Biosynthesis of Corrinoids and Porphyrinoids. XI. Source of Oxaloacetic Acid for Uroporphyrinogen III Biosynthesis in *Arthrobacter hyalinus*

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The source of oxaloacetic acid, required for the synthesis of porphyrins in *Arthrobacter hyalinus*, was examined by means of a feeding experiment with [1,3-¹³C₂]glycerol, which is transformed to pyruvic acid. A half of the carbon dioxide liberated from pyruvic acid in the formation of acetyl CoA was utilized for carboxylation of pyruvic acid to generate oxaloacetic acid.

Key words Arthrobacter hyalinus; tricarboxylic acid cycle; uroporphyrin III; [1,3-13C2]glycerol

Arthrobacter hyalinus synthesizes porphyrins,¹⁾ and we have already examined its biopathways using several ¹³C-labeled compounds.^{2,3)} ¹³C-Labeled isopropanol or sodium acetate was transformed into ¹³C-labeled acetyl CoA, which condensed with oxaloacetic acid to form ¹³C-labeled citric acid, which ultimately afforded ¹³C-labeled porphyrins. To identify the source of the oxaloacetic acid, we have conducted a feeding experiment with [1,3-¹³C₂]glycerol in A. hyalinus.

Results and Discussion

The 13 C-NMR spectrum (Fig. 1) of the octamethyl ester derived from 13 C-labeled uroporphyrinogen III biosynthesized from $[1,3^{-13}C_2]$ glycerol in *A. hyalinus* showed 13 C-enriched signals at 21.9 ppm (β -methylene carbons of propyl side-chains), 32.7 ppm (α -methylene carbons of acetyl side-chains), 37.1 ppm (α -methylene carbons of

propvl side-chains), 133.1 ppm (C-2, 7, 12, 18), 141.0 ppm (C-3, 8, 13, 17) and 143.8 ppm (C-1, 6, 11, 19). This ¹³C-enrichment pattern showed that [1,3-¹³C₂]glycerol had been converted into ¹³C-labeled porphyrin through the same route as that we reported previously for incorporation of $[2^{-13}C]$ sodium acetate²⁾ (Figs. 1, 2). However, additional ¹³C-enriched signals due to C-5, 10, 15 and 20 (Figs. 1, 2, Table 1) were observed. It was considered that [13C]carbon dioxide, liberated by decarboxylation of [1,3-13C₂]pyruvic acid that was derived from [1,3-13C₂]glycerol, condensed with pyruvic acid to produce [4-13C]oxaloacetic acid. This condensed with acetyl CoA, to give $\lceil 5^{-13} \text{C} \rceil \delta$ -aminolevulinic acid via $[1-^{13}C]\alpha$ -ketoglutaric acid and $[1-^{13}C]$ glutamic acid. Consequently, C-5, 10, 15 and 20 of uroporphyrin III octamethyl ester would also be ¹³C-labeled.

Further, carbon dioxide liberated from pyruvic acid

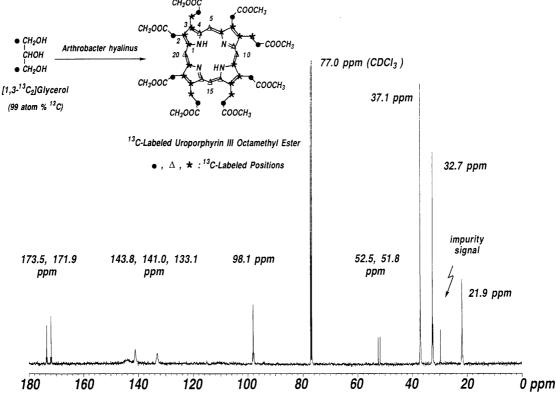


Fig. 1. Structure of Uroporphyrin III Octamethyl Ester and ¹³C-NMR Spectrum of ¹³C-Labeled Uroporphyrin III Octamethyl Ester Derived from [1,3-¹³C₂]Glycerol

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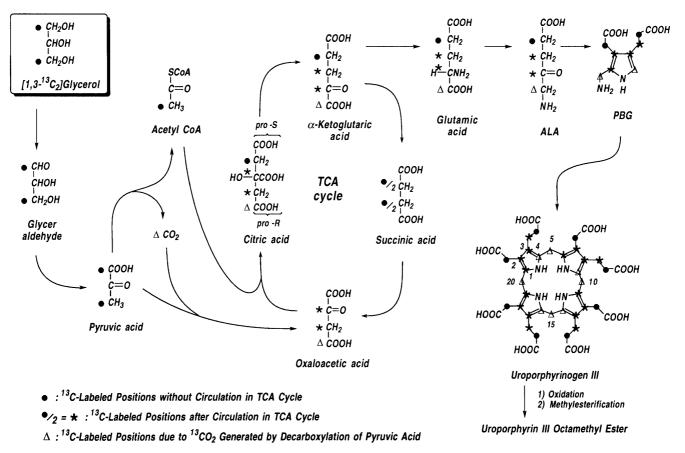


