Structures of New Eremophilane Derivatives from the Rhizomes of *Petasites japonicus* MAXIM.¹⁾

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Eight new eremophilenolides, 6β -angeloyloxy- 3β ,9α-dihydroxyeremophil-7(11)-en-12,8β-olide (1), 6β -angeloyloxy- 3β ,9β-dihydroxyeremophil-7(11)-en-12,8β-olide (2), 6β -angeloyloxy- 3β ,8β,9β-trihydroxyeremophil-7(11)-en-12,8α-olide (3), 6β -(3'-chloro-2'-hydroxy-2'-methylbutyloyloxy)- 3β -hydroxyeremophil-7(11)-en-12,8β-olide (4), 6β -epoxyangeloyloxy- 3β -hydroxyeremophil-7(11)-en-12,8β-olide (5), 3β -hydroxy- 6α -methoxyeremophil-7(11)-en-12,8α-olide (6), 3β -hydroxy- 6β -methoxyeremophil-7(11)-en-12,8α-olide (7) and 3β -hydroxy- 3β -oxoeremophil-7(11)-en-12,8α-olide (8) were isolated from the dried rhizomes of *Petasites japonicus* Maxim. (Compositae) together with a new eremophilane-type sesquiterpenoid, 3β -hydroxy- 3β -oxoeremophil-6-en-12-oic acid methyl ester (9). Compound 4 is the first chlorine-containing eremophilenolide derivative isolated from natural sources. The structures of these compounds were elucidated by spectroscopy.

Key words Petasites japonicus; Compositae; eremophilenolide; eremophilane-type sesquiterpenoid

The rhizomes of *Petasites japonicus* MAXIM. (Fuki in Japanese, Compositae) have been used for the treatment of tonsillitis, contusions and poisonous-snake bite in China.²⁾ In previous papers, we reported the structural elucidation of nor-sesquiterpenoid,¹⁾ eremophilenolides,³⁾ triterpenoids,⁴⁾ anthraquinones⁴⁾ and phenolic compounds.⁵⁾ We now wish to report the isolation and structural elucidation of eight new eremophilenolides, 6β -angeloyloxy- 3β , 9α -dihydroxyeremophil-7(11)-en-12, 8β -olide (1), 6β -angeloyloxy- 3β , 9β -dihydroxyeremophil-7(11)-en-12, 8β -olide (2), 6β -angeloyloxy- 3β , 8β , 9β -trihydroxyeremophil-7(11)-en-12, 8β -olide (3), 6β -(3'-chloro-2'-hydroxy-2'-methylbutyloyloxy)- 3β -hydroxyeremophil-7(11)-en-12, 8β -olide (5), 3β -hydroxy- 6α -methoxyeremophil-7(11)-en-12, 8β -olide (6), 3β -hydroxy- 6α -methoxyeremophil- α -methoxyeremophil-

methoxyeremophil-7(11)-en-12,8 α -olide (7) and 8 β -hydroxy-3-oxoeremophil-7(11)-en-12,8 α -olide (8), as well as a new eremophilane-type sesquiterpenoid, 3 β -hydroxy-8-oxoeremophil-6-en-12-oic acid methyl ester (9). Extraction and isolation were carried out as described in the Experimental section.

Compound 1 was isolated as a colorless oil, $[\alpha]_D - 32.4^\circ$. The molecular formula was determined to be $C_{20}H_{28}O_6$ by high-resolution (HR)-MS. The IR spectrum of 1 suggested the presence of a hydroxyl group (3423 cm⁻¹), an α,β -unsaturated- γ -lactone (1752, 1681 cm⁻¹) and an α,β -unsaturated ester (1719, 1646 cm⁻¹). The ¹H- (Table 1) and ¹³C-NMR (Table 2) spectra indicated the presence of a tertiary methyl group $[\delta_H 0.97 \text{ (s, H-15)}, \delta_C 20.8 \text{ (C-15)}]$, a secondary methyl group $[\delta_H 0.98 \text{ (d, } J=7.0 \text{ Hz, H-14)}, \delta_C 7.3 \text{ (C-14)}]$ and an olefinic methyl group $[\delta_H$

Chart 1

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Table 1. ¹H-NMR Chemical Shifts (CDCl₃, 400 MHz)

