and 9 was obtained as the main product after 2 h.

Experimental

Melting points were determined with a Yanagimoto micromelting apparatus and are uncorrected. Optical rotations were taken with a JASCO DIP-360 automatic polarimeter. The ¹H- and ¹³C-NMR spectra were measured with a JEOL GSX-500 spectrometer. Ultraviolet (UV) spectra were recorded on a Hitachi 323 spectrometer and infrared (IR) spectra on a Shimadzu IR-460 spectrometer. Mass spectra (MS) were measured with Hitachi M-80A and JEOL SX-102 spectrometers. GC was run on a Shimadzu GC-8A gas chromatograph using a Dexil 300GC column (30 m × 0.25 mm i.d.) at 300 °C. Nitrogen was used as the carrier gas.

Preparation of 28-p-Toluenesulfonyloxylupane (4) and 28-p-Toluenesulfonyloxylup-20(29)-ene (7) The sulfonates 4 and 7 were prepared by tosylation of lupan-28-ol and 20(29)-lupen-28-ol. 5) 4: colorless needles, mp 170—171 °C, $[\alpha]_D^{20}$ –11° $(c=1.0, \text{CHCl}_3)$. IR v_{max}^{KBr} cm⁻¹: 3450, 2950, 2860, 1600, 1460, 1380, 1360, 1180, 1170, 1100, 960, 840. UV λ_{max}^{MOH} nm (log ε): 338 (3.90). EI-MS m/z: 582 [M]⁺, 410, 231, 218, 191. HR-MS m/z: 582.4076. Calcd for C₃₇H₅₈O₃S: 582.4102. ¹H-NMR (CDCl₃) δ : 0.72 (3H, d, J = 6.6 Hz), 0.76 (3H, s), 0.79 (3H, s), 0.80 (3H, s), 0.80 (3H, s)d, $J = 6.6 \,\mathrm{Hz}$), 0.84 (3H, s), 0.89 (3H, s), 2.46 (3H, s), 3.73 (1H, d, J=9.5 Hz), 4.04 (1H, d, J=9.5 Hz), 7.35 (2H, d, J=8.4 Hz), 7.80 (2H, d, J=8.4 Hz). ¹³C-NMR (CDCl₃) δ : 144.6 (C), 132.9 (C), 129.8 (CH), 128.0 (CH), 69.5 (CH₂), 56.2 (CH), 49.9 (CH), 48.0 (CH), 46.9 (C), 44.4 (CH), 42.7 (C), 42.1 (CH₂), 40.9 (C), 40.2 (CH₂), 37.3 (C), 37.2 (C), 34.2 (CH₂), 34.1 (CH₂), 33.3 (CH₃), 33.2 (C), 29.4 (CH), 29.2 (CH₂), 26.7 (CH₂), 26.3 (CH₂), 22.8 (CH₃), 21.6 (CH₃), 21.3 (CH₂), 20.5 (CH₂), 18.6 (CH₂), 18.5 (CH₂), 16.0 (CH₃), 15.8 (CH₃), 14.8 (CH₃), 14.6 (CH₃). 7: a white amorphous powder, $[\alpha]_D^{20} + 50^\circ$ (c=1.0, CHCl₃). IR $v_{\text{max}}^{\text{KB}}$ cm⁻¹: 3050, 2940, 2880, 1640, 1595, 1460, 1390, 1365, 1190, 1175, 1100, 1020, 960. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 232 (4.09). EI-MS m/z: 580 [M]⁺, 408, 231, 216, 191. HR-MS m/z: 580.3949. Calcd for $C_{37}H_{56}O_3S$: 580.3947. ¹H-NMR (CDCl₃) δ : 0.75 (3H, s), 0.79 (6H, s), 0.83 (3H, s), 0. 91 (3H, s), 1.64 (3H, s), 2.46 (3H, s), 3.75 (1H, d, J=9.5 Hz), 4.06 (1H, dd, J=9.5 Hz) 9.5, 1.8 Hz), 4.56 (1H, dd, J=2.2, 1.5 Hz), 4.64 (1H, d, J=2.2 Hz), 7.35 (2H, d, J=8.0 Hz), 7.81 (2H, d, J=8.0 Hz). ¹³C-NMR (CDCl₃) δ : 149.7 (C), 144.7 (C), 132.8 (C), 129.9 (CH), 128.1 (CH), 110.0 (CH₂), 69.4 (CH₂), 56.3 (CH), 50.3 (CH), 48.7 (CH), 47.6 (CH), 46.7 (C), 42.6 (C), 42.1 (CH₂), 40.9 (C), 40.3 (CH₂), 37.6 (CH), 37.4 (C), 34.1 (CH₂), 34.0 (CH₂), 33.3 (CH₃), 33.2 (C), 29.3 (CH₂), 29.1 (CH₂), 26.5 (CH₂), 25.1 (CH₂), 21.6 (CH₃), 21.5 (CH₃), 20.5 (CH₂), 19.0 (CH₃), 18.7 (CH₂), 18.5 (CH₂), 16.0 (CH₃), 15.8 (CH₃), 14.7 (CH₃).