Fig. 2. Metabolic Pathways Leading from [1,3-13C2]Glycerol to 13C-Labeled Uroporphyrinogen III in Arthrobacter hyalinus

Table 1. ¹³C-Enrichment Ratios for Carbon Atoms in ¹³C-Labeled Uroporphyrin III Octamethyl Ester Derived from [1,3-¹³C₂]Glycerol

		23 3
Positions of carbon in ¹³ C-labeled uroporphyrin III octamethyl ester	Chemical shift (coupling pattern)	Ratio of ¹³ C-enrichment
β -Methylene carbons of Ps	21.9 ppm (s)	5.3
α-Methylene carbons of As	32.7 ppm (s)	11.2
α-Methylene carbons of Ps	37.1 ppm (s)	11.3
Methyl ester carbons of Ps	51.8 ppm (s)	1.0
Methyl ester carbons of As	52.5 ppm (s)	1.0
C-5, 10, 15, 20	98.1 ppm (s)	5.9
C-2, 7, 12, 18	133.1 ppm (br s)	N.C.
C-3, 8, 13, 17	141.0 ppm (br s)	N.C.
C-1, 4, 6, 9, 11, 14, 16, 19	143.8 ppm (br)	N.C.
Carbonyl carbons of As	171.9 ppm (s)	1.6
Carbonyl carbons of Ps	173.5 ppm (s)	1.4

The "A" indicates the acetyl side-chains and the "P" indicates the propyl side-chains in ¹³C-labeled uroporphyrin III octamethyl ester. The signals of methyl ester carbons of As and Ps are given for reference. The signals of carbon in ¹³C-labeled uroporphyrin III octamethyl ester were compared with natural abundance in uroporphyrin III octamethyl ester to obtain the enrichment ratio. N.C.=not calculable.

was immediately used for carboxylation of pyruvic acid. Based on the ratio of 13 C-enrichment at C-5, 10, 15 and 20 to 13 C-enrichment at the α -methylene carbons of the acetyl and propyl moieties (Table 1), approximately 50% of [13 C]carbon dioxide liberated from [1,3- 13 C₂]pyruvic acid was utilized in this way.

Experimental

Materials and Instrument [1,3- 13 C₂]Glycerol (99 atom% 13 C) was supplied by Icon. 13 C-{ 1 H} NMR spectra were recorded on a Varian Unity INOVA 500 (125 MHz) spectrometer with a nanoprobe in CDCl₃ solution referenced to the solvent peak. The spectral width was 25000 Hz with 65536 K data points, which corresponds to a resolution of 0.76 Hz per point. The determined 10° pulse width was $2.2\,\mu$ s, the acquisition time was 1.311 s, the pulse delay time was 0.689 s and the number of scans was 10000.

Feeding of [1,3- 13 C₂]Glycerol to *A. hyalinus* [1,3- 13 C₂]Glycerol (250 mg × 4) was added to the fermentation culture medium (pH 7.0, 200 ml × 4), which consisted of MgSO₄·7H₂O (5.0 g), CaCO₃ (5.0 g), NH₄NO₃ (3.0 g), peptone (3.0 g), Na₂HPO₄·12H₂O (1.5 g), L-cystine (0.6 g), yeast extract (1.0 g), KH₂PO₄ (0.4 g), ZnSO₄·7H₂O (10 mg), CuSO₄·5H₂O (50 μ g), MoO₃ (10 μ g) and monosodium L-glutamate (1.0 g) in ion-exchanged water (1.0 l), in 500 ml Erlenmeyer flasks. The flasks were shaken at 27 °C for 10 d on a rotary incubator (200 rpm).

Isolation of ¹³C-Labeled Uroporphyrin III Octamethyl Ester The method of isolation of ¹³C-labeled uroporphyrin III octamethyl ester (1.2 mg), which was produced by feeding of $[1,3^{-13}C_2]$ glycerol to *A. hyalinus*, has been detailed in the preceding papers. ^{1–3)}

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References

- Kojima I., Maruhashi K., Fujiwara Y., Saito H., Kajiwara M., Mizutani M., J. Ferment. Bioeng., 75, 353—358 (1993).
- 2) Kajiwara M., Mizutani M., Matsuda R., Hara K., Kojima I., *J. Ferment. Bioeng.*, **77**, 626—629 (1994).
- Kajiwara M., Hara K., Takatori K., Chem. Pharm. Bull., 42, 817—820 (1994).