Proton	1	2	3a)	4	5	6	7	8	9
1	α 1.39 dddd (13.6, 13.2, 13.2, 4.0)		***				/		
	β 2.17 dddd								
2	(13.6, 4.8, 4.0, 3.7) β 1.54 dddd								
2	(13.2, 12.8, 11.4, 4.0)								
3	4.12 ddd	4.08 ddd	3.85 ddd	4.03 ddd	4.15 ddd	4.74 ddd	3.79 ddd		3.74 ddd
,	(11.7, 4.4, 4.4)	(11.7, 4.4, 4.4)	(2.6, 2.6, 2.6)	(10.7, 4.4, 4.4)	(11.4, 4.4, 4.4)	(11.7, 4.8, 4.8)	(2.9, 2.9, 2.9)		(9.9, 4.4, 4.4)
4	1.93 m	(1117, 111, 111)	(2.0, 2.0, 2.0)	(,,	(,,	1.94 m	(===,===,===,	2.37q (6.6)	1.88 qd (7.0, 4.4)
6	6.19 br s	6.12 br s	5.56 s	5.96 br s	6.26 br s	3.88 s	4.01 s	$\alpha 2.74 d$	6.47 br s
								(13.9)	
								$\beta 2.28 dd$	
0	4.021 1.00.20	4.051		4.00	4.97	5.01 m	4.93 ddg	(13.9, 1.7)	
8	4.83 br d (9.2)	4.95 br s		4,89 m	4.87 m	3.01 m	4.93 daq (11.0, 7.0, 1.8)		
9	3.73 dd (9.2, 5.1)	4.24 dd	3.81 d	2.24 ddd	2.22 ddd	2.22 ddd	(11.0, 7.0, 1.0)		α 2.30 dd
,	3.73 dd (9.2, 3.1)	(4.4, 2.2)	(10.6)	(12.7, 6.4, 2.0)	(12.8, 6.6, 2.2)	(12.8, 6.6, 2.2)			(17.2, 4.0)
		(4.4, 2.2)	(10.0)	(12.7, 0.1, 2.0)	(12.0, 0.0, 2.2)	(12.0, 0.0, 2.2)			β 2.66 dd
									(17.2, 4.8)
10									2.03 m
11									3.62 qd
									(7.3, 1.1)
13	1.80 dd	1.84 dd	2.02 s	1.85 dd	1.91 dd	1.89 d (1.8)	1.89 d (1.8)	1.87 d	1.28 d (7.3)
	(1.5, 1.5)	(1.8, 1.8)		(1.5, 1.5)	(1.8, 1.8)			(1.7)	
14	0.98 d (7.0)	0.98 d (7.3)	1.06 d (7.0)	1.00 d (7.3)	0.98 d (7.3)	0.99 d (7.0)	1.01 d (7.0)	0.97 d (6.6)	0.99 d (7.0)
15	0.97 s	1.12 s	1.36 s	0.98 s	0.97 s	0.80 s	1.34 s	1.00 s	1.24 s
3′	6.29 gg (7.3, 1.5)	6.28 qq	6.13 gg	4.35 q (6.3)	3.17 q (5.1)				
	11() /	(7.3, 1.5)	(7.3, 1.5)	• ` '	• • •				
4′	2.07 dq (7.3, 1.5)	2.07 dq	2.02 dq	1.64 d (6.3)	1.39 d (5.1)				
		(7.3, 1.5)	(7.3, 1.5)						
5′	1.98 dq (1.5, 1.5)	1.99 dq	1.90 dq	1.54 s	1.66 s				
		(1.5, 1.5)	(1.5, 1.5)						
OCH_3						3.20 s	3.21 s		3.66 s

Coupling constants (J in Hz) are given in parentheses. a) In CDCl₃ with small amounts of CD₃OD.

Table 2. 13C-NMR Chemical Shifts (CDCl₃, 100 MHz)

Carbon	1	2	3 ^{a)}	4	5	6	7	8	9
1	20.6 ^{b)}	25.8	15.6	27.1	27.2	28.4	20.8	26.5	26.5
2	28.2	28.1	27.6	27.8	28.6	29.5	27.8	36.7	28.0
2 3	68.1	67.9	71.5	67.7	67.8	69.5	71.8	211,3	70.6
4	39.1	39.7	34.6	39.2	39.1	43.6	33.3	45.1	44.8
5	45.2	45.7	41.8	45.8	45.9	43.8	42.8	45.2	40.1
6	70.8	71.3	70.2	75.2	73.0	84.3	79.0	34.6	154.3
7	154.9	155.2	150.4	157.4	157.3	160.1	160.3	157.1	136.6
8	82.3	79.0	104.9	77.1°)	77.1	76.3	78.2	103.0	197.6
9	73.6	72.8	72.3	34.6	34.3	35.3	34.7	38.9^{f}	40.1
10	41.0	40.8	42.5	35.1	35.3	35.7	34.3	38.9^{f}	35.7
11	123.8	123.6	130.5	122.3	122.9	124.9	124.4	123.9	37.8
12	173.9	174.1	171.2	174.1^{d}	173.9	174.3	174.3	171.5	175.1
13	8.3	8.4	9.1	9.0	8.8	8.3	8.6	8.3	16.0
14	7.3	7.5	12.9	7.5	7.4	7.9	12.8	7.7	7.4
15	20.8	22.2	19.0	20.0	20.2	24.7	18.8	12.3	25.2
1′	166.6	166.7	167.1	174.1 ^{d)}	169.3				
2'	126.3	126.5	127.3	77.1°)	$60.4^{e)}$				
3′	142.2	142.0	139.8	63.2	60.4 ^{e)}				
4′	16.1	16.1	15.9	18.0	13.6				
5′	20.6^{b}	20.6	20.6	22.5	19.5				
OCH ₃						57.3	57.0		52.0

a) In CDCl₃ with small amounts of CD₃OD. b—f) Signals were overlapped.