Reaction of 4 in Acetic Acid Containing 20% Sodium Acetate A solution of the p-toluenesulfonate 4 (30 mg) in acetic acid containing

20% sodium acetate (10 ml) was heated under reflux for 3 h. After cooling, the reaction mixture was poured into ice-water and extracted with ether. The organic layer was washed successively with water, 10% sodium carbonate solution and water, dried, and evaporated *in vacuo*. The residue was crystallized from CHCl₃–MeOH to afford 5 (15 mg). Colorless needles, mp 185—186 °C, $[\alpha]_{\rm D}^{20}$ – 49° (c=1.0, CHCl₃). IR $v_{\rm max}^{\rm KB}$ cm $^{-1}$: 2950, 2880, 1460, 1445, 1380, 1370, 1360, 1345, 1195, 1080, 1040, 1010, 975, 800, 640. EI-MS m/z: 410 [M] $^+$, 395 [M – CH₃] $^+$, 367, 231, 191. HR-MS m/z: 410.3910. Calcd for C₃₀H₅₀: 410.3910. 1 H-NMR (CDCl₃) δ : 0.79 (3H, s), 0.83 (3H, s), 0.85 (3H, s), 0.87 (3H, d, J=6.7 Hz), 0.88 (3H, d, J=6.7 Hz), 0.95 (3H, s), 1.06 (3H, s), 5.32 (1H, d, J=4.3 Hz). 13 C-NMR (CDCl₃): Table I.

Reaction of 4 in Acetic Acid A solution of the 28-*p*-toluenesulfonate 4 (500 mg) in acetic acid (50 ml) was heated under reflux for 5 h. After cooling, the reaction mixture was poured into ice-water and extracted with ether. The extract was washed with water, 10% sodium carbonate solution and water, then dried and concentrated *in vacuo*. The residue was crystallized from acetone to afford 6 (220 mg). Colorless needles, mp 207—208 °C, $[\alpha]_D^{21}$ –92° (c=1.0, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 3000, 2930, 2850, 1460, 1440, 1380, 1360, 1210, 1160, 1115, 1080, 1030, 1000, 980. EI-MS m/z: 410 [M]⁺, 395 [M—CH₃]⁺, 367, 229, 218, 190. HR-MS m/z: 410,3900. Calcd for C₃₀H₅₀: 410.3910. ¹H-NMR (CDCl₃) δ: 0.68 (3H, d, J=6.6 Hz), 0.80 (3H, s), 0.86 (3H, s), 0.87 (3H, s), 9.19 (3H, s), 0.91 (3H, dd, J=5.6 Hz), 1.13 (3H, s), 2.36 (1H, ddd, J=10.3, 4.8, 2.2 Hz), 2.68 (1H, ddd, J=15.0, 4.8, 2.2 Hz). ¹³C-NMR (CDCl₃): Table I.

Reaction of 4 in Diethylene Glycol A solution of the 28-p-toluenesulfonate 4 (100 mg) in diethylene glycol (25 ml) was heated under reflux for 3 h. After cooling, the reaction mixture was diluted with water and extracted with ether. The extract was washed with water, dried, and concentrated *in vacuo*. The residue was crystallized from acetone to afford 6 (40 mg).

Reaction of 7 in Acetic Acid Containing 20% Sodium Acetate A solution of the 28-p-toluenesulfonate 7 (30 mg) in acetic acid containing 20% sodium acetate (10 ml) was heated under reflux for 3 h. After cooling, the reaction mixture was poured into ice-water and extracted with ether. The extract was washed successively with water, 10% sodium carbonate solution and water, then dried and concentrated in vacuo. The residue was crystallized from CHCl₃-MeOH to give 8 (13 mg). Colorless needles, mp 144—145 °C, $[\alpha]_D^{21} - 26^\circ$ (c=1.0, CHCl₃). IR $v_{max}^{\rm Em}$ cm⁻¹: 3070, 3000, 2930, 2880, 1640, 1445, 1380, 1370, 1040, 975, 885, 805. EI-MS m/z: 408 [M]⁺, 393 [M-CH₃]⁺, 231, 216, 191. HR-MS m/z: 408.3758. Calcd for $C_{30}H_{48}$: 408.3753. ¹H-NMR (CDCl₃) δ : 0.79 (3H, s), 0.83 (3H, s), 0.85 (3H, s), 0.95 (3H, s), 1.03 (3H, s), 1.77 (3H, s), 4.65 (1H, s), 4.73 (1H, s), 5.35 (1H, t, J=4 Hz). ¹³C-NMR (CDCl₃): Table I.