1.80 (dd, J=1.5, 1.5 Hz, H-13), $\delta_{\rm C}$ 8.3 (C-13)], which are characteristic of an eremophilenolide derivative.³⁾ Other signals in the ¹H- and ¹³C-NMR spectra, obtained with the aid of ¹H-¹H shift correlation spectroscopy (¹H-¹H COSY), ¹³C-¹H shift correlation spectroscopy (¹³C-¹H

COSY) and ¹H-detected heteronuclear multiple bond correlation (HMBC) spectrum, indicated to two hydroxybearing methines [$\delta_{\rm H}$ 3.73 (dd, J=9.2, 5.1 Hz, H-9 β), 4.12 (ddd, J=11.7, 4.4, 4.4 Hz, H-3 α), $\delta_{\rm C}$ 73.6 (C-9), 68.1 (C-3)], an oxygenated methine [$\delta_{\rm H}$ 4.83 (br d, J=9.2 Hz,

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H-8 α), $\delta_{\rm C}$ 82.3 (C-8)], an angeloyloxyl group [$\delta_{\rm H}$ 1.98 (dq, J = 1.5, 1.5 Hz, H-5'), 2.07 (dq, J = 7.3, 1.5 Hz, H-4'), 6.29 $(qq, J=7.3, 1.5 Hz, H-3'), \delta_C 16.1 (C-4'), 20.6 (C-5'), 126.3$ (C-2'), 142.2 (C-3'), 166.6 (C-1')], an angeloyloxy-bearing methine [δ_H 6.19 (br s, H-6 α), δ_C 70.8 (C-6)] and an α,β -unsaturated- γ -lactone [$\delta_{\rm C}$ 123.8 (C-11), 154.9 (C-7), 173.9 (C-12)]. These spectral data and molecular formula suggested that the most likely structure of this compound is 1, as illustrated. The stereostructure was determined from the nuclear Overhauser effect correlation spectroscopy (NOESY) spectrum. The NOESY cross-peaks observed between H-6 α and H-1 α : H-6 α and H-3 α : H-6 α and H-8α, and between H-8α and H-1α implied a cisjunction of the A/B rings which had a non-steroidal conformation (Fig. 1). The coupling patterns and constants for H-3 [$\delta_{\rm H}$ 4.12 (ddd, J=11.7, 4.4, 4.4 Hz)] and H-9 [$\delta_{\rm H}$ 3.73 (dd, J=9.2, 5.1 Hz)] suggested that the hydroxyl groups at C-3 and C-9 are β and α configurations, respectively, and thus was supported by the NOESY cross-peaks between H-3 α and H-6 α , and between H-9 β and H-15 (Fig. 1). The configuration of the angeloyloxyl group at C-6 was determined to be β from the NOESY spectrum, in which cross-peaks were observed between H-6 α and H-3 α , and between H-6 α and H-8 α (Fig. 1). On the basis of the above evidence, the structure of 1 was determined to be 6β -angeloyloxy- 3β , 9α -dihydroxyeremophil-7(11)-en-12,8 β -olide.

Compound 2 was isolated as a colorless oil, $[\alpha]_D - 39.9^\circ$. The molecular formula was determined to be C₂₀H₂₈O₆ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3442 cm⁻¹), an α,β -unsaturated- γ -lactone $(1756 \,\mathrm{cm}^{-1})$ and an α,β -unsaturated ester (1718, 1646) cm⁻¹). The ¹H- and ¹³C-NMR spectra of 2 were closely related to those of 1. Naya et al. 6) reported homoallylic coupling $(J=1.0-1.8 \,\mathrm{Hz})$ between the olefinic methyl group (H-13) and H-6α found in the 8α-methoxyeremophil-7(11)-en-12,8 β -olide derivatives, which had a non-steroidal conformation, while this long-range coupling was absent in 8β -methoxyeremophil-7(11)-en-12,8 α olide derivatives, which had a steroidal conformation. The ¹H-NMR spectrum of 2 showed the homoallylic coupling (J=1.8 Hz) of the olefinic methyl group (H-13) with H-6 α . Thus, 2 exists in a non-steroidal conformation. In the

¹H-NMR spectrum of **2**, the coupling pattern and constants for H-9 [$\delta_{\rm H}$ 4.24 (dd, J=4.4, 2.2 Hz)] suggested that the hydroxyl group at C-9 has a β configuration. In the ¹³C-NMR spectrum of **2**, the C-15 signal appeared at δ 22.2, 1.4 ppm lower field than that of **1**. This is due to the δ_1 -hydroxy substituent effect⁷⁾ of the 9 β -hydroxyl group, which adopts a 1,3-syn periplanar arrangement with respect to the C-15. Thus, **2** was the 9-epimer of **1**. Based on this evidences, the structure of **2** was determined to be 6 β -angeloyloxy-3 β ,9 β -dihydroxyeremophil-7(11)-en-12,8 β -olide.

Compound 3 was isolated as an amorphous powder, $[\alpha]_D$ +41.2°. The molecular formula was determined to be $C_{20}H_{28}O_7$ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3445 cm⁻¹), an α,β -unsaturated- γ -lactone (1757 cm⁻¹) and an α,β -unsaturated ester (1718, 1644 cm⁻¹). The ¹H- and ¹³C-NMR spectra of 3 were virtually identical to those of 6β -angeloyloxy- 3β , 8β -dihydroxyeremophil-7(11)-en-12, 8α -olide $^{3\alpha$, $^{8)}$ cept for the presence of one more hydroxyl group. In the ¹H-NMR spectrum of 3, the homoallylic coupling between the olefinic methyl group (H-13) and H-6 α is lacking. In the NOESY spectrum, nuclear Overhauser effects (NOEs) were seen between H-6 α and H-13; H-6 α and H-14; between H-6α and H-15. These data indicate that 3 exists in a steroidal conformation. The ¹³C-NMR spectrum of 3 showed a signal due to a hemi-ketal carbon [$\delta_{\rm C}$ 104.9 (C-8)], suggesting that the lactone was γ -hydroxylated. The hydroxyl group at C-8 has a β configuration, which was supported by the NOESY cross-peaks between H-6α and H-13; H-6 α and H-14; between H-6 α and H-15. In the HMBC spectrum, the hydroxy-bearing methine at δ 3.81 showed connectivity to C-1 at δ 15.6 and C-10 at δ 42.5, indicating that this hydroxyl group is attached at the C-9. The coupling pattern and constant for H-9 [$\delta_{\rm H}$ 3.81 (d, $J = 10.6 \,\mathrm{Hz}$)] suggested that the hydroxyl group at C-9 has a β configuration. From the above data, the structure of 3 was determined to be 6β -angeloyloxy- 3β , 8β , 9β trihydroxyeremophil-7(11)-en-12,8α-olide. Compounds 1-3 are the first eremophilenolide derivatives isolated from the genus Petasites plants having the 9-hydroxyl group.

Compound 4 was isolated as a colorless oil, $[\alpha]_D - 55.6^\circ$.

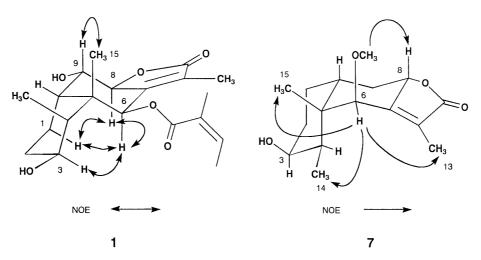


Fig. 1

The electron ionization (EI)-MS of 4 showed a pair of molecular ion peaks at m/z 400 and 402 in a 3:1 ratio, indicating the presence of a chlorine atom in the molecule. The molecular formula was determined to be C₂₀H₂₉ClO₆ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3447 cm⁻¹), a saturated ester (1750 cm⁻¹) and an α,β -unsaturated- γ lactone (1750 cm⁻¹). The ¹H- and ¹³C-NMR spectra of 4 were virtually identical to those of 6β -angeloyloxy- 3β -hydroxyeremophil-7(11)en-12,8 β -olide (10). The ¹H-NMR spectrum of 4 showed the homoallylic coupling (J=1.5 Hz) of the olefinic methyl group (H-13) with H-6 α . In the NOESY spectrum, NOEs were seen between H-3 α and H-6 α ; and between H-6 α and H-8 α . These data indicate that 4 exists in a non-steroidal conformation. The ¹H- and ¹³C-NMR spectra of 4 showed the presence of a 3'-chloro-2'-hydroxy-2'-methylbutyloyloxyl group $[\delta_{\rm H} 1.54 \text{ (s, H-5')}, 1.64 \text{ (d, } J=6.3 \text{ Hz, H-4'}),$ 4.35 (q, J = 6.3 Hz, H-3'), δ_C 18.0 (C-4'), 22.5 (C-5'), 63.2 (C-3'), 77.1 (C-2'), 174.1 (C-1')]⁹⁾ in place of the angeloyloxyl group of 10. The EI-MS gave a fragment ion peak at m/z 248, indicating loss of 3'-chloro-2'-hydroxy-2'-methylbutylic acid from the molecular ion peak at m/z400. The structure of 3'-chloro-2'-hydroxy-2'-methylbutyloyloxyl group was confirmed by the HMBC spectrum, in which the tertiary methyl group (H-5') showed connectivity to C-1', C-2' and C-3', and the secondary methyl group (H-4') showed connectivity to C-2' and C-3'. The absolute configuration of C-2' and C-3' could not be determined. Based on this evidence, the structure of 4 was determined to be 6β -(3'-chloro-2'-hydroxy-2'-methylbutyloyloxy)-3 β hydroxyeremophil-7(11)-en-12,8 β -olide. Compound 4 is the first chlorine-containing eremophilenolide derivative isolated from natural sources.