Reaction of 7 in Acetic Acid i) A solution of the 28-p-toluenesulfonate 7 (1 g) in acetic acid (140 ml) was heated under reflux for 1 h. After cooling, the reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was washed with 10% sodium carbonate solution and water, then dried and concentrated *in vacuo*. The residue was purified by chromatography on 20% AgNO₃-impregnated silica gel (n-hexane) followed by recrystallization (benzene–MeOH) to give 9 (180 mg) and 10 (39 mg). 9: colorless needles, mp 230—231.5 °C, $[\alpha]_b^{27}$ – 164° (c = 0.5, CHCl₃). IR v_{max}^{KBr} cm⁻¹: 2920, 1436, 1373, 1035, 855, 832,

803, 670. EI-MS m/z: 408 [M]⁺, 231, 201, 191, 148, 135. HR-MS m/z: 408.3749. Calcd for $C_{30}H_{48}$: 408.3752. ¹H-NMR (CDCl₃) δ : 0.79 (3H, s), 0.82 (3H, s), 0.85 (3H, s), 0.96 (3H, s), 1.13 (3H, s), 1.64 (3H, d, J=1.5 Hz), 1.69 (3H, s), 2.90 (1H, d, J=11.4 Hz), 5.33 (1H, d, J=5.5 Hz). ¹³C-NMR (CDCl₃): Table I. 10: colorless needles, mp 248—249.5°C, [α] $_{28}^{2B}$ -50° (c=0.52, CHCl₃). IR ν_{\max}^{KBr} cm⁻¹: 2900, 2830, 1440, 1360, 1195, 1160, 1115. UV $\lambda_{\max}^{n-hexane}$ nm (log ε): 214 (4.43). EI-MS m/z: 408 [M]⁺, 393 [M-CH₃]⁺, 365, 203, 191, 145. HR-MS m/z: 408.3749. Calcd for $C_{30}H_{48}$: 408.3752. ¹H-NMR (CDCl₃) δ : 0.80 (3H, s), 0.866 (3H, s), 0.869 (3H, s), 1.14 (3H, s), 1.44 (3H, d, J=1.8 Hz), 1.64 (3H, s), 2.31 (1H, ddd, J=14.7, 5.1, 1.8 Hz), 2.64 (1H, dt, J=12.5, 3.3 Hz). ¹³C-NMR (CDCl₃): Table I.

ii) A solution of the 28-p-toluenesulfonate 7 (1 g) in acetic acid (140 ml) was heated under reflux for 11 h. After cooling, the reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was washed with 10% sodium carbonate solution and water, then dried and concentrated in vacuo. The residue was purified by chromatography on 20% AgNO₃-impregnated silica gel (n-hexane) and preparative TLC (petroleum ether) to give 11 (20 mg), 12 (26 mg) and 13 (111 mg). 11: colorless needles, mp 212—214.5 °C, $[\alpha]_D^{20}$ –123° (c=0.4, CHCl₃). IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2910, 2850, 1450, 1370, 1360, 1205, 620. EI-MS m/z: 408 [M]⁺, 393 [M–CH₃]⁺, 365, 269, 227, 187. HR-MS m/z: 408.3749. Calcd for C₃₀H₄₈: 408.3753. UV $\lambda_{\max}^{n-\text{bexane}}$ nm (log ε): 243 (4.45), 252 (4.52), 261 (4.33). ¹H-NMR (CDCl₃) δ : 0.74 (3H, d, J=6.6Hz, H₃-30), 0.76 (3H, s, H₃-26), 0.81 (3H, s, H₃-24), 0.87 (3H, s, H₃-23), 0.90 (3H, s, H_3 -25), 0.93 (3H, s, H_3 -27), 0.93 (3H, d, J=6.6 Hz, H_3 -29), 2.46 (1H, ddd, J=10.7, 4.3, 2.7 Hz, H-19), 5.55 (1H, dd, J=10.5, 2.0 Hz, H-11), 6.39 (1H, dd, J=10.5, 3.0 Hz, H-12). ¹³C-NMR (CDCl₃): Table I. 12: a white amorphous powder, $[\alpha]_D^{24}$ +66.8° (c=0.52, CHCl₃). IR $\nu_{max}^{CHCl_3}$ cm⁻¹: 2930, 1451, 1375, 1360, 1110, 1086. EI-MS m/z: 408 [M]⁺, 393 $[M-CH_3]^+$, 365, 216, 173, 95. HR-MS m/z: 408.3747. Calcd for $C_{30}H_{48}$: 408.3753. UV $\lambda_{\text{max}}^{\text{n-hexane}}$ nm (log ε): 239 (4.14), 246 (4.19), 254 (4.04). ¹H-NMR (CDCl₃) δ : 0.78 (3H, d, J=7.0 Hz), 0.83 (3H, s), 0.88 (6H, s), 0.88 (3H, d, J = 7.0 Hz), 0.97 (3H, s), 1.00 (3H, s), 5.53 (1H, dd, J = 5.5,2.7 Hz). 13 C-NMR (CDCl₃) δ : 137.5 (C), 131.0 (C), 130.4 (C), 118.4 (CH), 56.5 (CH), 47.4 (CH), 42.0 (CH₂), 41.3 (C), 40.6 (CH₂), 39.1 (C), 37.2 (C), 36.7 (CH), 34.0 (CH₂), 33.5 (CH₃), 33.2 (C), 31.0 (CH₂), 30.1