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Compound 5 was isolated as a colorless oil, $[\alpha]_D - 96.2^\circ$. The molecular formula was determined to be C₂₀H₂₈O₆ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3423 cm⁻¹), a saturated ester (1750 cm⁻¹) and an α,β -unsaturated- γ -lactone (1750 cm⁻¹). In comparing the ¹H- and ¹³C-NMR spectra of 5 with those of 4, the main skeleton of 5 was in accord with 4. The ¹H-NMR spectrum of 5 showed homoallylic coupling (J=1.8 Hz) of the olefinic methyl group (H-13) with H-6α. Thus, 5 exists in a non-steroidal conformation. The ¹Hand ¹³C-NMR spectra of 5 showed the presence of an epoxyangeloyloxyl group [$\delta_{\rm H}$ 1.39 (d, $J=5.1\,{\rm Hz}, {\rm H}-4'$), 1.66 (s, H-5'), 3.17 (q, J = 5.1 Hz, H-3'), $\delta_{\rm C}$ 13.6 (C-4'), 19.5 (C-5'), 60.4 (C-2' and C-3'), 169.3 (C-1')]¹⁰⁾ in place of a 3'-chloro-2'-hydroxy-2'-methylbutyloyloxyl group of 4. The EI-MS of 5 showed a fragment ion peak at m/z 248, indicating loss of epoxyangelic acid from the molecular ion peak at m/z 364. Comparison of ¹H- and ¹³C-NMR data for epoxyangeloyloxyl group with publishid data¹⁰⁾ confirmed the structure of the epoxyangeloyloxyl moiety of 5. The structure of this acyl group was further confirmed by the HMBC spectrum, in which the tertiary methyl group (H-5') showed connectivity to C-1', C-2' and C-3', and the secondary methyl group (H-4') showed connectivity to C-2' and C-3'. The absolute configuration of C-2' and C-3' could not be determined. From the above data, the structure of 5 was determined to be 6β -epoxyangeloyloxy- 3β -hydroxyeremophil-7(11)-en-12,8 β -olide. Compound 5 is the first eremophilenolide derivative with an epoxyangeloyloxyl group isolated from natural sources.

Compound 6 was isolated as a colorless oil, $[\alpha]_D = 54.4^\circ$. The molecular formula was determined to be C₁₆H₂₄O₄ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3475 cm⁻¹) and an α,β -unsaturated- γ lactone (1751 cm⁻¹). Detailed analysis of the ¹H- and ¹³C-NMR spectra of 6 revealed a gross structure, including conformation, identical with that of 3β -hydroxy- 6β methoxyeremophil-7(11)-en-12,8 β -olide (11), which had a non-steroidal conformation. 3b) However, the chemical shift and coupling pattern for the olefinic methyl group (H-13) differ significantly between the two compounds. The ¹H-NMR spectrum of 11^{3b)} showed the homoallylic coupling of the olefinic methyl group [δ 1.97 (dd, J=1.5, 1.5 Hz, H-13)] with H-6α. In addition, in the ¹H-NMR spectrum of 6, the homoallylic coupling between the olefinic methyl group [δ 1.89 (d, J=1.8 Hz, H-13)] and H-6α is lacking. In the NOE difference spectra, the NOE between H-6 α and H-3 α was not observed, while NOEs were seen between H-6 β and H-4 α ; H-6 β and H-13; and between H-6 β and H-15. These data indicate that the methoxyl group at C-6 has an α configuration. Thus, the structure of 6 was determined to be 3β -hydroxy- 6α methoxyeremophil-7(11)-en-12,8 β -olide. Compound 6 is the first 6α -substituted eremophil-7(11)-en-12,8 β -olide derivative from natural sources.

Compound 7 was isolated as a colorless oil, $[\alpha]_D + 95.7^\circ$. The molecular formula was determined to be C₁₆H₂₄O₄ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3455 cm⁻¹) and an α,β -unsaturated- γ lactone (1747 cm⁻¹). The ¹H-NMR spectrum of 7 was closely related to that observed for 6 except for a signal due to the tertiary methyl group [δ 1.34 (s, H-15)]. Naya et al.⁶⁾ reported that, for 8β -methoxyeremophil-7(11)-en-12,8α-olide derivatives, which have a steroidal conformation, the chemical shifts due to the tertiary methyl group (H-15) are downfield from those due to the secondary methyl group (H-14), whereas this relationship is reversed in the 8α -series, which has a non-steroidal conformation. These variations in the chemical shifts may be explained similarly in terms of the effect due to the alteration in geometry of the skeleton observed in the steroid field.⁶⁾ The ¹H-NMR spectrum of 7 showed a singlet of the tertiary methyl group (H-15) at δ 1.34 and a doublet of the secondary methyl group (H-14) at δ 1.01 ($J=7.0\,\mathrm{Hz}$). Furthermore, in the NOE difference spectra, NOEs were observed between H-6α and H-13; H-6α and H-14; H-6α and H-15; and between the 6β -methoxyl group and H-8 β . These data indicate that 7 exists in a steroidal conformation (Fig. 1). The position of the hydroxyl group was determined to be at the C-3 β by comparing the chemical shift, coupling pattern and constants of the hydroxy-bearing methine proton of 7 with those of 3β hydroxy- 6β -acyleremophil-7(11)-en- $12,8\alpha$ -olides.⁸⁾ The position of the methoxyl group was determined to be at the C-6 β from the HMBC spectrum. Based on this evidence, the structure of 7 was determined to be 3β hydroxy- 6β -methoxyeremophil-7(11)-en- $12,8\alpha$ -olide.

Compound 8 was isolated as a colorless needles, mp 204—206 °C, $[\alpha]_D$ +48.4°. The molecular formula was

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determined to be $C_{15}H_{20}O_4$ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3346 cm⁻¹), an α,β -unsaturated- γ -lactone (1753 cm⁻¹) and a sixmembered ring ketone (1709 cm⁻¹). The ¹H- and ¹³C-NMR spectra showed signals due to a tertiary methyl group [$\delta_{\rm H}$ 1.00 (s, H-15), $\delta_{\rm C}$ 12.3 (C-15)], a secondary methyl group [$\delta_{\rm H}$ 0.97 (d, J = 6.6 Hz, H-14), $\delta_{\rm C}$ 7.7 (C-14)], an olefinic methyl group [$\delta_{\rm H}$ 1.87 (d, J=1.7 Hz, H-13), $\delta_{\rm C}$ 8.3 (C-13)], an AB-type methylene [$\delta_{\rm H}$ 2.28 (dd, J=13.9, 1.7 Hz, H-6 β), 2.74 (d, J = 13.9 Hz, H-6 α), $\delta_{\rm C}$ 34.6 (C-6)], an α,β -unsaturated- γ -lactone [δ_C 123.9 (C-11), 157.1 (C-7), 171.5 (C-12)], a hemi-ketal carbon [$\delta_{\rm C}$ 103.0 (C-8)] and a carbonyl carbon [$\delta_{\rm C}$ 211.3 (C-3)]. Thus, the structure of **8** was 8 (α or β)-hydroxy-3-oxoeremophilenolide. The HMBC spectrum supported this structure. In the NOESY spectrum, NOEs were seen between H-6α and H-13; H-6α and H-14; H-6α and H-15; and between H-13 and H-14. Based on this evidence, the structure of 8 was determined to be 8β -hydroxy-3-oxoeremophil-7(11)-en-12,8 α -olide, which had a steroidal conformation.

Compound 9 was isolated as a colorless oil, $[\alpha]_D - 9.3^\circ$. The molecular formula was determined to be C₁₆H₂₄O₄ by HR-MS. The IR spectrum suggested the presence of a hydroxyl group (3452 cm⁻¹), a saturated ester (1733 cm⁻¹) and an α,β -unsaturated ketone (1671 cm⁻¹). The ¹H- and ¹³C-NMR spectra showed signals due to two secondary methyl groups [δ_H 0.99 (d, $J=7.0\,\text{Hz}$, H-14), 1.28 (d, J = 7.3 Hz, H-13), $\delta_{\rm C}$ 7.4 (C-14), 16.0 (C-13)], a tertiary methyl group [$\delta_{\rm H}$ 1.24 (s, H-15), $\delta_{\rm C}$ 25.2 (C-15)], two secondary methyl-bearing methines [δ_H 1.88 (qd, J=7.0, 4.4 Hz, H-4 α), 3.62 (qd, J=7.3, 1.1 Hz, H-11), $\delta_{\rm C}$ 44.8 (C-4), 37.8 (C-11)], a methine [δ_H 2.03 (m, H-10), δ_C 35.7 (C-10)], a methylene [δ_H 2.30 (dd, J = 17.2, 4.0Hz, H-9 α), 2.66 (dd, J=17.2, 4.8 Hz, H-9 β), $\delta_{\rm C}$ 40.1 (C-9)], a carbomethoxyl group [$\delta_{\rm H}$ 3.66 (s, COCH₃), $\delta_{\rm C}$ 52.0 (OCH₃), 175.1 (C-12)], a hydroxy-bearing methine $[\delta_H]$ 3.74 (ddd, J=9.9, 4.4, 4.4 Hz, H-3 α , $\delta_{\rm C}$ 70.6 (C-3)], an olefin [$\delta_{\rm H}$ 6.74 (br s, H-6), $\delta_{\rm C}$ 154.3 (C-6)] and a carbonyl group [$\delta_{\rm C}$ 197.6 (C-8)]. These spectral data and molecular formula suggested that the most likely structure of this compound is 9, as illustrated. The stereostructure was determined by NOE difference spectra. An NOE was observed between H-6 and H-3a, suggesting that the

Fig. 2

hydroxyl group at C-3 has a β configuration. Furthermore, a *cis*-junction of the A/B rings which had a non-steroidal conformation, was suggested by the NOEs between H-15 and H-9 β ; H-15 and H-10; H-6 and H-3 α ; and between H-6 and H-4 α (Fig. 2). The absolute configuration of the secondary C-11 methyl group could not be determined. From the above data, the structure of **9** was determined to be 3 β -hydroxy-8-oxoeremophil-6-en-12-oic acid methyl ester.