(CH), 29.2 (CH₂), 27.7 (CH₂), 24.1 (CH₂), 23.5 (CH₂), 21.8 (CH₃), 21.7 (CH₃), 20.7 (CH₃), 19.9 (CH₃), 19.2 (CH₂), 18.7 (CH₂), 18.6 (CH₂), 17.1 (CH₃), 16.1 (CH₃). **13**: a white amorphous powder, $[\alpha]_b^{25} - 6.6^\circ$ (c = 0.71, CHCl₃). IR $v_{\rm max}^{\rm KB}$ cm⁻¹: 2910, 2860, 1608, 1459, 1380, 1362, 1205, 1060, 840, 775, 653. EI-MS m/z: 408 [M]⁺, 393 [M - CH₃]⁺, 365, 297, 269, 229, 203, 187, 145, 119, 95. HR-MS m/z: 408.3768. Calcd for C₃₀H₄₈: 408.3754. UV $\lambda_{\rm max}^{\rm n-hexane}$ nm (log ε): 245 (4.17), 252 (4.22), 262 (4.04). ¹H-NMR (CDCl₃) δ : 0.74 (3H, d, J = 6.7 Hz, H₃-30), 0.79 (3H, s, H₃-26), 0.81 (3H, s, H₃-24), 0.86 (3H, s, H₃-23), 0.90 (3H, s, H₃-25), 0.93 (3H, d, J = 6.7 Hz, H₃-29), 0.95 (3H, s, H₃-27), 2.45 (1H, dt, J = 10.0, 3.1 Hz, H₃-19), 5.54 (1H, dd, J = 10.5, 1.5 Hz, H-11), 6.44 (1H, dd, J = 10.5, 3.0 Hz, H-12). ¹³C-NMR (CDCl₃): Table I.

Catalytic Hydrogenation of Compound 11 A solution of 11 (39 mg) in AcOEt (10 ml) and EtOH (5 ml) was stirred with PtO₂ (20 mg) under reflux for 6 h and at room temperature for 7 h in an atmosphere of hydrogen. Then the mixture was filtered, and the filtrate was concentrated in vacuo. The residue was purified by chromatography on 20% AgNO₃-impregnated silica gel (n-hexane) to afford 6 (7 mg) and 11 (26 mg).

References and Notes

- E. W. H. Hayek, U. Jordis, W. Moche, F. Sauter, *Phytochemistry*, 28, 2229 (1989).
- 2) P. Jääskeläinen, Paperi ja puu, 10, 599 (1981).
- 3) D. H. R. Barton, N. I. Holness, J. Chem. Soc., 1952, 78.
- A. Vystrčil, V. Křeček, M. Buděšínský, Collection Czech. Chem. Commun., 39, 2494 (1974).
- A. Vystrčil, V. Křeček, M. Buděšínský, Collect. Czech. Chem. Commun., 48, 1499 (1983).
- A. S. R. Anjaneyulu, M. Narayana Rao, A. Sree, V. Suryanarayana Murty, *Indian J. Chem.*, 19B, 735 (1980).
- E. J. Corey, H. J. Hess, S. Proskow, J. Am. Chem. Soc., 85, 3979 (1963).
- 8) D. H. R. Barton, H. T. Cheung, P. J. L. Daniels, K. G. Lewis, J. F. Mcghie, J. Chem. Soc., 1962, 5163.
- E. Breitmaier, W. Voelter, "Carbon-13 NMR Spectroscopy," 3rd ed., VCH Publishers, Weinheim, 1987, p. 115.