Experimental

General Procedures Melting points were determined with a Yanagimoto micromelting apparatus and are uncorrected. Optical rotations were determined with a JASCO DIP-360 digital polarimeter. IR spectra were recorded with a Perkin-Elmer FT-IR 1725X IR spectrophotometer and UV spectra with a Beckman DU-64 spectrophotometer. ¹H- and ¹³C-NMR spectra were recorded with a JEOL JNM-GSX 400 (400 and 100 MHz, respectively) spectrometer. Chemical shifts were given on a δ (ppm) scale with tetramethylsilane as an internal standard (s, singlet; brs, broad singlet; d, doublet; brd, broad doublet; dd, double doublet; ddd, double double doublet; dddd, double double doublet; dq, double quartet; ddq, double double quartet; q, quartet; qd, quartet doublet; qq, quartet quartet; m, multiplet). The EI-MS and HR-MS were recorded on a JEOL JMS-DX 303 mass spectrometer. Column chromatography was carried out on Kieselgel 60 column (Merck; 230-400 mesh). Preparative HPLC was carried out on a Tosoh HPLC system (pump, CCPD; detector, UV-8011) using a TSK gel ODS-120T column (Tosoh).

Plant Material The dried rhizomes of *Petasites japonicus* were purchased from Tochimoto Tenkaido Co., Ltd. in 1990.

Extraction and Isolation The dried rhizomes of Petasites japonicus (3.0 kg) were extracted three times with MeOH at room temperature for 2 weeks. The MeOH extract was concentrated under reduced pressure and the residue was suspended in a small excess of water. This suspension was extracted, successively, with CHCl₃, Et₂O, AcOEt and n-BuOH. The CHCl₃-soluble fraction was concentrated under reduced pressure to afford a residue (112.5 g). This residue (60.0 g) was chromatographed on a silica-gel column using benzene-AcOEt (9:1, 8:2, 7:3) and CHCl₃-MeOH (8:2), and the eluate was separated into 4 fractions (frs. 1-4). Fraction 4 was rechromatographed on a silica-gel column using benzene-AcOEt (6:4, 5:5, 4:6, 3:7) and CHCl₃-MeOH (9:1, 8:2), and the eluate was separated into 4 fractions (frs. 1'-4'). Fraction 2' was rechromatographed on a silica-gel column using n-hexane-acetone (5:4, 5:5, 4:5, 3:6) and acetone, and the eluate was separated into 5 fractions (frs. 1''-5''). Fraction 3'' was rechromatographed on a silica gel column using benzene-AcOEt (3:2, 2:2), and the eluate was separated into 7 fractions (frs. 1"'-7"'). Fraction 6" was purified by preparative HPLC (Column, TSK gel ODS-120T, 21.5 mm i.d. × 30 cm; mobile phase, MeOH-H₂O (8:9); flow rate, 5.0 ml/min; UV detector. 220 nm) to give 5 (0.5 mg), a mixture of 6 and 7 (5.0 mg), 8 (1.2 mg) and 9 (2.9 mg). The mixture of 6 and 7 was purified by preparative HPLC (Column, TSK gel ODS-120T, 21.5 mm i.d. × 30 cm; mobile phase, MeOH-H₂O (2:3); flow rate, 4.0 ml/min; UV detector, 220 nm) to give 6 (2.4 mg) and 7 (1.9 mg). Fraction 4" was purified by preparative HPLC (Column, TSK gel ODS-120T, 21.5 mm i.d. × 30 cm; mobile phase, MeOH-H₂O (1:1); flow rate, 4.0 ml/min; UV detector, 220 nm) to give 1 (12.3 mg), 2 (6.0 mg), 3 (3.6 mg) and 4 (1.4 mg).

6β-Angeloyloxy-3β,9α-dihydroxyeremophil-7(11)-en-12,8β-olide (1) Colorless oil. $[\alpha]_D^{22} - 32.4^\circ$ (c = 1.2, CHCl₃). IR $\nu_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$: 3423, 1752, 1719, 1681, 1646. UV $\lambda_{\rm max}^{\rm MeOH}$ nm (log ε): 221 (4.0). HR-MS m/z: 364.1915 (M⁺, Calcd for C₂₀H₂₈O₆; 364.1886). ¹H-NMR: see Table 1. ¹³C-NMR: see Table 2.

6β-Angeloyloxy-3β,9β-dihydroxyeremophil-7(11)-en-12,8β-olide (2) Colorless oil. $[\alpha]_{\rm D}^{22}$ – 39.9° $(c=0.6,{\rm CHCl_3})$. IR $\nu_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$: 3442, 1756, 1718, 1646. UV $\lambda_{\rm max}^{\rm MeOH}$ nm $(\log \varepsilon)$: 218 (4.1). HR-MS m/z: 364.1839 (M⁺, Calcd for ${\rm C_{20}H_{28}O_6}$; 364.1886). 1 H-NMR: see Table 1. 13 C-NMR: see Table 2.

6β-Angeloyloxy-3β,8β,9β-trihydroxyeremophil-7(11)-en-12,8α-olide (3) Amorphous powder. $[\alpha]_{D}^{2^2}$ +41.2° (c=0.4, EtOH). IR ν_{\max}^{KBa} cm $^{-1}$: 3445, 1757, 1718, 1644. UV $\lambda_{\max}^{\text{MeOH}}$ nm ($\log \varepsilon$): 218 (4.2). HR-MS m/z: 380.1852 (M⁺, Calcd for $\text{C}_{20}\text{H}_{28}\text{O}_{7}$; 380.1835). $^{1}\text{H-NMR}$: see Table 1. $^{13}\text{C-NMR}$: see Table 2.

6β-(3'-Chloro-2'-hydroxy-2'-methylbutyloyloxy)-3β-hydroxyeremophil-7(11)-en-12,8β-olide (4) Colorless oil. $[\alpha]_{2}^{D1}$ –55.6° $(c=0.1, \text{ CHCl}_3)$. IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm $^{-1}$: 3447, 1750. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 220 (4.1). HR-MS m/z: 400.1684 (M⁺, Calcd for $C_{20}H_{29}^{35}\text{ClO}_6$; 400.1652), m/z: 402.1667 (M⁺+2, Calcd for $C_{20}H_{29}^{37}\text{ClO}_6$; 402.1624). $^1\text{H-NMR}$: see Table 1. $^1\text{C-NMR}$: see Table 2.

6β-Epoxyangeloyloxy-3β-hydroxyeremophil-7(11)-en-12,8β-oilde (5) Colorless oil. $[\alpha]_{\rm D}^{20}$ – 96.2° (c = 0.05, CHCl $_{\rm 3}$). IR $\nu_{\rm max}^{\rm CHCl}$ s cm $^{-1}$: 3423, 1750. UV $\lambda_{\rm max}^{\rm MoOH}$ nm (log ε): 224 (3.9). HR-MS m/z: 364.1912 (M $^+$, Calcd for C $_{\rm 20}$ H $_{\rm 28}$ O $_{\rm 6}$; 364.1886). 1 H-NMR: see Table 1. 13 C-NMR: see Table 2.

3β-Hydroxy-6α-methoxyeremophil-7(11)-en-12,8β-olide (6) Colorless oil. $[\alpha]_D^{26}$ – 54.4° (c = 0.2, CHCl $_3$). IR $\nu_{\rm max}^{\rm CHCl}$ cm $^{-1}$: 3475, 1751. UV $\lambda_{\rm max}^{\rm MeOH}$ nm (log ϵ): 220 (4.0). HR-MS m/z: 280.1650 (M $^+$, Calcd for C $_{16}$ H $_{24}$ O $_{4}$; 280.1674). 1 H-NMR: see Table 1. 13 C-NMR: see Table 2.

3β-Hydroxy-6β-methoxyeremophil-7(11)-en-12,8α-olide (7) Coloress oil. $[\alpha]_D^{25}$ +95.7° (c=0.2, CHCl₃). IR $\nu_{\max}^{\text{CHCl}_3}$ cm $^{-1}$: 3455, 1747. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 220 (4.1). HR-MS m/z: 280.1632 (M $^+$, Calcd for $C_{16}H_{24}O_4$; 280.1674). 1 H-NMR: see Table 1. 1 3C-NMR: see Table 2.

8β-Hydroxy-3-oxoeremophil-7(11)-en-12,8α-olide (8) Colorless needles (from AcOEt), mp 204—206 °C. [α] $_{\rm D}^{\rm 27}$ +48.4° (c=0.1, CHCl $_{\rm 3}$). IR $\nu_{\rm max}^{\rm CHCl}$ cm $^{-1}$: 3346, 1753, 1709. UV $\lambda_{\rm max}^{\rm McOH}$ nm (log ε): 219 (3.9). HR-MS m/z: 264.1351 (M $^+$, Calcd for C $_{15}$ H $_{20}$ O $_{4}$; 264.1361). 1 H-NMR: see Table 1. 13 C-NMR: see Table 2.

3β-Hydroxy-8-oxoeremophil-6-en-12-oic Acid Methyl Ester (9) Colorless oil. $[\alpha]_D^{24}$ -9.3° (c=0.2, MeOH). IR $\nu_{\max}^{\text{CHCl}_3}$ cm $^{-1}$: 3452, 1733, 1671. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 231 (3.9). HR-MS m/z: 280.1637 (M $^+$, Calcd for $C_{16}H_{24}O_4$; 280.1675). 1 H-NMR: see Table 1. 1 3C-NMR: see Table 2.

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